

The Effects of CO₂ on Respiratory Mechanics in Anesthetized Paralyzed Humans

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Background: There is little information concerning the carbon dioxide-related effects on respiratory mechanics in anesthetized, paralyzed subjects; however, hypocapnia or hypercapnia is often permitted in patients with severe brain injury or acute respiratory distress syndrome. Therefore, the carbon dioxide dependence of respiratory mechanics in healthy anesthetized, paralyzed subjects was investigated.

Methods: Interrupter resistance (Rint), additional tissue viscoelastic resistance (ΔR), and quasi-static elastance (Est) of lung (L) and chest wall were assessed by means of the rapid end-inspiratory occlusion method in two groups of seven healthy paralyzed subjects anesthetized with diazepam or isoflurane. They underwent ventilation with a fixed pattern and hyperoxic gas mixtures with different fractions of inspired carbon dioxide (FICO₂) to produce a partial pressures of arterial carbon dioxide (Paco₂) of 24.4 ± 3.4, 39.6 ± 3.2, and 62 ± 4.1 (SD) mmHg.

Results: Chest wall mechanics and Est,L were unaffected by Paco₂ changes. With diazepam anesthesia, Rint,L decreased linearly, with increasing Paco₂, from 2.3 to 1.4 cm H₂O · s · l⁻¹, whereas ΔR ,L decreased from 2 to 1.7 cm H₂O · s · l⁻¹, though not significantly. With isoflurane anesthesia, the decrease of Rint,L (0.2 ± 0.5 cm H₂O · s · l⁻¹) was not significant, and ΔR L remained unchanged. With diazepam, Rint,L was 45 (hypercapnia) to 110% (hypocapnia) greater than with isoflurane.

Conclusions: Changes of Paco₂ from 20–65 mmHg cause increasing bronchodilation in anesthetized, paralyzed subjects, this effect being attenuated or abolished by drugs (e.g., halogenated anesthetics) that depress smooth muscle tone substantially. The carbon dioxide bronchodilating effects are probably direct for peripheral structures and are paralleled by a tendency of lung tissue resistance to decrease. Because local Paco₂-related changes in bronchomotor tone promote \dot{V}_A/\dot{Q} matching, this mechanism should be impaired by anesthetics that cause bronchodilation.

ALTHOUGH patients with severe brain injury are routinely made to hyperventilate¹ and permissive hypercapnia is advocated for patients with acute respiratory distress syndrome,² there are no reports on the effects of changes in partial pressure of arterial carbon dioxide (Pco₂) on respiratory mechanics in patients undergoing mechanical ventilation and healthy subjects, except for the report by Don and Robson.³ Using the rapid end-inspiratory occlusion technique, they measured the in-

terrupter resistance (Rint,rs) of the respiratory system in healthy subjects anesthetized with nitrous oxide (N₂O) and found that, during hypocapnia, Rint,rs was significantly higher than during normocapnia. The high mean value of Rint,rs obtained during normocapnic conditions suggests that the anesthetic regimen used by these authors increases bronchomotor tone. Conversely, during inhalation of halogenated agents (halothane, enflurane, methoxyflurane), which markedly reduce bronchomotor tone, a progressive decrease of hypocapnic bronchoconstriction has been found with increasing concentration of the anesthetic agents in isolated dog lobes.⁴ To our knowledge, there are no reports on the effects of either hypocapnia or hypercapnia on Rint and the additional resistance (ΔR), which reflects pressure dissipation caused by viscoelastic behavior and time constant inequality, in humans anesthetized with halogenated agents.

Accordingly, in the current study, we assessed the effects of hypocapnia and hypercapnia on Rint and ΔR of the lung (L) and chest wall (w) on a group of healthy subjects anesthetized with isoflurane. In addition, we repeated the measurements on another group of healthy subjects anesthetized with diazepam. In contrast to isoflurane, diazepam, at least at subanesthetic doses, has been shown to enhance bronchomotor tone in healthy volunteers.⁵

Methods

Fourteen patients (10 men) undergoing general anesthesia for minor surgery were observed before undergoing intervention. None was obese or had a history or showed clinical evidence of cardiopulmonary disease. The institutional ethics committees (Azienda Ospedaliera di Lecco, Lecco and Istituti Clinici di Perfezionamento, Milan, Italy) approved the investigation, and informed consent was obtained from the patients. Depending on the type of anesthesia, they were classified into two groups (seven subjects, each; mean age (± SD), weight, and height were 27 ± 10 yr, 64 ± 10 kg, and 173 ± 6 cm, and 28 ± 5 yr, 69 ± 11 kg, and 171 ± 13 cm).

In the first group of patients, premedication and induction of anesthesia were obtained using diazepam (0.15 mg/kg intramuscular and 0.3–0.4 mg/kg intravenous, respectively); anesthesia was maintained with additional aliquots of diazepam (0.15 mg/kg intravenous) according to clinical requirements. The second group was premedicated with diazepam (0.2 mg/kg

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intramuscular) and anesthesia was induced using intravenous thiopental sodium (5–7 mg/kg); however, anesthesia was maintained with isoflurane (0.8–1%). In all cases, muscle relaxation was induced using pancuronium bromide (0.1 mg/kg) and maintained with additional aliquots (0.03 mg/kg), as needed. Placed in the supine position, the subjects were transorally intubated using a cuffed endotracheal (ET) tube (Mallinkrodt Medical, Athlow, Ireland; 7.5–8.5 mm ID; length, 30–36 cm) and underwent mechanical ventilation (Siemens Servo Ventilator 900C, Berlin, Germany). To reduce the resistance and compliance of the circuit, a single length of standard low-compliance tubing (2 cm ID; length, 110 cm) was used, and the humidifier was removed during the experiments.

Flow (\dot{V}) was measured using a heated pneumotachograph (Fleisch No. 2; Lausanne, Switzerland) connected to the breathing circuit *via* a cone and to a differential pressure transducer (Statham 270; Hewlett-Packard, Andover, MA). The response of the pneumotachograph was linear over the experimental range of flows. Tracheal pressure (Ptr) was measured by means of a pressure transducer (1290A; Hewlett-Packard) connected to a polyethylene catheter (1.5 mm ID; length, 50 cm), the tip of which, protected by a 2-mm thick ring to avoid the entrance of mucus, jutted 3 to 4 cm from the ET tube into the trachea. Esophageal pressure (Pes) was measured with a similar transducer connected to a thin-walled latex balloon (8 cm) filled with 0.5–1 ml of air through a polyethylene catheter (2 mm ID; length, 120 cm) with multiple holes in the last 5 cm near the closed tip. The validity of Pes measurements was verified before the induction of paralysis by use of the occlusion test.⁶ Transpulmonary pressure was obtained as $\text{Ptr} - \text{Pes}$. With this recording system, phase shift or alteration in amplitude up to 20 Hz did not affect pressure measurements. The signals from the transducers were amplified (Carrier 20-3615-45; Gould, Valley View, OH) and recorded on a personal computer *via* a 16-bit analog-to-digital converter at a sample rate of 200 Hz.

Arterial blood partial pressure of oxygen (Po₂), Pco₂ and pH were measured by means of a blood gas analyzer (IL 1620; Instrumentation Laboratory, Lexington, MA) on samples drawn at the beginning and at the end of each test. In the group of subjects during diazepam anesthesia, plasma obtained from these blood samples was also processed for assessment of catecholamine concentration.⁷ The heart rate (range: 80–105 min⁻¹), mean systemic arterial pressure (range: 90–105 mmHg), oxygen saturation (always $\geq 98\%$), and end-tidal concentration of carbon dioxide were continuously monitored in addition to the electrocardiogram (Siemens Monitor 7000).

Procedure and Data Analysis

The ventilator settings consisted of a fixed tidal volume (V_T; range: 0.77–1.02 l), inspiratory duration (T_i; range:

0.71–0.97 s) and flow (\dot{V}) range: 0.97–1.12 l/s). Respiratory frequency (range: 13–17 min⁻¹) was chosen to decrease the end-tidal carbon dioxide concentration while ventilating with a mixture of 60% N₂ in O₂. Two other mixtures, 5% CO₂–55% N₂ in O₂ and 9% CO₂–51% N₂ in O₂, were used sequentially to perform ventilation of the patients, before returning to the initial mixture. Each mixture was breathed for 9–10 min to ensure a steady end-tidal carbon dioxide concentration; thereafter a series of 25–30 breaths was obtained, in which an end-inspiratory pause, lasting 0.4–0.5 s, depending on the duration of inspiration, was automatically introduced by the ventilator, thus providing end-inspiratory occlusions on a breath-by-breath basis. Moreover, during 5 to 6 breaths, an end-inspiratory occlusion lasting 5 s was introduced by pressing the end-inspiratory hold button of the ventilator. A normally open, solenoid valve, with a closing time of 10 ms, placed between the Y piece of the breathing circuit and the pneumotachograph, was triggered by the inspiratory transistor-transistor logic signal of the ventilator. All measurements were performed at zero end-expiratory pressure applied by the ventilator.

Data analysis was performed as previously described in detail.^{8,9} Briefly, for each subject and gas mixture, 25 breaths with the short-lasting end-inspiratory pause and 5 to 6 breaths with the long-lasting end-inspiratory pause were ensemble averaged, and the signals of the individual breaths were superimposed at the onset of airway occlusion as detected on the flow trace. End-inspiratory airway occlusion was followed by a rapid initial decrease in transpulmonary pressure and Pes from the end-inspiratory (P_{max}) to a certain value (P₁), and by a slow decay that in breaths, with long-lasting occlusions eventually reached in approximately 4 s an apparent plateau value (P₂). The rapid pressure decreases (P_{max} – P₁) divided by the flow preceding the occlusion yield R_{int,L} and R_{int,w}, respectively. The slow pressure decreases (P₁ – P₂) divided by the flow preceding the occlusion yield ΔR_{L} and ΔR_{w} , respectively. Finally Est_L and Est_w were computed by dividing the corresponding P₂ value by V_T, obtained by numerical integration of the flow signal. In all instances, there was a pause (zero flow) at end-expiration, indicating absence of intrinsic positive end-expiratory pressure.

While R_{int,L} represents the flow resistance of the airways, ΔR_{L} reflects the additional pressure dissipation caused by viscoelastic behavior and time constant inequality.^{8,10} Similarly, R_{int,w} and ΔR_{w} represent, respectively, the ohmic resistance and the viscoelastic behavior of the chest wall tissues.^{8,9}

Statistical Analysis

Results are presented as the mean \pm SD. Paired Student *t* test was used to compare values of two samples from the same group of subjects. When a significant

Table 1. Arterial Blood Gases, pH, and Catecholamine Concentration at Different Inspired Carbon Dioxide Concentrations in Seven Normal Subjects Anesthetized with Diazepam (A) or Isoflurane (B)

| Fico ₂ | PaO ₂ (mmHg) | Paco ₂ (mmHg) | pHa | E (pg/ml) | NE (pg/ml) |
|-------------------|----------------------------|-----------------------------|-------------|-------------------|--------------------|
| 0 | | | | | |
| A | 233 ± 22 | 23.7 ± 2.8 | 7.53 ± 0.03 | 150 ± 26 (10–100) | 191 ± 72 (150–300) |
| B | 230 ± 32 | 25.1 ± 4.0 | 7.50 ± 0.06 | | |
| 0.05 | | | | | |
| A | 249 ± 44 | 40.6 ± 3.7 | 7.39 ± 0.03 | 143 ± 27 | 183 ± 68 |
| B | 246 ± 42 | 38.6 ± 2.5 | 7.37 ± 0.04 | | |
| 0.09 | | | | | |
| A | 222 ± 38 | 60.9 ± 4.8 | 7.27 ± 0.02 | 168 ± 58 | 203 ± 72 |
| B | 208 ± 26 | 63.1 ± 4.5 | 7.22 ± 0.05 | | |
| 0 | | | | | |
| A | 244 ± 52 | 25.0 ± 3.0 | 7.50 ± 0.03 | | |
| B | 235 ± 34 | 28.3 ± 6.0 | 7.45 ± 0.09 | | |

Values are mean ± SD. Normal values for plasma epinephrine (E) and norepinephrine (NE) concentration^{6,10} are shown in parentheses.

Fico₂ = fraction of inspired carbon dioxide; PaO₂ = arterial oxygen tension; Paco₂ = arterial carbon dioxide tension; pHa = arterial pH.

difference was found, the Bonferroni *t* test was performed to determine significant difference between different experimental conditions. Linear regression was computed using the least-squares method and statistical assessment was made by covariance analysis (ANCOVA). The criterion of statistical significance was $P < 0.05$.

Results

The average values of arterial Po₂, Pco₂, and pH obtained in the two groups of subjects breathing mixtures of various carbon dioxide concentrations (Fico₂) are shown in table 1. For any given Fico₂, none of these values differed significantly between subjects anesthetized with diazepam or isoflurane. In diazepam-anesthetized patients, catecholamine concentration did not change systematically with changes in Fico₂ or time. Norepinephrine concentration remained below the upper limits of healthy, awake resting subjects; however, that of epinephrine exceeded those limits.^{7,11}

In all subjects, the rapid decrease in transpulmonary pressure after airway occlusion was easily recognizable on the individual breath records, whereas no such decrease could be seen in the tracings of Pes. Conversely, the rapid decrease in Pes became evident only after ensemble averaging, whereas the decrease in transpulmonary pressure after airway occlusion became clearer (fig. 1). Thus, ensemble averaging proved effective in markedly reducing the cardiac artefact, allowed the measure of Rint,w, and provided a better definition of Rint,L.

The average values of Rint,w and Rint,L obtained in the two groups of subjects at various Fico₂ are shown in table 2. In all subjects, Rint,w did not change with Fico₂, and its mean value was independent of the type of anesthesia. However, in the subjects anesthetized with diazepam, Rint,L decreased significantly when Fico₂ was

increased to 0.05 ($\Delta R_{int,L} = -0.51 \pm 0.2 \text{ cm H}_2\text{O} \cdot \text{s} \cdot \text{l}^{-1}$; $P < 0.001$), and from 0.05 to 0.09 ($\Delta R_{int,L} = -0.46 \pm 0.16 \text{ cm H}_2\text{O} \cdot \text{s} \cdot \text{l}^{-1}$; $P < 0.001$). In all these subjects, Rint,L increased when breathing at Fico₂ = 0 was resumed; the average values of Rint,L of the first and last test were similar. A tendency for Rint,L to decrease with increasing Fico₂ was observed also in the group of subjects during isoflurane anesthesia, but even the difference in Rint,L for the largest Fico₂ changes ($\Delta R_{int,L} = -0.2 \pm 0.5 \text{ cm H}_2\text{O} \cdot \text{s} \cdot \text{l}^{-1}$) was not significant. Conversely, the values of Rint,L were significantly

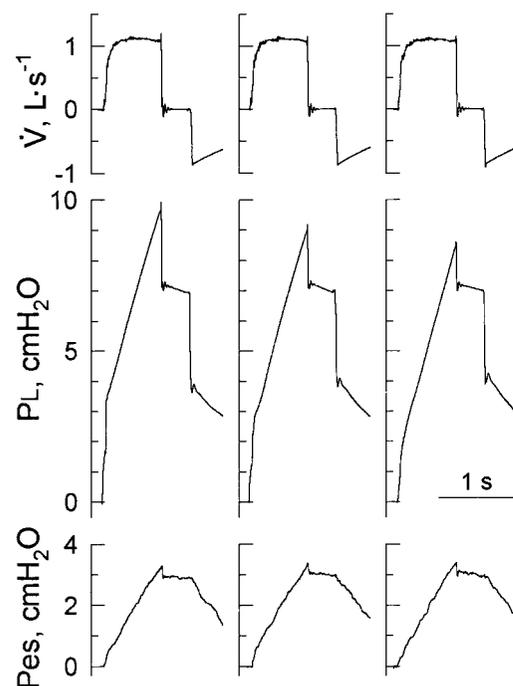


Fig. 1. Ensemble average of records of flow (\dot{V}) and changes in transpulmonary pressure (PL) and esophageal (Pes) pressure from 25 consecutive breaths in a representative subject during hypo- (Paco₂ = 21 mmHg; left), normo- (Paco₂ = 39 mmHg; middle) and hypercapnic conditions (Paco₂ = 66 mmHg; right).

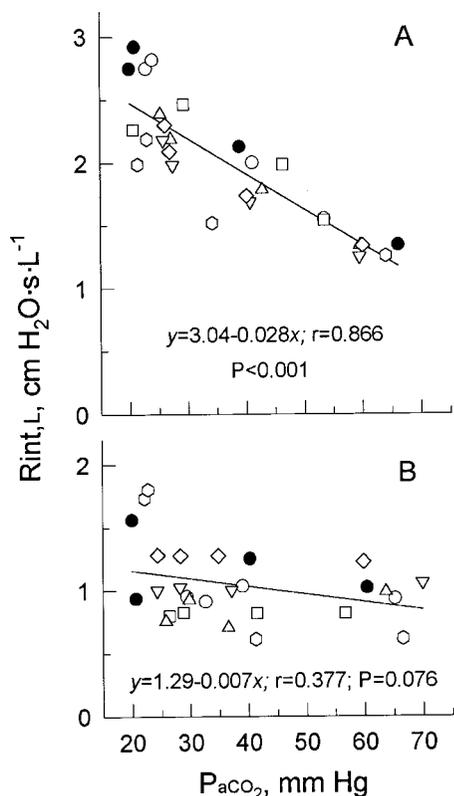


Fig. 2. Relation between arterial P_{aCO_2} and lung interrupter resistance ($R_{int,L}$) obtained in subjects anesthetized with (A) diazepam or isoflurane. Different symbols refer to different subjects.

greater in the group of subjects during diazepam anesthesia at all levels of F_{iCO_2} (table 2). Differences in the effects of carbon dioxide on $R_{int,L}$ between the two groups were not related to larger intersubject variability within the two groups; the sample standard deviation from the regression between individual values of $R_{int,L}$ and P_{aCO_2} (fig. 2) were similar in the group of subjects undergoing diazepam and isoflurane anesthesia (0.26 and 0.28 $\text{cm H}_2\text{O} \cdot \text{s} \cdot \text{L}^{-1}$, respectively).

The average values of ΔR_{w} and ΔR_{L} obtained in the two groups of subjects at various F_{iCO_2} are shown in table 2: neither ΔR_{w} nor ΔR_{L} changed with increasing F_{iCO_2} ; nor did the corresponding mean values differ significantly between the groups receiving diazepam or isoflurane anesthesia. When individual values of ΔR_{L} were plotted against those of P_{aCO_2} (fig. 3), a tendency for ΔR_{L} to decrease with increasing P_{aCO_2} was observed only in the subjects anesthetized with diazepam. Also shown in table 2 are the average values of Est_{w} and Est_{L} obtained in the two groups of subjects at the various F_{iCO_2} : none of these variables changed significantly with changes in F_{iCO_2} , nor did the corresponding mean values differ significantly between the two groups of subjects.

Discussion

Reported effects of P_{aCO_2} changes in respiratory mechanics of spontaneously breathing, unanesthetized subjects are controversial; with inhalation of carbon dioxide mixtures pulmonary resistance has been shown to increase,¹² decrease,¹³ or remain unchanged,¹⁴ whereas hypocapnia was found to cause bronchoconstriction¹⁵⁻¹⁷ and bronchodilation.¹⁸ Part of this variability probably reflects the multiple mechanisms and sites of action of carbon dioxide, especially the balance between contrasting systemic effects^{15,19-21} and that between antagonistic local and systemic effects,^{19,22} but part could be a result of the various methods used to measure resistance,²³ the changes in the breathing pattern that occur with changing P_{aCO_2} , because hyperventilation is itself bronchoconstricting, or individual differences in bronchial or parenchymal reactivity. Moreover, it has been suggested that in spontaneously breathing unanesthetized subjects the carbon dioxide-related changes of respiratory mechanics are entirely caused by upper airway resistance.¹⁴ On the basis of these results, it is not possible to predict the effects of carbon dioxide on airway resistance in anesthetized subjects undergoing artificial ventilation.

The current study is the first report in which the effects of hypocapnia and hypercapnia on respiratory mechanics have been assessed in healthy, anesthetized, paralyzed subjects. Although with diazepam and isoflurane there was no carbon dioxide-related effect on chest wall mechanics, the effect of carbon dioxide on lung mechanics differed between the two anesthetic agents.

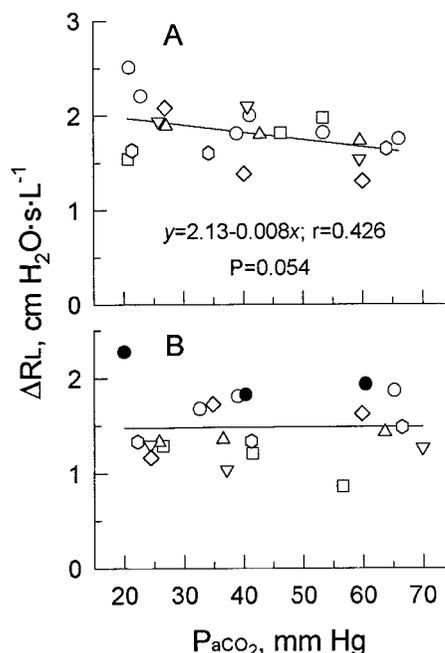


Fig. 3. Relation between arterial P_{aCO_2} and lung additional resistance (ΔR_{L}) obtained in subjects anesthetized with (A) diazepam or (B) isoflurane. Different symbols refer to different subjects.

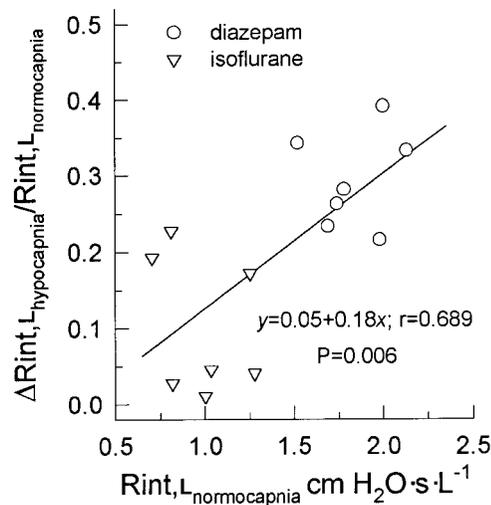


Fig. 4. Relation between changes in lung interrupter resistance ($\Delta R_{int,L}$) during hypocapnia, expressed as a fraction of $R_{int,L}$ during normocapnia, to normocapnic $R_{int,L}$ during different types of anesthesia.

With isoflurane, the changes in P_{aCO_2} had no significant effects on $R_{int,L}$, whereas with diazepam $R_{int,L}$ increased with hypocapnia and decreased with hypercapnia, the changes being inversely related to P_{aCO_2} in the range of 20–70 mmHg. This different response probably occurred because isoflurane can cause marked bronchodilation,^{4,24} whereas with diazepam some bronchomotor tone was still present, as indicated by the significantly higher normocapnic values of $R_{int,L}$ with diazepam as compared with isoflurane (table 2). The baseline bronchomotor tone during normocapnia seems to play an important role in determining the magnitude of the carbon dioxide-related changes in $R_{int,L}$, as shown in figure 4, which depicts the relation between the changes in $R_{int,L}$ caused by hypocapnia ($\Delta R_{int,L}$) and the corresponding normocapnic values. Indeed, $\Delta R_{int,L}$ correlates significantly with the normocapnic values of $R_{int,L}$. The relevance of the baseline bronchomotor tone is further supported by the results ob-

tained by Don and Robson³: their mean value of $R_{int,L}$ during normocapnia ($3 \pm 0.5 \text{ cm H}_2\text{O} \cdot \text{s} \cdot \text{l}^{-1}$) was markedly greater than that of the subjects anesthetized with diazepam (table 2), and the relative increase of $R_{int,L}$ with hypocapnia was also greater (~40 vs. ~30%). It should be noted that Don and Robson³ measured the interrupter resistance of the total respiratory system ($R_{int,rs} = R_{int,L} + R_{int,w}$). Because $R_{int,w}$ is relatively small and independent of both the level of carbon dioxide and the anesthetic used (table 2), $R_{int,L}$ was computed by subtracting $0.47 \text{ cm H}_2\text{O} \cdot \text{s} \cdot \text{l}^{-1}$ (the mean value of $R_{int,w}$ in table 2) from $R_{int,rs}$ given by Don and Robson.³

Associated to the changes of $R_{int,L}$ with changing P_{aCO_2} in diazepam-anesthetized subjects, there were parallel, though smaller, changes in $\Delta R_{int,L}$ (fig. 3A), likely reflecting changes in tissue viscoelastic properties. In fact, the greater decrease in $R_{int,L}$ with carbon dioxide administration observed in hypocapnic subjects by Don and Robson³ was paralleled by significant decrease in the difference between dynamic and static elastance of the respiratory system, *i.e.*, a significant decrease in $\Delta R_{int,L}$ because, as in the current study, inflation volume and flow were the same during hypo- and normocapnia, and Est,w was found to be independent of P_{aCO_2} (table 2). Conversely, $\Delta R_{int,L}$ with isoflurane anesthesia and Est,L with diazepam and isoflurane anesthesia did not change with changes in P_{aCO_2} (table 2). Furthermore, although Est,L was similar with diazepam and isoflurane at all levels of P_{aCO_2} studied, $\Delta R_{int,L}$ with hypocapnia was significantly larger with diazepam anesthesia (table 2).

Bronchoconstriction with hypocapnia and bronchodilation with hypercapnia can be ascribed to the direct action of carbon dioxide because these effects are observed in isolated preparations of airway smooth muscles.^{25,26} The decrease in $R_{int,L}$ with increasing P_{aCO_2} in hypocapnic subjects during diazepam anesthesia, which was probably attenuated by the increased plasma levels of catecholamines (table 1), can be therefore explained

Table 2. Lung and Chest Wall Mechanics at Different Inspired Carbon Dioxide Concentrations (F_{iCO_2}) in Seven Normal Subjects Anesthetized with Diazepam (A) or Isoflurane (B)

| F_{iCO_2} | $R_{int,L}$ (cm H ₂ O · s · l ⁻¹) | $R_{int,w}$ (cm H ₂ O · s · l ⁻¹) | $\Delta R_{int,L}$ (cm H ₂ O · s · l ⁻¹) | $\Delta R_{int,w}$ (cm H ₂ O · s · l ⁻¹) | Est,L (cm H ₂ O/l) | Est,w (cm H ₂ O/l) |
|-------------|--|--|---|---|---------------------------------|---------------------------------|
| 0 | | | | | | |
| A | 2.34 ± 0.26 | 0.45 ± 0.05 | 1.97 ± 0.26 | 0.82 ± 0.33 | 8.4 ± 4.8 | 5.3 ± 0.7 |
| B | 1.15 ± 0.38* | 0.51 ± 0.11 | 1.48 ± 0.38† | 0.85 ± 0.08 | 7.7 ± 2.4 | 5.6 ± 1.0 |
| 0.05 | | | | | | |
| A | 1.83 ± 0.21‡ | 0.44 ± 0.15 | 1.79 ± 0.24 | 0.85 ± 0.36 | 8.2 ± 4.9 | 5.4 ± 0.7 |
| B | 0.94 ± 0.28* | 0.50 ± 0.12 | 1.47 ± 0.31 | 0.82 ± 0.33 | 7.6 ± 2.5 | 5.5 ± 1.1 |
| 0.09 | | | | | | |
| A | 1.37 ± 0.12§ | 0.44 ± 0.16 | 1.68 ± 0.21 | 0.80 ± 0.39 | 8.6 ± 4.5 | 5.4 ± 0.6 |
| B | 0.95 ± 0.19* | 0.48 ± 0.12 | 1.50 ± 0.37 | 0.90 ± 0.21 | 7.9 ± 2.5 | 5.2 ± 0.9 |
| 0 | | | | | | |
| A | 2.41 ± 0.30 | 0.46 ± 0.15 | | | | |
| B | 1.11 ± 0.34* | 0.47 ± 0.12 | | | | |

Values are mean ± SD for interrupter (R_{int}) and additional resistance (ΔR) and quasi-static elastance (Est) of lung (l) and chest wall (w). Values are significantly different from A under iso- F_{iCO_2} conditions (* $P < 0.001$; † $P < 0.05$), at $F_{iCO_2} = 0$ (‡ $P < 0.01$), and at $F_{iCO_2} = 0.05$ (§ $P < 0.01$).

on the basis of the local effects of carbon dioxide. The latter can also explain the tendency of ΔR_{L} to decrease with increasing P_{aCO_2} (fig. 3A) and the finding that the greater decrease in $R_{int,L}$ observed by Don and Robson³ with carbon dioxide administration in hypocapnic subjects was paralleled by a significant decrease in ΔR_{L} . It is conceivable that a decrease in smooth muscle tone occurring in the most peripheral airways and lung parenchyma eventually affects the viscoelastic properties of lung tissue, manifested in ΔR_{L} changes. Involvement of systemic effects in the observed response to carbon dioxide cannot be ruled out because atropine was not administered in the current study. Systemic effects of carbon dioxide are mediated *via* vagal parasympathetic pathways and are therefore prevented by atropine administration.^{15,19-21} They could substantially affect $R_{int,L}$ because the larger bronchi are the main site of vagally mediated reflexes.¹⁹ However, the impact of the reflex effects of carbon dioxide on bronchomotor tone was probably very small during the current experimental conditions, partly because of the opposite effects caused by central²¹ and peripheral chemoreceptor stimulation²² and partly because of the depression exerted by general anesthetics on ganglionic and central nervous system synapses of the parasympathetic bronchomotor pathway.²⁷ The vagolytic effect of pancuronium bromide,²⁸ though modest, should have further contributed to minimize the role of parasympathetic reflexes. Finally, it should be noted that sympathetic responses were not involved in the carbon dioxide-related effects observed in diazepam-anesthetized subjects because plasma catecholamine concentrations were not affected by changes in P_{aCO_2} (table 1).

From a clinical standpoint, there are two aspects of interest that stem from the current study. (1) In anesthetized, paralyzed, healthy subjects, the carbon dioxide-related changes in resistance offered by intrathoracic airways ($R_{int,L}$) are either abolished, as with isoflurane, or relatively small, as with diazepam. Whether this is also the case in patients with acute respiratory distress syndrome undergoing mechanical ventilation or patients with severe brain injury undergoing hyperventilation must be determined. Our results, however, suggest that the high values of pulmonary resistance found in patients with brain injury¹ are not caused by hypocapnia *per se*. (2) Local changes in bronchomotor tone mediated by changes in P_{CO_2} are believed to play an important role in promoting \dot{V}_A/\dot{Q} matching.²⁹⁻³² Accordingly, the changes in bronchomotor tone reflected by those in $R_{int,L}$ with changing P_{CO_2} in subjects anesthetized with diazepam or as anesthetized in the Don and Robson³ study indicate that during these conditions this mechanism is preserved. In contrast, the regulation of \dot{V}_A/\dot{Q} *via* local changes in P_{CO_2} should be impaired in subjects anesthetized with isoflurane, in whom the carbon dioxide-related changes in $R_{int,L}$ are abolished.

In conclusion, this study has shown that, in healthy subjects anesthetized with isoflurane, changes in P_{aCO_2} have no effect on respiratory mechanics, whereas with diazepam pulmonary resistance decreases during hypercapnia and increases during hypocapnia, probably because of the local effects exerted on peripheral airways.

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References

1. Tantucci C, Corbeil C, Chassé M, Braidy J, Matar N, Milic-Emili J: Flow resistance in mechanically ventilated patients with severe neurological injury. *J Crit Care* 1993; 8:133-9
2. The ARDS Network: Ventilation with lower tidal volumes as compared with traditional tidal volumes for acute lung injury and the acute respiratory distress syndrome. *N Engl J Med* 2000; 342:1301-8
3. Don HF, Robson JG: The mechanics of the respiratory system during anesthesia. *ANESTHESIOLOGY* 1965; 26:168-78
4. Coon RL, Kampine JP: Hypocapnic bronchoconstriction and inhalation anesthetics. *ANESTHESIOLOGY* 1975; 43:635-45
5. Cottrell JE, Wolfson B, Siker ES: Changes in airway resistance following droperidol, hydroxyzine, and diazepam in normal volunteers. *Anesth Analg* 1977; 55:18-21
6. Baydur A, Behrakis PK, Zin WA, Jaeger M, Milic-Emili J: A simple method for assessing the validity of the esophageal balloon technique. *Am Rev Respir Dis* 1983; 126:788-91
7. Willemsen JJ, Ross HA, Jacobs MC, Lenders JW, Thien T, Swinkels LM, Benraad TJ: Highly sensitive and specific HPLC with fluorimetric detection for determination of plasma epinephrine and norepinephrine applied to kinetic studies in humans. *Clin Chem* 1995; 41:1455-60
8. D'Angelo E, Robatto FM, Calderini E, Tavola M, Bono D, Milic-Emili J: Pulmonary and chest wall mechanics in anesthetized paralyzed humans. *J Appl Physiol* 1991; 70:2602-10
9. D'Angelo E, Prandi E, Tavola M, Calderini E, Milic-Emili J: Chest wall interrupter resistance in anesthetized paralyzed humans. *J Appl Physiol* 1994; 77:883-7
10. Bates JHT, Ludwig MS, Sly PD, Brown K, Martin JG, Fredberg JJ: Interrupter resistance elucidated by alveolar pressure measurement in open-chest normal dogs. *J Appl Physiol* 1998; 65:408-14
11. Buhler HU, Da Prada M, Haefely W, Picotti GB: Plasma adrenaline, noradrenaline and dopamine in man and different animal species. *J Physiol (Lond)* 1978; 276:311-20
12. Sterling GM: The mechanism of decreased specific airway conductance in man during hypercapnia caused by inhalation of 7% CO₂. *Clin Sci* 1969; 37:539-48
13. Badr SM, Skatrud JB, Simon PM, Dempsey JA: Effect of hypercapnia on total pulmonary resistance during wakefulness and during NREM sleep. *Am Rev Respir Dis* 1991; 144:406-14
14. Rodarte JR, Hyatt RE: Effect of acute exposure to CO₂ on lung mechanics in normal man. *Respir Physiol* 1973; 17:135-45
15. Sterling GM: The mechanism of bronchoconstriction due to hypocapnia in man. *Clin Sci* 1968; 34:277-85
16. Newhouse MT, Becklake MR, Macklem PT, McGregor M: Effect of alteration in end-tidal CO₂ tension on flow resistance. *J Appl Physiol* 1964; 19:745-9
17. O'Cain CF, Hensley MJ, McFadden ER, Ingram RH: Pattern and mechanism of airway response to hypocapnia in normal subjects. *J Appl Physiol* 1979; 47:8-12
18. Elshout FVD, Herwaarden CV, Folgering H: Effects of hypercapnia and hypocapnia on respiratory resistance in normal and asthmatic subjects. *Thorax* 1991; 46:28-32
19. Ingram R: Effects of airway versus arterial CO₂ changes on lung mechanics in dogs. *J Appl Physiol* 1975; 38:603-7
20. Delpierre S, Yammes Y, Mei N, Mathiout M, Grimaud M: Mise en évidence de l'origine vagale reflexe des affets bronchoconstricteurs du CO₂ chez le chat. *J Physiol (Paris)* 1980; 76:889-91
21. Daly M de Burgh, Lambertsen C, Schweitzer A: The effects upon the bronchial musculature of altering the oxygen and carbon dioxide tensions of the blood perfusing the brain. *J Physiol (Lond)* 1953; 119:292-314

22. Daly M de Burgh, Schweitzer A: Reflex bronchomotor responses to stimulation of receptors in the region of the carotid sinus and arch of the aorta in the dog and cat. *J Physiol (Lond)* 1951; 113:442-62
23. Sly P, Bates J, Kochi T, Okubo S, Milic-Emili J: Frequency dependent effects of hypercapnia on respiratory mechanics of cats. *J Appl Physiol* 1987; 62:444-50
24. Parnass SM, Feld JM, Chamberlin WH, Segil LJ: Status asthmaticus treated with isoflurane and enflurane. *Anesth Analg* 1987; 66:193-5
25. Croxton T, Lande B, Hirschman C: Role of intracellular pH in relaxation of porcine tracheal smooth muscle by respiratory gases. *Am J Physiol* 1995; 268: L207-13
26. Twort C, Cameron I: Effects of P_{CO_2} , pH, and extracellular calcium on contraction of airway smooth muscles from rats. *Respir Physiol* 1986; 66:259-67
27. Holtzman MJ, Hahn HL, Sasaki K, Skoogh BE, Graf PD, Fabbri LM, Nadel JA: Selective effect of general anesthetics on reflex bronchoconstrictor responses in dogs. *J Appl Physiol* 1982; 53:126-33
28. Taylor P: Neuromuscular blocking agents, Goodman and Gilman's: The Pharmacological Basis of Therapeutics, 7th edition. New York, MacMillan Publishing, 1985, pp 229-30
29. Shirer H, Orr J, Loker J: Inspiratory airway CO_2 loading in the pony. *J Appl Physiol* 1984; 57:1097-103
30. Domino KB, Swenson ER, Polissar NL, Lu Y, Eisenstein BL, Hlastala MP: Effect of inspired CO_2 on ventilation and perfusion heterogeneity in hyperventilated dogs. *J Appl Physiol* 1993; 74:1306-14
31. Swenson E, Robertson H, Hlastala M: Effects of inspired carbon dioxide on ventilation-perfusion matching in normoxia, hypoxia, and hyperoxia. *Am J Respir Crit Care Med* 1994; 149:1563-9
32. Domino KB, Swenson ER, Emery M, Hlastala MP: Ventilation heterogeneity is increased in hypocapnic dogs but not in pigs. *Respir Physiol* 1998; 111:89-100