

Mechanisms of Cardioprotection by Volatile Anesthetics

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A RAPIDLY growing body of evidence indicates that volatile anesthetics protect myocardium against reversible and irreversible ischemic injury. Identifying the mechanisms by which volatile agents mediate these antiischemic actions is the subject of intense research. This objective has been difficult to accomplish because volatile anesthetics also profoundly affect cardiovascular function. Volatile agents reduce arterial and coronary perfusion pressure, cause dose-related depression of myocardial contractility, produce coronary vasodilation, affect electrophysiologic function, and modify autonomic nervous system activity to varying degrees. Therefore, the antiischemic effects of volatile anesthetics may be mediated, at least in part, by favorable alterations in myocardial oxygen supply-demand relations, preservation of energy-dependent cellular functions, and increased coronary blood flow. However, it seems unlikely that changes in myocardial metabolism and coronary perfusion caused by volatile anesthetics are solely responsible for protection against ischemic damage. Instead, several endogenous signal transduction pathways, acting through the adenosine triphosphate (ATP)-sensitive potassium (K_{ATP}) channel and involving the generation of reactive oxygen species (ROS), have been implicated in mediating the antiischemic actions of volatile anesthetics. The experimental and clinical findings documenting the phenomenon of volatile anesthetic pre-

conditioning against ischemic injury of myocardium are evaluated. Recent findings *in vitro* and *in vivo* that seek to define the intracellular mechanisms responsible for these beneficial actions are also summarized.

Historical Perspective

The antiischemic effects of volatile anesthetics were initially proposed more than 20 yr ago. Lowenstein *et al.*¹ demonstrated that halothane reduced ST segment elevation in a canine model of brief coronary artery occlusion. These data were consistent with the hypothesis that exposure to halothane reduced acute ischemic injury. A subsequent study by this research group also demonstrated that halothane reduced myocardial infarct size when administered before prolonged coronary artery occlusion in dogs.² Lactate production was decreased in the presence as compared with the absence of enflurane during demand-induced ischemia produced by a critical coronary artery stenosis and ventricular pacing when coronary perfusion pressure was maintained.³ These results suggested that myocardial metabolism may be improved by administration of a volatile agent during an ischemic episode independent of alterations in hemodynamics. The relative importance of these early findings was initially overshadowed⁴ by a series of reports published in the mid-1980s⁵⁻⁸ suggesting that isoflurane may be capable of producing an abnormal redistribution of coronary blood flow away from ischemic toward normal myocardium.⁹⁻¹¹ This "coronary steal" phenomenon was attributed to the coronary vasodilating properties of isoflurane that are known to occur primarily in arterioles of less than 100 μm in diameter.¹² Isoflurane was thought to be capable of directly producing myocardial ischemia in susceptible patients with "steal-prone" coronary artery anatomy under certain hemodynamic conditions¹³ in a fashion similar to that of potent coronary vasodilators (*e.g.*, adenosine, chromonar, dipyridamole).

The implication that isoflurane might produce myocardial ischemia through such a steal mechanism was subsequently dispelled by several investigations conducted in animal models¹⁴⁻¹⁷ and humans with coronary artery disease.¹⁸⁻²⁰ For example, isoflurane did not selectively redistribute blood flow away from the collateral-dependent region in a chronically instrumented canine model

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of multivessel coronary artery disease.¹⁵ In contrast, adenosine produced marked coronary steal by preferentially shunting blood flow away from collateral-dependent myocardium in this model.¹⁵ Other studies^{14,16-20} also suggested that isoflurane-induced hypotension may reduce myocardial perfusion, but true coronary steal did not occur when coronary perfusion pressure was maintained. Subsequent investigations with the newer volatile anesthetics sevoflurane^{21,22} and desflurane²³ showed that these drugs also did not reduce or abnormally redistribute coronary collateral blood flow. Therefore, despite initial concerns, volatile anesthetics were subsequently shown to be relatively weak coronary vasodilators that are incapable of causing coronary steal under the vast majority of clinical conditions.²⁴

Contrary to the hypothesis that the use of volatile anesthetics may be potentially deleterious in some patients with coronary artery disease, many laboratory and clinical investigations conducted since the resolution of the coronary steal controversy have convincingly shown that volatile anesthetics *protect* the heart against ischemia and reperfusion injury. In addition to previously cited studies suggesting that halothane^{1,2} and enflurane³ exerted antiischemic effects, halothane was also shown to preserve contractile function and ultrastructural integrity during cardioplegic arrest.²⁵ This latter study was of considerable interest because these data indicated that halothane was capable of exerting a cardioprotective effect completely independent of improvements in myocardial oxygen supply-demand balance. In addition, halothane,²⁶ enflurane,²⁶ desflurane,²⁶⁻²⁸ and sevoflurane²⁶⁻³¹ have been shown to reduce myocardial damage when administered during reperfusion after prolonged coronary occlusion or cardioplegic arrest. Another study showed that preservation of high-energy phosphate concentrations was coupled to the protective effects of enflurane.³² Isoflurane and desflurane did not depress but modestly enhanced left ventricular diastolic function during acute coronary occlusion in dogs.³³ Halothane,³⁴⁻³⁶ enflurane,^{34,37,38} isoflurane,^{34,35,37} and sevoflurane³⁹ were shown to improve the functional recovery of isolated hearts subjected to global ischemia and reperfusion. Halothane and isoflurane markedly augmented the recovery of regional contractile function of stunned myocardium *in vivo*.^{40,41} Halothane² and isoflurane⁴² reduced myocardial infarct size in dogs, and this beneficial action was found to persist despite discontinuation of the volatile anesthetic before coronary artery occlusion.⁴² The myocardium acted as if it had "remembered" the previous exposure to the volatile agent. This phenomenon was termed *anesthetic-induced preconditioning* (APC)⁴³ and was characterized by a short-term memory phase similar to that observed during ischemic preconditioning (IPC).

Anesthetic-induced preconditioning has also been described in other animal species, including rats⁴⁴ and

rabbits.⁴³ The efficacy of APC conferred by isoflurane to reduce infarct size has been shown to be dose dependent in rats,⁴⁴ an animal model with minimal coronary collateral flow.⁴⁵ High concentrations of isoflurane may also have greater efficacy to protect myocardium during conditions of low coronary collateral blood flow in dog myocardium.⁴⁶ Similarly, isoflurane and sevoflurane dose-dependently preserved the viability of isolated cardiac myocytes during ischemia.⁴⁷ Isoflurane has been shown to elicit cardioprotective effects after discontinuation for 15 min or 30 min before coronary artery occlusion in rats and rabbits^{43,44} or dogs,⁴² respectively. In contrast, sevoflurane did not exert antiischemic actions after a 30-min washout period.⁴⁸ Taken together, these data suggest that the memory period associated with APC may differ between volatile anesthetics and among species. Interestingly, recent findings showed that isoflurane reduced myocardial damage when administered 24 h before coronary artery occlusion and reperfusion in rabbit hearts *in vivo*.⁴⁹ Pretreatment with isoflurane also preserved endothelial and vascular smooth muscle cell viability 12-48 h after cytokine-induced injury.⁵⁰ Therefore, volatile anesthetics also produce a late phase (*i.e.*, a second window) of myocardial protection similar to IPC. In addition, sevoflurane reduced the duration of a brief ischemic episode required to protect against infarction during IPC.⁴⁸ Sevoflurane also enhanced cardioprotection when administered 24 h after an initial IPC stimulus.⁵¹ These important findings showed that administration of a volatile anesthetic combined with a brief ischemic event synergistically protects myocardium against subsequent damage as well.

Additional data about the effects of volatile agents on the coronary circulation also stand in contrast with the conclusions implicated by the coronary steal hypothesis. These results indicated that volatile agents are certainly not deleterious to but may instead exert beneficial actions on coronary collateral perfusion to ischemic myocardium. Volatile anesthetics have been shown to produce coronary vasodilation by activating K_{ATP} channels^{39,52-55} or by favorably affecting intracellular Ca^{2+} homeostasis in vascular smooth muscle.⁵⁶ Halothane attenuated reductions in coronary collateral perfusion associated with acute coronary occlusion and also improved the myocardial oxygen supply-demand relation in collateral-dependent myocardium.⁵⁷ In addition, halothane reduced cyclical changes in coronary blood flow and prevented the development of platelet thrombi in the presence of a critical coronary artery stenosis.⁵⁸ Sevoflurane increased collateral blood flow to ischemic myocardium when perfusion pressure was maintained.^{21,59} Sevoflurane also improved the functional recovery of coronary vascular reactivity and nitric oxide release in isolated hearts after global ischemia.³⁹ Lastly, volatile anesthetics attenuated neutrophil and platelet aggregation⁶⁰ and also inhibited cytokine-induced cell

death^{50,61} after ischemia-reperfusion injury *in vitro*. The results of these studies collectively show the protection against ischemia and reperfusion injury may be at least partially based on favorable effects of volatile agents on coronary perfusion.

The precise mechanisms responsible for volatile anesthetic-induced protection against ischemic injury remain unclear despite extensive study. Although it is clear that volatile anesthetics may indirectly improve myocardial oxygen supply-demand relations or enhance coronary collateral perfusion, it is equally clear that these actions are not entirely responsible for the antiischemic effects of these agents. This contention is emphasized by findings showing that volatile anesthetics conferred protection during cardioplegic arrest²⁵ and during reperfusion,²⁶⁻³⁰ conditions in which myocardial oxygen supply-demand relations play little if any role. Similarly, isoflurane and sevoflurane increased the viability of isolated cardiac myocytes,⁴⁷ and sevoflurane⁶² and desflurane⁶³ improved contractility of isolated cardiac muscle exposed to simulated ischemia. These results were initially attributed to reductions in excessive intracellular Ca^{2+} concentrations during ischemia and reperfusion⁶⁴ produced by partial inhibition of Ca^{2+} channel activity.⁶⁵⁻⁶⁸ However, this relatively generic Ca^{2+} hypothesis did not address the precise mechanisms or provide deeper insight into the intracellular processes by which volatile anesthetics exert protective effects in the intact heart.

K_{ATP} Channels

The signal transduction pathways involved in APC bear striking similarity to those responsible for IPC. It is hypothesized that volatile anesthetics stimulate a trigger that initiates a cascade of events leading to activation of an end-effector that is responsible for resistance to injury. To date, adenosine type 1 (A_1) receptors,^{34,69,70} protein kinase C (PKC),^{34,71,72} inhibitory guanine nucleotide binding (G_i) proteins,⁷³ ROS,⁷⁴⁻⁷⁶ and mitochondrial and sarcolemmal K_{ATP} (mito K_{ATP} and sarc K_{ATP} , respectively) channels^{42,77-79} have been shown to mediate APC (fig. 1). K_{ATP} channels are heteromultimeric complexes containing an inward-rectifying potassium (K_{ir}) channel and a sulfonylurea receptor (SUR).⁸⁰ Pharmacologic and recombinant techniques indicate that sarc K_{ATP} and mito K_{ATP} channels^{81,82} are composed of the $K_{ir}6.2/SUR2A$ and $K_{ir}6.1/SUR1$ isoforms,⁸³ respectively. K_{ATP} channel opening was initially implicated as the central end-effector during APC,⁸⁴ similar to the findings during studies of the mechanisms responsible for IPC.^{85,86} Isoflurane and sevoflurane preserved myocardial viability in a cellular model of ischemia, and this protective effect was abolished by the selective mito K_{ATP} channel antagonist 5-hydroxydecanoate (5-HD) but not the selective sarc K_{ATP} channel antagonist HMR-1098.⁴⁷ Isoflurane,⁶⁹ sevoflurane,⁶² and desflurane⁶³ but

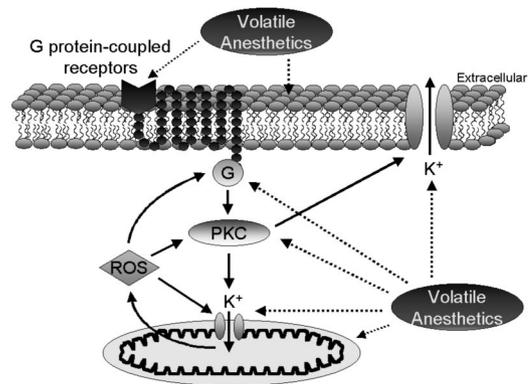


Fig. 1. Multiple endogenous signaling pathways mediate volatile anesthetic-induced myocardial protection. A trigger initiates a cascade of signal transduction events, resulting in the activation of an end-effector that promotes resistance against ischemic injury. Mitochondrial adenosine triphosphate-sensitive potassium (K_{ATP}) channels have been implicated as the end-effector in this protective scheme, but sarcolemmal K_{ATP} channels may also play a role. Volatile anesthetics signal through adenosine and opioid receptors, modulate G proteins, stimulate protein kinase C (PKC) and other intracellular kinases, or have direct effects on mitochondria to generate reactive oxygen species (ROS) that ultimately enhance K_{ATP} channel activity. Volatile anesthetics may also directly facilitate K_{ATP} channel opening. Dashed arrows delineate the intracellular targets that may be regulated by volatile anesthetics; solid arrows represent potential signaling cascades.

not halothane⁶⁹ enhanced the recovery of contractile force of isolated human right atrial trabeculae after hypoxia and reoxygenation. The nonselective K_{ATP} channel blocker glyburide (glibenclamide) or 5-HD inhibited this protective effect. HMR-1098 also attenuated the beneficial actions produced by sevoflurane in isolated human atria.⁶² Glyburide blocked the enhanced recovery of contractile function produced by isoflurane in stunned myocardium *in vivo*.^{41,87} Reductions in canine myocardial infarct size produced by isoflurane⁴² and the ATP-sparing effects of this agent⁸⁸ have been shown to be blocked by glyburide as well. 5-HD also inhibited preconditioning by isoflurane in rats⁴⁴ and rabbits.⁷⁷ Both 5-HD and HMR-1098 abolished the protective effects of desflurane against ischemia and reperfusion injury in dogs,⁷⁸ supporting a role for both mito K_{ATP} and sarc K_{ATP} channels in APC. In contrast, another study showed that HMR-1098 did not modify desflurane-induced preconditioning in isolated human right atria *in vitro*.⁶³ Therefore, some controversy continues to exist about the relative contribution of sarc K_{ATP} and mito K_{ATP} channels in APC.

Carefully conducted *in vitro* experiments suggest that volatile anesthetics are capable of modifying K_{ATP} channel activity. Isoflurane stimulated outward K^+ current through sarc K_{ATP} channels in isolated ventricular myocytes during patch clamping.^{89,90} Volatile anesthetics also reduced sarc K_{ATP} channel sensitivity to inhibition by ATP, thereby increasing open state probability.⁹¹ In contrast, other patch clamp results suggested that vola-

tile agents alone did not open K_{ATP} channels. Isoflurane did not affect sarc K_{ATP} channel current in human atrial cells⁶⁹ and also inhibited sarc K_{ATP} channel activity in rabbit ventricular myocytes.⁹¹ However, some volatile anesthetics were able to enhance sarc K_{ATP} channel current by facilitating channel opening after initial activation.^{89,90} Isoflurane enhanced sarc K_{ATP} channel opening in the presence of the mitochondrial uncoupler 2,4-dinitrophenol, the K_{ATP} channel opener pinacidil, and the protein tyrosine kinase (PTK) inhibitor genistein in a whole cell patch clamp model.^{89,92} Activation of PKC,⁹⁰ adenosine receptors,⁹³ and phosphatidylinositol kinase⁹³ seemed to be necessary for this process to occur. Isoflurane also directly opened sarc K_{ATP} channels during intracellular acidosis, a condition that is known to occur during ischemia.⁹⁴ These data suggest that volatile anesthetics may not directly interact with sarc K_{ATP} channels but instead may affect other signaling elements that modulate sarc K_{ATP} channel activity. In contrast with the findings with isoflurane, halothane had no effect on pinacidil-induced increases in sarc K_{ATP} channel current and even inhibited K_{ATP} channel current that had been maximally activated by 2,4-dinitrophenol.⁸⁹ The anesthetic specificity for APC remains to be well characterized, although studies such as these do suggest important differences in efficacy may exist among individual agents.

The ability of volatile anesthetics to directly open mito K_{ATP} channels has also been examined. Isoflurane and sevoflurane increased mitochondrial flavoprotein oxidation, an index of mito K_{ATP} channel activity, in guinea pig cardiac myocytes.⁹⁵ This process was inhibited by 5-HD.⁹⁵ Flavoprotein fluorescence may not be entirely specific for mito K_{ATP} channel opening,⁹⁶ but isoflurane has also been shown to directly activate mito K_{ATP} channels reconstituted in lipid bilayers.⁹⁷ In contrast with these intriguing findings,⁹⁷ Zaugg *et al.*⁴⁷ recently demonstrated that although isoflurane or sevoflurane did not directly enhance flavoprotein oxidation in rat ventricular myocytes, these volatile agents did potentiate increases in fluorescence produced by the selective mito K_{ATP} channel agonist diazoxide. These results suggested that volatile anesthetics may not directly open but instead act to prime mito K_{ATP} channels, thus enhancing their ability to open in response to an agonist. Sarc K_{ATP} channels may also be linked to the function of the mitochondrial inner membrane. For example, ROS generated by mitochondria may act to open sarc K_{ATP} channels.⁹⁸ 2,4-Dinitrophenol-induced activation of sarc K_{ATP} channel current was reversible and accompanied by nicotinamide adenine dinucleotide oxidation, suggesting the existence of cross-talk between mito K_{ATP} and sarc K_{ATP} channels.⁹⁹ Taken as a whole, the preponderance of evidence collected to date implies that volatile anesthetics do not necessarily directly open K_{ATP} channels but

instead prime the activation of these channels in both sarcolemmal and mitochondrial membranes.

Adenosine triphosphate-sensitive potassium channels in vascular smooth muscle cells have been shown to be essential regulators of coronary vascular tone when ATP production is reduced.¹⁰⁰ Volatile anesthetic-induced coronary vasodilation^{39,52-55} was attenuated by glyburide, indicating an important role for K_{ATP} channels in this process. These data suggest that the beneficial actions of volatile agents during myocardial ischemia may be partially attributed to increased oxygen supply mediated *via* K_{ATP} channel-dependent coronary vasodilation. However, sevoflurane increased coronary collateral blood flow in the presence of glyburide *in vivo*, indicating that volatile anesthetics enhance collateral perfusion independent of K_{ATP} channel activation.⁵⁹ In fact, sevoflurane-induced increases in collateral perfusion were recently shown to occur as a result of Ca^{2+} -regulated potassium and not K_{ATP} channel activation.¹⁰¹ Based on these findings and results obtained in isolated cardiac myocytes where blood flow is not a factor,⁴⁷ it seems highly unlikely that myocardial protection produced by volatile anesthetics is solely related to favorable alterations in coronary vascular tone mediated by K_{ATP} channels.

G Protein-coupled Receptors

Volatile anesthetics may activate parallel or redundant signaling pathways that involve K_{ATP} channel opening to generate a physiologically meaningful cellular response. The sequential activation of several intracellular elements within a given transduction pathway may facilitate signal amplification and interaction between other redundant signaling systems. For example, administration of isoflurane in the presence of the K_{ATP} channel openers nicorandil¹⁰² or diazoxide¹⁰³ markedly enhances protection against ischemic injury beyond that observed with either drug alone. Several receptor-mediated events and intracellular signaling elements that converge on the K_{ATP} channel have been implicated in APC. Pretreatment with pertussis toxin abolished any reduction in infarct size produced by isoflurane, indicating that G_i proteins are linked to the signal transduction pathways that mediate APC.⁷³ In contrast, pertussis toxin did not alter the beneficial effects of direct K_{ATP} channel opening produced by nicorandil. These data strongly support the contention that volatile anesthetics modulate K_{ATP} channel activity through second messenger signaling.

Halothane-induced protection against infarction was completely abolished by blockade of the adenosine A_1 receptor.³⁴ The nonselective adenosine receptor antagonist 8-(p-sulfophenyl)-theophylline abolished isoflurane-induced preconditioning in rabbits.⁷⁹ The selective A_1 receptor antagonist 8-cyclopentyl-1,3-dipropylxanthine partially attenuated the beneficial effects of isoflu-

rane in canine stunned myocardium.⁷⁰ A role for adenosine receptors in APC was also identified in isolated human right atrial myocardium *in vitro*.⁶⁹ Isoflurane eliminated increases in interstitial adenosine during repetitive periods of coronary artery occlusion and reperfusion using a microdialysis technique.⁷⁰ These findings suggest that ATP preservation and a subsequent reduction of adenosine released into the interstitium occur during isoflurane anesthesia.⁷⁰ In addition, the data imply that volatile agents may either directly activate A₁ receptors or indirectly enhance A₁ receptor sensitivity to diminished endogenous adenosine concentrations.⁷⁰ These results were also similar to those observed during IPC¹⁰⁴ or bimakalim-induced pharmacologic preconditioning.¹⁰⁵ The preservation of cardiac myocyte viability during ischemia produced by volatile anesthetics was also sensitive to adenosine receptor and G_i protein inhibition in rats.⁴⁷

Stimulation of the δ_1 -opioid receptor has been shown to produce a cardioprotective effect that is abolished by selective opioid antagonists¹⁰⁶⁻¹¹⁰ or K_{ATP} channel blockers.^{107,108,110-113} The acute and delayed phases of IPC are also mediated by activation of the δ_1 -opioid receptor.¹¹⁴ Recent results indicated that the combined administration of isoflurane and selective δ_1 -opioid receptor agonists TAN-67 or BW373U86 potentiated K_{ATP} channel opening and enhanced protection against myocardial ischemia and reperfusion injury.¹⁰³ Combined administration of isoflurane and morphine, a μ receptor agonist with δ_1 receptor agonist properties, also reduced the extent of myocardial infarction to a greater degree than either drug alone.⁴⁴ This beneficial effect was shown to be mediated by mito K_{ATP} channels and opioid receptors. Interestingly, the nonselective opioid antagonist naloxone abolished isoflurane-induced preconditioning.⁴⁴ These intriguing data suggest an important link between volatile anesthetics and the opioid family of G protein-coupled receptors. Another recent study also indicated that halothane competitively inhibited the ligand-binding site of G protein-coupled receptors.¹¹⁵ Adrenergic receptor blockade was shown to abolish desflurane-induced preconditioning in isolated human right atria⁶³ but had no effect on the antiischemic actions of sevoflurane in isolated rat cardiac myocytes.⁴⁷ Overall, APC seems to be associated with the activation of separate receptor-mediated pathways that are linked to G_i proteins.

Protein Kinases

Translocation and phosphorylation of multiple protein kinases are known to be involved in signal transduction pathways involved in protecting myocardium against cell death after ischemia and reperfusion.¹¹⁶⁻¹¹⁸ In particular, PKC is an essential component of the signaling pathways associated with preserving cellular viability.¹¹⁹ The diverse PKC isoform family is a large group of

serine/threonine protein kinases that are distinguished by variable regulatory domains and cofactors and also display diverse tissue and species distributions.¹²⁰ Activation of G protein-coupled receptors (e.g., A₁,^{121,122} bradykinin,^{123,124} δ_1 opioid^{125,126}) stimulate PKC during IPC. Volatile anesthetics have also been shown to stimulate PKC translocation and activity,¹²⁷ possibly by interacting with the regulatory domain of the enzyme.¹²⁸ Inhibition of PKC attenuated isoflurane-enhanced recovery of contractile function in canine stunned myocardium.⁷¹ The antiischemic actions of halothane were abolished by selective PKC antagonism in rabbits.³⁴ The δ and ϵ isoforms of PKC translocated to mitochondria and sarcolemma, respectively, 10 min after discontinuation of isoflurane in isolated rat hearts.¹²⁹ In contrast, isoflurane stimulated translocation of PKC- δ and - ϵ to sarcolemmal and mitochondrial membranes, respectively, in the *in vivo* rat heart.⁷² These discrepancies may be attributed to differences in experimental model or time of tissue sampling. The microtubule depolymerizing drug, colchicine, prevented isoflurane-induced reductions in myocardial infarct size in rabbits,¹³⁰ suggesting that an intact cytoskeleton is essential for translocation of these protein kinases.^{72,129}

Recent findings strongly suggest that volatile anesthetic-induced PKC activation is required to open K_{ATP} channels and produce myocardial protection. For example, the nonselective PKC antagonist chelerythrine abolished sevoflurane-induced increases in mito K_{ATP} channel activity in rat ventricular myocytes and prevented protection against simulated ischemia.⁴⁷ Patch clamp experiments showed that isoflurane did not facilitate K_{ATP} channel opening in excised membrane patches but enhanced K_{ATP} channel current in a whole cell configuration concomitant with PKC stimulation.⁹⁰ These observations were supported by other studies showing that adenosine and PKC increased K_{ATP} channel activity.^{103,131-134} Specific PKC consensus sites have been identified on K_{ATP} channels, indicating a molecular basis for phosphorylation and activation of the channel by the enzyme.¹³⁵ Mito K_{ATP} channel opening also occurred after PKC activation during IPC in isolated rabbit hearts.¹³⁶ In contrast, recent evidence indicates that 5-HD inhibited PKC translocation,⁷² suggesting that mito K_{ATP} channel opening may be upstream of PKC activation. Therefore, a possible feed-forward system between PKC and K_{ATP} channel activation may occur during APC.

Protein kinase C has been shown to stimulate PTK^{137,138} and mitogen-activated protein kinases (MAPKs),¹³⁹ and volatile anesthetics may modulate several of these critical intracellular signaling proteins independent of direct receptor activation as well. Ischemic and pharmacologic preconditioning have been shown to be mediated by activation of PKC,^{34,71,106,124,140} PTK,^{116,137,138,141,142} and MAPK.^{117,118,143-145} A recent investigation showed that the PTK inhibitor lavendustin

A and the Src-selective inhibitor PP1 abolished isoflurane-induced preconditioning in rats.⁷² The MAPK family plays an important role in signal transduction from the cell surface and the nucleus and has been strongly implicated in the initiation and progression of cell death (*i.e.*, apoptosis).¹⁴⁶⁻¹⁴⁸ The p38 MAPK subfamily mediated IPC of myocardium in dogs, and activation of p38 MAPK was also associated with phosphorylation and translocation of heat shock protein 27 *in vivo*.¹⁴⁴ In contrast with the findings during IPC, recent data suggest that p38 MAPK may not play a role in APC in isolated rat hearts.¹⁴⁹ Nonetheless, volatile anesthetics modulate activity of one or more intracellular kinases to produce APC, and it seems that activation of PKC is critical to cardioprotection.

Reactive Oxygen Species

Large quantities of ROS are released during reperfusion of ischemic myocardium that damage proteins responsible for intracellular homeostasis, depress contractile function, and produce membrane damage.¹⁵⁰⁻¹⁵² Halothane, isoflurane, and enflurane have been shown to attenuate the toxic effects of ROS on left ventricular pressure development in isolated hearts.¹⁵³ Isoflurane decreased hydroxyl radical generation in the ischemic rat heart,¹⁵⁴ and halothane had a similar effect in dogs.¹⁵⁵ The protective effects of sevoflurane were associated with reduced dityrosine formation, an indirect marker of ROS and reactive nitrogen species.⁷⁶ These results support the hypothesis that volatile anesthetics reduce the release of deleterious quantities of ROS associated with coronary artery occlusion and reperfusion. Isoflurane also inhibited superoxide anion production by activated neutrophils, an action that occurred independent of K_{ATP} channel opening.¹⁵⁶ In addition, isoflurane and sevoflurane have been shown to abolish activated neutrophil-induced myocardial dysfunction.¹⁵⁷ These effects were associated with reductions in superoxide anion production and neutrophil adherence to coronary vascular endothelium. Therefore, volatile anesthetics also seem to exert beneficial actions by inhibiting neutrophil-induced injury during reperfusion.

In contrast with data implicating a pathologic role of large amounts of ROS, other findings strongly suggest that a variety of preconditioning stimuli, including brief ischemia, direct mito K_{ATP} channel openers, opioids, and volatile anesthetics, stimulate a small burst of ROS that initiate downstream signaling events and produce protection from subsequent ischemic injury.¹⁵⁸ For example, pretreatment with low concentrations of ROS have been shown to mimic the beneficial actions of IPC.^{159,160} Free radical scavengers administered before or during brief ischemia markedly attenuated the protective effect of the preconditioning stimulus on infarct size.^{161,162} These findings indicate that IPC is mediated by small quantities of ROS released during the precon-

ditioning stimulus. The beneficial actions of sevoflurane against ischemic damage were abolished by scavengers of superoxide anion and inhibition of nitric oxide synthase.⁷⁶ These results suggest that superoxide anion may act to trigger APC and further indicated that nitric oxide may scavenge superoxide anion on reperfusion to reduce injury. ROS scavengers attenuated isoflurane-induced reductions in myocardial infarct size in rabbits^{74,75} and also inhibited the beneficial effects of direct mito K_{ATP} channel activation.¹⁶³ Isoflurane has been shown to directly increase superoxide anion formation *in vivo* independent of an ischemic episode by use of the fluorescent probe dihydroethidium and laser confocal microscopy.⁷⁵ These data⁷⁵ indicated for the first time that volatile anesthetics were capable of producing small amounts of ROS that were correlated with a reduction in myocardial infarct size after prolonged ischemia. Taken as a whole, these reports provide compelling evidence that small quantities of ROS also play a critical role in APC.

Reactive oxygen species have been shown to act as regulatory mediators in many signaling processes that protect the cells against oxidative stress.¹⁶⁴ ROS-induced activation of PKC¹⁵⁹ and MAPK¹⁶⁵⁻¹⁶⁷ have been implicated in both ischemic and pharmacologic preconditioning. Hydrogen peroxide activated all three MAPK subtypes in neonatal rat ventricular myocytes, but stimulation of the p38 MAPK family and the consequent phosphorylation of heat shock protein 25/27 seemed to be of critical importance during cardioprotection.¹⁶⁵ ROS have also been shown to activate $G\alpha_i$ and $G\alpha_o$ proteins.^{168,169} Recent findings showed that sevoflurane-induced ROS generation was unaffected by PKC inhibition,¹⁷⁰ but ROS scavengers inhibited isoflurane-induced PKC translocation.⁷² These findings provide indirect evidence linking ROS production by volatile anesthetics to subsequent activation of protein kinases implicated in the signal transduction responsible for APC.

A controversy continues to exist regarding the temporal relation between mito K_{ATP} channel opening and ROS production during ischemic or pharmacologic preconditioning.¹⁷¹ Although sarc K_{ATP} channel opening was initially assumed to be the end-effector of IPC,^{81,85} mito K_{ATP} channel activation may instead trigger preconditioning by generating ROS.^{163,172} Mito K_{ATP} channel opening produced by selective agonists generates ROS^{163,172} that seem to be essential for activation of MAPK¹⁶⁶ and are also required for beneficial effects on myocardium.¹⁷³ The mito K_{ATP} channel agonist diazoxide caused oxidation of the ROS probe MitoTracker® orange (Molecular Probes, Eugene, OR) and enhanced cell viability after hypoxia and reoxygenation *in vitro*.¹⁷² These actions were attenuated by pretreatment with 5-HD or ROS scavengers. Therefore, the protective effects of mito K_{ATP} channel agonists may occur as a consequence of triggering by ROS that subsequently

reduces myocyte injury, including the release of large quantities of these reactive intermediates during reperfusion injury. Morphine increased cardiomyocyte viability and the fluorescence intensity of the hydrogen peroxide-sensitive probe, 2',7'-dichlorofluorescein.¹¹¹ These actions were abolished by 5-HD pretreatment, suggesting that activation of mito K_{ATP} channels by opioids results in ROS production. Conversely, other studies^{174,175} have indicated that ROS modulate mito K_{ATP} channel activity¹⁷⁴ to provide a beneficial effect, indirectly suggesting that mito K_{ATP} channels may also function as an end-effector of preconditioning. For example, superoxide anion generated by xanthine oxidase activated mito K_{ATP} channels from bovine ventricular myocardium reconstituted from lipid bilayers.¹⁷⁴ Lebuffe *et al.*¹⁷⁵ demonstrated that ROS may serve as a trigger by opening mito K_{ATP} channels, which subsequently generates additional ROS and nitric oxide that are both required for preconditioning in isolated chick neonatal myocytes. Therefore, whether mito K_{ATP} channel opening serves as a trigger or end-effector of ischemic or pharmacologic preconditioning remains unclear.¹⁷⁶ Nevertheless, the apparently complimentary interaction between ROS and mito K_{ATP} channels suggest the intriguing possibility that positive feedback loops may exist between these elements that contribute to myocardial protection.

It also remains unclear whether volatile anesthetic-induced mito K_{ATP} channel opening precedes or follows ROS generation. Pretreatment with 5-HD or the ROS scavengers N-acetylcysteine or N-2-mercaptopyrionyl glycine before administration of isoflurane abolished ROS generation *in vivo*.¹⁷⁷ In contrast, administration of 5-HD after discontinuation of isoflurane but before prolonged ischemia only partially attenuated this effect. These data suggest that mito K_{ATP} channel opening acted as a trigger of APC by generating ROS. Conversely, another recent investigation conducted in isolated guinea pig hearts showed that sevoflurane-induced generation of ROS was not inhibited by 5-HD before ischemia.¹⁷⁸ Therefore, experimental findings remain equivocal in support of the hypothesis that mito K_{ATP} channel opening is the major trigger of APC.

Reactive oxygen species derived from the mitochondrial respiratory chain have been shown to play important roles during IPC or pharmacologic preconditioning.^{111,160,179,180} The complex III inhibitor myxothiazol blocked hypoxia-¹⁶⁰ or acetylcholine-induced¹⁷⁹ ROS generation and abolished preconditioning in isolated chick cardiac myocytes. The precise source of volatile anesthetic-induced production of ROS has yet to be finally established, but volatile agents have been previously shown to inhibit electron transport chain complexes I and II of cardiac mitochondria.^{181,182} Interestingly, sevoflurane-induced complex I inhibition was attenuated by the superoxide dismutase mimetic Mn(III)tetrakis(4-benzoic acid)porphyrin chloride.¹⁸²

These results suggested that ROS may inhibit mitochondrial respiration through a positive feedback mechanism to amplify the ROS signal for triggering APC. In contrast, a recent investigation showed that the complex III inhibitor myxothiazol, but not the complex I inhibitor diphenyleneiodonium, abolished isoflurane-induced reductions in myocardial infarct size and generation of ROS.¹⁸³ These preliminary data indicated that mitochondrial electron transport chain complex III may be the source of ROS production induced by isoflurane during APC. Taken together, it is possible that volatile anesthetics may modulate multiple sites of the electron transport chain either directly or indirectly *via* a ROS-mediated feedback mechanism. Despite these compelling results, other potential enzymatic sources of ROS (*i.e.*, nicotinamide adenine dinucleotide oxidase, cyclooxygenase, lipoxygenase, xanthine oxidase, nitric oxide synthase, cytochrome P450¹⁸⁴⁻¹⁸⁸) may play a role in APC and have yet to be excluded from this process.

The precise identities of the specific ROS involved in APC have yet to be defined, and the signaling pathways that may be modulated by these ROS are also largely unknown. ROS activated PKC, restored contractility, and reduced infarct size in rabbit hearts.¹⁵⁹ Superoxide anion also opened mito K_{ATP} channels.¹⁷⁴ Hydrogen peroxide stimulated PTK-dependent activation of phospholipase C in mouse embryonic fibroblasts, rendering these cells resistant to stress.¹⁸⁹ Hydrogen peroxide has also been shown to directly activate G_i and G_o proteins^{168,169} and other protein kinases involved in reducing cellular injury.^{166,190-192} Hydrogen peroxide may also be converted to other more reactive species that modify cysteine residues of specific G proteins, resulting in their selective activation.¹⁶⁹ Recent results showed that isoflurane administration produced ethidium fluorescence in rabbit myocardium.^{75,177} Dihydroethidium is oxidized by intracellular superoxide anion to produce ethidium that subsequently binds to DNA, further amplifying its fluorescence.¹⁹³ These results strongly suggest that superoxide anion is the particular ROS involved in isoflurane-induced preconditioning. Sevoflurane also generated superoxide anion before ischemia and reperfusion in isolated hearts.¹⁷⁸ Alternatively, different ROS may exert opposing actions on mito K_{ATP} channel activity. Dismutation of superoxide anion leads to production of secondary ROS, including hydrogen peroxide, hydroxyl radical, and peroxynitrite,¹⁹⁴ and these radicals may differentially alter channel activity. For example, superoxide anion and hydrogen peroxide enhanced but peroxynitrite decreased Ca^{2+} -regulated potassium channel activity.¹⁹⁵ Therefore, it remains possible that volatile anesthetics may also generate ROS other than superoxide that activate mito K_{ATP} channels, or these agents may inhibit the formation of intermediates such as peroxynitrite that adversely affect mito K_{ATP} channel function. Further research is needed to clarify this issue.

Mechanisms of Protection

Opening of sarc K_{ATP} channels was originally implicated in IPC and pharmacologic preconditioning by shortening the action potential duration,^{81,196} thereby reducing intracellular Ca^{2+} overload during ischemia.⁸¹ However, subsequent studies conducted after the discovery of mito K_{ATP} channels⁸² indicated that the anti-ischemic actions of K_{ATP} channel activation occurred independent of action potential duration.¹⁹⁷⁻¹⁹⁹ Nevertheless, IPC did not occur in $K_{ir6.2}$ -deficient mice, suggesting that the presence of the sarc K_{ATP} channel was still required for myocardial protection to occur.²⁰⁰ Despite these latter data, the majority of findings accumulated indicate that preservation of mitochondrial bioenergetic function that occurs as a consequence of mito K_{ATP} channel opening seems to be of critical importance for protection against ischemia.²⁰¹⁻²⁰⁴ Selective pharmacologic openers of mito K_{ATP} channels (e.g., diazoxide) maintain mitochondrial Ca^{2+} homeostasis and inhibit Ca^{2+} overload within the organelle.^{203,204} Alteration of the mitochondrial oxidation-reduction balance by mito K_{ATP} channel opening may also act to promote cellular protection.^{204,205} Membrane depolarization, matrix swelling, and uncoupling of ATP synthesis occur as a result of mito K_{ATP} channel opening that may mediate cellular viability during IPC.²⁰⁵ Mito K_{ATP} channel opening depolarizes the inner mitochondrial membrane and causes a transient swelling of the mitochondrial matrix,²⁰⁶ resulting from a shift in the ionic balance.²⁰⁷ These actions initially reduce ATP production²⁰⁴ but subsequently stimulate a compensatory increase in respiration that optimizes the efficacy of oxidative phosphorylation in part through energy-dependent matrix volume regulation.²⁰⁸ Therefore, the moderate disturbance of mitochondrial homeostasis caused by mito K_{ATP} channel opening (fig. 2) may promote tolerance to subsequent ischemic damage by reducing Ca^{2+} overload,^{203,204} preventing activation of necrotic or apoptotic pathways,^{209,210} or attenuating oxidative stress.²¹¹

Mitochondrial ATP synthesis has also been shown to be preserved after prolonged as compared with brief ischemia and reperfusion.²¹² This beneficial effect was abolished by 5-HD, suggesting that activation of mito K_{ATP} channels improves mitochondrial energy production.²¹³ Opening of mito K_{ATP} channels has been hypothesized to preserve outer mitochondrial membrane permeability to ATP precursors (e.g., adenosine, adenosine diphosphate) and cytochrome c. The structure of the intermembrane space may also be maintained as a consequence of mito K_{ATP} channel activation despite generalized swelling of the mitochondrial matrix.²⁰¹ Preservation of ATP substrates and mitochondrial structure may facilitate more efficient energy transfer between the mitochondria and the cytosol immediately after ischemia. Sevoflurane was recently shown to preserve ATP synthesis in isolated cardiac mitochondria obtained during

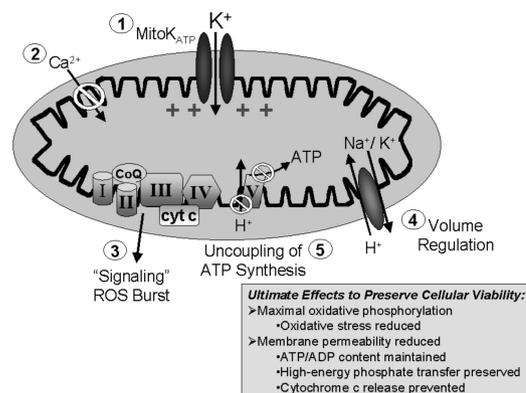


Fig. 2. Cellular preservation during ischemia-reperfusion injury may be mediated by the opening of mitochondrial adenosine triphosphate-sensitive potassium (mito K_{ATP}) channels concomitant with modest modulation of mitochondrial function. Depolarization of the inner mitochondrial membrane and matrix swelling occur as a consequence of mito K_{ATP} channel opening (1). Membrane depolarization due to a shift in the ionic balance prevents Ca^{2+} overload (2). Initial disruption of the electron transport chain may generate signaling amounts of reactive oxygen species (ROS; 3). Volume regulatory mechanisms (4) dissipate the proton gradient and uncouple adenosine triphosphate (ATP) synthesis (5) that ultimately maximizes oxidative phosphorylation and reduces oxidative stress. Mito K_{ATP} channel opening can also reduce membrane permeability and prevent apoptosis and/or necrosis by maintaining nucleotide content, preserving high-energy phosphate transfer, and reducing cytochrome c release. ADP = adenosine diphosphate.

early reperfusion after ischemia *in vivo*, and this beneficial effect was abolished by pretreatment with a ROS scavenger.²¹⁴ Sevoflurane-induced preconditioning improved mitochondrial bioenergetics through mito K_{ATP} channel activation in isolated guinea pig hearts as assessed using flavoprotein fluorescence.²¹⁵ Therefore, it seems likely that mito K_{ATP} channel opening by volatile anesthetics may be associated with preservation of mitochondrial function during reperfusion and, further, that this maintenance of mitochondrial performance contributes to cardioprotection.

Experiments conducted in isolated mitochondria have shown that a triggering quantity of ROS exceeding a critical threshold results in a transition in mitochondrial inner membrane permeability and the subsequent release of a burst of ROS in a process that has been termed *ROS-induced ROS release*.²¹⁶ This mitochondrial permeability transition (MPT) has been shown to precede necrotic or apoptotic cell death,²¹⁷ and glutathione is a primary defense against this event.^{216,218} These data suggest that volatile anesthetics or other mito K_{ATP} channel agonists may prevent MPT in an oxidant-sensitive fashion, but this intriguing hypothesis has yet to be tested. Opening of an MPT pore using the agonist atractyloside during reperfusion was recently shown to abolish IPC and diazoxide-induced preconditioning in isolated rat hearts.²¹⁹ These findings suggested that inhibition of MPT pore opening may represent a distal effector responsible for preconditioning, whereas mito K_{ATP} acti-

vation functions as a trigger or a mediator. Most recently, rabbit hearts pretreated with desflurane before ischemia and reperfusion exhibited resistance to MPT pore opening.²²⁰ Future investigations are necessary to delineate the role of MPT during APC.

Cytosolic and mitochondrial Ca^{2+} overload during prolonged ischemia and reperfusion have been shown to be associated with mitochondrial damage and myocardial cell death.²²¹⁻²²³ Ischemic and sevoflurane-induced preconditioning were shown to reduce cytosolic Ca^{2+} overload and improve the recovery of contractile function during reperfusion.⁶⁴ Administration of sevoflurane after ischemia also reduced cytosolic Ca^{2+} and myocardial damage.³¹ IPC and APC attenuated mitochondrial Ca^{2+} overload during ischemia in rat and guinea pig hearts,^{224,225} actions that were abolished by 5-HD. These data suggested that volatile anesthetics protect against ischemia-reperfusion injury, at least in part, by attenuating cytosolic and mitochondrial Ca^{2+} overload through a mito K_{ATP} channel-dependent mechanism. Volatile anesthetics have also been shown to suppress sarcoplasmic reticulum Ca^{2+} release^{226,227} and depress myofilament Ca^{2+} sensitivity.²²⁷ Therefore, modulation of the sarcoplasmic reticulum to reduce cellular Ca^{2+} overload and alterations of myofilament Ca^{2+} sensitivity under conditions of excess Ca^{2+} have been implicated in cardioprotection.^{228,229} The inhibitory actions of the volatile anesthetics on the voltage-dependent Ca^{2+} channel are also well known,⁶⁵⁻⁶⁸ and reductions in cytosolic and mitochondrial Ca^{2+} overload during ischemia and reperfusion injury may also occur through this mechanism.

Myocardial Protection in Clinical Conditions

Compelling experimental data in multiple animal models regarding the protective effects of volatile anesthetics remain to be translated into therapeutic approaches to reduce morbidity and mortality in patients with ischemic heart disease. However, evidence accumulated to date strongly suggests that APC occurs in human myocardium. Repetitive, brief (60- to 90-s) balloon inflations and deflations performed during percutaneous transluminal coronary angioplasty were associated with progressive reductions in the severity of chest pain and in the extent of ST segment elevation, decreases in myocardial lactate production, and declines in cardiac enzyme and troponin release.²³⁰⁻²³³ Adenosine²³⁴ or the mito K_{ATP} channel opener nicorandil²³⁵ administered before the first balloon inflation during percutaneous transluminal coronary angioplasty was also shown to reduce the severity of ST changes during subsequent occlusions. Pretreatment with nicorandil before percutaneous transluminal coronary angioplasty also attenuated the release of troponin T, an indicator of myocyte necrosis.²³⁶ These findings suggest that both ischemic and pharmacologic preconditioning can be elicited during percutaneous

transluminal coronary angioplasty. Consecutive exercise stress tests (separated by 15 min) performed in patients with critical left anterior descending coronary artery stenoses showed that anginal symptoms, ST segment depression, and myocardial oxygen consumption were reduced during the second as compared with the first exercise period for an equivalent amount of work.²³⁷ This "warm-up" phenomenon occurred independent of coronary vasodilation²³⁷ and also provides evidence of IPC in humans.^{238,239}

Ischemic preconditioning has also been shown during cardiac surgery in patients with coronary artery disease. Yellon *et al.*²³⁹ used intermittent aortic cross clamping to produce IPC during coronary artery bypass graft surgery (CABG) and found enhanced preservation of ATP content in preconditioned hearts as compared with those that did not receive preconditioning stimuli. These investigators also showed that less troponin T was released in the presence as compared with the absence of IPC in this model.²⁴⁰ The incidence of ventricular tachyarrhythmias was reduced after cardiopulmonary bypass in patients undergoing CABG when cold blood cardioplegia was used for myocardial preservation.²⁴¹⁻²⁴³ Regional IPC produced by brief coronary occlusion has also been shown to result in improved hemodynamic recovery and reduced release of cardiac troponin I during off-pump CABG.²⁴⁴ In contrast with the findings of these investigations, other studies have failed to show that IPC exerts beneficial effects during CABG in the presence of cardioplegia and cardiopulmonary bypass.²⁴⁵⁻²⁴⁷ Therefore, although the myocardial protective effects of IPC have been clearly identified in the experimental laboratory, further large scale clinical trials are needed to definitively demonstrate the beneficial actions of IPC in humans.

Documentation of APC in patients has been complicated by alterations in systemic and coronary hemodynamics; the use of other anesthetics, analgesics, or vasoactive drugs; preexisting disease states; and the acute influence of surgery on cardiovascular homeostasis. Nevertheless, isoflurane,⁶⁹ desflurane,⁶³ and sevoflurane⁶² enhanced the recovery of contractile function of human atrial trabeculae *in vitro* by stimulation of adenosine receptors and opening of K_{ATP} channels. Other studies have previously shown a role for adenosine receptors, MAPK,²⁴⁸ and ROS¹⁷² in other forms of preconditioning concomitant with opening of mito K_{ATP} channels in human atrial myocytes. Isoflurane increased the tolerance to pacing-induced ischemia in patients with coronary artery disease.²⁴⁹ Isoflurane also decreased postoperative release of troponin I and creatine kinase-MB in patients undergoing CABG.²⁵⁰ Although the aforementioned results²⁵⁰ were not statistically significant, these data suggest that reduction in the extent of myocardial necrosis had occurred. Administration of isoflurane immediately before aortic cross clamping in patients un-

dergoing CABG was shown to decrease the severity of subsequent ST segment changes and preserve cardiac index to a greater extent than that observed in patients who did not receive pretreatment with the volatile anesthetic.²⁵¹ Administration of enflurane before cardioplegic arrest enhanced recovery of postischemic contractile function assessed using pressure-area relations in CABG patients.²⁵² Sevoflurane²⁵³⁻²⁵⁵ and desflurane^{253,255} but not the intravenous anesthetic propofol was shown to preserve myocardial function in patients undergoing CABG as well as a reduction in troponin I release. Most recently, preconditioning with sevoflurane reduced a biochemical marker of myocardial dysfunction (*i.e.*, N-terminal pro-brain natriuretic peptide) in patients undergoing CABG concomitant with translocation of PKC- δ and $-\epsilon$.²⁵⁶ This compelling evidence strongly suggests that volatile anesthetics exert beneficial effects against ischemic injury in humans.

In contrast with the aforementioned results, no differences in PKC and p38 MAPK activity or peak troponin I release were observed between patients undergoing cardioplegic arrest in the presence or absence of sevoflurane pretreatment.²⁵⁷ The activities of PKC, PTK, and p38 MAPK were increased equally in both groups, suggesting that cardiopulmonary bypass and cardioplegic arrest may produce a preconditioning-like effect that obscured the antiischemic actions of sevoflurane in this setting. However, sevoflurane was recently shown to reduce myocardial injury to a greater degree than propofol in patients undergoing off-pump CABG,²⁵⁴ a clinical setting that does not require cardiopulmonary bypass. Most investigations conducted in humans seem to indicate that volatile anesthetics represent an important therapeutic modality to reduce the sequelae of perioperative myocardial ischemia and infarction.^{258,259} A large-scale, randomized clinical trial is clearly needed to firmly establish this conclusion. Given the recent data suggesting that cardiopulmonary bypass and cardioplegia may exert a protective effect,²⁵⁷ such a clinical trial may best be conducted in patients undergoing off-pump CABG²⁵⁴ or those with documented coronary artery disease undergoing noncardiac surgery.⁴ Furthermore, no clinical study has shown that the use of volatile anesthetics in patients with coronary artery disease contributes to reduced cardiac morbidity or perioperative mortality. Additional multicenter trials also need to be conducted to identify the relative impact of APC on clinical outcome.

Future Perspectives

Experimental evidence collected indicates that volatile anesthetics exert important cardioprotective effects that reduce the consequences of reversible and irreversible ischemia and reperfusion injury. Differences in the effi-

cacy of APC and timing of administration among volatile anesthetics alone and in combination with other cardioprotective drugs remain to be fully distinguished. Another important aspect is to determine the effect of aging on APC.^{255,260} Characterization of a late phase or second window of APC may be of special clinical significance to protect against ischemic events that frequently occur in the postoperative period. Several endogenous signaling elements seem to mediate APC, and mito K_{ATP} channels, PKC, and ROS have emerged as central features in this process. Future investigations are needed to further delineate and identify essential components in these complex signal transduction cascades that mediate the early and late phases of APC. In this regard, microarray technology may prove useful in ascertaining candidate genes that are responsible for APC.²⁶¹ Use of other fundamental molecular and biochemical tools is needed to determine whether preservation of mitochondrial integrity and metabolic homeostasis ultimately enhances tolerance to myocardial ischemia. Volatile anesthetics also act to elicit signaling that is probably present in many types of cells. Therefore, it is not surprising that these agents may reduce injury to other tissues.^{256,262} This may also be based on preventing cytokine-induced injury in endothelial and vascular smooth muscle cells.^{50,61} Finally, recent investigations have also strongly implied that APC occurs in humans and may represent an important therapeutic approach to reduce morbidity and mortality in patients with coronary artery disease. Nevertheless, further investigation is needed to firmly link this emerging body of clinical evidence to the already strongly established experimental data about the protective effects of volatile anesthetics in myocardium.

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References

1. Bland JH, Lowenstein E: Halothane-induced decrease in experimental myocardial ischemia in the non-failing canine heart. *ANESTHESIOLOGY* 1976; 45:287-93
2. Davis RF, DeBoer LW, Rude RE, Lowenstein E, Maroko PR: The effect of halothane anesthesia on myocardial necrosis, hemodynamic performance, and regional myocardial blood flow in dogs following coronary artery occlusion. *ANESTHESIOLOGY* 1983; 59:402-11
3. van Ackern K, Vetter HO, Bruckner UB, Madler C, Mittman U, Peter K: Effects of enflurane on myocardial ischaemia in the dog. *Br J Anaesth* 1985; 57:497-504
4. Wartier DC, Kersten JR, Pagel PS, Gross GJ: Anesthetic preconditioning: Serendipity and science (editorial). *ANESTHESIOLOGY* 2002; 97:1-3
5. Reiz S, Balfors E, Sorensen MB, Ariola S Jr, Friedman A, Truedsson H: Isoflurane: A powerful coronary vasodilator in patients with coronary artery disease. *ANESTHESIOLOGY* 1983; 59:91-7
6. Becker LC: Is isoflurane dangerous for the patient with coronary artery disease? *ANESTHESIOLOGY* 1987; 66:259-61
7. Buffington CW, Romson JL, Levine A, Duttlinger NC, Huang AH: Isoflurane induces coronary steal in a canine model of chronic coronary occlusion. *ANESTHESIOLOGY* 1987; 66:280-92
8. Priebe HJ, Foex P: Isoflurane causes regional myocardial dysfunction in dogs with critical coronary artery stenoses. *ANESTHESIOLOGY* 1987; 66:293-300
9. Gross GJ, Wartier DC: Coronary steal in four models of single or multiple vessel obstruction in dogs. *Am J Cardiol* 1981; 48:84-92
10. Wartier DC, Gross GJ, Brooks HL: Coronary steal-induced increase in

myocardial infarct size after pharmacologic coronary vasodilation. *Am J Cardiol* 1980; 46:83-90

11. Becker LC: Conditions for vasodilator-induced coronary steal in experimental myocardial ischemia. *Circulation* 1978; 57:1103-10

12. Sill JC, Bove AA, Nugent M, Blaise GA, Dewey JD, Grabau C: Effects of isoflurane on coronary arteries and coronary arterioles in the intact dog. *ANESTHESIOLOGY* 1987; 66:273-9

13. Buffington CW, Davis KB, Gillispie S, Pettinger M: The prevalence of steal-prone coronary anatomy in patients with coronary artery disease: An analysis of the Coronary Artery Surgery Study Registry. *ANESTHESIOLOGY* 1988; 69:721-7

14. Moore PG, Kien ND, Reitan JA, White DA, Safwat AM: No evidence for blood flow redistribution with isoflurane or halothane during acute coronary artery occlusion in fentanyl-anesthetized dogs. *ANESTHESIOLOGY* 1991; 75:854-65

15. Hartman JC, Kampine JP, Schmeling WT, Wartler DC: Steal-prone coronary circulation in chronically instrumented dogs: Isoflurane *versus* adenosine. *ANESTHESIOLOGY* 1991; 74:744-56

16. Hartman JC, Kampine JP, Schmeling WT, Wartler DC: Actions of isoflurane on myocardial perfusion in chronically instrumented dogs with poor, moderate, or well-developed coronary collaterals. *J Cardiothorac Anesth* 1990; 4:715-25

17. Hartman JC, Kampine JP, Schmeling WT, Wartler DC: Alterations in collateral blood flow produced by isoflurane in a chronically instrumented canine model of multivessel coronary artery disease. *ANESTHESIOLOGY* 1991; 74:120-33

18. Leung JM, Goehner P, O'Kelly BF, Hollenberg M, Pineda N, Cason BA, Mangano DT: Isoflurane anesthesia and myocardial ischemia: Comparative risk *versus* sufentanil anesthesia in patients undergoing coronary artery bypass graft surgery. The SPI (Study of Perioperative Ischemia) Research Group. *ANESTHESIOLOGY* 1991; 74:838-47

19. Pulley DD, Kirvassilis GV, Keleremos N, Kater K, Barzilai B, Genton RE, Efstathiou C, Lappas DG: Regional and global myocardial circulatory and metabolic effects of isoflurane and halothane in patients with steal-prone coronary anatomy. *ANESTHESIOLOGY* 1991; 75:756-66

20. Diana P, Tullock WC, Gorcsan J III, Ferson PF, Arvan S: Myocardial ischemia: A comparison between isoflurane and enflurane in coronary artery bypass patients. *Anesth Analg* 1993; 77:221-6

21. Kersten JR, Brayer AP, Pagel PS, Tessmer JP, Wartler DC: Perfusion of ischemic myocardium during anesthesia with sevoflurane. *ANESTHESIOLOGY* 1994; 81:995-1004

22. Kitahata H, Kawahito S, Nozaki J, Kimura H, Tanaka K, Kitagawa T, Oshita S: Effects of sevoflurane on regional myocardial blood flow distribution: Quantification with myocardial contrast echocardiography. *ANESTHESIOLOGY* 1999; 90:1436-45

23. Hartman JC, Pagel PS, Kampine JP, Schmeling WT, Wartler DC: Influence of desflurane on regional distribution of coronary blood flow in a chronically instrumented canine model of multivessel coronary artery obstruction. *Anesth Analg* 1991; 72:289-99

24. Kenny D, Coughlan MG, Kampine JP, Montgomery RR, Bosnjak ZJ, Wartler DC: Cultured endothelial cells restore vasodilator responses to coronary arteries with impaired endothelial function and alter the response to a nitric oxide donor. *Pharmacology* 1994; 49:249-56

25. Lochner A, Harper IS, Salie R, Genade S, Coetzee AR: Halothane protects the isolated rat myocardium against excessive total intracellular calcium and structural damage during ischemia and reperfusion. *Anesth Analg* 1994; 79:226-33

26. Schlack W, Preckel B, Stunneke D, Thamer V: Effects of halothane, enflurane, isoflurane, sevoflurane and desflurane on myocardial reperfusion injury in the isolated rat heart. *Br J Anaesth* 1998; 81:913-9

27. Preckel B, Thamer V, Schlack W: Beneficial effects of sevoflurane and desflurane against myocardial reperfusion injury after cardioplegic arrest. *Can J Anaesth* 1999; 46:1076-81

28. Preckel B, Schlack W, Comfere T, Obal D, Barthel H, Thamer V: Effects of enflurane, isoflurane, sevoflurane and desflurane on reperfusion injury after regional myocardial ischaemia in the rabbit heart *in vivo*. *Br J Anaesth* 1998; 81:905-12

29. Ebel D, Preckel B, You A, Mullenheim J, Schlack W, Thamer V: Cardioprotection by sevoflurane against reperfusion injury after cardioplegic arrest in the rat is independent of three types of cardioplegia. *Br J Anaesth* 2002; 88:828-35

30. Obal D, Preckel B, Scharbatke H, Mullenheim J, Hoterkes F, Thamer V, Schlack W: One MAC of sevoflurane provides protection against reperfusion injury in the rat heart *in vivo*. *Br J Anaesth* 2001; 87:905-11

31. Varadarajan SG, An J, Novalija E, Stowe DF: Sevoflurane before or after ischemia improves contractile and metabolic function while reducing myoplasmic Ca²⁺ loading in intact hearts. *ANESTHESIOLOGY* 2002; 96:125-33

32. Kanaya N, Fujita S: The effects of isoflurane on regional myocardial contractility and metabolism in "stunned" myocardium in acutely instrumented dogs. *Anesth Analg* 1994; 79:447-54

33. Pagel PS, Hettrick DA, Lowe D, Tessmer JP, Wartler DC: Desflurane and isoflurane exert modest beneficial actions on left ventricular diastolic function during myocardial ischemia in dogs. *ANESTHESIOLOGY* 1995; 83:1021-35

34. Cope DK, Impastato WK, Cohen MV, Downey JM: Volatile anesthetics protect the ischemic rabbit myocardium from infarction. *ANESTHESIOLOGY* 1997; 86:699-709

35. Marijic J, Stowe DF, Turner LA, Kampine JP, Bosnjak ZJ: Differential protective effects of halothane and isoflurane against hypoxic and reoxygenation injury in the isolated guinea pig heart. *ANESTHESIOLOGY* 1990; 73:976-83

36. Coetzee A, Brits W, Genade S, Lochner A: Halothane does have protective properties in the isolated ischemic rat heart. *Anesth Analg* 1991; 73:711-9

37. Coetzee A, Skein W, Genade S, Lochner A: Enflurane and isoflurane reduce reperfusion dysfunction in the isolated rat heart. *Anesth Analg* 1993; 76:602-8

38. Freedman BM, Hamm DP, Everson CT, Wechsler AS, Christian CM II: Enflurane enhances postischemic functional recovery in the isolated rat heart. *ANESTHESIOLOGY* 1985; 62:29-33

39. Novalija E, Fujita S, Kampine JP, Stowe DF: Sevoflurane mimics ischemic preconditioning effects on coronary flow and nitric oxide release in isolated hearts. *ANESTHESIOLOGY* 1999; 91:701-12

40. Wartler DC, al-Wathiqui MH, Kampine JP, Schmeling WT: Recovery of contractile function of stunned myocardium in chronically instrumented dogs is enhanced by halothane or isoflurane. *ANESTHESIOLOGY* 1988; 69:552-65

41. Kersten JR, Lowe D, Hettrick DA, Pagel PS, Gross GJ, Wartler DC: Glyburide, a K_{ATP} channel antagonist, attenuates the cardioprotective effects of isoflurane in stunned myocardium. *Anesth Analg* 1996; 83:27-33

42. Kersten JR, Schmeling TJ, Pagel PS, Gross GJ, Wartler DC: Isoflurane mimics ischemic preconditioning *via* activation of K_{ATP} channels: Reduction of myocardial infarct size with an acute memory phase. *ANESTHESIOLOGY* 1997; 87:361-70

43. Cason BA, Gamperl AK, Slocum RE, Hickey RF: Anesthetic-induced preconditioning: Previous administration of isoflurane decreases myocardial infarct size in rabbits. *ANESTHESIOLOGY* 1997; 87:1182-90

44. Ludwig LM, Patel HH, Gross GJ, Kersten JR, Pagel PS, Wartler DC: Morphine enhances pharmacological preconditioning by isoflurane: Role of mitochondrial K_{ATP} channels and opioid receptors. *ANESTHESIOLOGY* 2003; 98:705-11

45. Maxwell MP, Hearse DJ, Yellon DM: Species variation in the coronary collateral circulation during regional myocardial ischaemia: A critical determinant of the rate of evolution and extent of myocardial infarction. *Cardiovasc Res* 1987; 21:737-46

46. Kehl F, Krolkowski JG, Mraovic B, Pagel PS, Wartler DC, Kersten JR: Is isoflurane-induced preconditioning dose related? *ANESTHESIOLOGY* 2002; 96:675-80

47. Zaugg M, Lucchinetti E, Spahn DR, Pasch T, Schaub MC: Volatile anesthetics mimic cardiac preconditioning by priming the activation of mitochondrial K_{ATP} channels *via* multiple signaling pathways. *ANESTHESIOLOGY* 2002; 97:4-14

48. Toller WG, Kersten JR, Pagel PS, Hettrick DA, Wartler DC: Sevoflurane reduces myocardial infarct size and decreases the time threshold for ischemic preconditioning in dogs. *ANESTHESIOLOGY* 1999; 91:1437-46

49. Tanaka K, Ludwig LM, Krolkowski JG, Alcindor D, Pratt PF, Kersten JR, Pagel PS, Wartler DC: Isoflurane produces delayed preconditioning against myocardial ischemia and reperfusion injury: Role of cyclooxygenase-2. *ANESTHESIOLOGY* 2004; 100:525-31

50. de Klaver MJ, Buckingham MG, Rich GF: Isoflurane pretreatment has immediate and delayed protective effects against cytokine-induced injury in endothelial and vascular smooth muscle cells. *ANESTHESIOLOGY* 2003; 99:896-903

51. Mullenheim J, Ebel D, Bauer M, Otto F, Heinen A, Frassdorf J, Preckel B, Schlack W: Sevoflurane confers additional cardioprotection after ischemic late preconditioning in rabbits. *ANESTHESIOLOGY* 2003; 99:624-31

52. Cason BA, Shubayev I, Hickey RF: Blockade of adenosine triphosphate-sensitive potassium channels eliminates isoflurane-induced coronary artery vasodilation. *ANESTHESIOLOGY* 1994; 81:1245-55

53. Crystal GJ, Gurevicius J, Salem MR, Zhou X: Role of adenosine triphosphate-sensitive potassium channels in coronary vasodilation by halothane, isoflurane, and enflurane. *ANESTHESIOLOGY* 1997; 86:448-58

54. Zhou X, Abboud W, Manabat NC, Salem MR, Crystal GJ: Isoflurane-induced dilation of porcine coronary arterioles is mediated by ATP-sensitive potassium channels. *ANESTHESIOLOGY* 1998; 89:182-9

55. Crystal GJ, Zhou X, Gurevicius J, Czinn EA, Salem MR, Alam S, Piotrowski A, Hu G: Direct coronary vasomotor effects of sevoflurane and desflurane *in situ* canine hearts. *ANESTHESIOLOGY* 2000; 92:1103-13

56. Kersten JR, Wartler DC: The coronary circulation, Anesthesia: Biologic Foundations. Edited by Yaksh TL, Lynch C III, Zapol WM, Maze M, Biebuyck JF, Saidman LJ. Philadelphia, Lippincott-Raven, 1997, pp 1169-92

57. Smith G, Rogers K, Thorburn J: Halothane improves the balance of oxygen supply to demand in acute experimental myocardial ischaemia. *Br J Anaesth* 1980; 52:577-83

58. Bertha BG, Folts JD, Nugent M, Rusy BF: Halothane, but not isoflurane or enflurane, protects against spontaneous and epinephrine-exacerbated acute thrombus formation in stenosed dog coronary arteries. *ANESTHESIOLOGY* 1989; 71:96-102

59. Kersten JR, Schmeling T, Tessmer J, Hettrick DA, Pagel PS, Wartler DC: Sevoflurane selectively increases coronary collateral blood flow independent of K_{ATP} channels *in vivo*. *ANESTHESIOLOGY* 1999; 90:246-56

60. Kowalski C, Zahler S, Becker BF, Flaucher A, Conzen PF, Gerlach E, Peter K: Halothane, isoflurane, and sevoflurane reduce postischemic adhesion of neutrophils in the coronary system. *ANESTHESIOLOGY* 1997; 86:188-95

61. de Klaver MJ, Manning L, Palmer LA, Rich GF: Isoflurane pretreatment inhibits cytokine-induced cell death in cultured rat smooth muscle cells and human endothelial cells. *ANESTHESIOLOGY* 2002; 97:24-32

62. Yvon A, Hanouz JL, Haelewyn B, Terrien X, Massetti M, Babatasi G, Khayat A, Ducouret P, Bricard H, Gerard JL: Mechanisms of sevoflurane-induced myocardial preconditioning in isolated human right atria *in vitro*. *ANESTHESIOLOGY* 2003; 99:27-33
63. Hanouz JL, Yvon A, Massetti M, Lepage O, Babatasi G, Khayat A, Bricard H, Gerard JL: Mechanisms of desflurane-induced preconditioning in isolated human right atria *in vitro*. *ANESTHESIOLOGY* 2002; 97:33-41
64. An J, Varadarajan SG, Novalija E, Stowe DF: Ischemic and anesthetic preconditioning reduces cytosolic $[Ca^{2+}]_i$ and improves Ca^{2+} responses in intact hearts. *Am J Physiol Heart Circ Physiol* 2001; 281:H1508-23
65. Eskinder H, Rusch NJ, Supan FD, Kampine JP, Bosnjak ZJ: The effects of volatile anesthetics on L- and T-type calcium channel currents in canine cardiac Purkinje cells. *ANESTHESIOLOGY* 1991; 74:919-26
66. Bosnjak ZJ, Aggarwal A, Turner LA, Kampine JM, Kampine JP: Differential effects of halothane, enflurane, and isoflurane on Ca^{2+} transients and papillary muscle tension in guinea pigs. *ANESTHESIOLOGY* 1992; 76:123-31
67. Lynch C III: Effects of halothane and isoflurane on isolated human ventricular myocardium. *ANESTHESIOLOGY* 1988; 68:429-32
68. Hatakeyama N, Momose Y, Ito Y: Effects of sevoflurane on contractile responses and electrophysiological properties in canine single cardiac myocytes. *ANESTHESIOLOGY* 1995; 82:559-65
69. Roscoe AK, Christensen JD, Lynch C III: Isoflurane, but not halothane, induces protection of human myocardium *via* adenosine A1 receptors and adenosine triphosphate-sensitive potassium channels. *ANESTHESIOLOGY* 2000; 92:1692-701
70. Kersten JR, Orth KG, Pagel PS, Mei DA, Gross GJ, Warltier DC: Role of adenosine in isoflurane-induced cardioprotection. *ANESTHESIOLOGY* 1997; 86:1128-39
71. Toller WG, Montgomery MW, Pagel PS, Hettrick DA, Warltier DC, Kersten JR: Isoflurane-enhanced recovery of canine stunned myocardium: Role for protein kinase C? *ANESTHESIOLOGY* 1999; 91:713-22
72. Ludwig LM, Weihrauch D, Kersten JR, Pagel PS, Warltier DC: Protein kinase C translocation and Src protein tyrosine kinase activation mediate isoflurane-induced preconditioning *in vitro*: Potential downstream targets of mitochondrial adenosine triphosphate-sensitive potassium channels and reactive oxygen species. *ANESTHESIOLOGY* 2004; 100:532-9
73. Toller WG, Kersten JR, Gross ER, Pagel PS, Warltier DC: Isoflurane preconditions myocardium against infarction *via* activation of inhibitory guanine nucleotide binding proteins. *ANESTHESIOLOGY* 2000; 92:1400-7
74. Mullenheim J, Ebel D, Frassdorf J, Preckel B, Thamer V, Schlack W: Isoflurane preconditions myocardium against infarction *via* release of free radicals. *ANESTHESIOLOGY* 2002; 96:934-40
75. Tanaka K, Weihrauch D, Kehl F, Ludwig LM, LaDisa JF Jr, Kersten JR, Pagel PS, Warltier DC: Mechanism of preconditioning by isoflurane in rabbits: A direct role for reactive oxygen species. *ANESTHESIOLOGY* 2002; 97:1485-90
76. Novalija E, Varadarajan SG, Camara AK, An J, Chen Q, Riess ML, Hogg N, Stowe DF: Anesthetic preconditioning: Triggering role of reactive oxygen and nitrogen species in isolated hearts. *Am J Physiol Heart Circ Physiol* 2002; 283:H44-52
77. Piriou V, Chiari P, Knezynski S, Bastien O, Loufoua J, Lehot JJ, Foex P, Annat G, Ovize M: Prevention of isoflurane-induced preconditioning by 5-hydroxydecanoate and gadolinium: Possible involvement of mitochondrial adenosine triphosphate-sensitive potassium and stretch-activated channels. *ANESTHESIOLOGY* 2000; 93:756-64
78. Toller WG, Gross ER, Kersten JR, Pagel PS, Gross GJ, Warltier DC: Sarcolemmal and mitochondrial adenosine triphosphate-dependent potassium channels: Mechanism of desflurane-induced cardioprotection. *ANESTHESIOLOGY* 2000; 92:1731-9
79. Ismaeil MS, Tkachenko I, Gamperl AK, Hickey RF, Cason BA: Mechanisms of isoflurane-induced myocardial preconditioning in rabbits. *ANESTHESIOLOGY* 1999; 90:812-21
80. Inagaki N, Gono T, Clement JP IV, Namba N, Inazawa J, Gonzalez G, Aguilar-Bryan L, Seino S, Bryan J: Reconstitution of IKATP: An inward rectifier subunit plus the sulfonylurea receptor. *Science* 1995; 270:1166-70
81. Noma A: ATP-regulated K⁺ channels in cardiac muscle. *Nature* 1983; 305:147-8
82. Inoue I, Nagase H, Kishi K, Higuti T: ATP-sensitive K⁺ channel in the mitochondrial inner membrane. *Nature* 1991; 352:244-7
83. Liu Y, Ren G, O'Rourke B, Marban E, Seharaseyon J: Pharmacological comparison of native mitochondrial K(ATP) channels with molecularly defined surface K(ATP) channels. *Mol Pharmacol* 2001; 59:225-30
84. Kersten JR, Gross GJ, Pagel PS, Warltier DC: Activation of adenosine triphosphate-regulated potassium channels: Mediation of cellular and organ protection. *ANESTHESIOLOGY* 1998; 88:495-513
85. Gross GJ, Auchampach JA: Blockade of ATP-sensitive potassium channels prevents myocardial preconditioning in dogs. *Circ Res* 1992; 70:223-33
86. O'Rourke B: Myocardial K(ATP) channels in preconditioning. *Circ Res* 2000; 87:845-55
87. Kersten JR, Schmeling TJ, Hettrick DA, Pagel PS, Gross GJ, Warltier DC: Mechanism of myocardial protection by isoflurane: Role of adenosine triphosphate-regulated potassium (K_{ATP}) channels. *ANESTHESIOLOGY* 1996; 85:794-807
88. Nakayama M, Fujita S, Kanaya N, Tsuchida H, Namiki A: Blockade of ATP-sensitive K⁺ channel abolishes the anti-ischemic effects of isoflurane in dog hearts. *Acta Anaesthesiol Scand* 1997; 41:531-5
89. Kwok WM, Martinelli AT, Fujimoto K, Suzuki A, Stadnicka A, Bosnjak ZJ: Differential modulation of the cardiac adenosine triphosphate-sensitive potassium channel by isoflurane and halothane. *ANESTHESIOLOGY* 2002; 97:50-6
90. Fujimoto K, Bosnjak ZJ, Kwok WM: Isoflurane-induced facilitation of the cardiac sarcolemmal K_{ATP} channel. *ANESTHESIOLOGY* 2002; 97:57-65
91. Han J, Kim E, Ho WK, Earm YE: Effects of volatile anesthetic isoflurane on ATP-sensitive K⁺ channels in rabbit ventricular myocytes. *Biochem Biophys Res Commun* 1996; 229:852-6
92. Stadnicka A, Kwok WM, Warltier DC, Bosnjak ZJ: Protein tyrosine kinase-dependent modulation of isoflurane effects on cardiac sarcolemmal K_{ATP} channel. *ANESTHESIOLOGY* 2002; 97:1198-208
93. Gassmayr S, Stadnicka A, Suzuki A, Kwok WM, Bosnjak ZJ: Isoflurane sensitizes the cardiac sarcolemmal adenosine triphosphate-sensitive potassium channel to pinacidil. *ANESTHESIOLOGY* 2003; 98:114-20
94. Stadnicka A, Bosnjak ZJ: Isoflurane decreases ATP sensitivity of guinea pig cardiac sarcolemmal K_{ATP} channel at reduced intracellular pH. *ANESTHESIOLOGY* 2003; 98:396-403
95. Kohro S, Hogan QH, Nakae Y, Yamakage M, Bosnjak ZJ: Anesthetic effects on mitochondrial ATP-sensitive K channel. *ANESTHESIOLOGY* 2001; 95:1435-40
96. Hanley PJ, Mickel M, Loffler M, Brandt U, Daut J: K(ATP) channel-independent targets of diazoxide and 5-hydroxydecanoate in the heart. *J Physiol* 2002; 542:735-41
97. Nakae Y, Kwok WM, Bosnjak ZJ, Jiang MT: Isoflurane activates rat mitochondrial ATP-sensitive K⁺ channels reconstituted in lipid bilayers. *Am J Physiol Heart Circ Physiol* 2003; 284:H1865-71
98. Ichinari K, Kakei M, Matsuoka T, Nakashima H, Tanaka H: Direct activation of the ATP-sensitive potassium channel by oxygen free radicals in guinea-pig ventricular cells: Its potentiation by MgADP. *J Mol Cell Cardiol* 1996; 28:1867-77
99. Sasaki N, Sato T, Marban E, O'Rourke B: ATP consumption by uncoupled mitochondria activates sarcolemmal K(ATP) channels in cardiac myocytes. *Am J Physiol Heart Circ Physiol* 2001; 280:H1882-8
100. Daut J, Maier-Rudolph W, von Beckerath N, Mehrke G, Gunther K, Goedel-Meinen L: Hypoxic dilation of coronary arteries is mediated by ATP-sensitive potassium channels. *Science* 1990; 247:1341-4
101. Kehl F, Krolkowski JG, Tessmer JP, Pagel PS, Warltier DC, Kersten JR: Increases in coronary collateral blood flow produced by sevoflurane are mediated by calcium-activated potassium (BKCa) channels *in vivo*. *ANESTHESIOLOGY* 2002; 97:725-31
102. Piriou V, Ross S, Bastien O, Pigott D, Trivin F, Foex P: Cardiovascular effects of concomitant administration of isoflurane and nicorandil in dogs. *Br J Anaesth* 1998; 80:481-7
103. Patel HH, Ludwig LM, Fryer RM, Hsu AK, Warltier DC, Gross GJ: Delta opioid agonists and volatile anesthetics facilitate cardioprotection via potentiation of K(ATP) channel opening. *FASEB J* 2002; 16:1468-70
104. Van Wylen DG: Effect of ischemic preconditioning on interstitial purine metabolite and lactate accumulation during myocardial ischemia. *Circulation* 1994; 89:2283-9
105. Mizumura T, Nithipatikom K, Gross GJ: Bimacalim, an ATP-sensitive potassium channel opener, mimics the effects of ischemic preconditioning to reduce infarct size, adenosine release, and neutrophil function in dogs. *Circulation* 1995; 92:1236-45
106. Fryer RM, Wang Y, Hsu AK, Gross GJ: Essential activation of PKC-delta in opioid-initiated cardioprotection. *Am J Physiol Heart Circ Physiol* 2001; 280:H1346-53
107. Fryer RM, Hsu AK, Nagase H, Gross GJ: Opioid-induced cardioprotection against myocardial infarction and arrhythmias: Mitochondrial versus sarcolemmal ATP-sensitive potassium channels. *J Pharmacol Exp Ther* 2000; 294:451-7
108. Schultz Jel J, Hsu AK, Nagase H, Gross GJ: TAN-67, a delta 1-opioid receptor agonist, reduces infarct size via activation of Gi/o proteins and KATP channels. *Am J Physiol* 1998; 274:H909-14
109. Patel HH, Hsu A, Moore J, Gross GJ: BW373U86, a delta opioid agonist, partially mediates delayed cardioprotection via a free radical mechanism that is independent of opioid receptor stimulation. *J Mol Cell Cardiol* 2001; 33:1455-65
110. Fryer RM, Hsu AK, Eells JT, Nagase H, Gross GJ: Opioid-induced second window of cardioprotection: Potential role of mitochondrial KATP channels. *Circ Res* 1999; 84:846-51
111. McPherson BC, Yao Z: Morphine mimics preconditioning via free radical signals and mitochondrial K(ATP) channels in myocytes. *Circulation* 2001; 103:290-5
112. Liang BT, Gross GJ: Direct preconditioning of cardiac myocytes via opioid receptors and KATP channels. *Circ Res* 1999; 84:1396-400
113. Schultz JE, Hsu AK, Gross GJ: Morphine mimics the cardioprotective effect of ischemic preconditioning via a glibenclamide-sensitive mechanism in the rat heart. *Circ Res* 1996; 78:1100-4
114. Schultz JE, Hsu AK, Gross GJ: Ischemic preconditioning in the intact rat heart is mediated by delta-1- but not mu- or kappa-opioid receptors. *Circulation* 1998; 97:1282-9
115. Ishizawa Y, Pidikiti R, Liebman PA, Eckenhoff RG: G protein-coupled receptors as direct targets of inhaled anesthetics. *Mol Pharmacol* 2002; 61:945-52
116. Fryer RM, Schultz JE, Hsu AK, Gross GJ: Importance of PKC and tyrosine

- kinase in single or multiple cycles of preconditioning in rat hearts. *Am J Physiol Heart Circ Physiol* 1999; 276:H1229-35
117. Fryer RM, Patel HH, Hsu AK, Gross GJ: Stress-activated protein kinase phosphorylation during cardioprotection in the ischemic myocardium. *Am J Physiol Heart Circ Physiol* 2001; 281:H1184-92
118. Fryer RM, Pratt PF, Hsu AK, Gross GJ: Differential activation of extracellular signal regulated kinase isoforms in preconditioning and opioid-induced cardioprotection. *J Pharmacol Exp Ther* 2001; 296:642-9
119. Liu H, McPherson BC, Yao Z: Preconditioning attenuates apoptosis and necrosis: Role of protein kinase C epsilon and -delta isoforms. *Am J Physiol Heart Circ Physiol* 2001; 281:H404-10
120. Puceat M, Vassort G: Signalling by protein kinase C isoforms in the heart. *Mol Cell Biochem* 1996; 157:65-72
121. Liu GS, Thornton J, Van Winkle DM, Stanley AW, Olsson RA, Downey JM: Protection against infarction afforded by preconditioning is mediated by A1 adenosine receptors in rabbit heart. *Circulation* 1991; 84:350-6
122. Tsuchida A, Miura T, Miki T, Shimamoto K, Iimura O: Role of adenosine receptor activation in myocardial infarct size limitation by ischaemic preconditioning. *Cardiovasc Res* 1992; 26:456-61
123. Wall TM, Sheehy R, Hartman JC: Role of bradykinin in myocardial preconditioning. *J Pharmacol Exp Ther* 1994; 270:681-9
124. Goto M, Liu Y, Yang XM, Ardell JL, Cohen MV, Downey JM: Role of bradykinin in protection of ischemic preconditioning in rabbit hearts. *Circ Res* 1995; 77:611-21
125. Schultz JE, Rose E, Yao Z, Gross GJ: Evidence for involvement of opioid receptors in ischemic preconditioning in rat hearts. *Am J Physiol Heart Circ Physiol* 1995; 268:H2157-61
126. Schultz JJ, Hsu AK, Gross GJ: Ischemic preconditioning and morphine-induced cardioprotection involve the delta (delta)-opioid receptor in the intact rat heart. *J Mol Cell Cardiol* 1997; 29:2187-95
127. Hemmings HC Jr, Adamo AI: Activation of endogenous protein kinase C by halothane in synaptosomes. *ANESTHESIOLOGY* 1996; 84:652-62
128. Hemmings HC Jr: General anesthetic effects on protein kinase C. *Toxicol Lett* 1998; 100-101:89-95
129. Uecker M, da Silva R, Grampp T, Pasch T, Schaub MC, Zaugg M: Translocation of protein kinase C isoforms to subcellular targets in ischemic and anesthetic preconditioning. *ANESTHESIOLOGY* 2003; 99:138-47
130. Ismaeil MS, Tkachenko I, Hickey RF, Cason BA: Colchicine inhibits isoflurane-induced preconditioning. *ANESTHESIOLOGY* 1999; 91:1816-22
131. Sato T, O'Rourke B, Marban E: Modulation of mitochondrial ATP-dependent K⁺ channels by protein kinase C. *Circ Res* 1998; 83:110-4
132. Hu K, Duan D, Li GR, Nattel S: Protein kinase C activates ATP-sensitive K⁺ current in human and rabbit ventricular myocytes. *Circ Res* 1996; 78:492-8
133. Liu Y, Gao WD, O'Rourke B, Marban E: Synergistic modulation of ATP-sensitive K⁺ currents by protein kinase C and adenosine: Implications for ischemic preconditioning. *Circ Res* 1996; 78:443-54
134. Sato T, Sasaki N, O'Rourke B, Marban E: Adenosine primes the opening of mitochondrial ATP-sensitive potassium channels: A key step in ischemic preconditioning? *Circulation* 2000; 102:800-5
135. Light PE, Bladen C, Winkfein RJ, Walsh MP, French RJ: Molecular basis of protein kinase C-induced activation of ATP-sensitive potassium channels. *Proc Natl Acad Sci U S A* 2000; 97:9058-63
136. Ohnuma Y, Miura T, Miki T, Tanno M, Kuno A, Tsuchida A, Shimamoto K: Opening of mitochondrial K(ATP) channel occurs downstream of PKC-epsilon activation in the mechanism of preconditioning. *Am J Physiol Heart Circ Physiol* 2002; 283:H440-7
137. Baines CP, Wang L, Cohen MV, Downey JM: Protein tyrosine kinase is downstream of protein kinase C for ischemic preconditioning's anti-infarct effect in the rabbit heart. *J Mol Cell Cardiol* 1998; 30:383-92
138. Ping P, Zhang J, Zheng YT, Li RC, Dawn B, Tang XL, Takano H, Balafanova Z, Bolli R: Demonstration of selective protein kinase C-dependent activation of Src and Lck tyrosine kinases during ischemic preconditioning in conscious rabbits. *Circ Res* 1999; 85:542-50
139. Ping P, Zhang J, Cao X, Li RC, Kong D, Tang XL, Qiu Y, Manchikalapudi S, Auchampach JA, Black RG, Bolli R: PKC-dependent activation of p44/p42 MAPKs during myocardial ischemia-reperfusion in conscious rabbits. *Am J Physiol Heart Circ Physiol* 1999; 276:H1468-81
140. Ytrehus K, Liu Y, Downey JM: Preconditioning protects ischemic rabbit heart by protein kinase C activation. *Am J Physiol Heart Circ Physiol* 1994; 266:H1145-52
141. Tanno M, Tsuchida A, Nozawa Y, Matsumoto T, Hasegawa T, Miura T, Shimamoto K: Roles of tyrosine kinase and protein kinase C in infarct size limitation by repetitive ischemic preconditioning in the rat. *J Cardiovasc Pharmacol* 2000; 35:345-52
142. Hattori R, Otani H, Uchiyama T, Imamura H, Cui J, Maulik N, Cordis GA, Zhu L, Das DK: Src tyrosine kinase is the trigger but not the mediator of ischemic preconditioning. *Am J Physiol Heart Circ Physiol* 2001; 281:H1066-74
143. Fryer RM, Hsu AK, Gross GJ: ERK and p38 MAP kinase activation are components of opioid-induced delayed cardioprotection. *Basic Res Cardiol* 2001; 96:136-42
144. Sanada S, Kitakaze M, Papst PJ, Hatanaka K, Asanuma H, Aki T, Shinozaki Y, Ogita H, Node K, Takashima S, Asakura M, Yamada J, Fukushima T, Ogai A, Kuzuya T, Mori H, Terada N, Yoshida K, Hori M: Role of phasic dynamism of p38 mitogen-activated protein kinase activation in ischemic preconditioning of the canine heart. *Circ Res* 2001; 88:175-80
145. Mocanu MM, Baxter GF, Yue Y, Critz SD, Yellon DM: The p38 MAPK inhibitor, SB203580, abrogates ischaemic preconditioning in rat heart but timing of administration is critical. *Basic Res Cardiol* 2000; 95:472-8
146. Canman CE, Kastan MB: Signal transduction: Three paths to stress relief. *Nature* 1996; 384:213-4
147. Ichijo H, Nishida E, Irie K, ten Dijke P, Saitoh M, Moriguchi T, Takagi M, Matsumoto K, Miyazono K, Gotoh Y: Induction of apoptosis by ASK1, a mammalian MAPKKK that activates SAPK/JNK and p38 signaling pathways. *Science* 1997; 275:90-4
148. Xia Z, Dickens M, Raingeaud J, Davis RJ, Greenberg ME: Opposing effects of ERK and JNK-p38 MAP kinases on apoptosis. *Science* 1995; 270:1326-31
149. da Silva R, Grampp T, Pasch T, Schaub MC, Zaugg M: Differential activation of mitogen-activated protein kinases in ischemic and anesthetic preconditioning. *ANESTHESIOLOGY* 2004; 100:59-69
150. Zweier JL, Flaherty JT, Weisfeldt ML: Direct measurement of free radical generation following reperfusion of ischemic myocardium. *Proc Natl Acad Sci U S A* 1987; 84:1404-7
151. Bolli R, Patel BS, Jeroudi MO, Lai EK, McCay PB: Demonstration of free radical generation in "stunned" myocardium of intact dogs with the use of the spin trap alpha-phenyl N-tert-butyl nitron. *J Clin Invest* 1988; 82:476-85
152. Ambrosio G, Zweier JL, Duilio C, Kuppusamy P, Santoro G, Elia PP, Tritto I, Cirillo P, Condorelli M, Chiariello M: Evidence that mitochondrial respiration is a source of potentially toxic oxygen free radicals in intact rabbit hearts subjected to ischemia and reflow. *J Biol Chem* 1993; 268:18532-41
153. Tanguay M, Blaise G, Dumont L, Beique G, Hollmann C: Beneficial effects of volatile anesthetics on decrease in coronary flow and myocardial contractility induced by oxygen-derived free radicals in isolated rabbit hearts. *J Cardiovasc Pharmacol* 1991; 18:863-70
154. Nakamura T, Kashimoto S, Oguchi T, Kumazawa T: Hydroxyl radical formation during inhalation anesthesia in the reperfused working rat heart. *Can J Anaesth* 1999; 46:470-5
155. Glantz L, Ginosar Y, Chevion M, Gozal Y, Elami A, Navot N, Kitrossky N, Drenger B: Halothane prevents posts ischemic production of hydroxyl radicals in the canine heart. *ANESTHESIOLOGY* 1997; 86:440-7
156. Hu G, Vinten-Johansen J, Salem MR, Zhao ZQ, Crystal GJ: Isoflurane inhibits neutrophil-endothelium interactions in the coronary circulation: Lack of a role for adenosine triphosphate-sensitive potassium channels. *Anesth Analg* 2002; 94:849-56
157. Hu G, Vasiliauskas T, Salem MR, Rhone DP, Crystal GJ: Neutrophils pretreated with volatile anesthetics lose ability to cause cardiac dysfunction. *ANESTHESIOLOGY* 2003; 98:712-8
158. Ambrosio G, Tritto I, Chiariello M: The role of oxygen free radicals in preconditioning. *J Mol Cell Cardiol* 1995; 27:1035-9
159. Tritto I, D'Andrea D, Eramo N, Scognamiglio A, De Simone C, Violante A, Esposito A, Chiariello M, Ambrosio G: Oxygen radicals can induce preconditioning in rabbit hearts. *Circ Res* 1997; 80:743-8
160. Vanden Hoek TL, Becker LB, Shao Z, Li C, Schumacker PT: Reactive oxygen species released from mitochondria during brief hypoxia induce preconditioning in cardiomyocytes. *J Biol Chem* 1998; 273:18092-8
161. Baines CP, Goto M, Downey JM: Oxygen radicals released during ischemic preconditioning contribute to cardioprotection in the rabbit myocardium. *J Mol Cell Cardiol* 1997; 29:207-16
162. Tanaka M, Fujiwara H, Yamasaki K, Sasayama S: Superoxide dismutase and N-2-mercaptopyrionyl glycine attenuate infarct size limitation effect of ischaemic preconditioning in the rabbit. *Cardiovasc Res* 1994; 28:980-6
163. Pain T, Yang XM, Critz SD, Yue Y, Nakano A, Liu GS, Heusch G, Cohen MV, Downey JM: Opening of mitochondrial K(ATP) channels triggers the preconditioned state by generating free radicals. *Circ Res* 2000; 87:460-6
164. Droge W: Free radicals in the physiological control of cell function. *Physiol Rev* 2002; 82:47-95
165. Clerk A, Michael A, Sugden PH: Stimulation of multiple mitogen-activated protein kinase sub-families by oxidative stress and phosphorylation of the small heat shock protein, HSP25/27, in neonatal ventricular myocytes. *Biochem J* 1998; 333:581-9
166. Samavati L, Monick MM, Sanlioglu S, Buettner GR, Oberley LW, Hunninghake GW: Mitochondrial K(ATP) channel openers activate the ERK kinase by an oxidant-dependent mechanism. *Am J Physiol Cell Physiol* 2002; 283:C273-81
167. Kulisz A, Chen N, Chandel NS, Shao Z, Schumacker PT: Mitochondrial ROS initiate phosphorylation of p38 MAP kinase during hypoxia in cardiomyocytes. *Am J Physiol Lung Cell Mol Physiol* 2002; 282:L1324-9
168. Nishida M, Maruyama Y, Tanaka R, Kontani K, Nagao T, Kurose H: G alpha(i) and G alpha(o) are target proteins of reactive oxygen species. *Nature* 2000; 408:492-5
169. Nishida M, Schey KL, Takagahara S, Kontani K, Katada T, Urano Y, Nagano T, Nagao T, Kurose H: Activation mechanism of Gi and Go by reactive oxygen species. *J Biol Chem* 2002; 277:9036-42
170. Novalija E, Kevin LG, Camara AK, Bosnjak ZJ, Kampine JP, Stowe DF: Reactive oxygen species precede the epsilon isoform of protein kinase C in the anesthetic preconditioning signaling cascade. *ANESTHESIOLOGY* 2003; 99:421-8
171. Patel HH, Gross GJ: Diazoxide induced cardioprotection: What comes

- first, K(ATP) channels or reactive oxygen species? *Cardiovasc Res* 2001; 51: 633-6
172. Carroll R, Gant VA, Yellon DM: Mitochondrial K(ATP) channel opening protects a human atrial-derived cell line by a mechanism involving free radical generation. *Cardiovasc Res* 2001; 51:691-700
173. Forbes RA, Steenbergen C, Murphy E: Diazoxide-induced cardioprotection requires signaling through a redox-sensitive mechanism. *Circ Res* 2001; 88:802-9
174. Zhang DX, Chen YF, Campbell WB, Zou AP, Gross GJ, Li PL: Characteristics and superoxide-induced activation of reconstituted myocardial mitochondrial ATP-sensitive potassium channels. *Circ Res* 2001; 89:1177-83
175. Lebuffe G, Schumacker PT, Shao ZH, Anderson T, Iwase H, Vanden Hoek TL: ROS and NO trigger early preconditioning: Relationship to mitochondrial KATP channel. *Am J Physiol Heart Circ Physiol* 2003; 284:H299-308
176. Gross GJ, Fryer RM: Mitochondrial K(ATP) channels: Triggers or distal effectors of ischemic or pharmacological preconditioning? *Circ Res* 2000; 87: 431-3
177. Tanaka K, Weihrach D, Ludwig LM, Kersten JR, Pagel PS, Wartier DC: Mitochondrial adenosine triphosphate-regulated potassium channel opening acts as a trigger for isoflurane-induced preconditioning by generating reactive oxygen species. *ANESTHESIOLOGY* 2003; 98:935-43
178. Kevin LG, Novalija E, Riess ML, Camara AK, Rhodes SS, Stowe DF: Sevoflurane exposure regenerates superoxide but leads to decreased superoxide during ischemia and reperfusion in isolated hearts. *Anesth Analg* 2003; 96: 949-55
179. Yao Z, Tong J, Tan X, Li C, Shao Z, Kim WC, vanden Hoek TL, Becker LB, Head CA, Schumacker PT: Role of reactive oxygen species in acetylcholine-induced preconditioning in cardiomyocytes. *Am J Physiol Heart Circ Physiol* 1999; 277:H2504-9
180. Vanden Hoek TL, Shao Z, Li C, Schumacker PT, Becker LB: Mitochondrial electron transport can become a significant source of oxidative injury in cardiomyocytes. *J Mol Cell Cardiol* 1997; 29:2441-50
181. Hanley PJ, Ray J, Brandt U, Daut J: Halothane, isoflurane and sevoflurane inhibit NADH:ubiquinone oxidoreductase (complex I) of cardiac mitochondria. *J Physiol* 2002; 544:687-93
182. Riess ML, Stowe DF, Henry MM, Camara AK, Eells JT, Kevin LG: Concentration dependent attenuation of mitochondrial respiration by sevoflurane in isolated cardiac mitochondria is mediated in part by reactive oxygen species (on-line abstract available at: www.anesthesiology.org). *ANESTHESIOLOGY* 2003; 99:A1556
183. Tanaka K, Ludwig LM, Kersten JR, Pagel PS, Wartier DC: Reactive oxygen species released from mitochondrial electron transport chain complex III are involved in isoflurane-induced preconditioning *in vivo* (on-line abstract available at: www.anesthesiology.org). *ANESTHESIOLOGY* 2003; 99:A735
184. Mohazzab KM, Kaminski PM, Wolin MS: NADH oxidoreductase is a major source of superoxide anion in bovine coronary artery endothelium. *Am J Physiol Heart Circ Physiol* 1994; 266:H2568-72
185. Xia Y, Zweier JL: Superoxide and peroxynitrite generation from inducible nitric oxide synthase in macrophages. *Proc Natl Acad Sci U S A* 1997; 94:6954-8
186. Turrens JF: Superoxide production by the mitochondrial respiratory chain. *Biosci Rep* 1997; 17:3-8
187. Cai H, Harrison DG: Endothelial dysfunction in cardiovascular diseases: The role of oxidant stress. *Circ Res* 2000; 87:840-4
188. Abe J, Berk BC: Reactive oxygen species as mediators of signal transduction in cardiovascular disease. *Trends Cardiovasc Med* 1998; 8:59-64
189. Wang XT, McCullough KD, Wang XJ, Carpenter G, Holbrook NJ: Oxidative stress-induced phospholipase C-gamma 1 activation enhances cell survival. *J Biol Chem* 2001; 276:28364-71
190. Maulik N, Watanabe M, Zu YL, Huang CK, Cordis GA, Schley JA, Das DK: Ischemic preconditioning triggers the activation of MAP kinases and MAPKAP kinase 2 in rat hearts. *FEBS Lett* 1996; 396:233-7
191. Aikawa R, Komuro I, Yamazaki T, Zou Y, Kudoh S, Tanaka M, Shiojima I, Hiroi Y, Yazaki Y: Oxidative stress activates extracellular signal-regulated kinases through Src and Ras in cultured cardiac myocytes of neonatal rats. *J Clin Invest* 1997; 100:1813-21
192. Dabrowski A, Boguslowicz C, Dabrowska M, Tribillo I, Gabryelewicz A: Reactive oxygen species activate mitogen-activated protein kinases in pancreatic acinar cells. *Pancreas* 2000; 21:376-84
193. Carter WO, Narayanan PK, Robinson JP: Intracellular hydrogen peroxide and superoxide anion detection in endothelial cells. *J Leukoc Biol* 1994; 55: 253-8
194. Cuzzocrea S, Riley DP, Caputi AP, Salvemini D: Antioxidant therapy: A new pharmacological approach in shock, inflammation, and ischemia/reperfusion injury. *Pharmacol Rev* 2001; 53:135-59
195. Liu Y, Gutterman DD: Oxidative stress and potassium channel function. *Clin Exp Pharmacol Physiol* 2002; 29:305-11
196. Nichols CG, Lederer WJ: The regulation of ATP-sensitive K⁺ channel activity in intact and permeabilized rat ventricular myocytes. *J Physiol* 1990; 423:91-110
197. Yao Z, Gross GJ: Effects of the KATP channel opener bimakalim on coronary blood flow, monophasic action potential duration, and infarct size in dogs. *Circulation* 1994; 89:1769-75
198. Munch-Ellingsen J, Lokebo JE, Bugge E, Jonassen AK, Ravingerova T, Ytrehus K: 5-HD abolishes ischemic preconditioning independently of monophasic action potential duration in the heart. *Basic Res Cardiol* 2000; 95:228-34
199. Hamada K, Yamazaki J, Nagao T: Shortening of action potential duration is not prerequisite for cardiac protection by ischemic preconditioning or a KATP channel opener. *J Mol Cell Cardiol* 1998; 30:1369-79
200. Suzuki M, Sasaki N, Miki T, Sakamoto N, Ohmoto-Sekine Y, Tamagawa M, Seino S, Marban E, Nakaya H: Role of sarcolemmal K(ATP) channels in cardioprotection against ischemia/reperfusion injury in mice. *J Clin Invest* 2002; 109: 509-16
201. Dos Santos P, Kowaltowski AJ, Laclau MN, Seetharaman S, Paucek P, Boudina S, Thambo JB, Tariosse L, Garlid KD: Mechanisms by which opening the mitochondrial ATP-sensitive K(+) channel protects the ischemic heart. *Am J Physiol Heart Circ Physiol* 2002; 283:H284-95
202. Dzeja PP, Holmuhamedov EL, Ozcan C, Pucar D, Jahangir A, Terzic A: Mitochondria: Gateway for cytoprotection. *Circ Res* 2001; 89:744-6
203. Holmuhamedov EL, Wang L, Terzic A: ATP-sensitive K⁺ channel opens prevent Ca²⁺ overload in rat cardiac mitochondria. *J Physiol* 1999; 519:347-60
204. Holmuhamedov EL, Jovanovic S, Dzeja PP, Jovanovic A, Terzic A: Mitochondrial ATP-sensitive K⁺ channels modulate cardiac mitochondrial function. *Am J Physiol Heart Circ Physiol* 1998; 275:H1567-76
205. Minners J, Lacerda L, McCarthy J, Meiring JJ, Yellon DM, Sack MN: Ischemic and pharmacological preconditioning in Girardi cells and C2C12 myotubes induce mitochondrial uncoupling. *Circ Res* 2001; 89:787-92
206. Halestrap AP: The regulation of the matrix volume of mammalian mitochondria in vivo and in vitro and its role in the control of mitochondrial metabolism. *Biochim Biophys Acta* 1989; 973:355-82
207. Garlid KD: Cation transport in mitochondria: The potassium cycle. *Biochim Biophys Acta* 1996; 1275:123-6
208. Garlid KD: On the mechanism of regulation of the mitochondrial K⁺/H⁺ exchanger. *J Biol Chem* 1980; 255:11273-9
209. Green DR, Reed JC: Mitochondria and apoptosis. *Science* 1998; 281: 1309-12
210. Akao M, Ohler A, O'Rourke B, Marban E: Mitochondrial ATP-sensitive potassium channels inhibit apoptosis induced by oxidative stress in cardiac cells. *Circ Res* 2001; 88:1267-75
211. Ozcan C, Bienengraeber M, Dzeja PP, Terzic A: Potassium channel openers protect cardiac mitochondria by attenuating oxidant stress at reoxygenation. *Am J Physiol Heart Circ Physiol* 2002; 282:H531-9
212. Fryer RM, Eells JT, Hsu AK, Henry MM, Gross GJ: Ischemic preconditioning in rats: Role of mitochondrial K(ATP) channel in preservation of mitochondrial function. *Am J Physiol Heart Circ Physiol* 2000; 278:H305-12
213. Tanonaka K, Taguchi T, Koshimizu M, Ando T, Morinaka T, Yogo T, Konishi F, Takeo S: Role of an ATP-sensitive potassium channel opener, YM934, in mitochondrial energy production in ischemic/reperfused heart. *J Pharmacol Exp Ther* 1999; 291:710-6
214. Novalija E, Kevin LG, Eells JT, Henry MM, Stowe DF: Anesthetic preconditioning improves adenosine triphosphate synthesis and reduces reactive oxygen species formation in mitochondria after ischemia by a redox dependent mechanism. *ANESTHESIOLOGY* 2003; 98:1155-63
215. Riess ML, Novalija E, Camara AK, Eells JT, Chen Q, Stowe DF: Preconditioning with sevoflurane reduces changes in nicotinamide adenine dinucleotide during ischemia-reperfusion in isolated hearts: Reversal by 5-hydroxydecanoic acid. *ANESTHESIOLOGY* 2003; 98:387-95
216. Zorov DB, Filburn CR, Klotz LO, Zweier JL, Sollott SJ: Reactive oxygen species (ROS)-induced ROS release: A new phenomenon accompanying induction of the mitochondrial permeability transition in cardiac myocytes. *J Exp Med* 2000; 192:1001-14
217. Kroemer G, Reed JC: Mitochondrial control of cell death. *Nat Med* 2000; 6:513-9
218. Kowaltowski AJ, Castilho RF, Vercesi AE: Mitochondrial permeability transition and oxidative stress. *FEBS Lett* 2001; 495:12-5
219. Hausenloy DJ, Maddock HL, Baxter GF, Yellon DM: Inhibiting mitochondrial permeability transition pore opening: A new paradigm for myocardial preconditioning? *Cardiovasc Res* 2002; 55:534-43
220. Piriou V, Chiari P, Roesch OG, Lehot JJ, Ovize M: Effect of desflurane-induced preconditioning on mitochondrial transition pore opening (on-line abstract available at: www.anesthesiology.org). *ANESTHESIOLOGY* 2003; 99:A1538
221. Allard MF, Flint JD, English JC, Henning SL, Salamanca MC, Kamimura CT, English DR: Calcium overload during reperfusion is accelerated in isolated hypertrophied rat hearts. *J Mol Cell Cardiol* 1994; 26:1551-63
222. Miyamae M, Camacho SA, Weiner MW, Figueredo VM: Attenuation of postischemic reperfusion injury is related to prevention of [Ca²⁺]_i overload in rat hearts. *Am J Physiol Heart Circ Physiol* 1996; 271:H2145-53
223. Di Lisa F, Bernardi P: Mitochondrial function as a determinant of recovery or death in cell response to injury. *Mol Cell Biochem* 1998; 184:379-91
224. Wang L, Cherednichenko G, Hernandez L, Halow J, Camacho SA, Figueredo V, Schaefer S: Preconditioning limits mitochondrial Ca(2+) during ischemia in rat hearts: Role of K(ATP) channels. *Am J Physiol Heart Circ Physiol* 2001; 280:H2321-8
225. Riess ML, Camara AK, Novalija E, Chen Q, Rhodes SS, Stowe DF: Anesthetic preconditioning attenuates mitochondrial Ca²⁺ overload during ischemia in guinea pig intact hearts: Reversal by 5-hydroxydecanoic acid. *Anesth Analg* 2002; 95:1540-6

226. Siegmund B, Schlack W, Ladilov YV, Balsler C, Piper HM: Halothane protects cardiomyocytes against reoxygenation-induced hypercontracture. *Circulation* 1997; 96:4372-9
227. Davies LA, Gibson CN, Boyett MR, Hopkins PM, Harrison SM: Effects of isoflurane, sevoflurane, and halothane on myofilament Ca^{2+} sensitivity and sarcoplasmic reticulum Ca^{2+} release in rat ventricular myocytes. *ANESTHESIOLOGY* 2000; 93:1034-44
228. Zucchi R, Ronca F, Ronca-Testoni S: Modulation of sarcoplasmic reticulum function: A new strategy in cardioprotection? *Pharmacol Ther* 2001; 89:47-65
229. Piper HM, Meuter K, Schafer C: Cellular mechanisms of ischemia-reperfusion injury. *Ann Thorac Surg* 2003; 75:S644-8
230. Yellon DM, Dana A: The preconditioning phenomenon: A tool for the scientist or a clinical reality? *Circ Res* 2000; 87:543-50
231. Kloner RA, Jennings RB: Consequences of brief ischemia: Stunning, preconditioning, and their clinical implications: II. *Circulation* 2001; 104:3158-67
232. Deutsch E, Berger M, Kussmaul WG, Hirshfeld JW Jr, Herrmann HC, Laskey WK: Adaptation to ischemia during percutaneous transluminal coronary angioplasty: Clinical, hemodynamic, and metabolic features. *Circulation* 1990; 82:2044-51
233. Laskey WK: Beneficial impact of preconditioning during PTCA on creatine kinase release. *Circulation* 1999; 99:2085-9
234. Leesar MA, Stoddard M, Ahmed M, Broadbent J, Bolli R: Preconditioning of human myocardium with adenosine during coronary angioplasty. *Circulation* 1997; 95:2500-7
235. Saito S, Mizumura T, Takayama T, Honye J, Fukui T, Kamata T, Moriuchi M, Hibiya K, Tamura Y, Ozawa Y: Antiischemic effects of nicorandil during coronary angioplasty in humans. *Cardiovasc Drugs Ther* 1995; 9(suppl 2):257-63
236. Sakai K, Yamagata T, Teragawa H, Matsuura H, Chayama K: Nicorandil-induced preconditioning as evidenced by troponin T measurements after coronary angioplasty in patients with stable angina pectoris. *Jpn Heart J* 2002; 43:443-53
237. Okazaki Y, Kodama K, Sato H, Kitakaze M, Hirayama A, Mishima M, Hori M, Inoue M: Attenuation of increased regional myocardial oxygen consumption during exercise as a major cause of warm-up phenomenon. *J Am Coll Cardiol* 1993; 21:1597-604
238. Marber MS, Joy MD, Yellon DM: Is warm-up in angina ischaemic preconditioning? *Br Heart J* 1994; 72:213-5
239. Yellon DM, Alkhulaifi AM, Pugsley WB: Preconditioning the human myocardium. *Lancet* 1993; 342:276-7
240. Jenkins DP, Pugsley WB, Alkhulaifi AM, Kemp M, Hooper J, Yellon DM: Ischaemic preconditioning reduces troponin T release in patients undergoing coronary artery bypass surgery. *Heart* 1997; 77:314-8
241. Wu ZK, Tarkka MR, Pehkonen E, Kaukinen L, Honkonen EL, Kaukinen S: Ischaemic preconditioning has a beneficial effect on left ventricular haemodynamic function after a coronary artery bypass grafting operation. *Scand Cardiovasc J* 2000; 34:247-53
242. Wu ZK, Tarkka MR, Pehkonen E, Kaukinen L, Honkonen EL, Kaukinen S: Beneficial effects of ischemic preconditioning on right ventricular function after coronary artery bypass grafting. *Ann Thorac Surg* 2000; 70:1551-7
243. Wu ZK, Iivainen T, Pehkonen E, Laurikka J, Tarkka MR: Ischemic preconditioning suppresses ventricular tachyarrhythmias after myocardial revascularization. *Circulation* 2002; 106:3091-6
244. Laurikka J, Wu ZK, Iisalo P, Kaukinen L, Honkonen EL, Kaukinen S, Tarkka MR: Regional ischemic preconditioning enhances myocardial performance in off-pump coronary artery bypass grafting. *Chest* 2002; 121:1183-9
245. Kaukoranta PK, Lepojarvi MP, Ylitalo KV, Kiviluoma KT, Peuhkurinen KJ: Normothermic retrograde blood cardioplegia with or without preceding ischemic preconditioning. *Ann Thorac Surg* 1997; 63:1268-74
246. Cremer J, Steinhoff G, Karck M, Ahnelt T, Brandt M, Teebken OE, Hollander D, Haverich A: Ischemic preconditioning prior to myocardial protection with cold blood cardioplegia in coronary surgery. *Eur J Cardiothorac Surg* 1997; 12:753-8
247. Cremer J, Karck M, Ahnelt T, Steinhoff G, Brandt M, Hollander D, Teebken O, Zick G, Haverich A: Ischemic preconditioning as an adjunct to crystalloid or blood cardioplegia for myocardial protection in routine coronary surgery. *Thorac Cardiovasc Surg* 1998; 46(suppl 2):298-301
248. Carroll R, Yellon DM: Delayed cardioprotection in a human cardiomyocyte-derived cell line: The role of adenosine, p38MAP kinase and mitochondrial KATP. *Basic Res Cardiol* 2000; 95:243-9
249. Tarnow J, Marksches-Hornung A, Schulte-Sasse U: Isoflurane improves the tolerance to pacing-induced myocardial ischemia. *ANESTHESIOLOGY* 1986; 64:147-56
250. Belhomme D, Peynet J, Louzy M, Launay JM, Kitakaze M, Menasche P: Evidence for preconditioning by isoflurane in coronary artery bypass graft surgery. *Circulation* 1999; 100(suppl 19):II340-4
251. Haroun-Bizri S, Khoury SS, Chehab IR, Kassas CM, Baraka A: Does isoflurane optimize myocardial protection during cardiopulmonary bypass? *J Cardiothorac Vasc Anesth* 2001; 15:418-21
252. Penta de Peppo A, Polisca P, Tomai F, De Paulis R, Turani F, Zupancich E, Sommariva L, Pasqualetti P, Chiariello L: Recovery of LV contractility in man is enhanced by preischemic administration of enflurane. *Ann Thorac Surg* 1999; 68:112-8
253. De Hert SG, ten Broecke PW, Mertens E, Van Sommeren EW, De Blier IG, Stockman BA, Rodrigus IE: Sevoflurane but not propofol preserves myocardial function in coronary surgery patients. *ANESTHESIOLOGY* 2002; 97:42-9
254. Conzen PF, Fischer S, Dettler C, Peter K: Sevoflurane provides greater protection of the myocardium than propofol in patients undergoing off-pump coronary artery bypass surgery. *ANESTHESIOLOGY* 2003; 99:826-33
255. De Hert SG, Cromheecke S, ten Broecke PW, Mertens E, De Blier IG, Stockman BA, Rodrigus IE, Van der Linden PJ: Effects of propofol, desflurane, and sevoflurane on recovery of myocardial function after coronary surgery in elderly high-risk patients. *ANESTHESIOLOGY* 2003; 99:314-23
256. Julier K, da Silva R, Garcia C, Bestmann L, Frascarolo P, Zollinger A, Chassot PG, Schmid ER, Turina MI, von Segesser LK, Pasch T, Spahn DR, Zaugg M: Preconditioning by sevoflurane decreases biochemical markers for myocardial and renal dysfunction in coronary artery bypass graft surgery: A double-blinded, placebo-controlled, multicenter study. *ANESTHESIOLOGY* 2003; 98:1315-27
257. Pouzet B, Lecharny JB, Dehoux M, Paquin S, Kitakaze M, Mantz J, Menasche P: Is there a place for preconditioning during cardiac operations in humans? *Ann Thorac Surg* 2002; 73:843-8
258. Wartier DC, Pagel PS, Kersten JR: Approaches to the prevention of perioperative myocardial ischemia. *ANESTHESIOLOGY* 2000; 92:253-9
259. Van Der Linden PJ, Daper A, Trenchant A, De Hert SG: Cardioprotective effects of volatile anesthetics in cardiac surgery. *ANESTHESIOLOGY* 2003; 99:516-7
260. Sniecinski R, Liu H: The effect of myocardial function protection of sevoflurane preconditioning in senescent rats (on-line abstract available at: www.anesthesiology.org). *ANESTHESIOLOGY* 2003; 99:A1548
261. Zaugg M, da Silva R, Sergeev PV, Pasch T, Schaub MC: Unraveling the gene expression patterns in ischemic and anesthetic preconditioning by gene chip analysis (on-line abstract available at: www.anesthesiology.org). *ANESTHESIOLOGY* 2003; 99:A1544
262. Homi HM, Mixco JM, Sheng H, Grocott HP, Pearlstein RD, Warner DS: Severe hypotension is not essential for isoflurane neuroprotection against forebrain ischemia in mice. *ANESTHESIOLOGY* 2003; 99:1145-51