

Prostaglandin $F_{2\alpha}$ Improves Oxygen Tension and Reduces Venous Admixture during One-lung Ventilation in Anesthetized Paralyzed Dogs

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The authors investigated the effect of prostaglandin $F_{2\alpha}$ infused into the pulmonary artery of an acutely atelectatic lung in dogs. Seven dogs were anesthetized with piritramid and pentobarbital and intubated with a Kottmeier canine endobronchial tube. Cardiac output, pulmonary arterial, capillary wedge, and systemic arterial pressure were measured via indwelling catheters. Ventilating both lungs with 66% O_2 , Pa_{O_2} was 327 ± 15 mmHg (mean \pm SD) and venous admixture (\dot{Q}_{sp}/\dot{Q}_t) was $11 \pm 3\%$. One-lung atelectasis reduced Pa_{O_2} to 91 ± 12 mmHg and increased \dot{Q}_{sp}/\dot{Q}_t to $40 \pm 4\%$. Prostaglandin $F_{2\alpha}$ in doses of 0.4, 0.6, 1.2, and $1.8 \mu\text{g} \cdot \text{kg}^{-1} \cdot \text{min}^{-1}$ was infused into the pulmonary artery of the atelectatic lung through a second pulmonary artery catheter. Up to a dose of $1.2 \mu\text{g} \cdot \text{kg}^{-1} \cdot \text{min}^{-1}$ there was a dose-dependent reduction in \dot{Q}_{sp}/\dot{Q}_t to a minimum of $25 \pm 4\%$ and an increase in Pa_{O_2} to 168 ± 25 mmHg, which could be explained by enhanced pulmonary vasoconstriction in the atelectatic lung with increased blood flow diversion toward the ventilated lung. Infusion of $1.8 \mu\text{g} \cdot \text{kg}^{-1} \cdot \text{min}^{-1}$ decreased Pa_{O_2} to 156 ± 32 mmHg and increased \dot{Q}_{sp}/\dot{Q}_t to $32 \pm 9\%$. Increased systemic effects of prostaglandin $F_{2\alpha}$ were observed and presumably were related to saturation of prostaglandin-dehydrogenase leading to vasoconstriction in both lungs and thus reduced blood flow diversion toward the ventilated lung. (Key words: Lung; hypoxic pulmonary vasoconstriction; shunting. Pharmacology: Prostaglandin $F_{2\alpha}$. Ventilation: one lung.)

IN A RECENT CLINICAL INVESTIGATION very low levels of prostaglandin $F_{2\alpha}$ ($\text{PGF}_{2\alpha}$) were reported during two- and one-lung ventilation.¹ This experimental study was devised to test the hypothesis that exogenous $\text{PGF}_{2\alpha}$ infused into the pulmonary artery of an atelectatic lung will potentiate hypoxic pulmonary vasoconstriction (HPV) of that lung and improve oxygenation by redistribution of blood flow to the ventilated lung. Although $\text{PGF}_{2\alpha}$ is not a direct mediator of HPV,²⁻⁴ being a potent pulmonary vasoconstrictor⁵⁻⁷ it seems to be a suitable drug to elicit this response.

As 70–98% of $\text{PGF}_{2\alpha}$ is inactivated within one passage through the lung,^{8,9} it is assumed that careful dosage of $\text{PGF}_{2\alpha}$ would prevent saturation of the inactiva-

tion mechanism and would limit the effect to the atelectatic lung.

Materials and Methods

Seven mongrel dogs weighing 23.4 ± 2.3 kg (mean \pm SD) and free from overt heart and lung disease were anesthetized with intravenous piritramid 15 mg**, pancuronium bromide 4 mg, and atropine 0.25 mg; thereafter with $66 \pm 1.2\%$ oxygen (mean \pm SD) in nitrogen. Anesthesia was maintained with piritramid 15 mg/h and pancuronium bromide 2 mg/h. Because piritramid is only an analgesic agent, pentobarbital 25 mg/h was added as an hypnotic agent. The animals were ventilated mechanically with a Dräger ventilator UV-1 at constant rate (10 breaths/min), adjusting tidal volume between 12 and 15 ml/kg to maintain arterial P_{CO_2} at 35 ± 4 mmHg. Fluid status was maintained by continuous intravenous infusion of 5% dextrose in water containing sodium chloride 20 mmol/l and potassium chloride 40 mmol/l (250–500 ml/h). A heating lamp and humidified inspired gas were utilized to maintain body temperature at $36.8 \pm 0.6^\circ\text{C}$ (mean \pm SD). After the dog was secured in the supine position, a cannula and a 5F catheter incorporating a Clark-type polarographic oxygen electrode (Fresenius) were placed into the right and left femoral artery to measure the time course of changes in Pa_{O_2} , induced by $\text{PGF}_{2\alpha}$ infusion. Under fluoroscopic control, balloon-tipped 7F triple lumen Swan-Ganz[®] thermodilution catheters were introduced into the right and left main pulmonary artery through the external jugular veins.

A subcricoid tracheostomy was performed and the endotracheal tube was replaced by a Rüscher[®]-Kottmeier tube. By its design this divided airway with two short tubes protruding from a common cuff does not obstruct ventilation of both upper lobes of the lungs. Complete separation of the two lungs was assured by statically inflating one lung to 50 mmHg pressure through one lumen while the other was attached to a tube submerged a few millimeters in a beaker of water with no air bubbles escaping.

Following baseline measurements during two-lung

** Piritramid is a tertiary amine of the diphenyl prophyllamine group; 20 mg piritramid are equianalgesic to 15 mg of morphine, causing less cardiovascular depression than morphine.¹⁰

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TABLE 1. Hemodynamic and Respiratory Parameters in Seven Dogs before, during, and after One-lung Ventilation and PGF_{2α} Infusion (Mean Values ± SD)

Type of Ventilation (Phase)	TLV 1	OLV						TLV 5	Significance
		2	3a	3b	3c	3d	4		
PGF _{2α} infusion (μg · kg ⁻¹ · min ⁻¹)	—	—	0.4	0.6	1.2	1.8	—	—	
PaO ₂ (mmHg)	327 ± 15	91 ± 12	108 ± 20	140 ± 15	168 ± 25	156 ± 32	99 ± 14	312 ± 30	<i>P</i> ≤ 0.001
Pv̄O ₂ (mmHg)	55 ± 6	46 ± 6	47 ± 3	47 ± 4	49 ± 4	55 ± 7	48 ± 5	56 ± 6	<i>P</i> ≤ 0.001
Q _{sp} /Q _t (%)	11 ± 3	40 ± 4	35 ± 7	27 ± 3	25 ± 4	32 ± 9	39 ± 5	12 ± 3	<i>P</i> ≤ 0.001
HR (beats/min)	79 ± 9	85 ± 13	91 ± 16	91 ± 17	103 ± 18	109 ± 18	103 ± 11	89 ± 12	<i>P</i> ≤ 0.001
CO (l/min)	4.1 ± 8	4.5 ± 1.2	4.2 ± 9	4.5 ± 1.2	4.7 ± 1.2	5.2 ± 1.1	4.6 ± 6	4.4 ± 7	<i>P</i> ≤ 0.01
MAP (mmHg)	94 ± 14	94 ± 10	99 ± 8	101 ± 9	106 ± 10	113 ± 11	99 ± 8	90 ± 11	<i>P</i> < 0.001
MPAP (mmHg)	9 ± 2	11 ± 3	11 ± 3	12 ± 4	15 ± 4	16 ± 4	13 ± 2	9 ± 2	<i>P</i> ≤ 0.001
PCWP (mmHg)	4 ± 1	4 ± 1	5 ± 1	4 ± 1	5 ± 1	5 ± 1	5 ± 1	4 ± 1	ns
PVR (dyn · s · cm ⁻⁵)	85 ± 23	120 ± 44	126 ± 50	145 ± 65	163 ± 70	170 ± 56	126 ± 57	91 ± 33	<i>P</i> ≤ 0.001
AWP _{peak} (mmHg)	20 ± 2	31 ± 2	31 ± 2	32 ± 3	32 ± 2	32 ± 3	31 ± 2	2 ± 1	ns
AWP _{plateau} (mmHg)	9 ± 1	13 ± 2	12 ± 2	13 ± 2	14 ± 2	14 ± 2	13 ± 1	9 ± 1	ns

* Significance related to analysis of variance, considering the block

of phase 2, 3a, 3b, 3c, 3d. For analysis of blocks 1, 2 and 1, 2, 4, 5 and multiple comparisons see text.

ventilation (TLV) (phase 1) nitrogen was washed out by ventilating the right lung in four animals and the left lung in three animals with 100% oxygen for 15 min prior to discontinuing ventilation to that lung. Atelectasis was induced by clamping the endobronchial tube and allowing the oxygen to be absorbed. The other lung was ventilated with 66 ± 1.2% oxygen. One lung atelectasis was verified by chest roentgenogram. After 60 min of one-lung ventilation (OLV) (phase 2), PGF_{2α} was infused into the pulmonary artery of the non-ventilated lung via an electronic infusion pump (Braun, West Germany). The initial dose of 0.4 μg · kg⁻¹ · min⁻¹ gradually was increased every 15 min to 0.6, 1.2, and 1.8 μg · kg⁻¹ · min⁻¹ (phase 3a–3d). OLV control measurements were taken 30 min after prostaglandin infusion was stopped (phase 4); the atelectatic lung then was hyperinflated manually. Following 30 min of TLV (phase 5), a final set of baseline measurements was taken and cardiac arrest was induced by an intravenous injection of 20 mmol potassium chloride. At each phase, cardiac output (CO), mean pulmonary artery (MPAP), and pulmonary capillary wedge (PCWP) pressures were determined using the pulmonary artery catheter in the ventilated lung. Heart rate (HR), airway pressure (AWP), mean arterial (MAP), and central venous pressure were measured at end-expiration and recorded on a five-channel polygraph recorder. Arterial and mixed venous blood gas analysis and determinations of oxygen concentration in the inspired gas were done immediately using a Radiometer® ABL-2. All measurements were made following two-point calibration.

Venous admixture (Q_{sp}/Q_t) was calculated using the traditional shunt equation.¹¹ Canine blood oxyhemo-

globin saturation was derived from the dissociation curve corrected for pH and temperature.¹² Hemoglobin was determined with an Eppendorf® Digital Photometer 6114 S. Alveolar oxygen tension was calculated. Pulmonary vascular resistance (PVR) was calculated using standard formulas.

STATISTICAL METHOD

To test the global influence of treatments, analysis of variance was used for each variable.

The main effects on the factor treatment were tested on the 0.05, 0.01, or 0.001 level of significance. If global differences of treatments were significant, multiple comparisons were performed with a posteriori probability of 0.05 based on the Bonferroni α -adjustment procedure.¹³ Statistical calculations were made using program INSTAT.¹⁴

Results

The results of the hemodynamic and respiratory measurements are shown in table 1.

EFFECT OF ONE-LUNG ATELECTASIS

With the changeover from TLV to OLV the arterial PO₂ decreased significantly (*P* ≤ 0.001). Mixed venous PO₂ also decreased (*P* ≤ 0.01), while venous admixture (Q_{sp}/Q_t) increased (*P* ≤ 0.001). Heart rate, cardiac output, MAP, and PCWP did not change, while MPAP and PVR increased (*P* ≤ 0.05). With the beginning of OLV peak airway pressure increased (*P* ≤ 0.001) and so did end-inspiratory plateau pressure (*P* ≤ 0.01).

EFFECT OF PROSTAGLANDIN INFUSION

Prostaglandin infusion had a significant effect on Pa_{O₂}. Infusion of 0.4, 0.6, 1.2, and 1.8 μg · kg⁻¹ · min⁻¹ of PGF_{2α} increased Pa_{O₂} significantly (*P* ≤ 0.05) when compared with the preinfusion OLV period (phase 2). Multiple comparisons showed that there was a significant (*P* ≤ 0.05) increase in Pa_{O₂} following each successive dose increment up to a dosage of 1.2 μg · kg⁻¹ · min⁻¹ of PGF_{2α}. Pa_{O₂} reached 168 ± 25 mmHg in phase 3c (fig. 1). The increase in Pa_{O₂} following infusion of PGF_{2α} occurred rapidly reaching a new equilibrium within 62 ± 24 s (mean ± SD) (fig. 2). Mixed venous PO₂ did not increase significantly before 1.8 μg · kg⁻¹ · min⁻¹ of PGF_{2α} was given. PGF_{2α} significantly reduced Q_{sp}/Q_t (*P* ≤ 0.001). When compared with phase 2, significant reductions in Q_{sp}/Q_t occurred at 0.6 and 1.2 μg · kg⁻¹ · min⁻¹ PGF_{2α}, maximum reduction was observed at 1.2 μg · kg⁻¹ · min⁻¹. With infusion of 1.8 μg · kg⁻¹ · min⁻¹, Pa_{O₂} decreased slightly and Q_{sp}/Q_t increased when compared with phase 3c (fig. 1). Heart rate and cardiac output were significantly influenced by PGF_{2α} infusion, but in multiple comparisons a significant increase was observed only between phases 2 and 3d (table 1).

MAP increased significantly during PGF_{2α} infusion, but there were no significant differences between the four dosages given. MPAP was significantly (*P* ≤ 0.05) increased during phase 3c and 3d when compared with the preinfusion period and 0.4 and 0.6 μ · kg⁻¹ · min⁻¹ of PGF_{2α}.

Likewise, PVR increased significantly when phase 2 and 3a were compared with 3d. The increase in peak airway pressure and end-inspiratory plateau pressure was not significant.

CONTROL MEASUREMENTS AFTER PGF_{2α} INFUSION

Thirty minutes after PGF_{2α} infusion was stopped, Pa_{O₂}, P_vO₂, Q_{sp}/Q_t, cardiac output, MAP, MPAP, PCWP, PVR, and AWP were not significantly different from their respective preinfusion levels. Heart rate still was significantly above preinfusion levels.

With reexpansion of the atelectatic lung, all measured variables returned to preatelectatic control values.

During the entire 3-h study period, arterial pH was 7.39 ± 0.03 and hemoglobin decreased from 12.1 ± 1.6 g/dl to 11.1 ± 0.9 g/dl. Central venous pressure was 2.6 ± 0.4 mmHg.

Discussion

The major findings in the present study show that arterial and mixed venous oxygen tension and calcu-

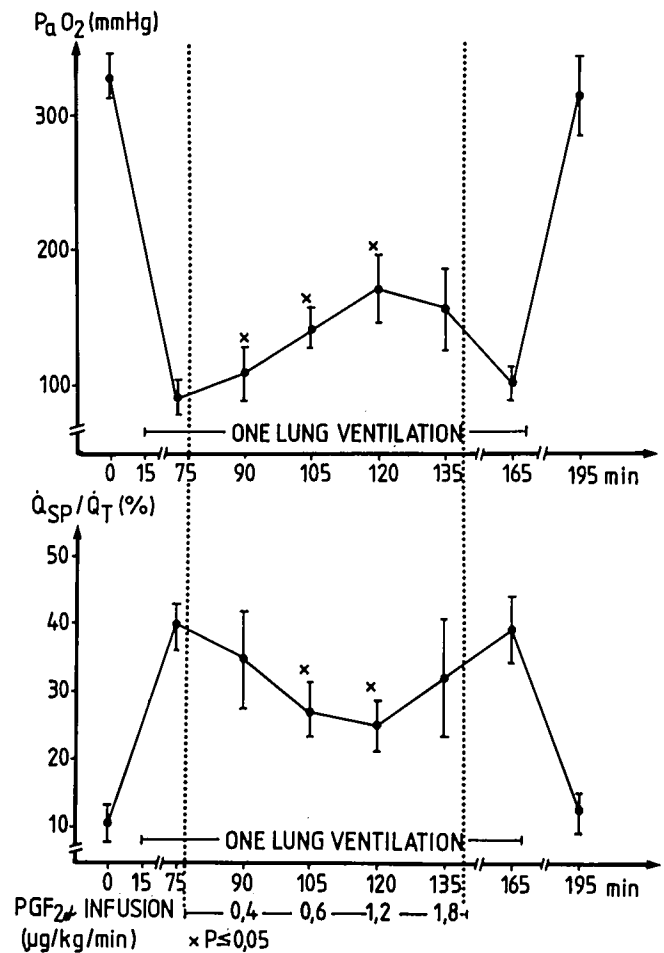


FIG. 1. Changes of arterial P_{O₂} and venous admixture (Q_{sp}/Q_t) during two-lung ventilation and one-lung ventilation and the effect of prostaglandin F_{2α} infusion in seven dogs (mean values ± SD). During prostaglandin infusion, Pa_{O₂} increased significantly (*P* ≤ 0.05) when compared with each preceding step; Q_{sp}/Q_t decreased (*P* ≤ 0.05) when compared with step 2 at 75 min.

lated venous admixture during stable one-lung atelectasis in mechanically ventilated dogs can be improved by infusion of PGF_{2α} into the pulmonary artery of the atelectatic lung. These findings are discussed in detail, after some methodologic considerations.

Induction of atelectasis resulted in a small but consistent increase in mean pulmonary artery pressure and an increase in venous admixture, comparable with results of other investigators.^{15,17} The increase in MPAP is typical for HPV in a large hypoxic segment, resulting in limited blood flow redistribution to the ventilated lung.¹⁸ Intermittent challenges with hypoxic gas mixtures potentiated HPV in an open-chest preparation of the left lower lobe in the dog.^{19,20} In the open chest the increase in pulmonary vascular resistance in response to acute atelectasis is maximal after 60 min and stable thereafter.²¹ In a closed-chest preparation, flow diver-

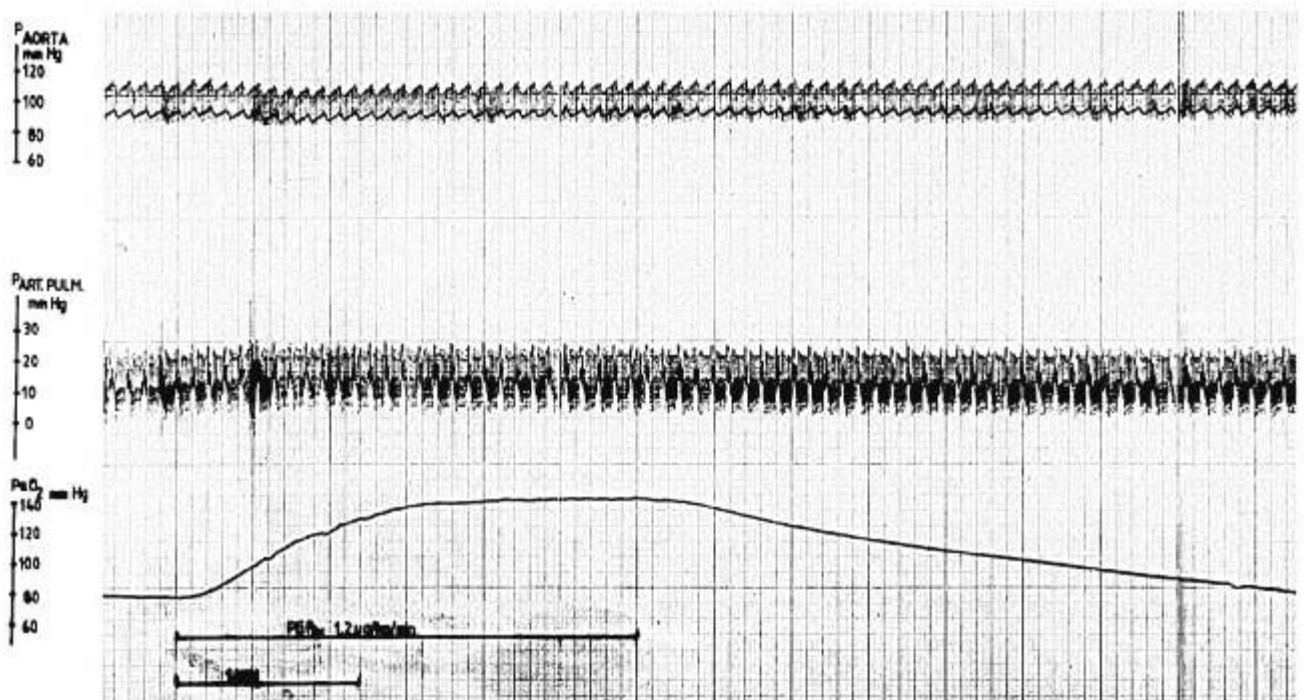


FIG. 2. Original, recording of aortic pressure, pulmonary artery pressure, and arterial oxygen tension measured by indwelling intraarterial oxygen probe. In little more than 1 min after $\text{PGF}_{2\alpha}$ infusion ($1.2 \mu\text{g} \cdot \text{kg}^{-1} \cdot \text{min}^{-1}$) was started, Pa_{O_2} reached a new stable level. There was a small increase in pulmonary artery pressure.

sion was maximal after the first hypoxic exposure compared with the fourth hypoxic trial in an open-chest experiment.²²

Therefore, in the present study repeated challenges of the pulmonary vasculature were not performed in this closed-chest model before HPV of the atelectatic lung was considered to have reached near steady state conditions.

In contrast to other studies,^{17,20,21} the nonatelectatic lung was ventilated with 66% instead of 100% oxygen. Using an FI_{O_2} of 0.66, no severe hypoxemia occurred during OLV in a previous human study¹ nor in the present study. Severe hypoxemia can induce HPV in the ventilated lung counteracting blood flow redistribution from the atelectatic lung.¹⁵ Increasing FI_{O_2} to 95 or 100% may result in a release of HPV in regions of the ventilated lung with critically low \dot{V}_A/Q ratios and increase venous admixture.²³

The effect of $\text{PGF}_{2\alpha}$ on arterial and mixed venous oxygen tension and shunt only can be explained by an increase in HPV in the atelectatic lung, resulting in increased blood flow redistribution toward the ventilated lung. Pa_{O_2} was improved despite significant increases in pulmonary perfusion pressure, estimated by MPAP-PCWP, and cardiac output. Increased pulmonary blood volume and increased intraluminal vascular

pressures normally are considered an important factor inhibiting effective blood flow redistribution by HPV.^{18,24}

The largest increase in Pa_{O_2} and associated decrease in $\dot{Q}_{\text{sp}}/\dot{Q}_t$ was observed at a PGF infusion rate of $1.2 \mu\text{g} \cdot \text{kg}^{-1} \cdot \text{min}^{-1}$. The oxygen content was estimated only by oxygen tension and hemoglobin, assuming a carboxyhemoglobin of zero and using the hemoglobin saturation derived from the Rossing nomogram for dogs,¹¹ therefore, the calculated venous admixture likely yielded falsely high shunt values.²⁵ However, the errors attributable to the common clinical practice of neglecting carboxyhemoglobin and methemoglobin measurements are smaller when saturation is calculated than when it is directly measured by an oxygen analyzer.²⁶

There was a slight decrease in Pa_{O_2} and increase in $\dot{Q}_{\text{sp}}/\dot{Q}_t$ at $\text{PGF}_{2\alpha}$ infusion rate of $1.8 \mu\text{g} \cdot \text{kg}^{-1} \cdot \text{min}^{-1}$. This decrease in response to $\text{PGF}_{2\alpha}$ presumably can be related to saturation of PG dehydrogenase, the enzyme responsible for PG metabolism.⁹ $\text{PGF}_{2\alpha}$ is inactivated almost completely during one passage through the lung,^{8,9} however, if PG infusion rate surpasses enzyme capacity, $\text{PGF}_{2\alpha}$ will reach the systemic circulation. Systemic effects of $\text{PGF}_{2\alpha}$ include an increase in mean arterial pressure, cardiac output, and heart rate.^{6,7} The

significant increases in heart rate and cardiac output in phase 3d, when compared with phase 2, coincide with the maximum increase in MPAP and PVR.

PGF_{2α} escaping hepatic metabolism^{3,4,6} will affect pulmonary vessels in both the ventilated and atelectatic lung. A slight vasoconstricting effect in the ventilated lung would diminish blood flow diversion away from the atelectatic lung.

In contrast to other investigators,^{27,28} PG doses given in the present study did not produce any major bronchoconstriction, as there was a nonsignificant increase in airway peak pressure and no significant increase in end-inspiratory plateau pressure. The latter could be attributed to decreased compliance caused by small airway constriction as normal PCWP values exclude pulmonary venous hypertension.²⁸

A few minutes after the PG infusion was stopped, all variables except heart rate returned to base line values (phase 2). The persisting moderately increased heart rate may be in large measure a reflex consequence of a decrease in blood pressure. Higher doses of PGF_{2α} in the systemic circulation may induce a persisting increase in sympathetic outflow from the central nervous system.⁶ However, the effects of PGF_{2α} are not mediated by sympathetic stimulation, because of the reported complete resistance of its pulmonary vasoconstricting effect to enhancement or blunting by alpha- and beta-adrenergic blocking agents.^{29,30} An activation of histamine receptors or the release of histamine does not mediate this action of PGF_{2α} as well.³⁰

Norepinephrine and histamine are unlikely to provide the same selective pulmonary vasoconstriction in the atelectatic lung as PGF_{2α}; both produce variable degrees of pulmonary vasoconstriction, but histamine, like epinephrine and dopamine, is not eliminated from the pulmonary circulation to any appreciable extent, and only 20–35% of exogenously administered norepinephrine is metabolized on passage through the lung.³¹

Experimental evidence in animals is difficult to apply to humans because of considerable species differences in response to PGs, but PGF_{2α} is a pulmonary vasoconstrictor in humans³² as well, and human pulmonary circulation metabolizes PGF_{2α} in much the same way as other mammalian lungs.³³ Therefore, PGF_{2α} infusion into the pulmonary artery of an acutely atelectatic lung also may prove to be a practical and safe method to improve oxygenation during one-lung ventilation in humans. PGF_{2α} can be titrated to achieve an optimal effect without causing major systemic reactions. There was no persisting PG action or cardiopulmonary dysfunction at the end of the study period.

References

1. Scherer RW, Van Aken H, Schlegel W, Lawin P: Prostaglandins during one lung ventilation in esophageal surgery. *ANESTHESIOLOGY* 59:A502, 1983
2. Fishman AP: Hypoxia on the pulmonary circulation: How and where it acts. *Cir Res* 38:221–231, 1976
3. Weir EK, Groves RF: The role of endogenous prostaglandins in the pulmonary circulation. *ANESTHESIOLOGY* 48:201–212, 1978
4. Hyman AL, Mathé AA, Lippton HL, Kadowitz PJ: Prostaglandins and the lung. *Med Clin North Am* 65:789–808, 1981
5. Ducharme DW, Weeks JR, Montgomery RG: Studies on the mechanism of the hypertensive effect of prostaglandin F_{2α}. *J Pharmacol Exp Ther* 160:1–10, 1968
6. Nakano J, Cole S: Effects of PGE₁, F_{2α} on systemic pulmonary and splanchnic circulations in dogs. *Am J Physiol* 217:222–227, 1969
7. Kadowitz PJ, Joiner PD, Hyman AL: The hypertensive effect of PGF_{2α} on the pulmonary circulation of swine, lamb and dog, *Progress in Respiratory Research*. Vol 9. Basel Karger, 1975, pp 285–292
8. Ferreira SH, Vane JR: Prostaglandins: Their disappearance from the release into the circulation. *Nature* 216:868–873, 1967
9. Piper PJ, Vane JR, Wyllie JH: Inactivation of PG by the lungs. *Nature* 225:600–604, 1970
10. Kettler D, Cott L, Hensel J, Martel J, Bretschneider HJ: Combination of piritramide and N₂O, a new anesthetic method for studies of the cardiovascular function in dogs. *Pfluegers Arch* 319:42–49, 1970
11. Berggren S: The oxygen deficit of arterial blood caused by non-ventilating parts of the lung. *Acta Physiol Scand* 11(Suppl):1–92, 1942
12. Rossing RG, Cain SM: A nomogram relating PO₂, pH, temperature and hemoglobin saturation in the dog. *J Appl Physiol* 21:195–201, 1966
13. John JA, Quenouille MH: *Experiments, Design and Analysis*. London, Charles Griffin & Co, 1977
14. Hultsch E: *INSTAT-Programmbeschreibung*. Interaktives Programmsystem für statistische Berechnungen. Schriftenreihe des Instituts für Medizinische Informatik und Bio-mathematik, Vol. 7. Münster, University Press, 1983
15. Newell JC, Levitzky MG, Krasney JA, Dutton RE: Phasic reflux of pulmonary blood flow in atelectasis: influence of systemic PO₂. *J Appl Physiol* 40:883–888, 1975
16. Alfery DD, Zamost BG, Benumof JL: Unilateral lung lavage: Blood flow manipulation by ipsilateral pulmonary artery balloon inflation in dogs. *ANESTHESIOLOGY* 55:376–380, 1981
17. Alfery DD, Benumof JL, Trousdale FR: Improving oxygenation during one-lung ventilation in dogs: The effects of PEEP and blood flow restriction to the non-ventilated lung. *ANESTHESIOLOGY* 55:381–385, 1981
18. Marshall BE, Marshall C: Continuity of response to hypoxic pulmonary vasoconstriction. *J Appl Physiol* 49:189–196, 1980
19. Pirlo AF, Benumof JL, Trousdale FR: Potentiation of lobar hypoxic pulmonary vasoconstriction by intermittent hypoxia in dogs. *ANESTHESIOLOGY* 55:226–230, 1981
20. Benumof JL: Intermittent hypoxia increases lobar hypoxic pulmonary vasoconstriction. *ANESTHESIOLOGY* 58:399–404, 1983
21. Glasser SA, Domino KB, Lindgren L, Parcella P, Marshall BE:

- Pulmonary blood pressure and flow during atelectasis in dog. *ANESTHESIOLOGY* 58:225–231, 1983
22. Chen L, Williams JJ, Domino KB, Alexander CM, Ray RJ, Marshall BE: Circumstances for potentiation of hypoxic pulmonary vasoconstriction. *ANESTHESIOLOGY* 59:A 497, 1983
 23. Lundh R, Hedenstierna G: Ventilation–perfusion relationship during halothane anaesthesia and mechanical ventilation. Effects of varying inspired oxygen concentration. *Acta Anaesthesiol Scand* 28:191–198, 1984
 24. Benumof JL, Wahrenbrock EA: Blunted hypoxic pulmonary vasoconstriction by increased lung vascular pressures. *J Appl Physiol* 38:846–850, 1975
 25. Cane RD, Shapiro BA, Harrison RA, Steiner MC, Kavanaugh J: Minimizing errors in intrapulmonary shunt calculation. *Crit Care Med* 8:294–297, 1980
 26. Maffeo CJ, Hoyt JW, Swain RF: Venous admixture: Errors and clinical decisions. *ANESTHESIOLOGY* 55:A 80, 1980
 27. Spannake EW, Hyman AL, Kadowitz PJ: Dissimilar in vivo effects of arachidonic acid on canine pulmonary vascular bed and airway. *Adv Prostaglandin Thromboxane Leukotriene Res* 7:937–942, 1980
 28. Wassermann MA: Bronchopulmonary effect of $\text{PGF}_{2\alpha}$ and three of its metabolites in the dog. *Prostaglandin* 9:958–967, 1975
 29. Okpako DT: The actions of histamine and $\text{PGF}_{2\alpha}$ and E_2 on the pulmonary vascular resistance of the lung of the guinea-pig. *J Pharm Pharmacol* 24:40–50, 1972
 30. Bergofsky EH: Active control of the pulmonary circulation, *Pulmonary Vascular Diseases*. Edited by Moser KM. New York, Basel, Marcel Dekker 1979, pp 233–277
 31. Fishman AP, Pietra GG: Handling of bioactive materials by the lung. Part 1 and 2. *N Engl J Med* 291:884–890, 953–959, 1974
 32. Karim SSM, Somers K, Hilker K: Cardiovascular and other effects of prostaglandins E_2 , $\text{F}_{2\alpha}$ in man. *Cardiovasc Res* 5:255–262, 1971
 33. Jose P, Niederhauser U, Piper PJ, Robinson C, Smith AP: Degradation of prostaglandin $\text{F}_{2\alpha}$ in the human pulmonary circulation. *Thorax* 31:713–719, 1976