

## Cardiopulmonary Bypass in a Patient with Factor XII Deficiency

MARKKU SALMENPERÄ, M.D.,\* VESA RASI, M.D.,† SEVERI MATTILA, M.D.‡

Factor XII (F XII), or Hageman factor, is the first enzyme of the intrinsic loop of blood coagulation cascade. Despite marked laboratory abnormalities, congenital deficiency of F XII is not associated with abnormal bleeding diathesis.<sup>1,2</sup> However, standard tests to monitor heparin activity intraoperatively depend on the activation of F XII. We describe a patient who required open heart surgery and in whom standard preoperative hemostatic evaluation and perioperative assessment of heparin activity could not be performed because of his congenital F XII deficiency.

## CASE REPORT

A 56-yr-old man with a 6-yr history of stable angina increasing to the grade of New York Heart Association III or IV within the preceding 6 months was scheduled for coronary artery bypass grafting. His medications included metoprolol, nifedipine, and isosorbide dinitrate. Coronary arteriography showed triple-vessel disease and well-preserved left ventricular function (ejection fraction 70%). Chest x-ray showed bilateral diffuse infiltrates compatible with sarcoidosis, but this diagnosis had not been verified. Pulmonary function was normal. The patient reported no history of bleeding, but he had not undergone surgery before. The family history was noteworthy, in that a sister had been advised to request therapy with fresh frozen plasma before any surgical procedures.

Routine preoperative blood coagulation scan showed platelet count 221,000 platelets per ml, template bleeding time 5.5 min, prothrombin time 14 s, and activated partial thromboplastin time (aPTT; Cephotest reagent, Nycomed, Oslo) > 240 s. Because of prolonged aPTT a more thorough coagulation factor evaluation was performed and yielded the following additional results: factor V 125%, factor VII 115%, factor VIII:C 111%, von Willebrand factor antigen 124%, von Willebrand factor ristocetin cofactor 96%, factor IX 111%, factor X 107%, and F XII < 1%. A diagnosis of F XII deficiency was made, and according to the hematologist consultation, no substitution therapy was planned before the operation.

Anesthesia was induced with fentanyl 30  $\mu\text{g} \cdot \text{kg}^{-1}$  and maintained with an infusion of 0.15  $\mu\text{g} \cdot \text{kg}^{-1}$  thereafter. Pancuronium was used

for muscle relaxation; anesthesia was supplemented with diazepam as an anesthetic adjuvant in divided doses (total dose up to 0.3  $\text{mg} \cdot \text{kg}^{-1}$ ) and brief periods of enflurane. Cardiopulmonary bypass (CPB) was performed with a membrane oxygenator (Capiiox-E<sup>®</sup>, Terumo). The circuit was primed with Ringer's acetate, and the patient was cooled to 28° C. A three-vessel bypass was performed using two saphenous vein grafts and left internal mammary artery as conduits. Subsequent separation from CPB was uneventful, and no vasoactive medications were needed. A cell saver was used preoperatively, but no attempts to retransfuse shed mediastinal blood postoperatively were made.

A specimen of arterial blood obtained before anesthesia induction demonstrated a hemoglobin concentration of 15.6 g/100 ml and a hematocrit of 45%. Activated clotting time (ACT, Hemochron<sup>®</sup>) using standard celite tubes was 541 and 560 s, performed with two separate machines. Clotting time performed with the tube without an activator (Hemochron<sup>®</sup>) was 840 s. A thromboelastographic tracing (fig. 1) was grossly abnormal, with a reaction time of 36 min (95% confidence limits for normal values in coronary artery bypass graft patients at our institution = 6–11 min), an  $\alpha$ -angle of 34° (95% confidence limits = 41–46°), and a maximum amplitude of 42 mm (95% confidence limits = 46–52 mm)—values indicative of hypocoagulability *in vitro*. Heparin (porcine mucosa heparin) was given in a dose of 300 U/kg.

Thereafter, and in the second sample, drawn during CPB, no clotting was detected in ACT tubes within a 30-min observation period. No additional heparin was given during the 82-min bypass time. After removal of the cannulas heparin was antagonized with 3 mg/kg of protamine sulfate. ACT after protamine was 386 s. Two hours after protamine ACT was 382 s, APTT was > 240 s, platelet count was 121 000 platelets per ml, and template bleeding time was 8.5 min. In the thromboelastographic tracing, the R value, 24 min, was shorter after CPB. The tracing was otherwise nearly indistinguishable from the preoperative tracing (fig. 1). An assay for fibrin (and fibrinogen) degradation products was negative. Postoperative blood loss was 860 ml up to the first postoperative day, when the chest tubes were removed. Two units of packed red cells were used to increase the hematocrit to greater than 30% postoperatively.

The patient made an uneventful recovery and was discharged from the hospital on the eighth postoperative day.

## DISCUSSION

F XII is a single-chain glycoprotein with a molecular weight of approximately 80,000 D.<sup>3</sup> Deficiency of F XII is a rare congenital disorder first identified by Ratnoff and Colopy<sup>4</sup>; subsequently, the requirement of two abnormal alleles for the complete deficiency of this factor has been documented.<sup>5</sup> F XII undergoes autocatalytic activation upon contact with artificial surfaces like glass and particularly silicates like celite and kaolin.<sup>6</sup> The physiologic activator has been postulated to be subendothelial basement membrane. The intrinsic coagulation pathway is initiated when the activated serine protease (F XIIa) acts on its substrate factor XI.<sup>7</sup> Factor XIIa also is involved

\* Associate Professor of Anesthesia, Department of Anesthesia, Helsinki University Central Hospital, University of Helsinki.

† Associate Director, Medical Services, Coagulation Laboratory, Finnish Red Cross Blood Transfusion Service.

‡ Professor and Chairman, Cardiothoracic Surgery, Department of Cardiothoracic Surgery, Helsinki University Central Hospital, University of Helsinki.

Received from Helsinki University Central Hospital, University of Helsinki and the Finnish Red Cross Blood Transfusion Service, Helsinki, Finland.

Address reprint requests to Dr. Salmenperä: Department of Anesthesia, Helsinki University Central Hospital, Haartmanink. 4, SF-00290 Helsinki, Finland.

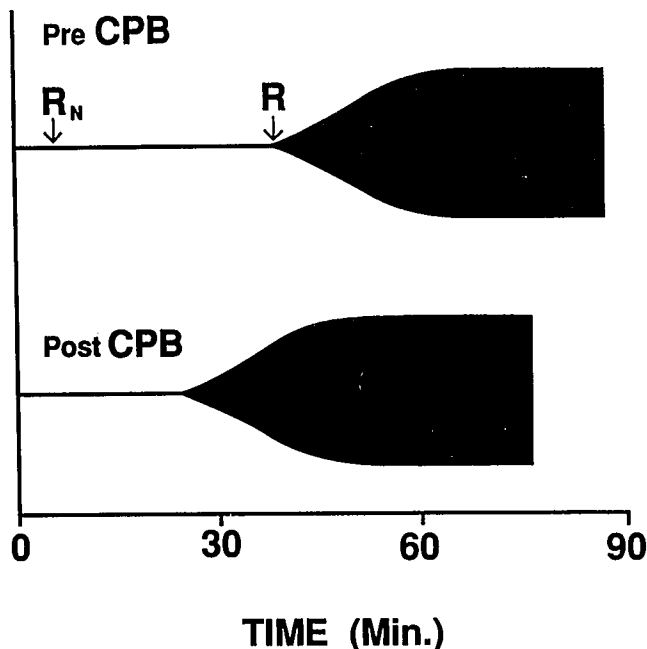


FIG. 1. Thromboelastographic tracings before (pre-CPB) and after (post-CPB) cardiopulmonary bypass. Arrows indicate the reaction time of the sample blood from our patient (R) with factor XII deficiency and the mean reaction time in our laboratory ( $R_N$ ).

in the generation of kinins, the activation of complement cascade, and (with the plasma proactivator) in the conversion of plasminogen to plasmin.

Patients with F XII deficiency do not have excessive bleeding after trauma or with surgery.<sup>1,2</sup> The discrepancy between the grossly prolonged clotting *in vitro* and the normal hemostasis *in vivo* in F XII deficiency is not yet fully understood. A disorder in the next protein in the intrinsic cascade, factor XI, is associated with delayed postoperative bleeding.<sup>8</sup> Several experiments suggest that vessel injury, besides liberating tissue factor and initiating extrinsic coagulation, can directly activate factor XI in an F XII-independent reaction that requires platelets.<sup>9</sup> In addition, the first circulating protease of the extrinsic cascade, factor VII, can directly activate factor IX, bypassing the contact factors.<sup>2</sup> Paradoxically, patients with F XII deficiency may be prone to thrombotic complications because of impaired plasminogen activation.<sup>10</sup>

Measurements to assess heparin activity take advantage of the contact activation to reach shorter coagulation endpoints and thereby to make possible its use in guiding therapy. This is done by providing a large, negatively charged surface area like ellagic acid, which is used in most aPTT systems, or celite or kaolin, which are coagulation promoting surfaces in ACT tests. Prolongation of aPTT and ACT values and the decreased *in vitro* coagulability in thromboelastographic tracing in our patient

are compatible with impaired contact activation and malfunction of the intrinsic coagulation pathway. Of course, these tests have no value in monitoring heparin in patients with F XII deficiency. It is surprising that, in the only earlier report of cardiac surgery in a patient with F XII deficiency, ACT was used to assess adequacy of heparinization and its reversal, although the authors do not document any actual ACT values.<sup>11</sup>

The test of the extrinsic system, prothrombin time, and the final common pathway, thrombin time, are prolonged over their measurement range with the high doses of heparin needed for anticoagulation. If there is doubt about unneutralized heparin after protamine in patients with F XII deficiency, thrombin time measurement is the best way to detect residual heparin, as it is in patients with a normal coagulation cascade. Heparin concentration can be detected indirectly using protamine titration *in vitro*. The clinically available devices (Hepcon A-10<sup>®</sup> and HMS<sup>®</sup>, Hemotec, Inc.) use exogenous thromboplastin to accelerate the coagulation process. This approach relies only on the extrinsic and common pathways, and will work normally in F XII-deficient patients. Heparin concentration monitoring should be helpful in these patients in order to ascertain stable heparin concentrations and completeness of reversal of heparin by protamine. Unfortunately, this methodology was unavailable to us at the time we took care of this patient.

The optimal heparin dose for CPB in the patients with F XII deficiency is not known. The capacity of heparin to inhibit kallikrein and factor XIIa is limited.<sup>12</sup> Since these steps are not operational in a F XII-deficient patient, there might be fewer activated clotting factors consuming heparin cofactors like antithrombin III. A heparin-sparing effect by this mechanism has been recently proposed after pharmacologic kallikrein inhibition by aprotinin.<sup>13</sup> The inhibition of early steps of the intrinsic coagulation cascade, however, do not allow for reduced heparin requirements in clinical practice, because the compensatory capacity of the alternate pathways is unpredictable.

Patients with a congenital F XII deficiency usually are identified by routine preoperative blood coagulation screening. An isolated markedly prolonged aPTT with a negative bleeding history strongly suggests an F XII defect. The equally rare deficiencies of the two other contact activation factors, high-molecular-weight kininogen and prekallikrein, can be responsible for a similar clinical presentation. An acquired circulating anticoagulant must be excluded and the diagnosis confirmed with a specific assay using F XII-deficient plasma. The kindred should be screened with aPTT to identify the homozygotes. Heterozygotes show abnormal F XII values, but these do not always affect the routine coagulation tests. No substitution therapy is needed for these patients even in cardiac sur-

gery, with inevitable further compromise to the coagulation system, as shown by the uneventful course of our patient. If perioperative bleeding occurs in these patients during their cardiac surgery, its cause and therapeutic approach should be no different from those of patients with normal secondary hemostasis.

Surface activation of F XII is an essential requirement for many *in vitro* assays used for coagulation monitoring in cardiac surgical patients. Such tests are unable to assess clinical hemostasis and heparin activity in F XII-deficient patients, because *in vivo* alternate modes of activation can compensate for the defect. In the absence of a clinically feasible method to assess surgical anticoagulation rapidly in these patients, heparin must be given according to weight-based protocols, and a normal dose-response relationship must be assumed. Measuring heparin concentrations during and after CPB could assist in the maintenance of anticoagulation and its reversal in the F XII-deficient patient.

#### REFERENCES

1. Wintrobe MM, Lee GR, Boggs DR, Bithell TC, Foerster J, Athens JW, Lukens JN: Blood coagulation, Clinical Hematology. Philadelphia, Lea & Febiger, 1981, pp 405-452
2. Larson L: Disorders of hemostasis, Textbook of Hematology. Edited by McKenzie SB. Philadelphia, Lea & Febiger, 1988, pp 453-454
3. Revak SD, Cochrane CG, Johnston AR, Hugli TE: Structural changes accompanying enzymatic activation of Hageman factor. J Clin Invest 54:619-627, 1974
4. Ratnoff OD, Colopy JE: A familial hemorrhagic trait associated with a deficiency of a clot promoting fraction of plasma. J Clin Invest 34:602-613, 1955
5. Margolius AJ, Jr, Ratnoff OD: Observations of the hereditary nature of Hageman trait. Blood 11:565-569, 1956
6. Hubbard D, Lucas GL: Ionic charges of glass surfaces and other materials, and their possible role in the coagulation of blood. J Appl Physiol 15:265-270, 1960
7. Ratnoff OD, Davie EW, Mallett DI: Studies on the action of Hageman factor: Evidence that activated Hageman factor in turn activates plasma thromboplastin antecedent. J Clin Invest 40:803-819, 1961
8. Kaufman J: Prostatectomy in Factor XII deficiency. J Urol 117:75-78, 1977
9. Walsh PN, Griffin JH: Contributions of human platelets to the proteolytic activation of blood coagulation factors XII and XI. Blood 57:106-118, 1981
10. Dyerberg J, Stofferson E: Recurrent thrombosis in a patient with Factor XII deficiency. Acta Haematol 63:278-282, 1980
11. Kelsey PR, Bottomley J, Grotte GJ, Maciver JE: Congenital factor XII deficiency: Successful open heart surgery and anticoagulation. Clin Lab Haematol 7:379-381, 1985
12. Pixly RA, Shapira M, Colman RW: Effect of heparin on the inactivation rate of human activated Factor XII by antithrombin III. Blood 66:198-203, 1985
13. deSmet AAEA, Joen MCN, van Oeveren W, Roozendaal KJ, Harder MP, Eijssman L, Wildevuur CRH: Increased anticoagulation during cardiopulmonary bypass. J Thorac Cardiovasc Surg 100:520-527, 1990

Anesthesiology  
75:541-543, 1991

## Epidural Opioid Analgesia Does Not Obscure Diagnosis of Compartment Syndrome Resulting from Prolonged Lithotomy Position

C. J. MONTGOMERY, M.D., F.R.C.P.,\* L. B. READY, M.D., F.R.C.P.†

Extensive genitourinary or gynecologic surgery often requires prolonged anesthesia and the lithotomy position. Many anesthesiologists prefer to conduct these cases under combined epidural and general anesthesia and then to use epidural analgesia for postoperative pain. A rare

complication of prolonged lithotomy position under general anesthesia is lower-extremity compartment syndrome.<sup>1</sup> It has been suggested that postoperative epidural analgesia may obscure symptoms of compartment syndrome and delay diagnosis and treatment.<sup>2</sup> We report two cases of patients who underwent prolonged genitourinary surgery while in the lithotomy position under combined general and epidural anesthesia. Postoperatively, while receiving intermittent epidural morphine, both patients developed readily diagnosed unilateral compartment syndrome requiring fasciotomies.

\* Acting Assistant Professor of Anesthesiology.

† Professor of Anesthesiology.

Received from the University of Washington School of Medicine, Children's Hospital and Medical Centre and University Hospital, Seattle, Washington. Accepted for publication May 22, 1991.

Address reprint requests to Dr. Montgomery: Children's Hospital and Medical Center, Department of Anesthesiology, 4800 Sand Point Way N.E., P.O. Box C5371, Seattle, Washington 98105.

Key words: Analgesia: epidural; postoperative; opioid. Complications: compartment syndrome. Position: lithotomy.

#### CASE REPORTS

Case 1. A 52-yr-old, 85-kg woman, ASA physical status 2, with carcinoma of the bladder underwent a radical cystectomy with urinary