

Effects of Phenobarbital on Cerebral Blood Flow and Metabolism in Young and Aged Rats

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The cerebrovascular and cerebral metabolic changes produced by intraperitoneal injection of phenobarbital (50, 150, and 250 mg/kg) were studied in young adult (6-month) and senescent (28-month) Wistar rats. Cerebral blood flow (CBF) was measured using radioactive microspheres and cerebral oxygen consumption (CMR_{O_2}) was obtained by multiplying cortex CBF by the arterial-sagittal sinus oxygen content difference. Control values for blood pressure, blood gas tensions, CBF, and CMR_{O_2} were similar in the young and aged animals during 70% $N_2O/30\%$ O_2 . Intraperitoneal phenobarbital produced dose-dependent decreases in CBF with no significant difference between young and aged rats at each phenobarbital dose. At the highest phenobarbital dose (250 mg/kg) CBF was reduced by 49% in the young rats and 52% in the aged rats ($P > 0.10$). CMR_{O_2} was also depressed in a dose-dependent fashion in both young and aged animals with each phenobarbital dose. However, the decrease produced by the highest phenobarbital dose was significantly greater in the aged rats (55%) than the young rats (43%, $P < 0.05$), even though the EEG was isoelectric in both groups. The difference in CMR_{O_2} between young versus aged rats at a time when the EEG is isoelectric suggests that high-dose phenobarbital may depress nonelectrical cerebral metabolic processes more in aged rats. (Key words: Age factors. Anesthetics, intravenous: phenobarbital. Brain: blood flow; metabolism.)

IT IS WELL established that decreases in cerebral blood flow (CBF) and cerebral oxygen consumption (CMR_{O_2}) produced by barbiturates correlate closely with the degree of anesthesia and EEG suppression.¹ CBF and CMR_{O_2} are maximally depressed with barbiturate doses that produce

a quiescent EEG. Nonelectrical cerebral metabolic activity is resistant to further depression with higher barbiturate doses.^{1,2} Little is known about how the cerebrovascular and cerebral metabolic responses induced by barbiturate anesthesia may be altered with aging. Young and aged rats were tested using a model that has previously demonstrated phenobarbital-induced decreases in CBF, CMR_{O_2} , and EEG activity.²

Methods

Thirty-seven young (6-month) and 32 aged (28-month) Wistar rats were anesthetized with halothane and, following tracheostomy, were artificially ventilated with 1% inspired halothane in 70% nitrous oxide/30% oxygen using a small animal respirator. Catheters were inserted into both femoral arteries and a femoral vein for fluid and drug administration, blood sampling, and hemodynamic monitoring. A catheter for microsphere injections was inserted into the left ventricle *via* the right carotid artery. Proper placement of the left ventricular catheter was assured by observing a change in the pressure tracing from an arterial to a left ventricular pattern. The animal was then placed in the prone position, the skull was exposed, and a small burr hole was drilled over the posterior sagittal sinus. A needle-tipped catheter was fixed into the hole for withdrawal of blood samples used to determine cortical venous oxygen content. Stainless steel screw electrodes were applied bilaterally over the parietal cortices for continuous bipolar EEG monitoring. A third screw electrode over the frontal cortex served as a ground electrode. All leads were shielded to prevent 60-cycle interference. The EEG was recorded using a Grass Instruments® P15 differential AC amplifier and a Hewlett-Packard® strip chart recorder. Filter settings were 1 Hz at the low range and 30 Hz at the high range.

Following surgical preparation halothane was discontinued and the rat was immobilized with 1 mg/kg tubocurarine and allowed to stabilize for 45 min while being ventilated with 70% $N_2O/30\%$ O_2 . Arterial P_{CO_2} was adjusted to 35–40 mmHg and arterial P_{O_2} was maintained greater than 100 mmHg. Rectal temperature was measured with a thermistor probe and maintained at 37° C using an overhead heat lamp. Mean arterial blood pressure

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was recorded continuously from a femoral arterial catheter. Heart rate was measured from arterial pressure pulses periodically throughout the experiment.

At the end of the stabilization period each rat received an intraperitoneal injection of one of three phenobarbital doses (50, 150, or 250 mg/kg) or sham treatment (normal saline). CBF was measured 90 min later using radioactive microspheres. Arterial and sagittal sinus blood samples (0.2 ml each) were taken immediately after the microsphere test for measurement of blood gas tension and pH using an IL 1303 (Instrument Laboratories, Lexington, MA) blood gas analyzer and for determination of arterial and sagittal sinus (venous) oxygen content using an IL 282 co-oximeter. Sagittal sinus samples were drawn over 10–15 s. Because the sagittal sinus drains primarily cerebral cortex, these venous samples reflect cortical oxygen extraction. Cortical CMR_{O₂} was calculated by multiplying cortical blood flow by the arterial–sagittal sinus oxygen content difference (AV_{O₂}).² EEG was recorded throughout the 90-min phenobarbital treatment period. Preliminary studies indicated that the maximum effect of ip phenobarbital was produced by 90 min, with little to no additional EEG suppression thereafter.

To measure CBF, 15-μm microspheres labeled with cobalt-57 (New England Nuclear, Boston, MA) were suspended in a stock solution of isotonic saline with 0.01% Tween-80®. After the solution was vortexed, 0.2 ml (100,000 microspheres) was injected into the left ventricular catheter and flushed in with 0.2 ml isotonic saline over 20 s. Starting immediately before the microsphere injection, blood was withdrawn from a femoral artery catheter at a rate of 0.4 ml/min using a Harvard infusion/withdrawal pump. A withdrawal period of 45 s was chosen to ensure the removal of all circulating microspheres. At the end of each microsphere test arterial and sagittal sinus blood samples for blood gas tension, pH, and oxygen content were obtained. Blood pressure was continuously monitored throughout the microsphere testing to ensure no hemodynamic changes prior to CMR_{O₂} determination. The rat was then killed, and the brain was removed, divided into left and right cortical and subcortical samples, weighed, and placed in counting tubes for analysis of radioactivity. The activity of the microsphere label was analyzed in blood and brain tissues with a Nuclear Chicago® 600 multichannel analyzer. CBF was calculated as follows:

$$\text{CBF(ml/100 g/min)} = \frac{\text{Tissue activity}}{\text{Blood sample activity}} \times \frac{\text{Blood sample withdrawal rate}}{\text{Tissue weight}} \times 100$$

Cerebrovascular resistance was calculated from CBF and

mean blood pressure measured at the time of microsphere injection.

When the higher phenobarbital doses produced hypotension, methoxamine was infused intravenously to maintain mean arterial blood pressure higher than 100 mmHg. Separate experiments were first carried out to ensure that methoxamine had no direct cerebral vasoconstrictor effect. Cortical and subcortical blood flow was measured in four rats during 70% N₂O/30% O₂ using cobalt-57-labeled microspheres and then during a methoxamine infusion that raised mean blood pressure approximately 25 mmHg using a second microsphere labeled with tin-113.

Data are reported as mean ± SE. The effect of phenobarbital anesthesia on young and aged rats was compared using a two-way analysis of variance. Multiple tests comparing the means of young and aged rats at each phenobarbital dose were performed using unpaired *t* tests with a Bonferroni correction. This correction reflected the four tests of pairs for each parameter measured.

Results

In preliminary validation studies, methoxamine infusion increased mean blood pressure from 133 ± 4 to 158 ± 6 mmHg (mean ± SE; n = 4) with no significant change in CBF. Cortical and subcortical blood flows were 125 ± 16 and 69 ± 5 ml/100 g/min, respectively, in control state and 125 ± 7 and 71 ± 5 ml/100 g/min, respectively, during the methoxamine infusion. Based on these data we concluded that methoxamine could be used to maintain blood pressure during phenobarbital treatment with little effect on cerebral resistance vessels.

The effects of phenobarbital on blood pressure, arterial blood gases, and pH are shown in table 1. Arterial blood pressure of young and aged rats was similar during nitrous oxide control conditions and decreased in both groups with increasing phenobarbital doses. A methoxamine infusion was necessary to maintain a mean arterial pressure greater than 100 mmHg in all young and aged rats treated with 150 mg/kg and 250 mg/kg phenobarbital. Pa_{CO₂} was maintained between 35 and 40 mmHg with mechanical ventilation, and Pa_{O₂} remained higher than 100 mmHg in all treatment groups. There were no significant intergroup differences in blood pressure and blood gas tensions. Arterial pH and P_{O₂} increased in both young and aged rats following treatment with phenobarbital.

The effects of phenobarbital on EEG are shown in figure 1. EEG changed from low-amplitude, high-frequency activity during nitrous oxide to high-amplitude, low-frequency waves with 50 mg/kg phenobarbital. This activity was further depressed with 150 mg/kg phenobarbital,

TABLE 1. Arterial Blood Pressure, Blood Gases, and pH in Phenobarbital-anesthetized Young and Aged Rats

Phenobarbital (mg/kg)	n	Blood Pressure (mmHg)	P _a CO ₂ (mmHg)	P _a O ₂ (mmHg)	pH
Young					
0	9	138 ± 6	37.7 ± 1.2	121 ± 9	7.37 ± .02
50	10	126 ± 1	37.8 ± 0.3	155 ± 2	7.44 ± .01*
150	9	122 ± 2*	35.2 ± 0.1	173 ± 2*	7.43 ± .01
250	9	115 ± 1*	36.0 ± 0.2	159 ± 6*	7.41 ± .01
Aged					
0	7	137 ± 4	35.8 ± 0.8	116 ± 7	7.39 ± .01
50	8	123 ± 6	40.2 ± 0.9	148 ± 18	7.45 ± .01*
150	9	127 ± 3	36.9 ± 1.0	181 ± 13*	7.45 ± .01*
250	8	118 ± 4*	35.8 ± 0.9	181 ± 9*	7.41 ± .01

There were no intergroup differences in any of the parameters measured.

* $P < 0.05$ compared with zero dose control for each age group.

and the EEG was isoelectric with the 250 mg/kg dose in both age groups. Little difference between young and aged rats was seen in the development of EEG changes over the 90-min phenobarbital treatment period or in the EEG records at the time of testing.

CBF changes produced by phenobarbital paralleled the changes in EEG (fig. 2). Control CBF values were similar in young and aged rats. Phenobarbital produced a dose-dependent decrease in cortical and subcortical CBF in both age groups. This depression was significant in both young and aged rats as compared with nitrous oxide control values. The maximal decrease in cortical blood flow was approximately 50%, whereas the maximal depression of subcortical flow was 33%, with no difference in response between the two age groups ($P > 0.10$). Cerebrovascular resistance (CVR) was also similar between young and aged rats. Cortical CVR increased from 1.18 ± 0.10 mmHg · ml⁻¹ · 100 g⁻¹ · min⁻¹ under control conditions to a maximum of 2.08 ± 0.07 mmHg · ml⁻¹ · 100 g⁻¹ · min⁻¹ at the highest phenobarbital dose in young rats. In aged rats, cortical CVR increased from 1.15

± 0.16 to 2.00 ± 0.10 mmHg · ml⁻¹ · 100 g⁻¹ · min⁻¹. Analyses of variance indicated that cortical and subcortical CVR increased significantly during phenobarbital treatment ($P < 0.01$) and that these increases were not significantly different between age groups ($P > 0.10$).

Cerebral metabolism was also depressed in both young and aged rats following phenobarbital administration (fig. 3). As with CBF, phenobarbital-induced metabolic depression reached a plateau in both young and aged rats at the 150–250 mg/kg doses. Analysis of variance indicated that significant decreases in CMR_{O₂} were produced with phenobarbital in both age groups ($P < 0.01$), and this depression was greater in aged (55%) as compared with young rats (43%; $P < 0.01$). A difference in response to phenobarbital in young versus aged rats was also suggested by changes in AV_{O₂}. AV_{O₂} was not significantly different between young and aged rats during 70% N₂O or following 50 mg/kg phenobarbital (70% N₂O, young = 5.8 ± 0.4 ml O₂/100 ml, and aged = 6.0 ± 0.9 ml O₂/100 ml [$P > 0.10$]; 50 mg/kg phenobarbital, young = 5.9 ± 0.1 ml O₂/100 ml, and aged = 5.9 ± 0.6 ml O₂/100

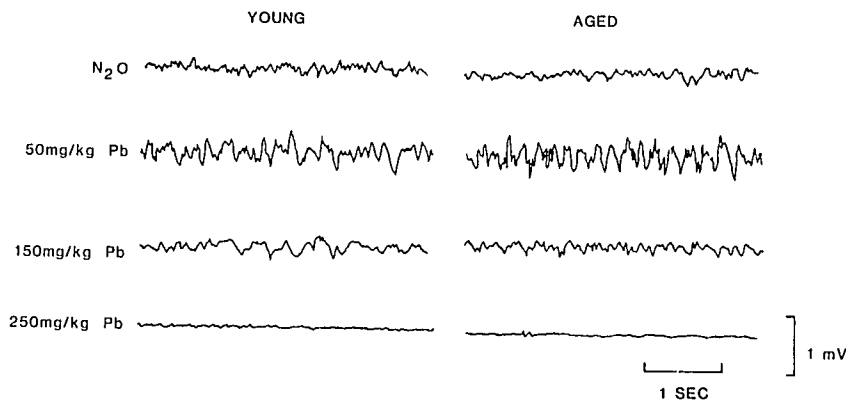


FIG. 1. EEG changes during phenobarbital (Pb) treatment in young and aged rats (mV = millivolts). Each EEG strip represents a separate rat and treatment condition. Phenobarbital produced dose-related depression of EEG activity in both young and aged rats and complete suppression of electrical activity at the 250 mg/kg dose. The time sequence of EEG changes, not shown here, was also similar between young and aged rats.

ml [$P > 0.10$]). Cerebral oxygen extraction increased in young compared with aged rats with 150 and 250 mg/kg phenobarbital (150 mg/kg, young = 7.3 ± 0.1 ml O₂/100 ml, and aged = 5.7 ± 0.3 ml O₂/100 ml [$P < 0.05$]; 250 mg/kg, young = 6.8 ± 0.1 ml O₂/100 ml, and aged = 5.5 ± 0.3 ml O₂/100 ml [$P < 0.05$]).

Discussion

Twenty-eight-month-old rats have been physiologically characterized as senescent in previous studies, approximately equivalent to a 70-yr-old human when the life spans of rats and humans are equated.³⁻⁵ The 6-month-old rats used in this study may be considered as a young adult.³ In these studies, ip phenobarbital produced similar dose- and time-related EEG suppression that reached a maximum in 90 min in young and aged rats. This suggests that brain uptake of the barbiturate was similar between

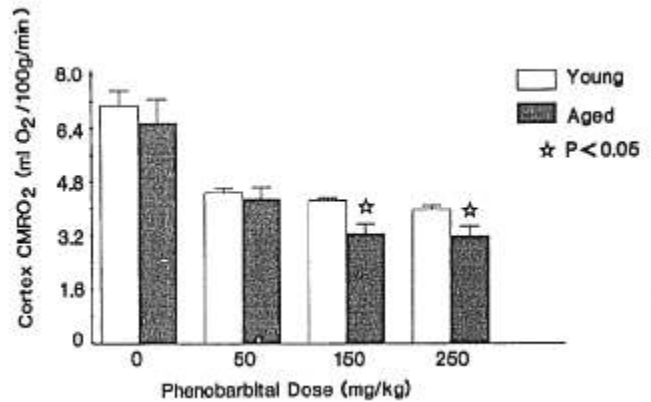


FIG. 3. Cortical CMRO₂ changes during phenobarbital anesthesia in young and aged rats. Significant differences between young and aged, as determined by *t* test and indicated by \star , are shown at 150 and 250 mg/kg phenobarbital doses. Analysis of variance indicates a significant decrease in CMRO₂ in both young and aged rats as compared with control with increasing phenobarbital doses ($P < 0.01$), with a greater decrease in the aged vs. young rats ($P < 0.01$).

the two age groups over the 90-min interval. This agrees with the report of Kapetanovic *et al.*⁶ that plasma and brain phenobarbital concentrations peak at approximately 90 min and are similar in young and aged rats up to 2 h after ip injection. CBF was decreased in a similar manner in young and aged rats following phenobarbital, but CMRO₂ decreased more in aged rats with higher phenobarbital doses. Because EEG was isoelectric at the highest phenobarbital dose, a difference in neuronal electrical activity between the two age groups is not likely.

Nitrous oxide was used as the control anesthetic because it provides analgesia in rats with no depression or stimulation of CBF or brain metabolism.^{7,8} Although the work of Seyde and Longnecker⁹ suggests that CBF in unanesthetized rats may be lower than values reported here with N₂O, others have reported that cortical CBF in unanesthetized rats is not different from N₂O-ventilated animals and is similar to our values.⁷ N₂O apparently produces little stress in rats because CBF and CMRO₂ are not elevated compared with unstimulated adrenalectomized rats or rats pretreated with beta blockers.¹⁰ Withdrawal of N₂O from paralyzed rats results in stress-related increases in both CBF and CMRO₂.¹⁰ It is also possible that N₂O may alter the cerebrovascular or metabolic response to phenobarbital. In a recent report, Sakabe *et al.*¹¹ found in rats that N₂O attenuates the decrease in cerebral glucose consumption produced by pentobarbital (30 mg/kg), but it does not alter cerebral metabolic depression or EEG flattening produced by 125 mg/kg pentobarbital. This suggests that N₂O may produce a small stimulation of CMRO₂ in rats during low-dose barbiturate anesthesia;

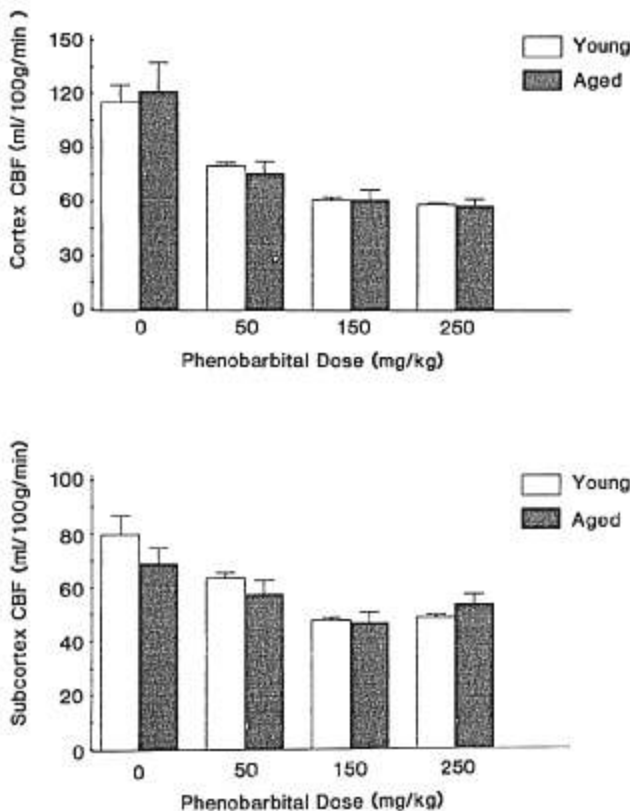


FIG. 2. Cortical and subcortical blood flow in young and aged phenobarbital-anesthetized rats. Analysis of variance indicates significant blood flow depression with increasing phenobarbital doses ($P < 0.01$) in both young and old rats as compared with control. There was no significant difference in CBF between young and aged rats at control or any phenobarbital dose ($P > 0.10$).

however, N₂O probably has little effect on cerebral metabolism and EEG depression produced by high-dose phenobarbital.

Both young and aged rats showed dose-related decreases in CMR_{O₂} that were not different under control conditions and with 50 mg/kg phenobarbital. These results were consistent with EEG changes and suggest that sensitivity to phenobarbital was similar between the two age groups. This agrees with reports that the brain sensitivity to thiopental does not change with age.¹² The aged rats, however, showed more cerebral metabolic depression than young rats at higher phenobarbital doses. Because electrical neuronal activity is absent with the highest phenobarbital dose in both age groups, the lower CMR_{O₂} observed in aged rats may represent a decrease in metabolic activity devoted to nonelectrical neuronal function. This suggests that elderly patients may be more sensitive to barbiturates, based not only on pharmacokinetic differences such as decreased drug clearance, changes in body composition, and drug volume of distribution,^{12,13} but that there may also be a pharmacodynamic effect of greater cerebral metabolic depression at high doses.

There are two ways to interpret the cerebrovascular effects of phenobarbital observed in this study. The first is to analyze the direct cerebrovasoconstrictor effects associated with increasing doses of the drug. By this analysis, phenobarbital decreased CBF and increased cerebrovascular resistance to the same extent in both young and aged rats. The second way is to relate decreases in CBF during phenobarbital treatment to simultaneous decreases in CMR_{O₂}. Since the early report of Roy and Sherrington¹⁴ it has been generally observed that both regional and global changes in CBF are tightly coupled to changes in brain metabolism. However, it has been observed previously² as well as in this study that CBF decreases more than CMR_{O₂} in young phenobarbital-treated rats, producing an increase in cerebral oxygen extraction. This may be interpreted as an additional cerebrovasoconstrictor effect, which is greater than that required by the decrease in brain metabolism.

There are two possible mechanisms by which phenobarbital could mediate the additional cerebrovasoconstriction that is seen in young but not aged rats. The first is by a direct vasoconstrictor effect of the drug on cerebral resistance vessels, which has been suggested previously.¹⁵ This appears unlikely since *in vitro* experiments on rat aorta strips by Altura and Altura¹⁶ and on isolated cerebral vessels by Marin *et al.*¹⁷ and Edvinsson and McCulloch¹⁸ have shown that barbiturates produce vascular smooth muscle relaxation, not constriction. Recently, however, Fukuda *et al.*¹⁹ reported that thiopental potentiated alpha-adenoreceptor-induced vasoconstriction in

artery smooth muscle preparations. Similar vascular adrenoceptors have been identified in human and rat cerebral vessels.²⁰ It is also possible that the *in vivo* effects of phenobarbital do not correspond to *in vitro* actions.²¹

The second possible mechanism by which barbiturates may produce vasoconstriction is by decreasing the sensitivity of vascular smooth-muscle receptors to endogenous mediators of local vasodilation, such as potassium, H⁺, or adenosine.²² Such an effect would decrease local cerebral perfusion and increase oxygen extraction requirements for the perfused tissue, as was observed in young rats. If phenobarbital produces cerebrovasoconstriction by one of the earlier mentioned mechanisms, a lack of this effect in aged rats may be due to a change in sensitivity to this phenobarbital-induced vascular action.^{23,24} This may occur independently from the direct cerebral metabolic depressant effects of phenobarbital in young *versus* old animals.

In summary, while both young and aged rats show dose-related decreases in CBF and CMR_{O₂} to phenobarbital, specific differences are apparent. First, aged rats show more cerebral metabolic depression to large doses of phenobarbital than young rats. Second, young rats decrease CBF more than CMR_{O₂} during phenobarbital treatment, a relative vasoconstrictor effect which was not seen in senescent rats. These effects represent an age-induced change in responsiveness to the cerebral metabolic and cerebrovascular actions of phenobarbital that may be mediated by separate mechanisms.

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References

1. Michenfelder JD: The interdependency of cerebral function and metabolic effects following massive doses of thiopental in the dog. *ANESTHESIOLOGY* 41:231-236, 1974
2. Nilsson L, Siesjo BK: The effect of phenobarbitone anaesthesia on blood flow and oxygen consumption in the rat brain. *Acta Anaesthesiol Scand [Suppl]* 57:18-24, 1975
3. Masoro EJ, Bertrand H, Liepa G, Yu BP: Analysis and exploration of age related changes in mammalian structure and function. *Fed Proc* 38:1956-1961, 1979
4. Coleman GL, Barthold SW, Osbaldiston GW, Foster SJ, Jonas AM: Pathologic changes during aging in barrier-reared Fisher-344 male rats. *J Gerontol* 32:258-278, 1977
5. Barrows CH, Kokkonen GC: Diet and life extension in animal model systems. *Age* 1:131-143, 1978
6. Kapetanovic IM, Sweeney DJ, Rapoport SI: Phenobarbital pharmacokinetics in rat as a function of age. *Drug Metab Dispos* 10:586-589, 1982
7. Dahlgren N, Ingvar M, Yokoyama H, Siesjo BK: Influence of nitrous oxide on local cerebral blood flow in awake, minimally restrained rats. *J Cereb Blood Flow Metab* 1:211-218, 1981

8. Ingvar M, Siesjo BK: Effect of nitrous oxide on local cerebral glucose utilization in rats. *J Cereb Blood Flow Metab* 2:481-486, 1982
9. Seyde WC, Longnecker DE: Anesthetic influences on regional hemodynamics in normal and hemorrhaged rats. *ANESTHESIOLOGY* 61:686-698, 1984
10. Carlsson C, Hagerdal M, Kaasik AE, Siesjo BK: A catecholamine-mediated increase in cerebral oxygen uptake during immobilization stress in rats. *Brain Res* 119:223-231, 1977
11. Sakabe T, Tsutsui T, Mackawa T, Ishikawa T, Takeshita H: Local cerebral glucose utilization during nitrous oxide and pentobarbital anesthesia in rats. *ANESTHESIOLOGY* 63:262-266, 1985
12. Homer TD, Stanski DR: The effect of increasing age on thiopental disposition and anesthetic requirement. *ANESTHESIOLOGY* 62:714-724, 1985
13. Christensen JH, Andreasen F, Jansen JA: Influence of age and sex on the pharmacokinetics of thiopentone. *Br J Anaesth* 53:1189-1195, 1981
14. Roy CS, Sherrington CS: On the regulation of blood supply to the brain. *J Physiol (Lond)* 11:85-108, 1890
15. Marshall LF, Smith RW, Shapiro HM: The outcome with aggressive treatment in severe head injuries. II. Acute and chronic barbiturate administration in the management of head injury. *J Neurosurg* 50:26-30, 1979
16. Altura BT, Altura BM: Pentobarbital and contraction of vascular smooth muscle. *Am J Physiol* 229:1635-1640, 1975
17. Marin J, Lobato RD, Rico ML, Salaices M, Benitez J: Effect of pentobarbital on the reactivity of isolated human cerebral arteries. *J Neurosurg* 54:521-524, 1981
18. Edvinsson L, McCulloch J: Effects of pentobarbital on contractile responses of feline cerebral arteries. *J Cereb Blood Flow Metab* 1:437-440, 1981
19. Fukuda S, Inomata I, Tsuji T, Takeshita H: Thiopental potentiation of isolated rabbit pulmonary artery contractions with alpha receptor agonists. *ANESTHESIOLOGY* 60:187-192, 1984
20. Edvinsson L, Hogestatt ED, Skarby T, Andersson KE: Alpha adrenoceptor heterogeneity in cerebral blood vessels. *Blood Vessels* 18:212-216, 1982
21. Bolton TB: Mechanisms of action of transmitters and other substances on smooth muscle. *Physiol Rev* 59:606-718, 1979
22. Berne RM: Metabolic regulation of blood flow. *Circ Res* 14:261-268, 1964
23. Lakatta EG: Alterations in the cardiovascular system that occur in advanced age. *Fed Proc* 38:163-167, 1979
24. Fleisch JH: Age related changes in the sensitivity of blood vessels to drugs. *Pharmacol Ther* 8:477-487, 1980