

# Recirculatory Pharmacokinetics and Pharmacodynamics of Rocuronium in Patients

## The Influence of Cardiac Output

Jette A. Kuipers, M.Sc.,\* Fred Boer, M.D., Ph.D.,† Erik Olofson, M.Sc.,‡ James G. Bovill, M.D., Ph.D., F.F.A.R.C.S.I.,§ Anton G. L. Burm, Ph.D.||

**Background:** Recirculatory models are capable of accurately describing first-pass pharmacokinetics and the influence of cardiac output (CO), which is important for drugs with a fast onset of effect. The influence of CO on pharmacokinetic and pharmacodynamic parameters of rocuronium in patients was evaluated using a recirculatory pharmacokinetic model.

**Methods:** Fifteen patients were included to study rocuronium pharmacokinetics and pharmacodynamics. Bolus doses of rocuronium (0.35 mg/kg) and indocyanine green (25 mg) were injected simultaneously *via* a peripheral intravenous catheter. Blood samples were taken for 240 min from the radial artery. The force of contraction of the adductor pollicis after a train-of-four at 2 Hz every 12 s was measured. Arterial concentration-time curves of rocuronium and indocyanine green were analyzed using a recirculatory model. Pharmacodynamics were described using a sigmoid maximum effect ( $E_{max}$ ) model.

**Results:** The CO of the patients varied from 2.43 to 5.59 l/min. Total distribution volume of rocuronium was  $17.3 \pm 4.8$  l (mean  $\pm$  SD). The CO showed a correlation with the fast tissue clearance ( $Cl_{T,f}$ ;  $r^2 = 0.51$ ), with the slow tissue clearance ( $Cl_{T,s}$ ;  $r^2 = 0.31$ ) and with the mean transit times of rocuronium except for the mean transit time of the slow tissue compartment. The blood-effect site equilibration constant ( $k_{e0}$ ) was strongly correlated with CO ( $r^2 = 0.70$ ).

**Conclusions:** Cardiac output influences the pharmacokinetics, including  $k_{e0}$ , for rocuronium in patients. For drugs with a fast onset of effect, a recirculatory model, which includes CO, can give a good description of the relation between concentration and effect, in contrast to a conventional compartmental pharmacokinetic model.

THE effect of anesthetic drugs is generally assumed to depend on the drug concentration-time profile at the site of action. After an intravenous bolus injection or rapid intravenous infusion, the initial drug effects are likely to depend highly on blood flow, which is the major determinant of the initial distribution.<sup>1</sup> With muscle relaxants, this could result in differences in the onset of action. In the setting of a rapid sequence induction, this could result in differences in the time to reach reliable intubation conditions.

Because it is impossible to measure drug concentrations at the site of action, drug effects are commonly

related to blood or plasma concentrations using a hypothetical effect site compartment. Theoretical effect site concentrations are linked to blood concentrations by the parameter  $k_{e0}$ , the blood-effect site equilibration rate constant.<sup>2,3</sup> When concentrations at the sites of action are dependent on tissue perfusion,  $k_{e0}$  would also be expected to depend on blood flow. However, in conventional pharmacokinetic-pharmacodynamic modeling, blood flow is not accounted for.<sup>4,5</sup> In contrast to conventional pharmacokinetic models, recently developed recirculatory pharmacokinetic models incorporate cardiac output (CO).<sup>6</sup> Furthermore, in contrast to conventional compartmental models, which assume the drug concentration to peak at time zero and then decrease monotonically, recirculatory models do not assume instantaneous mixing of administered drug and blood and as such provide a much better description of the concentration-time profile during the initial mixing phase. This in itself may affect the estimation of  $k_{e0}$ , in particular when drug input is rapid and the onset of action is fast.<sup>7</sup> The importance of adequate description of the initial part of the concentration-time curve in pharmacokinetic-pharmacodynamic modeling has been demonstrated previously.<sup>8,9</sup> In these studies, semiparametric approaches were used. These do not account for blood flow but are capable of solving some of the shortcomings of compartmental models.

In this study we examined the pharmacokinetics and pharmacodynamics of rocuronium using a recirculatory pharmacokinetic model and compared the results with those obtained with conventional modeling. In addition we examined the relation between CO and the  $k_{e0}$ , as well as the other pharmacokinetic and pharmacodynamic parameters. Rocuronium was selected as a model drug because its onset of action is fast, its effects can be quantified easily and reliably, and because muscle blood flow is likely to be linked to CO.

## Materials and Methods

### Experimental Protocol

The study was approved by the Medical Ethics Committee of the Leiden University Medical Center. The study comprised 15 female patients who were scheduled for eye surgery with general anesthesia. Informed consent was obtained from each patient. Exclusion criteria were as follows: (1) obesity (Quetelet index  $> 28$ );

\* Research Fellow, † Staff Anesthesiologist, ‡ Research Associate, § Professor of Anesthesiology.

Received from the Department of Anesthesiology, Leiden University Medical Center, Leiden, The Netherlands. Submitted for publication November 29, 1999. Accepted for publication July 25, 2000. Support was provided solely from institutional and/or departmental sources.

Address reprint requests to Dr. Burm: Department of Anesthesiology (P-5), Leiden University Medical Center, P.O. Box 9600, 2300 RC Leiden, The Netherlands. Address electronic mail to: A.G.L.Burm@LUMC.nl. Individual article reprints may be purchased through the Journal Web site, www.anesthesiology.org.

(2) history of cardiovascular disease, *i.e.*, hypertension, heart failure, recent myocardial infarction (< 6 months) or the use of cardiovascular medication; (3) diabetes mellitus; (4) immobility, because this may artificially decrease CO; and (5) participation in other studies.

Patients were premedicated with midazolam, 7.5 mg orally, 60 min before administration of anesthesia. In the operating room, electrocardiogram electrodes were placed, and a peripheral intravenous infusion was established. A pulse oximeter was connected for the measurement of arterial oxygen saturation. During local anesthesia with lidocaine 1%, a catheter was placed in a radial artery.

Anesthesia was induced and maintained with a target-controlled infusion of propofol and remifentanyl. The target propofol concentration was set at 4 mg/ml. The remifentanyl target concentration was adjusted according to the surgical requirement. After loss of consciousness, rocuronium 0.35 mg/kg was injected intravenously for muscle relaxation. To allow adequate measurement of muscle relaxation and CO in the period between induction of anesthesia and intubation of the trachea, intubation was delayed until 2 min after the administration of rocuronium. If the train-of-four ratio during the experiment recovered to more than 80%, vecuronium bromide was given as required to maintain adequate muscle relaxation for surgery. Vecuronium bromide does not interfere with the rocuronium assay.<sup>10</sup>

Rocuronium was given as a mixture with indocyanine green (ICG) and autologous blood. The mixture was prepared by adding 0.0385 ml/kg rocuronium (10 mg/ml; 0.3 mg/kg is ED<sub>95</sub><sup>11</sup>) to ICG, 25 mg in 2.5 ml of its solvent, and autologous blood to make a total volume of 10 ml. One milliliter of the mixture was stored for later measurement of the injectate concentrations, and the other 9 ml was put in a 10-ml syringe. The syringes were weighed before and after the experiment to allow calculation of the injected volume.

Sampling was performed from the radial artery catheter. Before the experiments, a 20-ml blood sample was drawn for construction of the calibration curves of ICG (5 ml) and rocuronium (15 ml). During the first 10 min of the experiment, sampling was performed with the aid of a specially constructed computer-controlled syringe pump and a fraction collector. Thereafter, samples were taken manually. The sample size was initially 1.5 ml (0.3 ml for ICG measurement and 1.2 ml for rocuronium measurement). After 10 min, the sample size was 3 ml (for rocuronium measurement only). Sampling started 3 s before injection of the drugs (blank sample) and continued for 240 min. The first minute's sampling was performed once every 3 s, the second minute's once every 10 s, and thereafter samples were taken at 2.5, 3, 4, 7, 10, 15, 30, 60, 120, 180, and 240 min. When the sampling device was used, *i.e.*, during the first 10 min of sampling, waste samples were taken to clear the system

between the samples. Sampling volume was equal to the sum of the dead space volumes in the catheter and the extension lines to the sampling device.

Neuromuscular block was measured by applying a supramaximal stimulus to the ulnar nerve at the wrist and monitoring the response of the adductor pollicis (thumb adduction), using a force transducer (UC3, Gould Instrument Systems, Valley View, OH) connected to an amplifier (Datascope 2000A, Datascope Corporation, Paramus, NJ). The arm and the hand were immobilized. The nerve was stimulated with a train-of-four: four stimuli at 2.0 Hz were applied every 12 s. The ratios between the T1 and the baseline value of T1 before rocuronium administration were determined to describe the effect. Patients reaching less than 80% twitch depression were excluded from the study. Twitch depression measurements were performed according to predefined guidelines.<sup>12,13</sup>

The amplifier of the force transducer and the sampling machine were connected to a data acquisition computer (3T model PS1600, Twente Technology Transfer, Enschede, The Netherlands) for the registration of the events during the experiment.

#### Analytical Methods

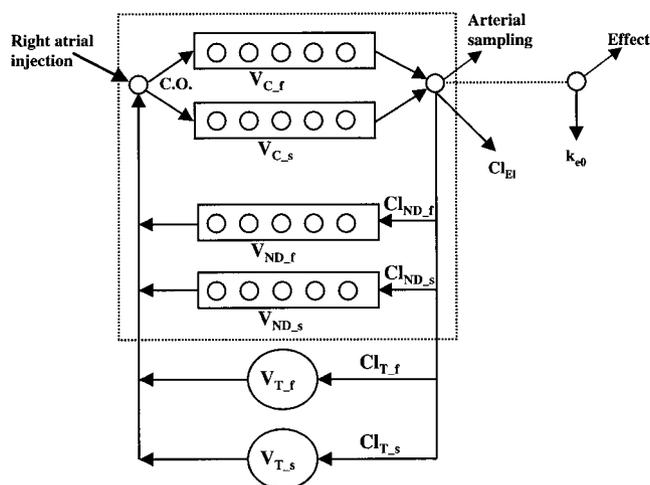
The concentration of both the ICG and rocuronium were measured in all samples collected during the first 10 min; in samples collected after 10 min, only rocuronium concentrations were measured. Blood ICG concentrations were measured spectrophotometrically at 805 nm. For each experiment, a reference line was constructed from whole blank blood from the patient and known amounts of ICG. The absorption at 805 nm caused by ICG was considered to be the measured absorption minus the absorption measured in the blank sample collected before the experiment.

Whole-blood rocuronium concentrations were measured by high-performance liquid chromatography with a fluorescence detector, according to the method described by Kleef *et al.*<sup>10</sup> These measurements were performed at the Research Laboratory of the Research Group for Experimental Anesthesiology and Clinical Pharmacology, University Hospital Groningen, The Netherlands.

#### Data Analysis

For the pharmacokinetic analysis, a recirculatory model described by Krejcie *et al.*<sup>6</sup> (fig. 1), was used. In this model, the central assumption is that ICG is confined to the intravascular space and thereby defines the intravascular kinetics of other simultaneously injected drugs, in this case rocuronium.

The central intravascular part of the model, representing blood flow through the heart and the lungs, was described by two combined parallel pathways, a fast and a slow central pathway.<sup>14</sup> The shape of the first-pass concentration-time curve of ICG (*i.e.*, data before evidence of ICG recirculation) was described by the sum of



**Fig. 1.** The recirculatory model used to analyze indocyanine green and rocuronium pharmacokinetics and the rocuronium pharmacodynamics. The parts in the dashed box represent the model for indocyanine green. The complete model is used to describe rocuronium kinetics and dynamics.  $Cl_{EI}$  = elimination clearance;  $Cl_{ND,f}$  = fast nondistributive clearance;  $Cl_{ND,s}$  = slow nondistributive clearance;  $Cl_{T,f}$  = fast tissue clearance;  $Cl_{T,s}$  = slow tissue clearance; CO = cardiac output;  $k_{e0}$  = blood-effect site equilibration rate constant;  $V_{C,f}$  = fast central volume;  $V_{C,s}$  = slow central volume;  $V_{ND,f}$  = fast peripheral nondistributive volume;  $V_{ND,s}$  = slow peripheral nondistributive volume;  $V_{T,f}$  = fast peripheral tissue volume;  $V_{T,s}$  = slow peripheral tissue volume.

two Erlang distribution functions, each representing the convolution of  $n$  1-compartment models connected in series<sup>15</sup>:

$$C(t) = A_1 \cdot \frac{k_1^{n_1} t^{n_1-1}}{(n_1 - 1)!} e^{-k_1 t} + A_2 \cdot \frac{k_2^{n_2} t^{n_2-1}}{(n_2 - 1)!} e^{-k_2 t}$$

where  $n_1$  and  $n_2$  are the number of compartments in series in the central delay elements,  $k_1$  and  $k_2$  are the rate constants between the compartments in series,  $n_1/k_1$  and  $n_2/k_2$  are the mean transit times of the central delay elements, and  $A_1$  and  $A_2$  the areas under the first-pass concentration-time curves. The CO was determined by dividing the dose of ICG by the area under the first-pass ICG concentration-time curve ( $A_1 + A_2$ ). The two Erlang functions were fitted to the data using the solver function in Quattro Pro (Borland, Scotts Valley, CA). The data were weighted uniformly during the first-pass fitting.

The parameters obtained from the Erlang functions were used as fixed parameters in a complete recirculatory model for ICG, including parallel fast and slow peripheral nondistributive circuits (each characterized by a volume and clearance:  $V_{ND,f}$  and  $Cl_{ND,f}$ , and  $V_{ND,s}$  and  $Cl_{ND,s}$ , respectively) and elimination clearance ( $Cl_{EI}$ ). The sum of the clearances through the parallel fast and slow peripheral nondistributive circuits for ICG (for which clearances equal blood flows) equals the CO.<sup>6</sup> Estimates of the model parameters were obtained by an iterative fitting procedure using the SAAM II program (SAAM Institute,

Seattle, WA). In this analysis, the data were iteratively re-weighted over the predicted values.

The intravascular pharmacokinetic parameters of ICG were used to evaluate the arterial rocuronium data by adding to the central intravascular model a fast and a slow distributive peripheral tissue compartment (characterized by  $V_{T,f}$  and  $Cl_{T,f}$ , and  $V_{T,s}$  and  $Cl_{T,s}$ , respectively). For rocuronium, the ratio between fast and slow peripheral nondistributive clearances was set equal to that for ICG, but absolute values were not the same as for ICG.

The mean transit time (MTT) determines whether a nondistributive pathway is either a fast or slow pathway and similarly whether a distributive pathway represents a rapidly or slowly equilibrating tissue compartment. The MTTs equal the (blood) volumes of the compartments or compartments in series divided by the (blood) flow through the compartments. The total peripheral tissue MTT ( $MTT_T$ ) is taken as the average of the fast peripheral MTT ( $MTT_{T,f}$ ) and the slow peripheral MTT ( $MTT_{T,s}$ ), weighted for the percentage of total blood flow through the fast and the slow peripheral distributive pathways. The fast nondistributive MTT ( $MTT_{ND,f}$ ) and the slow nondistributive MTT ( $MTT_{ND,s}$ ) are the same for ICG and rocuronium.

The relation between the measured arterial concentrations and the percentage neuromuscular block was described by connecting an effect compartment to the pharmacokinetic model (fig. 1) with a link parameter. The relation between effect-compartment concentration and effect was described with a sigmoid maximum effect ( $E_{max}$ ) function with the baseline fixed at 0 % and the  $E_{max}$  fixed at 100 %, using the SAAM II program. Data with twitch height exceeding the control twitch height were not included in the analysis.

For the pharmacokinetic analysis, we also used a conventional compartmental model to be able to compare these parameters with the recirculatory parameters. Both three- and two-compartmental models were fitted. The optimal model was selected on the basis of the Akaike criterion.<sup>16</sup> The concentrations at 1, 2, 3, 4, 7, 10, 15, 30, 60, 120, 180, and 240 min were used for the compartmental analysis. The pharmacokinetic parameters obtained with this model were also used for a pharmacokinetic-pharmacodynamic analysis as described by Sheiner *et al.*<sup>17</sup> This analysis was also performed in SAAM II, with iteratively reweighting over the predicted concentrations.<sup>17</sup>

**Statistics**

Patient characteristics and pharmacokinetic and pharmacodynamic parameters are presented as mean  $\pm$  SD. The correlations between CO and the pharmacokinetic and pharmacodynamic parameters and  $k_{e0}$ , were evaluated using linear regression. The pharmacodynamic parameters, derived with the recirculatory and compartmental models, were compared with the paired  $t$  test.

Downloaded from http://ahajournals.org/ at National Institute of Health on December 20, 2014



**Table 2. Volumes (l) for Recirculatory ICG and Rocuronium Pharmacokinetics and Their Relation with CO**

		V <sub>C</sub>	V <sub>ND,f</sub>	V <sub>ND,s</sub>	V <sub>T,f</sub>	V <sub>T,s</sub>	V <sub>T</sub>	V <sub>SS</sub>
ICG	Mean	1.52	0.34	1.48				3.35
	SD	0.40	0.24	0.37				0.63
	Slope	0.34	0.0033	0.069				0.412
	Intercept	0.21	0.33	1.214				1.76
	r <sup>2</sup>	0.68*	0.00	0.03				0.40*
Rocuronium	Mean	1.52	0.18	0.82	4.92	9.85	14.77	17.29
	SD	0.40	0.17	0.46	3.86	4.47	4.85	4.82
	Slope	0.34	-0.053	-0.083	-0.03	1.68	1.64	1.85
	Intercept	0.21	0.383	1.14	5.04	3.39	8.42	10.16
	r <sup>2</sup>	0.68*	0.09	0.03	0.001	0.13	0.11	0.14

\* P < 0.05. Note that values of V<sub>C</sub> were estimated from ICG concentration–time data and included as fixed parameters in the full recirculatory model for rocuronium (see Methods).

ICG = indocyanine green; CO = cardiac output; V<sub>C</sub> = central volume; V<sub>ND,f</sub> = fast peripheral nondistributive volume; V<sub>ND,s</sub> = slow peripheral nondistributive volume; V<sub>T,f</sub> = fast peripheral tissue volume; V<sub>T,s</sub> = slow peripheral tissue volume; V<sub>T</sub> = peripheral tissue volume; V<sub>SS</sub> = total distribution volume.

and age and k<sub>co</sub> of the recirculatory model (r<sup>2</sup> = 0.36; P = 0.029) were found to be correlated. No correlation between age and k<sub>co</sub> of the compartmental model was found.

**Discussion**

Until now, recirculatory models, the models described in this study, have not been used in pharmacokinetic–pharmacodynamic modeling in patients. In this study we demonstrated that the recirculatory pharmacokinetic model adequately describes the first-pass pharmacokinetics of rocuronium in patients. We also demonstrated that the recirculatory model can be used for pharmacokinetic–pharmacodynamic modeling and is capable of revealing that the k<sub>co</sub> of rocuronium, a drug with a fast onset of action, is strongly dependent on CO. The results of this study indicate that there is a significant difference between k<sub>co</sub> and CE<sub>50</sub>, estimated with the aid of a recirculatory model *versus* a compartmental pharmacokinetic model after a bolus injection of rocuronium.

The importance of the pharmacokinetics and pharmacodynamics during the first minutes after a bolus injection, especially for drugs with a fast onset of effect, has

been emphasized.<sup>18</sup> The inability of conventional compartmental models to describe the initial mixing of drug was described by Chiou in 1979.<sup>19</sup> Since then, recirculatory models have been used as an alternative to describe the pharmacokinetics of various drugs in animals.<sup>20,21</sup> Minimal recirculatory models<sup>22</sup> and a precursor model of the more extensive recirculatory models have been used to describe pharmacokinetics in patients. However, to our knowledge these specific recirculatory models with their ability to account for the influence of blood flow have not previously been used for integrated pharmacokinetic–pharmacodynamic modeling in patients.

The k<sub>co</sub> and CE<sub>50</sub> of the recirculatory model differ significantly from the values found with a compartmental model (see table 6). This is undoubtedly related to the inability of compartmental models to characterize the concentration–time relation during the first few minutes after a bolus injection, *e.g.*, the front-end kinetics. A compartmental model misspecifies the early time course of drug concentration because it assumes the body to be homogeneous rather than a complicated system of tissues and organs in series and parallel.<sup>23</sup> Depending on the model, *i.e.*, two- or three-compartmental, initial drug concentrations may be either seriously underestimated

**Table 3. Clearances (l/min) for Recirculatory ICG and Rocuronium Pharmacokinetics and Their Relation with CO**

		Cl <sub>EI</sub>	Cl <sub>ND,f</sub>	Cl <sub>ND,s</sub>	Cl <sub>T,f</sub>	Cl <sub>T,s</sub>	Cl <sub>T</sub>	ΣCl
ICG	Mean	0.69	1.87	1.29				3.86
	SD	0.19	0.64	0.44				0.97
	Slope	0.13	0.58	0.28				
	Intercept	0.19	-0.35	0.20				
	r <sup>2</sup>	0.43*	0.76*	0.40*				
Rocuronium	Mean	0.45	1.02	0.68	1.47	0.23	1.70	3.86
	SD	0.11	0.57	0.38	0.94	0.20	1.13	0.97
	Slope	-0.006	0.13	0.081	0.70	0.11	0.81	
	Intercept	0.47	0.50	0.36	-1.23	-0.20	-1.43	
	r <sup>2</sup>	0.003	0.051	0.04	0.51*	0.31*	0.49*	

\* P < 0.05.

ICG = indocyanine green; CO = cardiac output; Cl<sub>EI</sub> = elimination clearance; Cl<sub>ND,f</sub> = fast nondistributive clearance; Cl<sub>ND,s</sub> = slow nondistributive clearance; Cl<sub>T,f</sub> = fast tissue clearance; Cl<sub>T,s</sub> = slow tissue clearance; Cl<sub>T</sub> = peripheral tissue clearance; ΣCl = sum of the elimination clearance, fast and slow nondistributive clearances, and, when applicable, fast and slow tissue clearances.

Downloaded from http://ajph.aphspublishers.org/ at guest on December 10, 2012

**Table 4. Mean Transit Times (min) for Recirculatory ICG and Rocuronium Pharmacokinetics and Their Relation with CO**

		MTT <sub>C</sub>	MTT <sub>ND,f</sub>	MTT <sub>ND,s</sub>	MTT <sub>T,f</sub>	MTT <sub>T,s</sub>	MTT <sub>T</sub>
ICG	Mean	0.40	0.20	1.22			
	SD	0.07	0.13	0.36			
	Slope	-0.012	-0.075	-0.22			
	Intercept	0.45	0.49	2.06			
	r <sup>2</sup>	0.031	0.31*	0.33*			
Rocuronium	Mean	0.40	0.20	1.22	4.84	54.94	11.47
	SD	0.07	0.13	0.36	3.05	22.27	6.47
	Slope	-0.012	-0.075	-0.22	-2.26	-9.09	-4.49
	Intercept	0.45	0.49	2.06	13.55	90.02	28.79
	r <sup>2</sup>	0.031	0.31*	0.33*	0.51*	0.16	0.45*

\*  $P < 0.05$ .

ICG = indocyanine green; CO = cardiac output; MTT<sub>C</sub> = central mean transit time; MTT<sub>ND,f</sub> = fast nondistributive mean transit time; MTT<sub>ND,s</sub> = slow nondistributive mean transit time; MTT<sub>T,f</sub> = fast peripheral mean transit time; MTT<sub>T,s</sub> = slow peripheral mean transit time; MTT<sub>T</sub> = total peripheral tissue mean transit time.

or overestimated. In contrast, recirculatory models are capable of accurately describing the front-end kinetics.<sup>7</sup> Obviously, inadequate characterization of the plasma concentration-time relation also has consequences for the pharmacodynamic analysis. When initial plasma concentrations are either underestimated or overestimated, the plasma concentration-effect relation will be distorted. This does not only affect the estimation of  $k_{e0}$ , but also the estimated pharmacodynamic parameters, as has been demonstrated by Ducharme *et al.*<sup>9</sup> and Fisher *et al.*<sup>8</sup> These investigators examined the pharmacodynamics of vecuronium using nonparametric and semiparametric models, respectively, and demonstrated that estimation of  $k_{e0}$  and pharmacodynamic parameters depends on adequate characterization of the plasma concentration-time curve, including the initial mixing phase. In this study, the  $k_{e0}$  estimated with a compartmental model was nearly twofold larger than that estimated with a recirculatory model, and the CE<sub>50</sub> was approximately 22% lower. A recent study by Beaufort *et al.*<sup>24</sup> showed that including early blood samples in the pharmacokinetic-pharmacodynamic analysis of rocuronium and other muscle relaxants in pigs resulted in a significantly higher CE<sub>50</sub> and a significantly smaller  $k_{e0}$  compared with the results obtained with a conventional sampling schedule.

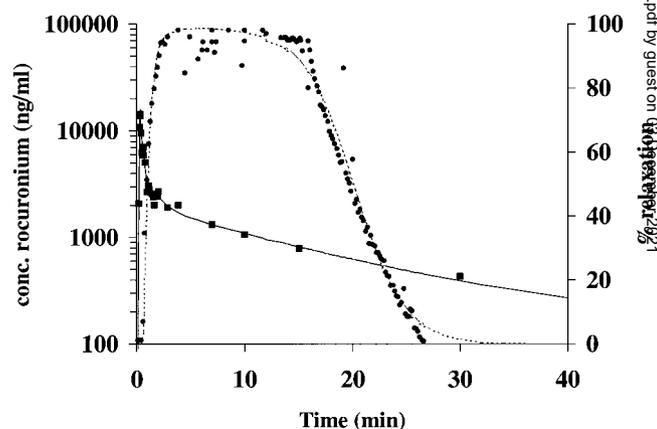
This study showed a significant correlation between CO and  $k_{e0}$  for both the recirculatory and the compartmental model. However, the correlation between CO and  $k_{e0}$  was much stronger with the recirculatory model

**Table 5. The Pharmacokinetic Parameters for Rocuronium Obtained with a Two-compartmental Model**

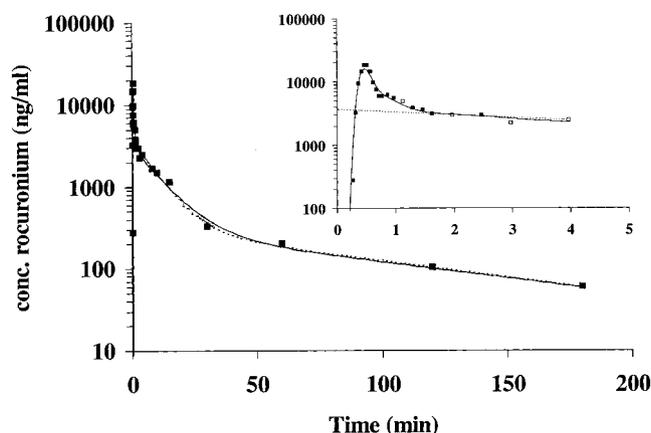
Parameters	Two-compartmental (n = 15)
V <sub>1</sub> (l)	6.76 ± 1.69
V <sub>2</sub> (l)	3.73 ± 1.98
V <sub>SS</sub> (l)	10.50 ± 3.54
Cl <sub>EI</sub> (l/min)	0.50 ± 0.14
Cl <sub>12</sub> (l/min)	0.43 ± 0.20

V<sub>SS</sub> = total distribution volume; Cl<sub>EI</sub> = elimination clearance.

( $r^2 = 0.70$ ; fig. 6). When CO was corrected for body weight (CI), the relation between CI and  $k_{e0}$  still showed a very strong correlation for the recirculatory model ( $r^2 = 0.60$ ,  $P = 0.002$ ), whereas the relation for the compartmental model was no longer significant ( $P > 0.05$ ). This indicates that the relation is probably not weight-dependent. CO is known to play an important role in the pharmacokinetics and the time course of action of muscle relaxants, including the onset of action during the initial distribution phase.<sup>25</sup> This is especially important for muscle relaxants that are used for rapid sequence induction, because the time to intubation must be as short as possible. The onset time of paralysis after intravenous bolus administration of a neuromuscular blocking drug depends mainly on the speed at which the relaxant reaches the postsynaptic acetylcholine receptors. Because muscle blood flow is related to CO, the high correlation between CO and  $k_{e0}$  strongly suggests a crucial role of muscle blood flow, *i.e.*, that the distribution to the site of action is perfusion-limited. This in al



**Fig. 4.** The concentration-time and effect-time relations of an individual patient during the first 40 min after the bolus injection. The straight line describes the fit of the recirculatory model, and the dashed line describes the accompanying pharmacodynamic fit. The squares represent the measured rocuronium concentrations, and the dots the measured muscle relaxation.



**Fig. 5.** The concentration–time relation of rocuronium for one individual patient for the whole experiment. The straight line represents the fit with the recirculatory model, and the dashed line represents the fit with the two-compartmental model. (Inset) The concentration–time relation is shown for the same patient for the first 4 min after bolus injection, and the measured concentrations that were used for the compartmental fit are left open. Note that samples collected during the first 20 s contained no or very small (< 10 ng/ml) concentrations of rocuronium and are therefore not shown.

likelihood constitutes the physiological basis of  $k_{e0}$  for drugs that have no diffusion limitation in their transfer to the effect sites.

In the present study,  $k_{e0}$  and pharmacodynamic parameters were derived after a bolus injection of rocuronium. It is conceivable that if these parameters were studied during and after a short infusion, the values derived by compartmental analysis would approach those derived by recirculatory analysis, because they would depend much less on the initial concentration–time data. On the other hand, the parameters thus derived may have a limited predictive value when the drug is administered as a bolus injection. Plaud *et al.*<sup>26</sup> studied rocuronium pharmacokinetics and their relation with pharmacodynamics after a 5-min infusion. The parameters they found

( $k_{e0} = 0.168 \text{ min}^{-1}$ ;  $CE_{50} = 823 \text{ } \mu\text{g/ml}$ ) are comparable with the parameters obtained with the recirculatory pharmacokinetic model in the present study ( $k_{e0} = 0.129 \text{ min}^{-1}$ ;  $CE_{50} = 876 \text{ } \mu\text{g/ml}$ ).

Our results showed a significant correlation between age and CO ( $r^2 = 0.39$ ;  $P = 0.013$ ) and between age and  $k_{e0}$  of the recirculatory model, whereas  $k_{e0}$  of the compartmental model was not age-dependent. Previous studies showed a relation between the  $k_{e0}$  and age for propofol<sup>27,28</sup> and for remifentanyl,<sup>5</sup> both drugs with a fast onset of effect. Schnider *et al.*<sup>28</sup> also showed a relation between the time to peak effect of propofol and age, in which the time to peak effect was longer in the elderly. In our opinion, these observations may be, among other reasons, a result of age-related changes in CO. Propofol and remifentanyl are fast-acting drugs, and therefore the onset is likely to be flow-dependent. When blood–effect-site concentration equilibration is rapid, it is more likely to be flow-dependent, and therefore this could also be caused by age-dependent changes in CO.

The initial concentrations predicted (fitted) by the compartmental models depend heavily on the timing of first sampling and the number of compartments. The dependence on the number of compartments is illustrated in figure 7. The first concentration included in the compartmental fits was the 1-min sample, as this is the first sample included in most compartmental analyses. As illustrated, the three-compartmental model fitted the first data point much better than the two-compartmental model. However, based on an objective criterion (Akaike) and in view of the uncertainty in, and high correlation between, some of the model parameters, inclusion of a third compartment was not warranted.

These recirculatory pharmacokinetic models have been previously used only for analysis of animal data.<sup>6,22</sup> A difference with the use in patients lies in the fact that the venous injection site and the arterial sampling sites were much more peripherally located in patients than in the animal studies. As a result, the central part of the recirculatory model represents more than just the heart and lungs. In our patients, the total central blood volume ( $V_C$ ) was 1.52 l, representing 45% of the total blood volume, whereas in previous animal studies,  $V_C$  represented a smaller fraction of the total blood volume: 31%,<sup>29</sup> 34%,<sup>6</sup> and 32%.<sup>30</sup> Another implication of the peripheral bolus injection is the lack of correlation between  $MTT_c$  and CO.

The pharmacokinetic parameters found by Plaud *et al.*,<sup>26</sup> who used a two-compartmental pharmacokinetic model, can be compared with our results of the compartmental parameters but also with some recirculatory parameters. They found a total distribution volume ( $V_{ss}$ ) of 14.8 l, which is in the same order as the value we found for the compartmental model ( $V_{ss} = 10.5 \text{ l}$ ) and with the recirculatory model ( $V_{ss} = 17.3 \text{ l}$ ). The elimination clearance of the two different models found in

**Table 6.** Pharmacodynamic Parameters Obtained with Recirculatory and Compartmental Pharmacokinetic Modeling and Their Relation with CO

Parameter		Recirculatory	Two-compartmental
$k_{e0}$ ( $\text{min}^{-1}$ )	Mean $\pm$ SD	$0.129 \pm 0.036$	$0.239 \pm 0.104^*$
	Slope	0.033	0.071
	Intercept	0.0042	-0.028
	$r^2$	0.70†	0.38‡
$\gamma$	Mean $\pm$ SD	$8.75 \pm 2.31$	$8.96 \pm 9.22$
	Slope	0.392	1.767
	Intercept	7.28	2.33
	$r^2$	0.024	0.030
$CE_{50}$ ( $\mu\text{g/l}$ )	Mean $\pm$ SD	$876 \pm 118$	$684 \pm 97^*$
	Slope	44.26	46.83
	Intercept	710	508
	$r^2$	0.12	0.19

\*  $P < 0.001$  versus recirculatory model. †  $P < 0.001$ . ‡  $P = 0.025$ .  
CO = cardiac output;  $k_{e0}$  = blood–effect-site equilibration constant;  $CE_{50}$  = effect compartment concentration at 50% of the maximum effect.

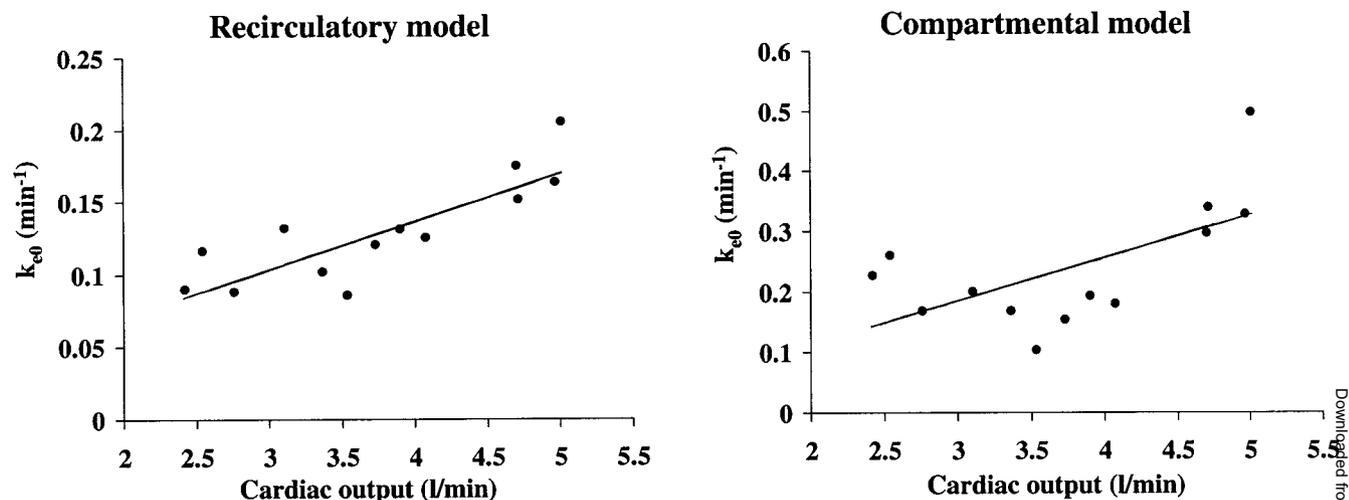


Fig. 6. Relation between the cardiac output and the blood-effect site equilibration rate constant ( $k_{e0}$ ) for the recirculatory model and the compartmental model. The lines represent the linear relation.

this study (0.45 l/min and 0.50 l/min) are also comparable to the value of 0.58 l/min reported by Plaud *et al.*<sup>26</sup>

Some of the pharmacokinetic parameters were correlated to CO (tables 2-4). The influence of increasing CO on the tissue clearance or distribution rate is obvious, at least when tissue uptake is perfusion limited, as is the case with rocuronium. The influence on total distribution volumes is less obvious but has been reported previously.<sup>29</sup> The elimination clearance of rocuronium was not correlated with CO because rocuronium has a relatively low hepatic extraction ratio, and elimination clearance is probably more dependent on protein binding and hepatic enzyme activity (intrinsic clearance).<sup>31</sup>

Our observations have some clinical implications. As expected from the smaller  $k_{e0}$  in patients with a lower CO, these patients generally required a longer time before a twitch depression of more than 90% was obtained. For example, in the patient with the lowest CO (2.4 l/min) 90% twitch depression was obtained after 2.5 min, whereas in

the patient with the highest CO (5.0 l/min), who was included in the pharmacodynamic evaluation, this was obtained after 1.5 min. In the context of a rapid-sequence induction, this indicates that not only the selection and the dose of the muscle relaxant are important, but the physiologic condition of the patient should also be taken into account.

In conclusion, this study demonstrated that recirculatory pharmacokinetic modeling adequately characterized the plasma concentration-time relation of rocuronium including the initial mixing phase, after a bolus dose. In addition, the study illustrated the role of the CO with respect to the pharmacokinetics, including the blood-effect site equilibration rate. Furthermore, it was shown that inadequate characterization of the initial concentrations using conventional pharmacokinetic models affected the estimated  $k_{e0}$  and  $CE_{50}$ . Adequate characterization of the initial phase using recirculatory modeling results in more realistic, physiologically based estimation of these parameters.

The authors thank Dr. J. Swen, M.D., Ph.D. (Department of Anesthesiology, Leiden University Medical Center, Leiden, The Netherlands), for help with the muscle relaxation measurements, M. J. Geerts, B.Sc. (Department of Anesthesiology, Leiden University Medical Center), for assistance during the experiments, and U. W. Kleef, B.Sc., and J. Roggevelde (Research Group for Experimental Anesthesiology and Clinical Pharmacology, University Hospital Groningen, Groningen, The Netherlands) for the determination of the rocuronium concentrations.

## References

1. Upton RN, Ludbrook GL, Grant C, Martinez AM: Cardiac output is a determinant of the initial concentrations of propofol after short-infusion administration. *Anesth Analg* 1999; 89:545-52
2. Sheiner LB, Stanski DR, Vozeh S, Miller RD, Ham J: Simultaneous modeling of pharmacokinetics and pharmacodynamics: Application to d-turbocurarine. *Clin Pharmacol Ther* 1979; 25:358-71
3. Hull CJ, van Beem HBH, McLeod K, Sibbald A, Watson MJ: A pharmacodynamic model for pancuronium. *Br J Anaesth* 1978; 50:1113-23
4. Wierda JMKH, Proost JH: The pharmacokinetics and pharmacokinetic-dynamic relationship of rocuronium bromide. *Anaesthetic Pharmacol Rev* 1995; 3:192-201
5. Minto CF, Schnider TW, Egan TD, Youngs E, Lemmens HJM, Gambus PL, Billard V, Hoke JF, Moore KHP, Hermann DJ, Muir KT, Mandema JW, Shafer SL:

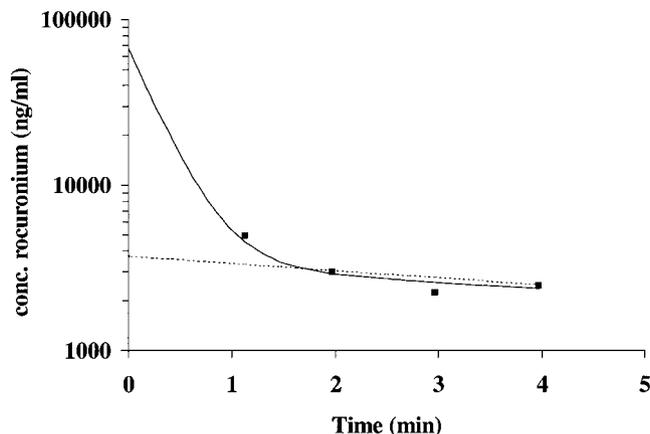


Fig. 7. The concentration-time relation of one individual patient for the first 4 min after bolus injection. The straight line represents the fit with a three-compartmental model, and the dashed line represents the fit with a two-compartmental model.

Influence of age and gender on the pharmacokinetics and pharmacodynamics of remifentanyl. I. Model development. *ANESTHESIOLOGY* 1997; 86:10-23

6. Krejcie TC, Henthorn TK, Niemann CU, Klein C, Gupta DK, Gentry WB, Shanks CA, Avram MJ: Recirculatory pharmacokinetic models of markers of blood, extracellular fluid and total body water administered concomitantly. *J Pharmacol Exp Ther* 1996; 278:1050-7

7. Krejcie TC, Avram MJ: What determines anesthetic induction dose? It's the front-end kinetics, doctor! [editorial]. *Anesth Analg* 1999; 89:541-4

8. Fisher DM, Szenohradszy J, Wright PMC, Lau M, Brown R, Sharma M: Pharmacodynamic modeling of vecuronium-induced twitch depression. *ANESTHESIOLOGY* 1997; 86:558-66

9. Ducharme J, Varin F, Bevan DR, Donati F: Importance of early blood sampling on vecuronium pharmacokinetic and pharmacodynamic parameters. *Clin Pharmacokinet* 1993; 24:507-18

10. Kleef UW, Proost JH, Roggevel J, Wierda JMKH: Determination of rocuronium and its putative metabolites in body fluids and tissue homogenates. *J Chromatogr* 1993; 621:65-76

11. Mellinghoff H, Diefenbach C, Bischoff A, Grond S, Buzello W: Dose-response relationship of rocuronium bromide during intravenous anaesthesia. *Eur J Anaesthesiol Suppl* 1994; 9:20-4

12. Viby-Mogensen J, Engbæk J, Eriksson LI, Gramstad L, Jensen E, Jensen FS, Koscielniak-Nielsen Z, Skovgaard LT, Ostergaard D: Good clinical research practice (GCRP) in pharmacodynamic studies of neuromuscular blocking agents. *Acta Anaesthesiol Scand* 1996; 40:59-74

13. Beemer GH, Goonetilleke PH: Monitoring neuromuscular transmission. *Curr Anaesth Crit Care* 1996; 7:101-6

14. Krejcie TC, Jacquez JA, Avram MJ, Niemann CU, Shanks CA, Henthorn TK: Use of parallel erlang density functions to analyze first-pass pulmonary uptake of multiple indicators in dogs. *J Pharmacokinet Biopharm* 1996; 24:569-88

15. Avram MJ, Krejcie TC, Niemann CU, Klein C, Gentry WB, Shanks CA, Henthorn TK: The effect of halothane on the recirculatory pharmacokinetics of physiologic markers. *ANESTHESIOLOGY* 1997; 87:1381-93

16. Akaike H: A new look at the statistical model identification. *IEEE Trans Automat Contr* 1974; 19:716-23

17. Gabrielsson J, Weiner D: Mathematics of parameter estimation, Pharmacokinetic and Pharmacodynamic Data Analysis: Concepts and Applications. Edited by Gabrielsson, Weiner. Stockholm, Thony Börk, 1994, pp 9-28

18. Runciman WB, Upton RN: Pharmacokinetics and pharmacodynamics: What is of value to the practising anaesthetist? *Anaesthetic Pharmacol Rev* 1994; 2:280-93

19. Chiou WL: Potential pitfalls in the conventional pharmacokinetic studies: Effects of the initial mixing of drug in blood and the pulmonary first-pass elimination. *J Pharmacokinet Biopharm* 1979; 7:527-36

20. Krejcie TC, Henthorn TK, Shanks CA, Avram MJ: A recirculatory pharmacokinetic model describing the circulatory mixing, tissue distribution and elimination of antipyrine in dogs. *J Pharmacol Exp Ther* 1994; 269:609-16

21. Henthorn TK, Avram MJ, Krejcie TC, Shanks CA, Asada A, Kaczynski DA: Minimal compartmental model of circulatory mixing of indocyanine green. *Am J Physiol* 1992; 262:H903-10

22. Weiss M, Hübner GH, Hübner IG, Teichmann W: Effects of cardiac output on disposition kinetics of sorbitol: recirculatory modelling. *Br J Clin Pharmacol* 1996; 41:261-8

23. Fisher DM: (Almost) Everything you learned about pharmacokinetics was (somewhat) wrong! *Anesth Analg* 1996; 83:901-3

24. Beaufort TM, Proost JH, Kuizenga K, Houwertjes MC, Kleef UW, Wierda JMKH: Do plasma concentrations obtained from early arterial blood sampling improve pharmacokinetic/pharmacodynamic modeling? *J Pharmacokinet Biopharm* 1999; 27:173-90

25. Lee C, Katz RL: Neuromuscular pharmacology: A clinical update and commentary. *Br J Anaesth* 1980; 52:173-88

26. Plaud B, Proost JH, Wierda JMKH, Barre J, Debaene B, Meistelman C: Pharmacokinetics and pharmacodynamics of rocuronium at the vocal cords and the adductor pollicis in humans. *Clin Pharmacol Ther* 1995; 58:185-91

27. Kazama T, Ikeda K, Morita K, Kikura M, Doi M, Ikeda T, Kurita T, Nakajima Y: Comparison of the effect-site  $k(e)_{0.5}$ s of propofol for blood pressure and EEG bispectral index in elderly and younger patients. *ANESTHESIOLOGY* 1999; 90:1517-27

28. Schnider TW, Minto CF, Shafer SL, Gambus PL, Andresen C, Goodale DR, Youngs EJ: The influence of age on propofol pharmacodynamics. *ANESTHESIOLOGY* 1999; 90:1502-16

29. Kuipers JA, Boer F, Olofsen E, Olieman W, Vletter AA, Burm AGL, Bovill JG: Recirculatory and compartmental pharmacokinetic modeling of alfentanil in pigs: The influence of cardiac output. *ANESTHESIOLOGY* 1999; 90:1146-57

30. Krejcie TC, Avram MJ, Gentry WB, Niemann CU, Janowski MP, Henthorn TK: A recirculatory model of the pulmonary uptake and pharmacokinetics of lidocaine based on analysis of arterial and mixed venous data from dogs. *J Pharmacokinet Biopharm* 1997; 25:169-90

31. Blaschke TF: Protein binding and kinetics of drugs in liver diseases. *Clin Pharmacokinet* 1977; 2:32-44