

## Review

# Dietary Strategies to Counteract the Effects of Mycotoxins: A Review

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### ABSTRACT

We reviewed various dietary strategies to contain the toxic effects of mycotoxins using antioxidant compounds (selenium, vitamins, provitamins), food components (phenolic compounds, coumarin, chlorophyll and its derivatives, fructose, aspartame), medicinal herbs and plant extracts, and mineral and biological binding agents (hydrated sodium calcium aluminosilicate, bentonites, zeolites, activated carbons, bacteria, and yeast). Available data are primarily from in vitro studies and mainly focus on aflatoxin B<sub>1</sub>, whereas much less information is available about other mycotoxins. Compounds with antioxidant properties are potentially very efficacious because of their ability to act as superoxide anion scavengers. Interesting results have been obtained by food components contained in coffee, strawberries, tea, pepper, grapes, turmeric, *Fava tonka*, garlic, cabbage, and onions. Additionally, some medicinal herbs and plant extracts could potentially provide protection against aflatoxin B<sub>1</sub> and fumonisin B<sub>1</sub>. Activated carbons, hydrated sodium calcium aluminosilicate, and bacteria seem to effectively act as binders. We conclude that dietary strategies are the most promising approach to the problem, considering their limited or nil interference in the food production process. Nevertheless, a great research effort is necessary to verify the in vivo detoxification ability of the purposed agents, their mode of action, possible long-term drawbacks of these detoxification-decontamination procedures, and their economical and technical feasibility.

Mycotoxins are highly toxic secondary products of the metabolism of some fungi mainly belonging to *Aspergillus*, *Penicillium*, and *Fusarium* spp. Toxic syndromes caused by mycotoxin ingestion by humans and animals are indicated as mycotoxicosis. It has been estimated that at least 300 fungal metabolites are potentially toxic for humans and animal, and it can be realistically assumed that other mycotoxins are likely to be discovered. Devegowda et al. (28) reported that as much as 25% of the world's cereals are contaminated with known mycotoxins. Mycotoxins can enter the food chain through contaminated cereals and foods (e.g., milk, meat, and eggs) obtained from animals fed mycotoxin-contaminated feeds.

The most notorious and extensively investigated mycotoxins are aflatoxin B<sub>1</sub> (AFB<sub>1</sub>), ochratoxin A (OTA), fumonisin B<sub>1</sub> (FB<sub>1</sub>), zearalenon (ZEN), deoxynivalenol (DON), and T2 toxin. However, recently the interest of researchers in other toxins, such as citrinin, sterigmatocystin, diacetoxyscirpenol, and the group of recently discovered *Fusarium* toxins, including beauvericin, fusaproliferin, moniliformin, and fusaric acid, has been growing.

Mycotoxins' chemical, biological, and toxicological properties are diverse. Hence, their toxic effects are ex-

tremely variable, depending on the intake level, duration of exposure, animal species, age, sex, physiological status, and eventual synergism between mycotoxins simultaneously present in feed or foods. However, the main toxic effects are carcinogenicity, genotoxicity, teratogenicity, nephrotoxicity, hepatotoxicity, reproductive disorders, and immunosuppression. Some mycotoxins are specifically indicated or strongly suspected as the cause of severe human and animal diseases, such as Reye's disease (caused by AFB<sub>1</sub>) (12), equine leukoencephalomalacia and porcine pulmonary edema (caused by FB<sub>1</sub>), human alimentary toxic aleukia (caused by T2 toxin), and Balkan endemic nephropathy (caused by OTA). The positive correlation between the consumption of AFB<sub>1</sub>-contaminated foods and the increased incidence of liver cancer in several Asian and African populations has led to the classification of AFB<sub>1</sub> as a group IA carcinogen by the International Agency for Research on Cancer (56). In 1997, the Joint Food and Agricultural Organization/World Health Organization Expert Committee on Food Additives provided qualitative and quantitative information on aflatoxins and concluded that aflatoxins should be treated as carcinogenic food contaminants, the intake of which should be reduced to levels as low as reasonably achievable (60). Although human health risk assessment involves complete knowledge of toxicological, epidemiological, and exposure data, in the risk management

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of mycotoxins it is necessary to take action before all this information is available (66). However, the lack of this information actually determines some uncertainties in assessing effective human exposure and health risk and in establishing causal relationships between incidence of mycotoxins in foods and human diseases (100).

From a regulatory standpoint, different countries have enforced different thresholds to limit the passage of mycotoxins along the food chain. In the United States, it is required by law that aflatoxin M<sub>1</sub> in milk should be less than 0.5 ppb, whereas in western Europe the regulations are more stringent, and maximum levels are set at 0.05 ppb (15). Currently, the U.S. Food and Drug Administration regulates only aflatoxins among the mycotoxins, and it established an action level ranging from 20 to 300 ppb of AFB<sub>1</sub> in animal feeds. Denmark, on the other hand, has decided that a level of 15 ppb of OTA in the liver or kidney of pigs results in their confiscation and that levels exceeding 25 ppb would result in the confiscation of the entire carcass.

No doubt exists concerning the economic impact of mycotoxins. Recent studies (20, 39) evidenced that economic losses occur at all levels of food and feed production, including crop and animal production, processing, and distribution. Even during favorable climatic periods, millions of dollars are lost as a consequence of crop contamination. For all these reasons, prevention, decontamination, and detoxification of mycotoxins are issues of great importance.

Generally, any approach aimed to reduce the toxic and economic impact of mycotoxins should be respondent to as much as possible of the following requisites: (i) prevent, destroy, remove, or detoxify mycotoxins in foods and feeds; (ii) not produce or leave toxic and/or carcinogenic or mutagenic residues in the final products; (iii) not significantly alter the important technologic and nutritional properties of the food or feed; and (iv) be technically and economically feasible (86).

A wide range of chemical, physical, and biological approaches has been experienced in the attempt to reduce the toxicity of mycotoxins. Although some chemical detoxification methods (i.e., ammonia, sodium bisulfite, and calcium hydroxide treatments) are effective, they do not fulfill all the requirements, especially those concerning the safety of reaction products and the safeguarding of the nutritional characteristics of the treated foods and feeds (29, 86).

For these reasons, nutritional approaches, such as supplementation of nutrients, food components, or additives with protective properties against mycotoxin toxicity and addition of nonnutritive sorbents or bacteria, yeast, and modified yeast cells capable of reducing the bioavailability of mycotoxins, are assuming increasing interest and are critically reviewed in this paper.

## ANTIOXIDANT SUBSTANCES

Since some mycotoxins (i.e., AFB<sub>1</sub>, FB<sub>1</sub>, OTA, and T2 toxin) are known to produce membrane damage through increased lipid peroxidation (8, 9, 25, 52, 53), the protective properties of antioxidant substances have been extensively

investigated. Selenium, some vitamins (A, C, and E), and their precursors have marked antioxidant properties that act as superoxide anion scavengers. For these reasons, these substances have been investigated as protecting agents against toxic effects of mycotoxins.

**Selenium.** In a controlled study, Lin et al. (72) observed that selenium is able to reduce in vitro toxic effects of T2 toxin on cultured chicken embryonic chondrocytes. When Na<sub>2</sub>SeO<sub>3</sub> was added in the culture in the presence of T2 toxin, there was no decrease in collagen microfibril, intramembrane particle numbers, and enzymatic (cytochrome *c*, oxidase, and H<sup>+</sup>-ATPase) activities.

In an in vivo study on rats, Shi et al. (96) demonstrated that selenium inhibits AFB<sub>1</sub>-DNA binding and adduct formation. The same authors (97) in an in vitro study on cultured hamster ovary cells found that sodium selenite and selenium-enriched yeast extract protect cells from AFB<sub>1</sub> cytotoxicity but not from mutagenicity.

However, McLeod et al. (80) reported that rats fed a selenium-deficient diet were more resistant to AFB<sub>1</sub> than those fed a selenium-sufficient diet. According to the authors, the protection conferred by selenium deficiency against AFB<sub>1</sub> is associated with the hepatic expression of an aldo-keto reductase and a glutathione S-transferase subunit that efficiently metabolizes the mycotoxin.

Based on a study on rats, Atroshi et al. (8) concluded that selenium, vitamin E, and vitamin C act as an antioxidant system and free radical scavenger that protects spleen and brain against membrane damage caused by T2 toxin and DON.

**Vitamins.** Further evidence of protective effects of some vitamins and/or their precursors against mycotoxin-induced damages arises from numerous in vivo and in vitro studies.

Grosse et al. (46) observed that vitamins A, C, and E reduced DNA adducts in kidney and liver of mice exposed to OTA and ZEN from 70 to 90%.

Vitamin C is also able to protect guinea pigs from AFB<sub>1</sub> hepatotoxicity (83).

In mice exposed to OTA, vitamin C reduced abnormalities in both mitotic and meiotic chromosomes and morphologies of the sperm head (14). Analogous protective actions have also been attributed to vitamin E (54) and vitamin A toward exposure both to OTA (67) and AFB<sub>1</sub> (98).

Supplementary vitamin E administered to chickens partially counteracts the formation of lipid peroxides due to single and combined exposure to OTA and T2 toxin (52).

As demonstrated by Webster et al. (108), the different vitamin A status is strongly related to the hepatocarcinogenicity of AFB<sub>1</sub> in rats. In fact, the authors observed an enhancement of AFB<sub>1</sub>-induced DNA damages during vitamin A deficiency, whereas damages were reversed on vitamin A supplementation.

Coelho (22) demonstrated that vitamin supplementations exceeding up to 25% of the standard requirements can reduce the negative effects of AFB<sub>1</sub> and environmental stress on turkeys.

Also, riboflavin has a potential chemopreventive action against AFB<sub>1</sub>-induced DNA damages in rats (109).

Carotenoids (carotene and xanthophylls) are excellent antioxidants with antimutagenic and anticarcinogenic properties. They occur naturally in some foods, such as carrots, red tomatoes, butter, cheese, paprika, palm oil, corn kernels, and red salmon. Dietary carotenoids inhibit AFB<sub>1</sub>-induced liver DNA damage in rats, as demonstrated by Gradelet et al. (44, 45). They concluded that beta-apo-8'-carotenol, canthaxanthin, and astaxanthin exert their protective effect through the deviation of AFB<sub>1</sub> metabolism toward detoxification pathways, leading to the formation of aflatoxin M<sub>1</sub>, a less toxic metabolite. Regarding beta-carotene, since it does not alter metabolism, its protective action should be mediated by other mechanisms, whereas no protective effects were observed by administration of supplementary lycopene and excess of vitamin A.

Okotie Eboh et al. (85) conducted a study on broilers to confirm in vivo the protective properties of beta-carotene and canthaxanthin against AFB<sub>1</sub>. The authors concluded that beta-carotene was not effective in ameliorating aflatoxicosis in broiler chickens, whereas canthaxanthin was shown to be somewhat effective with respect to certain blood chemistry indicators.

An overall inhibition of biochemical and cellular events thought to be precursors of AFB<sub>1</sub>-induced hepatocarcinogenesis in rats was observed by He et al. (49) as a consequence of dietary administration of carotenoid-rich extracts. Another study on solvent-extracted carotenoids (i.e., alpha-carotene, beta-carotene, and lycopene) and xanthophylls (beta-cryptoxanthin) from carotenoids-rich foods showed both in vitro and in vivo ability to reduce mutagenic effects of AFB<sub>1</sub> (92).

In a study on antimutagenic activity of natural xanthophylls against AFB<sub>1</sub> in *Salmonella* Typhimurium, González de Mejía et al. (43) reported that xanthophylls inhibited the mutagenicity of AFB<sub>1</sub> in a dose-dependent manner.

Yu et al. (111), using woodchuck hepatocytes as a model to investigate the effects of vitamin A, C, and E and beta-carotene on AFB<sub>1</sub>-DNA adducts, reported contrasting results. In fact, they showed that vitamin C and, particularly, vitamin A were effective in reducing AFB<sub>1</sub>-DNA binding, whereas vitamin E and beta-carotene enhanced it. The authors concluded that additional studies are needed to understand the mechanism of enhanced adduct formation.

Two vitamin A<sub>2</sub> compounds (3-dehydroretinol and 3-dehydroretinyl palmitate) mainly present in freshwater fish have been demonstrated to be very effective in inhibiting the microsome-catalyzed formation of DNA-AFB<sub>1</sub> adduct (3). The inhibition should be due to modulation of microsomal enzymes, which activate the carcinogen, hence suggesting a potential chemopreventive role of these compounds against carcinogenesis induced by AFB<sub>1</sub>.

In a study conducted on *Bacillus subtilis* cells, vitamin E was able to prevent the genotoxicity of ZEN (40). The authors attributed the specificity of the prevention to the structural similarity of vitamin E and ZEN.

## FOOD COMPONENTS AND ADDITIVES

Numerous food components, ingredients, or additives, with or without antioxidant properties, have been investigated with the aim of verifying their chemoprotective properties.

Lu et al. (76) indicated that the toxic effect of FB<sub>1</sub> can be eliminated by a reaction between FB<sub>1</sub> and fructose. When heated to 80°C for 48 h, fructose reacts with the amino terminus of FB<sub>1</sub>, resulting in more than 95% FB<sub>1</sub> conjugation to fructose.

Ellagic acid is a phenolic compound that occurs naturally in some foods, such as strawberries, raspberries, and grapes. It has both antimutagenic and anticarcinogenic activity as demonstrated in a wide range of in vitro and in vivo assays. Loarca Piña et al. (74, 75) in in vitro tests on *Salmonella* cells showed that ellagic acid inhibited AFB<sub>1</sub> direct-acting mutagenicity, particularly when incubated with metabolic enzymes. The result of sequential incubation indicated that the formation of an AFB<sub>1</sub>-ellagic acid chemical complex should be the involved mechanism of inhibition.

Another study emphasized the role of phenolic compounds in the activation and detoxification processes and hence in modulating the carcinogenicity of AFB<sub>1</sub> (2). In tests performed on rats fed a synthetic diet containing various food-associated phenolic compounds each at the 0.5% level, the authors observed a marked decrease in the ability of liver microsomes to catalyze reactions of AFB<sub>1</sub>, leading to its activation and DNA adduct formation. The phenolic compounds tested were several flavonoids (fisetin, kaempferol, morin, naringin, and catechin), phenolic acids (caffeic acid and chlorogenic acid), and other phenolic (eugenol, vanillin) and synthetic phenolics (butylated hydroxyanisole and butylated hydroxytoluene) antioxidants. Some phenolic compounds (naringin, catechin, eugenol, vanillin, and butylated hydroxyanisole) were also found to induce cytosolic glutathione S-transferase activity that stimulated the formation of specific AFB<sub>1</sub>-glutathione conjugate. Rompelberg et al. (95) found that eugenol does not modify the unscheduled DNA synthesis in hepatocytes exposed to AFB<sub>1</sub>.

Williams and Iatropoulos (110) and Manson et al. (78) confirmed that butylated hydroxyanisole and butylated hydroxytoluene inhibited the initiation of hepatocarcinogenesis by AFB<sub>1</sub> in rats. However, Allameh (5) demonstrated that the permitted dose of butylated hydroxytoluene, added to processed food as preservative, plays no role in the biotransformation of AFB<sub>1</sub>. Skrinjar et al. (102) confirmed that ellagic acid and butylated hydroxyanisole ameliorate aflatoxin-induced mutagenicity and carcinogenicity. The same authors also found that certain food additives and/or active ingredients with general antioxidant properties, such as turmeric (*Curcuma longa*), curcumin (diferuloyl methane), and garlic, have the same properties. Firozi et al. (33) demonstrated that curcumin reduces the formation of AFB<sub>1</sub>-DNA adducts by modulating cytochrome P-450 function.

Takahashi et al. (103, 104) and Im et al. (55) demonstrated that two flavonoids, alpha- and beta-naphthoflavone,

strongly inhibit microsome-catalyzed AFB<sub>1</sub>-DNA binding in trout.

In an *in vivo* study on rats, Hendrich et al. (50) reported that soy isoflavone extract has a marked protective action against FB<sub>1</sub> hepatotoxicity by the suppression of FB<sub>1</sub>-stimulated prostaglandin production.

Natural phenolics such as quercetin and kaempferol, ellagic acid, and curcumin generally reduced the *in vitro* enzyme activity consequent to AFB<sub>1</sub> treatment (82). According to the authors, the suppression of protein kinase C activity by phenolic compounds could be a way to control AFB<sub>1</sub> carcinogenicity.

A range of natural dietary constituents, including garlic oil, ethoxyquine, indole-3-carbinol, and phenethyl isothiocyanate, have *in vitro* chemoprotective actions toward AFB<sub>1</sub> (78).

Some natural polyphenolic compounds, i.e., polyhydroxylated flavonoids and phenolic acids, were found to be effective in reducing AFB<sub>1</sub>-DNA adducts (34).

S-methyl methanethiosulfonate, a compound present in the juice of cabbage and onion, has been demonstrated to have a suppressive effect on AFB<sub>1</sub>- or methyl methanesulfonate-induced chromosome aberrations in rat bone marrow cells (58). Also, the precursor of S-methyl methanethiosulfonate S-methyl-L-cysteinesulfoxide significantly suppressed AFB<sub>1</sub>- or methyl methanesulfonate-induced chromosome aberrations. According to the authors, although other mechanisms are not excluded, the protective properties of S-methyl methanethiosulfonate may result from its ability to modify -SH groups in proteins.

Cavin et al. (19) identified two diterpenes, cafestol and kahweol, present in green and roasted coffee beans as potentially chemoprotective agents against the covalent binding of AFB<sub>1</sub> metabolites to DNA of rats. It has been postulated that these compounds may act as blocking agents by producing a coordinated modulation of multiple enzymes involved in carcinogen detoxification. Manson et al. (78) found that caffeic acid also has *in vitro* chemoprotective actions toward AFB<sub>1</sub>. On the contrary, caffeine has been demonstrated to potentiate the *in vitro* genotoxicity of AFB<sub>1</sub> (63).

Theafulvins, compounds isolated from black tea aqueous infusions, despite their overall ability in reducing the mutagenicity of other food carcinogens, enhanced the mutagenicity of AFB<sub>1</sub> (18).

Jesval (59) attributed protective abilities against OTA-induced hepatoma and renal carcinoma in mice to leaf juice of the common grape (*Vitis vinifera*).

L-Methionine supplementation was found to inhibit the developmental toxicity induced by OTA in pregnant female rats and provided partial protection for renal and liver tissue (1).

Propionic acid and potassium sorbate used as preservatives in bread making in France destroyed AFB<sub>1</sub> from approximately 52 to 71% (6).

**Piperine.** According to Reen et al. (93), piperine (1-piperoylpiperidine), the major alkaloid constituent of pepper (*Piper nigrum*), is potentially a protective agent against

carcinogenic effects of AFB<sub>1</sub>. It is well known that the AFB<sub>1</sub> toxicity is bioactivated by the cytochrome P-450 monooxygenases (CYP450). In rat cultured cells, piperine reduced dramatically CYP450B1 activity and counteracted CYP450B1-mediated toxicity of AFB<sub>1</sub>, thus offering a chemopreventive effect against procarcinogens activated by CYP450B1.

**Coumarins.** More than 300 coumarins with general pharmacological and biochemical properties have been identified from green plants (53). Coumarin (1,2-benzopyrone), a natural food constituent especially present in *Fava tonka*, has a chemoprotective action against AFB<sub>1</sub>. As demonstrated by Goeger et al. (42) in *in vitro* studies on hamster ovary cells, liver cells from rats, and chick embryos, coumarin decreased cytotoxicity and mutagenicity of AFB<sub>1</sub>, although with marked species differences in chemoprotection. However, it must be considered that coumarin also has toxic properties and, because of their structural similarity, counteracts vitamin K absorption. Raj et al. (90) showed that different oxygenated substituents on 4-methylcoumarins (i.e., acetoxy>hydroxy>methoxy, in order of inhibition ability) also have chemopreventive properties on AFB<sub>1</sub>-DNA binding *in vitro*.

**Chlorophyll and its derivatives.** Dashwood et al. (27) demonstrated that chlorophyllin (a food grade, water-soluble derivative of the green plant pigment chlorophyll) has chemopreventive properties against wide classes of mutagens, including AFB<sub>1</sub>. Breinholt et al. (16) showed that chlorophyllin acts as an interceptor molecule by forming a strong noncovalent complex with AFB<sub>1</sub>, reducing hepatic AFB<sub>1</sub>-DNA adducts and liver tumors. In particular, it has been demonstrated that the complex formation occurs between the porphyrinlike structure of chlorophyllin and the planar molecular surface of AFB<sub>1</sub>. Arimoto Kobayashi et al. (7) demonstrated that chlorophyllin mixed with chitosan, a polyglucosamine, can form an insoluble saltlike material able to trap AFB<sub>1</sub>. The chemoprotective effect of chlorophyllin was confirmed in an *in vivo* study of Breinholt et al. (17), who showed that 2,000 to 4,000 ppm of chlorophyllin reduced AFB<sub>1</sub>-DNA adduction up to 77% in rainbow trout.

**Aspartame.** Four studies (9, 10, 23, 24) reported that aspartame (L-aspartyl-L-phenylalanine methyl ester) has a wide protective action against OTA-induced subchronic effects. Studies on monkey kidney cells showed that aspartame prevents or partially protects against some typical cytotoxic effects of OTA, such as inhibition of protein synthesis, lipid peroxidation, and leakage of certain enzymes, such as lactate dehydrogenase, gamma-glutamyl transferase, and alkaline phosphatase. *In vitro*, aspartame prevented OTA bindings to plasma proteins. When given to rats, aspartame prevented OTA genotoxicity and nephrotoxicity. When given after intoxication of animals with OTA, aspartame eliminated the toxin efficiently from the body. The protective action should be due to its structural similarity to OTA and phenylalanine. According to the detailed account by Creppy et al. (25), the molecular mechanism me-

diating the preventive effect of aspartame is not only the delivery of phenylalanine by cleavage of the peptide but also the direct effect of the peptide on the bending capacity and transport of the toxin in vivo and in vitro. To the authors, aspartame is the best candidate for preventing OTA-induced subchronic effects, also considering the absence of adverse effects in humans and animals.

**Cyproheptadine.** Cyproheptadine, a serotonin antagonist with appetite-stimulant properties, has been tested to reduce feed refusal due to the presence of DON (87). The authors observed that dosing levels, including various combinations of cyproheptadine and DON, offset the reduction of feed intake. It was concluded that, although serotonergic mechanism is involved in reducing DON-induced feed refusal, further investigations are needed to better understand the reasons of anorectic effect.

**Allyl sulfides.** Le Bon et al. (70) showed that the dietary administration of diallyl sulfide, diallyl disulfide, and allyl mercaptan to rats strongly reduced hepatic DNA breaks induced by AFB<sub>1</sub> and, to a lesser extent, its mutagenicity.

## MEDICINAL HERBS AND PLANT EXTRACTS

Some studies highlighted the capability of several extracts from medicinal herbs and plants to counteract the AFB<sub>1</sub> toxicity.

Al Dakan et al. (4) reported that an ethanol extract from a concentrate of *Cassia senna* (a medicinal herb commonly used as vegetable laxative) inhibits the in vitro mutagenic effect of AFB<sub>1</sub> at low concentrations but not at higher ones. Anthraquinone aglycones and naphthopyrone glycosides, two compounds detected in methanol extract of *Cassia tora*, showed a marked in vitro antimutagenic activity against AFB<sub>1</sub> (21). *Semecarpus anacardium* nut extract was demonstrated to be effective in reducing AFB<sub>1</sub>-induced hepatocarcinoma (88). The administration of a methanol extract of the leaves of *Piper argyrophyllum* normalized the genotoxic effect of AFB<sub>1</sub> on rat cells (89).

A study on rats showed that an extract from *Thonninga sanguinea* is able to protect against acute hepatotoxicity caused by AFB<sub>1</sub> (47). Carnosol and carnosic acid, two natural polyphenols found in *Rosmarinus officinalis* L., are potent inhibitors of in vitro AFB<sub>1</sub>-induced DNA adduct formation (84).

A reduction of in vitro metabolic activation of AFB<sub>1</sub> by *Azadirachta indica* var. *siamensis* and *Momordica charantia* L. was observed by Kusamran et al. (68). In a study on rats, a lignin-enriched extract of the fruit of *Schisandra chinensis* was found to provide hepatoprotective action against AFB<sub>1</sub> by enhancing the hepatic antioxidant and detoxification system (57).

Majonoside-R2, a saponin extracted from rhizomes and roots of *Panax vietnamensis* (also referred to as Vietnamese ginseng), showed a potent anti-tumor-promoting activity against AFB<sub>1</sub> applied to mouse skin (65).

## MYCOTOXIN-BINDING AGENTS

Addition of nutritionally inert sorbents is one of the most recent approaches to reduce mycotoxin toxicity in animals and carryover of mycotoxin from contaminated feeds to animal products (milk, meat, and eggs). After a generalized initial skepticism, the interest of researchers about sorbents has increased in the last years. Sorbents act by reducing the bioavailability of mycotoxins by adsorption on their surface. Indeed, if a stable sorbent-mycotoxin complex is formed, the absorption of mycotoxins in the gastrointestinal tract can be reduced, decreasing both toxic effects for the animal and carryover in animal products for human consumption.

With this aim, numerous sorbents from different sources have been tested, such as hydrated sodium calcium aluminosilicate (HSCAS), zeolites, bentonites, clays, and activated carbons (86, 91).

**HSCAS.** HSCAS, a phyllosilicate derived from natural zeolite, is perhaps the most extensively investigated sorbent. Evidence of a high in vitro and in vivo affinity of HSCAS for AFB<sub>1</sub> arises from numerous studies reviewed by Piva et al. (86) and Ramos et al. (91). However, to these latter authors, enthusiasm for the efficacy of HSCAS must be tempered by the fact that other studies demonstrated its ineffectiveness in binding dangerous mycotoxins other than AFB<sub>1</sub>. Indeed, its protective properties are very low toward OTA and ZEN and nil toward trichothecenes.

**Zeolites.** Zeolites are hydrated aluminosilicates of alkali and alkaline-earth cations characterized by infinite three-dimensional structure (91). Although contrasting results are present in the literature, an overall efficacy of zeolite in binding AFB<sub>1</sub> and ZEN has been reported (86, 91). As evidenced by Piva et al. (86), the origin of zeolite can widely affect results of adsorption tests. In fact, the pore size distribution of synthetic zeolites, as opposed to natural ones, varies very little, being generally concentrated within a narrow diameter range. If the size of the pores is compatible with those of the mycotoxin molecules, adsorption can occur. On the contrary, adsorption can be low or nil because of the absence of intermediate-sized pores. The use of a zeolite, the clinoptilolite, was shown to reduce liver accumulation also when administered to laying hens exposed to AFB<sub>1</sub>, although it had no effect on liver mixed-function oxygenase activities (113). In contrast, Mayura et al. (79) observed dangerous synergetic toxic effects between AFB<sub>1</sub> and clinoptilolite, resulting in severe liver lesions in female rats.

**Bentonites.** Bentonites are sorbents with layered (lamellar) crystalline microstructure and variable composition. Their adsorption properties mainly depend on the interchangeable cations (Na<sup>+</sup>, K<sup>+</sup>, Ca<sup>++</sup>, and Mg<sup>++</sup>) present in the layers (91).

Bentonite has been demonstrated to be able to bind efficaciously AFB<sub>1</sub> in vitro and reduce its toxic effects in trout and swine and also to reduce toxic effects of T2 toxin in rats, as referenced by Ramos et al. (91). Sodium benton-

ite and a synthetic zeolite mixture (80:20 ratio) was reported not to depress feed intake or apparent nutrient digestibility (94) but to prevent AFB<sub>1</sub> accumulation in the liver of growing lambs and decrease by several folds AFB<sub>1</sub> recovery in urine (112). To the authors, the detoxifying properties of bentonite could be enhanced by its ability to reduce the transit time of digestion through the gastrointestinal tract, thus increasing the fecal losses of the toxins. However, this ability was not observed concerning ZEN and nivalenol (91).

**Clays.** Other clays, such as kaolin, sepiolite, and montmorillonite, have a variable ability to reduce toxic effects of AFB<sub>1</sub> as reviewed by Ramos et al. (91). However, their efficacy is limited to AFB<sub>1</sub> and is lower than that of HSCAS and bentonite.

**Activated carbons.** Activated carbons (AC) are an important group of sorbents. They are a family of carbonaceous substances obtained by pyrolysis of several organic compounds and manufactured by activation processes aimed at developing a highly porous structure (38). Generally, the adsorption properties of AC are strictly dependent on the source materials and physicochemical parameters, such as surface area and pore size distribution. Preparation methods and chemical treatments can strongly modify the surface characteristics of AC. Because of the numerous possible combinations between typology of carbonaceous substances and activation processes, many AC with different adsorbing properties exist. This fact could explain some contrasting results reported in a recent review concerning the ability of AC in binding mycotoxins (91).

Since there is no reliable, universal test of adsorptive properties of AC, they must be selected under application conditions. However, to characterize AC well, several physicochemical parameters can be considered (38). Indeed, it is known that the adsorption properties are roughly correlated with the total surface area measured by adsorption of a very small molecule (nitrogen). The pore size distribution of AC is another important characteristic that affects the accessibility of the internal carbon surface. Since diffusion effects inside the pores can slow the adsorption process, the effective pore size distribution of AC can influence it as a function of the molecular size of the adsorbate. Information about mesopores and macropores can be obtained by mercury porosimetry, which allows determination of the size distribution of pores of inside diameter greater than 75 Å. Iodine number is a relative indicator of the microporosity of AC, and it is often used as an approximation of surface area, whereas methylene blue index is a test that established the medium-sized pore (mesopores) range and is an important indicator in practice of the ability of AC to adsorb organic molecules of medium-to-large size from a solution (38).

In a series of preliminary *in vitro* tests (Table 1), we investigated the adsorption ability of 19 experimental AC from different source materials (i.e., exhausted olive residues, peach stones, and almond shells) obtained with laboratory equipment by several experimental activation pro-

TABLE 1. *In vitro* adsorption of mycotoxins from standard solution: saturation limit of activated carbons and HSCAS for aflatoxin B<sub>1</sub>, fumonisin B, aflatoxin B<sub>1</sub> plus fumonisin B<sub>1</sub> (simultaneously), Ochratoxin A, and Deoxynivalenol

Sorbents	Saturation limit <sup>b</sup>				
	Aflatoxin B <sub>1</sub>	Fumonisins B <sub>1</sub>	Aflatoxin B <sub>1</sub> + fumonisin B <sub>1</sub>	Ochratoxin A	Deoxynivalenol
CAC1	123.0	3.8	25.0 + 2.1	115.0	2.0
CAC2	123.0	9.9	25.0 + 5.0	119.0	1.8
CAC4	95.0	9.6	25.0 + 4.7	91.0	1.9
AF32	112.0	3.4	25.0 + 1.8	80.0	2.0
AF48	>125.0	9.8	25.0 + 5.0	121.0	>2.0
HSCAS	79.0	0.3	NA	1.3	0.8

<sup>a</sup> Data are from Galvano et al. (36). HSCAS, hydrated sodium calcium aluminosilicate; CAC, commercial activated carbon; AF, olive residue CAC; NA, not assayed.

<sup>b</sup> Values indicate µg of mycotoxin per mg of sorbent.

cesses appropriately selected to obtain the desired physicochemical parameters and four commercial AC produced in industrial processing equipment.

Overall evidence of the high ability of AC in binding mycotoxins *in vitro* arises from our studies (35, 36, 38). The highest abilities have been observed in the adsorption of AFB<sub>1</sub> and OTA, whereas the lowest in the adsorption of DON (Table 1). AC have been demonstrated to adsorb efficiently FB<sub>1</sub> simultaneously with AFB<sub>1</sub>. When compared with HSCAS, AC showed much higher adsorption abilities toward all the tested mycotoxins. Thus, AC are capable of binding *in vitro* several mycotoxins, and it is reasonable to consider their potential use as multimycotoxin-sequestering agents differently from other extensively studied sorbents, such as HSCAS and bentonite, which are not capable of adsorbing efficiently mycotoxins other than AFB<sub>1</sub>.

The molecular size and physicochemical properties of the mycotoxins clearly affect the efficiency of the binding action. For this reason, further studies on the mechanism of the binding action (i.e., performing studies on chemisorption indices) are needed to clarify the mechanism of the binding process and improve the adsorption performance. We performed two studies with the aim of verifying the *in vivo* ability of one of the AC that showed the highest *in vitro* adsorption abilities. In one experiment on dairy cows, we compared the abilities of AC and HSCAS in reducing carryover of AFB<sub>1</sub> from dairy cows' feed to milk. AC reduced carryover up to 50%, whereas HSCAS reduction of carryover was 36% (37). In a study performed on rats fed an FB<sub>1</sub>-contaminated diet, the bioavailability of the toxin was indirectly monitored by measuring the sphinganine concentration and the sphinganine/sphingosine ratio in urine, liver, and kidney. The addition of 2% AC was effective in avoiding the increase of liver weight. AC also reduced the sphinganine concentration in liver and both sphinganine concentration and sphinganine/sphingosine ratio in kidney (101). Even though the results of the *in vivo* study were lower than those expected based on the *in vitro*

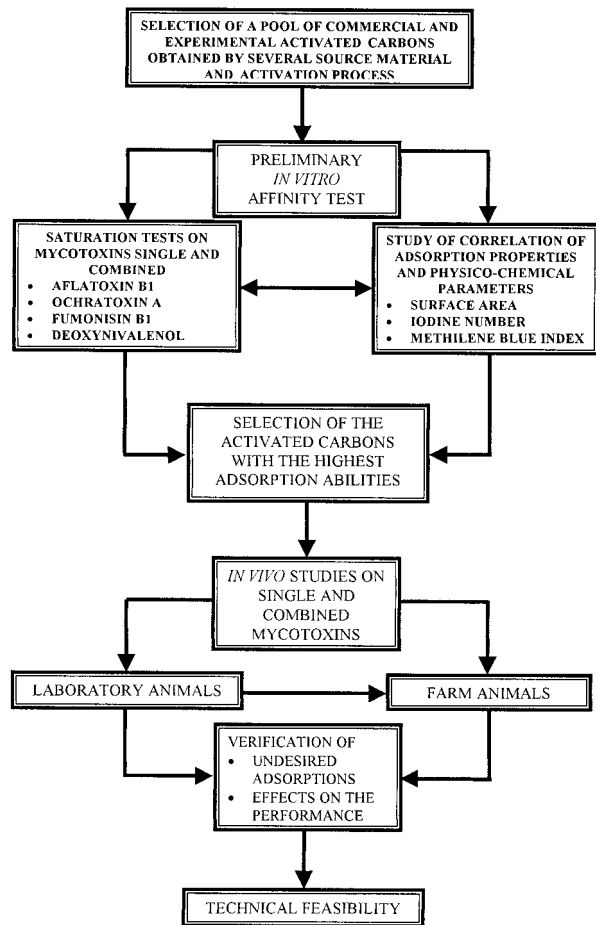


FIGURE 1. Methodological approach to the study of activated carbons as mycotoxin-sequestering agents.

results, data are promising, and further studies are in progress in our laboratory. In any case, that is the first report, to our knowledge, of an in vivo protective action of a sorbent toward  $FB_1$ .

Although the referenced tests are strongly encouraging, we are extremely conscious that additional in vivo studies are required to confirm the efficacy of AC in preventing or reducing the toxic effects of mycotoxins. We are sure that the in vivo efficacy of AC will be lower, since the practical conditions are widely different from the experimental ones, e.g., the binding sites of AC can be occupied by many other compounds present in the feed.

Assuming that in vivo studies would confirm the effectiveness of AC in detoxifying mycotoxins, three questions on whether AC could be added to feeds remain open.

The first is the possible long-term undesired adsorption of essential nutrients (i.e., vitamins and minerals). If long-term in vivo studies should confirm it, two strategies could be adopted: (i) addition of supplemental essential nutrients demonstrated to be excessively adsorbed onto AC or (ii) increase of the selectivity of AC toward mycotoxins by modulating the activation process and the physicochemical properties. The second question, as indicated by Ramos et al. (91), is related to the property of AC to blacken the environment, the animals, and the feed. Some manufactur-

ers have overcome this problem by producing AC that contain up to 65% water and have the consistency of brown sugar, thus eliminating the problems associated with the use of the powder form. We suggested that some of these problems could be also eliminated by pelleting the feed. Furthermore, we observed that, at least for  $AFB_1$ , the pelleting of the feed could increase the efficacy of AC (37). The third is the economic evaluation of the addition of AC to feed. Today the price of AC is perhaps prohibitive to the feed industry. However, the possibility to include AC in feed should largely increase the demand and, consequently, decrease the price. In any case, evaluation of the cost-benefit balance is needed.

**Cholestyramine.** In a study on rats, cholestyramine, a bile acid-binding resin, was tested as protective agent against OTA-induced nephrotoxicity (62). Cholestyramine decreased the concentration of OTA in plasma and the excretion of OTA and its metabolites in urine and bile and increased OTA excretion in feces. These results agree with those of Madhyastha et al. (77). The authors attributed the decreasing of OTA nephrotoxicity to the reduction of its bioavailability and/or enterohepatic circulation. Cholestyramine was also capable of binding efficiently ZEN (106). However, to the authors, its high cost would make its commercial use economically prohibitive.

**Polyvinylpyrrolidone.** Polyvinylpyrrolidone, a synthetic resin, can bind  $AFB_1$  from feed (105). A total of 0.4 g/kg of polyvinylpyrrolidone can bind up to 50  $\mu\text{g/kg}$  of  $AFB_1$  contained in feed. Polyvinylpyrrolidone plus bentonite partially ameliorated some hematological parameters altered by  $AFB_1$  administration to broiler chickens (61).

**Bovine serum albumin.** Hirano et al. (51) demonstrated that bovine serum albumin provides protection from  $AFB_1$  toxic effects. In studies on 1-day-old chicks, the authors observed a marked reduction of histological and biochemical symptoms of exposure to  $AFB_1$  and of  $AFB_1$  levels in the plasma and liver. The authors noted that bovine serum albumin is able to bind  $AFB_1$  in the intestinal tract and is excreted with it. The binding mechanism occurring between bovine serum albumin and  $AFB_1$  was highlighted by Vyjayanthi et al. (107).

**Microbiological-binding agents.** Increasing interest has also been generated by the possibility of using microorganisms to reduce the bioavailability of mycotoxins to farm animals, trying to overcome the inherent drawbacks associated with the use of described sorbents.

*Saccharomyces cerevisiae* 1026, initially used as a performance promoter in the early 1990s, was found to have beneficial effects on weight gain and immune response in broilers exposed to  $AFB_1$  (29). In vitro studies showed an  $AFB_1$  dose-dependent binding capacity of *S. cerevisiae* up to 77% (28). Interestingly, modified mannanoligosaccharide derived from the cell wall of *S. cerevisiae* was reported to have even higher binding capacity (95%  $AFB_1$ , 80% ZEN,  $FB_1$  up to 59%, and DON up to 12%) (28). These indica-

tions were further confirmed by the addition of 0.11% of modified mannanoligosaccharide to the diet of layers receiving 2.5 ppm of AFB<sub>1</sub>. AFB<sub>1</sub> did not contaminate the egg, but a 46% decrease of AFB<sub>1</sub> level in the liver was observed (4.13 versus 2.21 ppb) (113). The transformation of mycotoxins on fermentation has been repeatedly reported. DON and ZEN were degraded in vitro by the normal bacterial gut flora from the distal sections (cecum, colon, and rectum) of the gastrointestinal tract of pigs (64), whereas microorganisms from the cranial segments showed no transformation activity. DON was deepoxidated and ZEN was hydrolyzed to alpha-zearalenol and an unknown metabolite. These observations appear to be confined to the type of mycotoxin considered. In fact, various gram-positive and gram-negative bacteria, incubated with up to 1,000 mM of FB<sub>1</sub>, were shown not to be affected in the growth rate or to be associated with any reduction of FB<sub>1</sub> concentration during the incubation time (11).

*Flavobacterium aurantiacum* was the only microorganism that was shown to significantly remove AFB<sub>1</sub> from liquid medium and food products without the production of toxic by-products (71, 73), although another study conducted by Dsouza and Brackett (30) observed that the presence of trace divalent metal ions, such as Cu, Mn, Zn, and Co, strongly inhibits the binding properties of *F. aurantiacum*.

More recently, some dairy strains of lactic acid bacteria were found capable of removing AFB<sub>1</sub> from contaminated liquid media via a rapid process involving the removal of approximately 80% of AFB<sub>1</sub> immediately on contact without further incubation (31). Heat-treated bacteria had the same ability to remove AFB<sub>1</sub> as viable bacteria; consequently, metabolic degradation by viable bacteria has been ruled out as a possible mode of action under the experimental conditions tested. All the gram-positive strains tested were more efficient than *Escherichia coli*, suggesting that the bacterial ability to remove AFB<sub>1</sub> is dependent on bacterial cell wall structure. The process is temperature and bacterial concentration dependent, whereas no difference was observed due to pH variation across the range of 4 to 6. Furthermore, treatment with hydrochloric acid, autoclaving, or boiling enhanced the binding activity of the bacterial pellets, confirming that the type and structure of the cell wall is crucial for an effective binding of mycotoxins (32).

Megharaj et al. (81) demonstrated the ability of a mixed culture of bacteria to remove ZEN from culture media. Bolognani et al. (13), in a test aimed to determine whether viable cultures of lactic acid-producing organism can bind dietary carcinogens, observed that *Bifidobacterium longum* and *Lactobacillus acidophilus* poorly bound AFB<sub>1</sub>, whereas Lankaputhra and Shah (69) reported that live cells of several strains of the same bacteria bound or inhibited AFB<sub>1</sub>.

*Pseudomonas aeruginosa* was able to inhibit the growth of *Penicillium citrininum* and the production of citrinin (41).

Some strains of yogurt bacteria and bifidobacteria (*Streptococcus salivarius* subsp. *thermophilus*, *Lactobacillus delbrueckii* subsp. *bulgaricus*, and *Bifidobacterium bi-*

*fidum*) were found to reduce the content of OTA in milk by fermentation (99).

## CONCLUSIONS

Encouraging results have been obtained in studies on the protective action of a large number of nutrients, food components, additives, microorganism, and sorbents. The increasing interest of researchers in this field is demonstrated by a constantly growing literature. Nevertheless, most studies were conducted in vitro and confined to AFB<sub>1</sub>. Much less information is available from studies on other mycotoxins, such as OTA, FB<sub>1</sub>, T2 toxin, ZEN, and citrinin. No studies have been performed on recently discovered toxins such as beauvericin, fusaproliferin, moniliformin, and fusaric acid. In addition, most of the studies have been conducted on laboratory animals, whereas only a few were conducted on farm animals. Furthermore, there is a limited knowledge about the efficacy or drawbacks of any of the investigated strategies to prevent extended exposure to mycotoxins. This is comprehensibly due to the preliminary approach research stage in this field. Several natural (vitamins, provitamins, carotenoids, chlorophyll and its derivatives, phenolics, and selenium) and synthetic (butylated hydroxyanisole and butylated hydroxytoluene) compounds with antioxidant properties seem to be potentially very efficacious. The protective properties of antioxidants are probably due to their ability to act as superoxide anion scavengers, thereby protecting cell membrane by mycotoxin-induced damages. However, other more complex mechanisms involving modulation of metabolic detoxification pathways (i.e., microsomal enzymatic activation of AFB<sub>1</sub>), intercepting action and formation of stable nontoxic complexes (i.e., formation of noncovalent complex between chlorophylline and AFB<sub>1</sub>, ellagic acid and AFB<sub>1</sub>, and the reaction between fructose and FB<sub>1</sub>), and structural similarity between mycotoxin and protective agent molecules (i.e., ZEN and vitamin K) have been proposed and are currently being investigated.

Interesting results have also been reported regarding other food components with or without antioxidant properties. Several compounds contained in foods such as coffee, strawberries, tea, pepper, grapes, turmeric, *F. tonka*, garlic, cabbage, onions, medicinal herbs, and plant extract seem to be potentially capable of protecting against mycotoxins. However, further investigations are needed to better understand the modes of action.

Aspartame, a sweetener without adverse effects for humans and animals, has been proposed as a protective agent against the toxic effects of OTA.

A curious fact is that in some cases the sources of potential protective agents are also a potential way to the assumption of mycotoxins, especially AFB<sub>1</sub> and OTA (48).

These are the cases of coffee, pepper, tea, grapes, and medicinal herbs. Thus, caution should be used in promoting antimycotoxin action of discussed substances, since some may be carcinogenic (i.e., quercetin) or toxic (selenium and coumarin) properties.

Use of mycotoxin-binding agents seems to be a very



promising approach to the detoxification of mycotoxins, as demonstrated by 23 studies and two extensive reviews. The positive outcome from the initial investigations of AFB<sub>1</sub> binding to HSCAS and bentonite fostered additional interest in this approach, and further studies were conducted on other sorbents and mycotoxins. Concomitantly, a number of products arrived on the food market claiming multimycotoxin-binding capabilities. It is unfortunate that only a limited number of sorbents were peer reviewed and found capable of binding effectively only AFB<sub>1</sub>, making it difficult to objectively assess the capacity of other sorbents to bind different mycotoxins. None of the sorbents has been scientifically proven to bind simultaneously more than one mycotoxin in *in vivo* studies. Therefore, as emphasized by Dale (26), it is time for real science where mycotoxin binders are concerned.

Figure 1 shows the scientific approach we are using to study AC as mycotoxin binders. Although obviously our approach could be modified, depending on the type of sorbent, we propose it as a general model for the study of sorbents as mycotoxin binders.

Interesting results have been obtained by using microorganisms (i.e., *F. aurantiacum* and some *Lactobacillus* spp.) to remove and/or destroy mycotoxins, even though procedures have not yet been applied.

We think that dietary approaches to reduce the toxic and economic impact of mycotoxins are the most promising approach to the problem, considering their potential ability to fulfill the efficacy, safety, safeguarding of nutritional elements, and cost requisites of a satisfactory detoxification-decontamination process.

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## REFERENCES

- Abdel Wahhab, M. A., S. A. Nada, and M. S. Arbid. 1999. Ochratoxicosis: prevention of developmental toxicity by L-methionine in rats. *J. Appl. Toxicol.* 19:7–12.
- Aboobaker, V. S., A. D. Balgi, and R. K. Bhattacharya. 1994. *In vivo* effect of dietary factors on the molecular action of aflatoxin B<sub>1</sub>: role of non-nutrient phenolic compounds on the catalytic activity of liver fractions. *In Vivo* 8:1095–1098.
- Aboobaker, V. S., N. Sarma, U. C. Goswami, and R. K. Bhattacharya. 1997. Inhibition of microsomal activation of aflatoxin B<sub>1</sub> by 3-dehydroretinol and 3-dehydroretinyl palmitate. *Indian J. Exp. Biol.* 35:1125–1127.
- Al Dakan, A. A., M. Tuffail, and M. A. Hannan. 1995. *Cassia senna* inhibits mutagenic activities of benzo[a]-pyrene, aflatoxin B<sub>1</sub>, shamma and methyl methanesulfonate. *Pharmacol. Toxicol.* 77: 288–292.
- Allameh, A. 1997. Comparison of the effect of low and high dose dietary butylated hydroxytoluene on microsome-mediated aflatoxin B<sub>1</sub>-DNA binding. *Cancer Lett.* 114:217–220.
- Amra, H. A., S. A. Z. Mahmoud, A. H. Taha, and M. A. El-Azab. 1966. Destruction of aflatoxin B<sub>1</sub> and G<sub>1</sub> in bread making. *Mycotox. Res.* 12:73–78.
- Arimoto Kobayhashi, S., N. Narada, R. Tokunaga, J. Odo, and H. Hayatsu. 1997. Adsorption of mutagens to chlorophyllin-chitosan, an insoluble form of chlorophyllin. *Mutat. Res.* 381:243–249.
- Atroschi, F., A. Rizzo, I. Biese, M. Salonen, L. A. Lindberg, and H. Saloniemi. 1995. Effects of feeding T-2 toxin and deoxynivalenol on DNA and GSH contents of brain and spleen of rats supplemented with vitamin E and C and selenium combination. *J. Anim. Phys. Anim. Nutr.* 74:157–164.
- Baudrimont, I., R. Ahouandjivo, and E. E. Creppy. 1997. Prevention of lipid peroxidation induced by ochratoxin A in Vero cells in culture by several agents. *Chem. Biol. Interact.* 104:29–40.
- Baudrimont, I., A. M. Betbeder, and E. E. Creppy. 1997. Reduction of the ochratoxin A-induced toxicity in Vero cells by aspartame. *Arch. Toxicol.* 71:5, 290–298.
- Becker, B., H. Bresch, U. Schillinger, and P. G. Thiel. 1997. The effect of fumonisin B<sub>1</sub> on the growth of bacteria. *World J. Microb. Biot.* 13:539–543.
- Becroft, D. M. W., and D. R. Webster. 1972. Aflatoxins and Reye's disease. *Br. Med. J.* 4:117.
- Bolognani, F., C. J. Rumney, and I. R. Rowland. 1997. Influence of carcinogen binding by lactic acid-producing bacteria on tissue distribution and *in vivo* mutagenicity of dietary carcinogens. *Food Chem. Toxicol.* 35:535–545.
- Bose, S., and S. P. Sinha. 1994. Modulation of ochratoxin-produced genotoxicity in mice by vitamin C. *Food Chem. Toxicol.* 32:533–537.
- Boutrif, E., and C. Canet. 1998. Mycotoxin prevention and control: FAO programmes. *Rev. Med. Vet.* 149:681–694.
- Breinholt, V., J. Hendricks, C. Pereira, D. Arbogast, and G. Bailey. 1995. Dietary chlorophyllin is a potent inhibitor of aflatoxin B<sub>1</sub> hepatocarcinogenesis in rainbow trout. *Cancer Res.* 55:57–62.
- Breinholt, V., M. Schimerlik, R. Dashwood, and G. Bailey. 1995. Mechanisms of chlorophyllin anticarcinogenesis against aflatoxin B<sub>1</sub>: complex formation with the carcinogen. *Chem. Res. Toxicol.* 8: 506–514.
- Catterall, F., E. Copeland, M. N. Clifford, and C. Joannes. 1998. Contribution of theafulvins to antimutagenicity of black tea: their mechanism of action. *Mutagenesis* 13:631–636.
- Cavin, C., D. Holzhäuser, A. Constable, A. C. Huggett, and B. Schilter. 1998. The coffee-specific diterpenes cafestol and kahweol protect against aflatoxin B<sub>1</sub>-induced genotoxicity through a dual mechanism. *Carcinogenesis* 19:1369–1375.
- Charmley, L. L., H. L. Trenholm, D. B. Prelusky, and A. Roseburg. 1995. Economic losses and decontamination. *Nat. Toxins* 3: 199–203.
- Choi, J. S., H. J. Lee, K. Y. Park, J. O. Ha, and S. S. Kang. 1997. *In vitro* antimutagenic effects of anthraquinone glycosides and naphthopyrone glycosides from *Cassia tora*. *Planta Med.* 63:11–14.
- Coelho, M. 1996. Optimum vitamin supplementation needed for turkey performance and profitability. *Feedstuffs* 68:13–21.
- Creppy, E. E., I. Baudrimont, A. Belmadani, and A. M. Betbeder. 1996. Aspartame as a preventive agent of chronic toxic effects of ochratoxin A in experimental animals. *Food Addit. Contam.* 13:51–52.
- Creppy, E. E., I. Baudrimont, and A. M. Betbeder. 1995. Prevention of nephrotoxicity of ochratoxin A, a food contaminant. *Toxicol. Lett.* 71:869–877.
- Creppy, E. E., I. Baudrimont, and A. M. Betbeder. 1998. How aspartame prevents the toxicity of ochratoxin A. *J. Toxicol. Sci.* 2: 165–172.
- Dale, N. 1998. Mycotoxin binders: it's time for real science. *Poult. Digest.* 57:38–39.
- Dashwood, R., T. Negishi, H. Hayatsu, V. Breinholt, J. Hendricks, and G. Bailey. 1998. Chemopreventive properties of chlorophylls toward aflatoxin B<sub>1</sub>: a review of the antimutagenicity and anticarcinogenicity data in rainbow trout. *Mutat. Res.* 399:245–514.
- Devegowda, G., B. I. R. Aravind, and M. G. Morton. 1996. *Saccharomyces cerevisiae* and mannanoligosaccharides to counteract aflatoxicosis in broilers. *Proc. Aust. Poult. Sci. Symp.* 8:103–106.
- Devegowda, G., M. V. L. N. Raju, and H. V. L. N. Swamy. 1998.

- Mycotoxins: novel solutions for their counteraction. *Feedstuffs* 70: 12–15.
30. Dsouza, D. H., and R. E. Brackett. 1998. The role of trace metal ions in aflatoxin B<sub>1</sub> degradation by *Flavobacterium aurantiacum*. *J. Food Prot.* 61:1666–1669.
  31. El-Nezami, H., P. Kankaanpaa, S. Salminen, and J. Ahokas. 1998. Ability of dairy strains of lactic acid bacteria to bind a common food carcinogen, aflatoxin B<sub>1</sub>. *Food Chem. Toxicol.* 36:321–326.
  32. El-Nezami, H., P. Kankaanpaa, S. Salminen, and J. Ahokas. 1998. Physicochemical alterations enhance the ability of dairy strains of lactic acid bacteria to remove aflatoxin from contaminated media. *J. Food Prot.* 61:466–468.
  33. Firozi, P. F., V. S. Aboobaker, and R. K. Bhattacharya. 1996. Action of curcumin on the cytochrome P450-system catalyzing the activation of aflatoxin B<sub>1</sub>. *Chem. Biol. Interact.* 100:41–51.
  34. Firozi, P. F., and R. K. Bhattacharya. 1995. Effects of natural polyphenols on aflatoxin B<sub>1</sub> activation in a reconstituted microsomal monooxygenase system. *J. Biochem. Toxicol.* 10:25–31.
  35. Galvano, F., A. Pietri, T. Bertuzzi, M. Bognanno, A. De Angelis, L. Chies, and M. Galvano. 1997. Activated carbons: in vitro affinity for fumonisin B<sub>1</sub> and relation of adsorption ability to physicochemical parameters. *J. Food Prot.* 60:985–991.
  36. Galvano, F., A. Pietri, T. Bertuzzi, A. De Angelis, A. Piva, L. Chies, and M. Galvano. 1998. Activated carbons: in vitro affinity for Ochratoxin A and Deoxynivalenol and relation of adsorption ability to physicochemical parameters. *J. Food Prot.* 61:469–475.
  37. Galvano, F., A. Pietri, T. Bertuzzi, G. Fusconi, M. Galvano, A. Piva, and G. Piva. 1996. Reduction of carry over of aflatoxin from cow feed to milk by addition of activated carbons. *J. Food Prot.* 59:551–554.
  38. Galvano, F., A. Pietri, B. Fallico, T. Bertuzzi, S. Scirè, M. Galvano, and R. Maggiore. 1996. Activated carbons: in vitro affinity for aflatoxin B<sub>1</sub> and relation of adsorption ability to physicochemical parameters. *J. Food Prot.* 59:545–550.
  39. Garcia, R. P., C. L. Padilla, M. Sidik, B. M. Rejesus, R. P. Garcia, B. R. Champ, M. Bengston, O. S. Dharmaputa, and H. Halid. 1997. Mycotoxins and its significance in the implementation of general agreement on tariff and trade (GATT). In *Proceedings of the Symposium on Pest Management for Stored Food and Feed*, Bogor, Indonesia, September 5–7 1995. BIOTROP Special Publication 59: 33–51.
  40. Ghédira Chékir, L., K. Maaroufi, A. Zakhama, F. Ellouz, S. Dhouib, E. E. Creppy, and H. Bacha. 1998. Induction of a SOS repair system in lysogenic bacteria by zearalenone and its prevention by vitamin E. *Chem. Biol. Interact.* 113:15–25.
  41. Giridar, P., and S. M. Reddy. 1997. Efficacy of some bacteria in the control growth and citrinin production by *Penicillium citrinum*. *Indian Phytopathol.* 50:1–5.
  42. Goeger, D. E., K. E. Anderson, and A. W. Hsie. 1998. Coumarin chemoprotection against aflatoxin B<sub>1</sub>-induced mutation in a mammalian cell system: a species difference in mutagen activation and protection with chick embryo and rat liver S9. *Environ. Mol. Mutagen.* 32:64–74.
  43. González de Mejía, E., M. Ramos Gómez, and G. Loarca Piña. 1997. Antimutagenic activity of natural xanthophylls against aflatoxin B<sub>1</sub> in *Salmonella typhimurium*. *Environ. Mol. Mutagen.* 30: 346–353.
  44. Gradelet, S., P. Astorg, A. M. Le Bon, R. Bergès, and M. Suschetet. 1997. Modulation of aflatoxin B<sub>1</sub> carcinogenicity, genotoxicity and metabolism in rat liver by dietary carotenoids: evidence for a protective effect of CYP1A inducers. *Cancer Lett.* 114:221–223.
  45. Gradelet, S., A. M. Le Bon, R. Bergès, M. Suschetet, and P. Astorg. 1998. Dietary carotenoids inhibit aflatoxin B<sub>1</sub>-induced liver preneoplastic foci and DNA damage in the rat: role of the modulation of aflatoxin B<sub>1</sub> metabolism. *Carcinogenesis* 19:403–411.
  46. Grosse, Y., L. Ghédira Chékir, A. Huc, S. Obrecht Pflumio, G. Dirheimer, H. Bacha, and A. Pfohl Leszkowicz. 1997. Retinol, ascorbic acid and alpha-tocopherol prevent DNA adduct formation in mice treated with the mycotoxins ochratoxin A and zearalenone. *Cancer Lett.* 114:225–229.
  47. Gyamfi, M. A., and Y. Aniya. 1998. Medicinal herb *Thunninga sanguinea* protects against aflatoxin B<sub>1</sub> acute hepatotoxicity in Fisher 344 rats. *Hum. Exp. Toxicol.* 17:418–423.
  48. Halt, M. 1998. Moulds and mycotoxins in herb tea and medicinal plants. *Eur. J. Epidemiol.* 14:269–274.
  49. He, Y., M. M. Root, R. S. Parker, and T. C. Campbell. 1997. Effect of carotenoid-rich food extracts on the development of preneoplastic lesions in rat liver and on in vivo and in vitro antioxidant status. *Nutr. Cancer* 27:238–244.
  50. Hendrich, S., Z. Lu, H. J. Wang, E. C. Hopmans, and P. A. Murphy. Soy isoflavone extract suppresses fumonisin B<sub>1</sub>-promoted rat hepatocarcinogenesis. Presented at Second International Symposium on the Role of Soy in Preventing and Treating Chronic Disease, Brussels, Belgium, 15–18 September 1996.
  51. Hirano, K., Y. Adachi, and S. Ishibashi. 1994. Possible role of bovine serum albumin for the prevention of aflatoxinB<sub>1</sub>-adsorption from the intestinal tract in young chicks. *J. Vet. Med. Sci.* 56:281–286.
  52. Hoehler, D., and R. R. Marquardt. 1996. Influence of vitamins E and C on the toxic effects of ochratoxin A and T-2 toxin in chicks. *Poult. Sci.* 75:1508–1515.
  53. Hoult, J. R., and M. Payá. 1996. Pharmacological and biochemical actions of simple coumarins: natural products with therapeutic potential. *Gen. Pharmacol.* 27:713–722.
  54. Ibeh, I. N., and D. K. Saxena. 1998. Effect of alpha-tocopherol supplementation on the impact of aflatoxin B<sub>1</sub> on the testes of rats. *Exp. Toxicol. Pathol.* 50:221–224.
  55. Im, S. H., M. W. Bolt, R. K. Stewart, and T. E. Massey. 1996. Modulation of aflatoxin B<sub>1</sub> biotransformation by beta-naphthoflavone in isolated rabbit lung cells. *Arch. Toxicol.* 71:72–79.
  56. International Agency for Research on Cancer (IARC). 1993. Some naturally occurring substances: food items and constituent, heterocyclic aromatic amines and mycotoxins. IARC Monographs on the Evaluation of Carcinogenic Risks to Humans, vol. 56. IARC, Lyon.
  57. Ip, S. P., D. H. Mak, P. C. Li, M. K. Poon, and K. M. Ko. 1999. Effect of a lignan-enriched extract of *Schisandra chinensis* on aflatoxin B<sub>1</sub> and cadmium chloride-induced hepatotoxicity in rats. *Pharmacol. Toxicol.* 78:413–416.
  58. Ito, Y., and Y. Nakamura. 1997. Suppression of aflatoxin B<sub>1</sub>- or methyl methanesulfonate-induced chromosome aberrations in rat bone marrow cells after treatment with S-methyl methanethiosulfonate. *Mutat. Res.* 393:307–316.
  59. Jesval, P. 1998. Antidotal effect of grape juice (*Vitis vinifera*) on ochratoxin A caused hepatorenal carcinogenesis in mice (*Mus musculus*). *Cytobios* 93:123–128.
  60. Joint Food and Agricultural Organization/World Health Organization Expert Committee on Food Additives. 1997. Toxicological evaluation of certain food additives. WHO Feed Additives Series. Joint Food and Agricultural Organization/World Health Organization Expert Committee on Food Additives, Geneva.
  61. Keçeci, T., H. Oguz, V. Kurtoglu, and O. Demet. 1998. Effects of polyvinylpyrrolidone, synthetic zeolite and bentonite on serum biochemical and haematological characters of broiler chickens during aflatoxicosis. *Br. Poult. Sci.* 39:452–458.
  62. Kerkadi, A., C. Barriault, B. Tuchweber, A. A. Frohlich, R. R. Marquardt, G. Bouchardand, and I. M. Yousef. 1998. Dietary cholestyramine reduces ochratoxin A-induced nephrotoxicity in the rat by decreasing plasma levels and enhancing fecal excretion of the toxin. *J. Toxicol. Environ. Health* 3:231–250.
  63. Kiefer, F., and F. J. Wiebel. 1998. Caffeine potentiates the formation of micronuclei caused by environmental chemical carcinogens in V79 Chinese hamsters cells. *Toxicol. Lett.* 96–97:131–136.
  64. Kollarczik, B., M. Gareis, and M. Hanelt. 1994. In vitro transformation of the *Fusarium* mycotoxins deoxynivalenol and zearalenone by the normal gut microflora of pigs. *Nat. Toxins* 2:105–110.
  65. Konoshima, T., M. Takasaki, H. Tokuda, H. Nishino, N. M. Duc, R. Kasai, and K. Yamasaki. 1998. Anti-tumor-promoting activity of majonoside-R2 from Vietnamese ginseng, *Panax vietnamensis* Ha et Grushv. (L). *Biol. Pharm. Bull.* 21:834–838.

66. Kuiper Goodman, T. 1995. Mycotoxins: risk assessment and legislation. *Toxicol. Lett.* 82–83:853–859.
67. Kumari, D., and S. P. Sinha. 1994. Effect of retinol on ochratoxin-induced genotoxicity in mice. *Food Chem. Toxicol.* 32:471–475.
68. Kusamran, W. R., A. Ratanavila, and A. Tepsuwan. 1998. Effect of neem flowers, Thai and Chinese bitter melon fruits and sweet basil leaves on hepatic monooxygenases and glutathione S-transferase activities, and in vitro metabolic activation of chemical carcinogens in rats. *Food Chem. Toxicol.* 36:475–484.
69. Lankaputhra, W. E., and N. P. Shah. 1998. Antimutagenic properties of probiotic bacteria and of organic acids. *Mutat. Res.* 397:169–192.
70. Le Bon, A. M., C. Roy, C. Dupont, and M. Suschetet. 1997. In vivo antigenotoxic effects of dietary allyl sulfides in the rat. *Cancer Lett.* 114:131–134.
71. Lillehoj, E. B., A. Ciegler, and H. H. Hall. 1967. Aflatoxin B<sub>1</sub> uptake by *Flavobacterium aurantiacum* and resulting toxic effects. *J. Bacteriol.* 93:464–471.
72. Lin, Z. H., S. G. Li, L. Y. Wu, S. Sun, and Q. W. Lu. 1994. Antagonistic effect of Se on the T-2 toxin-induced changes in the ultrastructure and mitochondrial function of cultured chicken embryonic chondrocytes. *J. Clin. Biochem. Nutr.* 17:119–132.
73. Line, J. E., and R. E. Brackett. 1995. Factors affecting aflatoxin B<sub>1</sub> removal by *Flavobacterium aurantiacum*. *J. Food Prot.* 58:91–94.
74. Loarca Piña, G., P. A. Kuzmicky, E. González de Mejía, and N. Y. Kado. 1998. Inhibitory effects of ellagic acid on the direct-acting mutagenicity of aflatoxin B<sub>1</sub> in the *Salmonella* microsuspension assay. *Mutat. Res.* 398:183–187.
75. Loarca Piña, G., P. A. Kuzmicky, E. González de Mejía, N. Y. Kado, and D. P. Hsieh. 1996. Antimutagenicity of ellagic acid against aflatoxin B<sub>1</sub> in the *Salmonella* microsuspension assay. *Mutat. Res.* 360:15–21.
76. Lu, Z., W. R. Dantzer, E. C. Hopmans, V. Prisk, J. E. Cunnick, P. A. Murphy, and S. Hendrich. 1997. Reaction with fructose detoxifies fumonisin B<sub>1</sub> while stimulating liver-associated natural killer cell activity in rats. *J. Agric. Food Chem.* 45:803–809.
77. Madhyastha, M. S., A. A. Frohlich, and R. R. Marquardt. 1992. Effects of dietary cholestyramine on the elimination pattern of ochratoxin A in rats. *Food Chem. Toxicol.* 30:709.
78. Manson, M. M., H. W. Ball, M. C. Barrett, H. L. Clark, D. J. Judah, G. Williamson, and G. E. Neal. 1997. Mechanism of action of dietary chemoprotective agents in rat liver: induction of phases I and II drug metabolizing enzymes and aflatoxin metabolism. *Carcinogenesis* 18:1729–1738.
79. Mayura, K., M. A. Adel Wahhab, K. S. McKenzie, A. B. Sarr, J. F. Edwards, K. Naguib, and T. D. Phillips. Prevention of maternal and developmental toxicity in rats via dietary inclusion of common aflatoxins sorbents: potential for hidden risks. *Toxicol. Sci.* 41:175–182.
80. McLeod, R., E. M. Ellis, J. R. Arthur, G. E. Neal, D. J. Judah, M. M. Manson, and J. D. Hayes. 1997. Protection conferred by selenium deficiency against aflatoxin B<sub>1</sub> in the rat associated with the hepatic expression of an aldo-keto reductase and glutathione S-transferase subunit that metabolize the mycotoxin. *Cancer Res.* 57:4257–4266.
81. Megharaj, M., I. Garthwaite, and J. H. Thiele. 1997. Total biodegradation of the oestrogenic mycotoxin zearalenone by a bacterial culture. *Lett. Appl. Microbiol.* 24:329–333.
82. Mistry, K. J., M. Krishna, and R. K. Battacharya. 1997. Modulation of aflatoxin B<sub>1</sub> activated protein kinase C by phenolic compounds. *Cancer Lett.* 121:99–104.
83. Netke, S. P., M. W. Roomi, C. Tsao, and A. Niedzwiecki. 1997. Ascorbic acid protects guinea pigs from acute aflatoxin toxicity. *Toxicol. Appl. Pharmacol.* 143:429–435.
84. Offord, E. A., K. Mace, O. Avanti, and A. M. Pfeifer. 1997. Mechanism involved in the chemoprotective effects of rosemary extract studied in human liver and bronchial cells. *Cancer Lett.* 114:275–281.
85. Okotie Eboh, G. O., L. F. Kubena, A. D. Chinnah, and C. A. Bayles. 1997. Effects of beta-carotene and canthaxanthin on aflatoxicosis in broilers. *Poult. Sci.* 76:1337–1341.
86. Piva, G., F. Galvano, A. Pietri, and A. Piva. 1995. Detoxification methods of aflatoxins: a review. *Nutr. Res.* 5:689–715.
87. Prelusky, D. B., B. A. Rotter, B. K. Thompson, and H. L. Trenholm. 1997. Effect of the appetite stimulant cyproheptadine on deoxynivalenol-induced reductions in feed consumption and weight gain in the mouse. *J. Environ. Sci. Health* 32:429–448.
88. Premalatha, B., V. Sujatha, and P. Sachdanandam. 1997. Modulating effect of *Semecarpus anacardium* Linn. Nut extract on glucose metabolizing enzymes in aflatoxin B<sub>1</sub>-induced experimental hepatocellular carcinoma. *Pharmacol. Res.* 36:187–192.
89. Raj, H. G., K. Gupta, V. Rohil, M. Bose, G. Biswas, S. K. Singh, S. C. Jain, V. S. Parmar, C. E. Olsen, and J. Wengel. 1998. Aflatoxin B<sub>1</sub>-induced micronuclei and cell cycle alterations in lung and bone marrow cells and their modulation by *Piper argyrophyllum* extract. *Teratogen Carcin. Mutat.* 18:249–261.
90. Raj, H. G., S. Gupta, G. Biswas, S. Singh, A. Singh, A. Jha, K. S. Bisht, S. K. Sharma, S. C. Jain, and V. S. Parmar. 1996. Chemoprevention of carcinogen-DNA binding: the relative role of different oxygenated substituents on 4-methylcoumarins in the inhibition of aflatoxin B<sub>1</sub>-DNA binding in vitro. *Bioorg. Med. Chem.* 4:2225–2228.
91. Ramos, A. J., J. Fink-Gremmels, and E. Hernández. 1996. Prevention of toxic effects of mycotoxins by means of nonnutritive adsorbent compounds. *J. Food Prot.* 59:631–641.
92. Rauscher, R., R. Edenharder, and K. L. Platt. 1998. In vitro anti-mutagenic and in vivo anticlastogenic effects of carotenoids and solvent extracts from fruits and vegetables rich in carotenoids. *Mutat. Res.* 413:129–142.
93. Reen, R. K., F. J. Wiebel, and J. Singh. 1997. Piperine inhibits aflatoxin B<sub>1</sub>-induced cytotoxicity and genotoxicity in V79 Chinese hamster cells genetically engineered to express rat cytochrome P4502B1. *J. Ethnopharmacol.* 58:165–173.
94. Rizzi, L., L. Lambertini, L. Marchesini, and A. Zaghini. 1995. Affinity sorbent of aluminosilicate for aflatoxin B<sub>1</sub>: effects on digestibility in growing lambs, p. 591–592. *In Proceedings of the 50th Congresso Nazionale S.I.S. Vet. Grafiche Scuderi, Messina, Italy.*
95. Rempelberg, C. J., S. J. Evertz, G. C. Buijntjes-Rozier, P. D. van den Heuvel, and H. Verhagen. 1996. Effect of eugenol on the genotoxicity of established mutagens in the liver. *Food Chem. Toxicol.* 34:33–42.
96. Shi, C. Y., S. C. Chua, H. P. Lee, and C. N. Ong. 1994. Inhibition of aflatoxin B<sub>1</sub>-DNA binding and adduct formation by selenium in rats. *Cancer Lett.* 82:203–208.
97. Shi, C. Y., Y. C. Hew, and C. N. Ong. 1995. Inhibition of aflatoxin B<sub>1</sub>-induced cell injury by selenium: an in vitro study. *Hum. Exp. Toxicol.* 14:55–60.
98. Sinha, S. P., and K. Darmshila. 1994. Vitamin A ameliorates the genotoxicity in mice of aflatoxin B<sub>1</sub>-containing *Aspergillus flavus* infested food. *Cytobios* 79:85–95.
99. Skrinjar, M., J. L. Rasic, and V. Stojicic. 1996. Lowering of ochratoxin A level in milk by yogurt bacteria and bifidobacteria. *Folia Microbiol. (Praha)* 41:26–28.
100. Smith, J. E., G. Solomons, C. Lewis, and J. G. Anderson. 1995. Role of mycotoxins in human and animal nutrition and health. *Nat. Toxins.* 3:187–192.
101. Solfrizzo, M., G. Avanti, M. R. Carratu, F. Galvano, A. Pietri, and A. Visconti. 1998. The use of biomarkers to assess the in vivo effect of activated carbon on fumonisins fed through diets contaminated with *Fusarium moniliforme*. *Rev. Med. Vet.* 149:667.
102. Soni, K. B., M. Lahiri, P. Chackradeo, S. V. Bhide, and R. Kuttan. 1997. Protective effect of food additives on aflatoxin-induced mutagenicity and hepatocarcinogenicity. *Cancer Lett.* 115:129–133.
103. Takahashi, N., U. Hartig, D. E. Williams, and G. S. Bailey. 1996. The model Ah-receptor agonist beta-naphthoflavone inhibits aflatoxin B<sub>1</sub>-DNA binding in vivo in rainbow trout at dietary levels that do not induce CYP1A enzymes. *Carcinogenesis* 17:79–87.
104. Takahashi, N., C. L. Miranda, M. C. Henderson, D. R. Buhler, D. E. Williams, and G. S. Bailey. 1995. Inhibition of in vitro aflatoxin

- B<sub>1</sub>-DNA binding in rainbow trout by CYP1A inhibitors: alpha-naphthoflavone, beta-naphthoflavone and trout CYP1A1 peptide antibody. *Comp. Biochem. Physiol. C*. 110:273–280.
105. Thalib, A. 1995. Detoxification of aflatoxin in feed with a binder of polyvinylpyrrolidone. *J Ilmiah Penelitian Ternak Klepu (Indonesia)* 1:43–48.
106. Trenholm, H. L., L. L. Charmley, K. L. Underhill, and D. B. Prelusky. 1996. Reducing toxicity of mycotoxins in feed, p. 69–86. *In Proceedings of the 4th International Feed Production Conference*. Catholic University of Sacred Heart, Piacenza, Italy.
107. Vyjayanthi, V., A. K. Kapoor, and R. B. Sashidhar. 1995. Binding characteristics of bovine serum albumin-aflatoxin B<sub>1</sub> to polystyrene microtiter plates: importance of hapten to carrier protein molar ratio. *Indian J. Exp. Biol.* 33:329–332.
108. Webster, R. P., M. D. Gawde, and R. K. Bhattacharya. 1996. Effect of different vitamin A status on carcinogen-induced DNA damage and repair enzymes in rats. *In Vivo* 10:113–118.
109. Webster, R. P., M. D. Gawde, and R. K. Bhattacharya. 1996. Modulation of carcinogen-induced DNA damage and repair enzyme activity by dietary riboflavin. *Cancer Lett.* 98:129–135.
110. Williams, G. M., and M. J. Iatropoulos. 1996. Inhibition of the hepatocarcinogenicity of aflatoxin B<sub>1</sub> in rats by low levels of the phenolic antioxidants butylated hydroxyanisole and butylated hydroxytoluene. *Cancer Lett.* 104:49–53.
111. Yu, M. W., Y. J. Zhang, W. S. Blaner, and R. M. Santella. 1994. Influence of vitamins A, C, and E and beta-carotene on aflatoxin B<sub>1</sub> binding to DNA in woodchuck hepatocytes. *Cancer* 73:596–604.
112. Zaghini, A., L. Lambertini, L. Rizzi, and P. Roncada. 1993. Effects of a bentonite supplemented diet on prevention of aflatoxicosis in growing lambs, p. 1329–1333. *In Proceedings of the 47th Congresso Nazionale S.I.S. Vet. Grafiche Scuderi, Messina, Italy.*
113. Zaghini, A., P. Roncada, P. Anfossi, and L. Rizzi. 1998. Aflatoxin B<sub>1</sub> oral administration to laying hens: effects on hepatic MFO activities and efficacy of a zeolite to prevent aflatoxicosis. *Rev. Med. Vet.* 6:668.