

Identification of Genes Upregulated in *ALK*-Positive and *EGFR/KRAS/ALK*-Negative Lung Adenocarcinomas

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Abstract

Activation of the *EGFR*, *KRAS*, and *ALK* oncogenes defines 3 different pathways of molecular pathogenesis in lung adenocarcinoma. However, many tumors lack activation of any pathway (triple-negative lung adenocarcinomas) posing a challenge for prognosis and treatment. Here, we report an extensive genome-wide expression profiling of 226 primary human stage I–II lung adenocarcinomas that elucidates molecular characteristics of tumors that harbor *ALK* mutations or that lack *EGFR*, *KRAS*, and *ALK* mutations, that is, triple-negative adenocarcinomas. One hundred and seventy-four genes were selected as being upregulated specifically in 79 lung adenocarcinomas without *EGFR* and *KRAS* mutations. Unsupervised clustering using a 174-gene signature, including *ALK* itself, classified these 2 groups of tumors into *ALK*-positive cases and 2 distinct groups of triple-negative cases (groups A and B). Notably, group A triple-negative cases had a worse prognosis for relapse and death, compared with cases with *EGFR*, *KRAS*, or *ALK* mutations or group B triple-negative cases. In *ALK*-positive tumors, 30 genes, including *ALK* and *GRIN2A*, were commonly overexpressed, whereas in group A triple-negative cases, 9 genes were commonly overexpressed, including a candidate diagnostic/therapeutic target *DEPDC1*, that were determined to be critical for predicting a worse prognosis. Our findings are important because they provide a molecular basis of *ALK*-positive lung adenocarcinomas and triple-negative lung adenocarcinomas and further stratify more or less aggressive subgroups of triple-negative lung ADC, possibly helping identify patients who may gain the most benefit from adjuvant chemotherapy after surgical resection. *Cancer Res*; 72(1):100–11. ©2011 AACR.

Introduction

Lung cancer is the leading cause of cancer death worldwide (1, 2). Adenocarcinoma, which accounts for more than 50% of non-small-cell lung cancers (NSCLC), is the most frequent type and is increasing. Lung adenocarcinoma has a heterogeneous nature in various aspects, including clinicopathologic features

(3). Recent molecular studies have revealed at least 3 major molecular pathways for the development of lung adenocarcinoma (4–8). A considerable fraction (30%–60%) of lung adenocarcinomas develops through acquisition of mutations either in the *EGFR*, *KRAS*, or *ALK* genes in a mutually exclusive manner, and the remaining lung adenocarcinomas, that is, those without *EGFR*, *KRAS*, and *ALK* mutations (herein designated "triple-negative adenocarcinomas"), develop with mutations of several other genes. *HER2*, *BRAF*, etc. are known to be mutated also mutually exclusively with the *EGFR*, *KRAS*, and *ALK* genes; however, frequencies of their mutations are very low (<5%; refs. 4–7). Therefore, genes responsible for the development of triple-negative adenocarcinomas are largely unknown.

Mutations in the *EGFR* gene are prevalent in females and never-smokers, and the frequencies are considerably higher in Asians (40%–60%) than in Europeans/Americans (~10%; refs. 5–7, 9). *EGFR* mutations make tumor cells dependent on epidermal growth factor receptor (EGFR) signaling and define patients who respond to EGFR tyrosine kinase inhibitors (TKI), such as gefitinib (10, 11). On the other hand, mutations in the *KRAS* gene occur predominantly in males and ever-smokers, and their frequencies are higher in Europeans/Americans (>15%) than in Asians (10%; ref. 9). Specific inhibitors against *KRAS* activity are being developed (12). Therefore, clinicopathologic features of lung adenocarcinomas with *EGFR* mutations (herein designated "*EGFR*-positive adenocarcinomas") and

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those with *KRAS* mutations (herein designated "*KRAS*-positive adenocarcinomas") are considerably different from each other. Recently, a small subset of *EGFR*- and *KRAS*-negative lung adenocarcinomas (~5%) was shown to have rearrangements of the *ALK* gene generating gene fusion transcripts (13), and patients with *ALK* rearrangements tend to be younger and have little or no smoking histories (4, 6–8). Because lung adenocarcinoma cells with *ALK* rearrangements (herein designated "*ALK*-positive adenocarcinomas") are specifically sensitive to ALK TKIs, *ALK*-positive adenocarcinomas have been recently considered to be another subset of adenocarcinomas by considering the differences in therapeutic targets (4, 6–8). In contrast, clinicopathologic features of triple-negative lung adenocarcinomas have not been precisely characterized because of the lack of sufficient genetic information in these adenocarcinomas.

There have been several studies which attempted to characterize gene expression profiles in particular types of lung adenocarcinoma, including *EGFR*-positive and *KRAS*-positive adenocarcinomas (14–17). However, such information is limited for *ALK*-positive adenocarcinomas and triple-negative adenocarcinomas. Therefore, in this study, we aimed to elucidate clinicopathologic features and gene expression profiles of *ALK*-positive adenocarcinomas and triple-negative adenocarcinomas in comparison with those of *EGFR*-positive adenocarcinomas and *KRAS*-positive adenocarcinomas. We conducted a genome-wide gene expression profiling of 226 lung adenocarcinomas, consisting of 127 *EGFR*-positive adenocarcinomas, 20 *KRAS*-positive adenocarcinomas, 11 *ALK*-positive adenocarcinomas, and 68 triple-negative adenocarcinomas. To identify genes useful for molecular diagnosis and applicable to targeted therapy of *ALK*-positive adenocarcinomas and triple-negative adenocarcinomas, we focused on genes that were upregulated in these adenocarcinomas by selecting genes with low expression in *EGFR*-positive and *KRAS*-positive adenocarcinomas. Several genes were identified as being specifically and significantly upregulated in *ALK*-positive adenocarcinomas. In particular, the *ALK* gene itself was highly expressed exclusively in *ALK*-positive adenocarcinomas. More importantly, a distinct group of triple-negative adenocarcinomas with unfavorable outcome was identified. This group of triple-negative adenocarcinomas showed much worse prognosis than the other group of triple-negative adenocarcinomas, *EGFR*-positive adenocarcinomas, *KRAS*-positive adenocarcinomas, and *ALK*-positive adenocarcinomas. Several genes were identified as being upregulated and critical for predicting prognosis of patients in this group of adenocarcinomas.

Materials and Methods

Patients

The tumors were pathologically classified according to the TNM classification of malignant tumors (18). A total of 226 lung adenocarcinoma cases subjected to expression profiling were selected from 393 stage I–II cases who underwent potential curative resection between 1998 and 2008 at the National Cancer Center Hospital as follows (ref. 19; Supplementary Fig. S1). Among the 393 cases, 363 cases, consisting of 305 stage I

and 58 stage II cases, were eligible by the criteria of cases who did not receive any neoadjuvant therapies before surgery and had not been diagnosed with cancer in the 5 years before lung adenocarcinoma diagnosis. All 58 stage II cases were subjected to expression profiling. The 305 stage I cases included 37 cases with relapse and 268 cases without relapse. To improve statistical efficiency, all the 37 relapsed cases and 131 matched unreleased cases selected by the incidence density sampling method (20, 21) were subjected to expression profiling. In total, 226 cases, consisting of 168 stage I and 58 stage II cases, were subjected to the expression profiling. Among the 226 cases, 204 who received complete resection (i.e., free resection margins and no involvement of mediastinal lymph nodes examined by mediastinal dissection) and did not receive postoperative chemotherapy and/or radiotherapy, unless relapsed, were subjected to survival analyses. This study was approved by the Institutional Review Boards of the National Cancer Center.

Microarray experiments and data processing

Total RNA was extracted using TRIzol reagent (Invitrogen), purified by an RNeasy kit (Qiagen), and qualified with a model 2100 Bioanalyzer (Agilent). All samples showed RNA Integrity Numbers more than 6.0 and were subjected to microarray experiments. Two micrograms of total RNA were labeled using a 5X MEGAscript T7 Kit (Ambion) and analyzed by Affymetrix U133Plus2.0 arrays. The data were processed by the MAS5 algorithm, and the mean expression level of a total of 54,675 probes was adjusted to 1,000 for each sample. Microarray data are available at National Center for Biotechnology Information Gene Expression Omnibus (GSE31210).

Probe selection for unsupervised clustering

One hundred and seventy-four genes (190 probes), preferentially expressed in *ALK*-positive and triple-negative adenocarcinomas, were selected by the following criteria; probes whose expression levels were less than 1,000 in any adenocarcinomas with *EGFR* or *KRAS* mutations, and probes whose averaged expression levels in *ALK*-positive and triple-negative adenocarcinomas were more than 1.5-fold higher than those in *EGFR*-positive and *KRAS*-positive adenocarcinomas with *P* values less than 0.05 by *t* test. Expression levels for these 190 probes were log-transformed and median-centered, both for probes and samples, and were subjected to an unsupervised hierarchical clustering. The clustering was done by the centroid linkage method using the Cluster 3.0 program, and the results were visualized using the Java Treeview program (22).

Mutation analyses

Genomic DNAs from all 226 lung adenocarcinomas were analyzed for *EGFR* and *KRAS* mutations by the high-resolution melting method as described (23, 24). Total RNAs from the 226 adenocarcinomas were examined for expression of fusion transcripts between *ALK* and *EML4* or *KIF5* using a multiplex reverse transcription PCR (RT-PCR) method (25).

Statistics

Cumulative survival was estimated by the Kaplan–Meier method, and differences in the survivals between 2 groups were

analyzed by log-rank test. Influences of variables on relapse-free survival (RFS) and overall survival (OS) were evaluated by uni- and multivariate analyses of the Cox proportional hazard model. For all analyses, smoking status was polarized as never-smokers (0 pack years) and ever-smokers (>0 pack years). Pathologic TNM staging was categorized as stage I versus stage II. For multivariate analysis, all variables were included that were moderately associated ($P < 0.1$) with RFS or OS in any of the analyses.

Bioinformatics

Associations of gene expression levels with prognosis of NSCLC patients in 7 other expression profile studies were obtained from the PrognosScan database (26). In the PrognosScan database, association of gene expression with survival of patients was evaluated by the minimum P value approach. Briefly, patients were first arranged by expression levels of a given gene. They were then divided into high- and low-expression groups at all possible cutoff points, and the risk differences of any 2 groups were estimated by the log-rank test. Finally, the cutoff point that gave the most pronounced P value was selected.

Results

***EGFR/KRAS/ALK* mutations and clinicopathologic characteristics of lung adenocarcinomas subjected to gene expression profiling**

Among 226 stages I and II lung adenocarcinomas, *EGFR* and *KRAS* mutations were mutually exclusively detected in 127 (56%) and 20 (9%) cases, respectively, and an *EML4-ALK* fusion gene was expressed in 11 (4.9%) cases (Table 1). *EGFR* or *KRAS* mutations were not detected in any of the 11 cases with *EML4-ALK* fusion expression; thus, the occurrence of *ALK* rearrange-

ments in a mutually exclusive manner with *EGFR* and *KRAS* mutations in lung adenocarcinoma was confirmed. The incidence and the fraction of *EGFR*-, *KRAS*-, and *ALK*-positive cases in this study were consistent with those in previous studies (5–7, 9, 13). Accordingly, the remaining 68 (30%) cases were defined as "triple-negative adenocarcinomas" because of the absence of *EGFR*, *KRAS*, and *ALK* mutations. Clinicopathologic features of *EGFR*-positive adenocarcinomas and *KRAS*-positive adenocarcinomas in this study are well consistent with those in previous studies of Japanese populations (27, 28). Patients with *ALK*-positive adenocarcinomas were younger and more likely to be never-smokers, as previously indicated (4, 6–8). Triple-negative adenocarcinomas showed similar clinicopathologic features to those of *KRAS*-positive adenocarcinomas, that is, a predominance of males, ever-smokers, and advanced stages.

Expression profile unique to *ALK*-positive lung adenocarcinomas

All 226 cases were subjected to genome-wide expression profiling using Affymetrix U133Plus2.0 arrays. One hundred and seventy-four genes, evaluated with 190 probes (Supplementary Table S1), were selected as those preferentially expressed in either *ALK*-positive adenocarcinomas or triple-negative adenocarcinomas under the criteria described in Materials and Methods. In particular, 10 genes evaluated with 11 probes were markedly upregulated according to the criteria of fold-differences more than 2.0 with P values less than 0.05 (Supplementary Table S2). It was noted that 2 probes for the *ALK* gene were present among them, and 1 of them (probe ID = 208212_s_at) showed the highest fold-difference of 8.7 between *ALK*-positive/triple-negative adenocarcinomas and *EGFR*-positive/*KRAS*-positive adenocarcinomas among the 190 probes. This result indicated that there is a subset of adenocarcinomas in which *ALK* was overexpressed. Therefore, an unsupervised

Table 1. Clinicopathologic characteristics of 226 lung adenocarcinomas subjected to expression profile analysis

Variable	All	Mutation				Expression profile	
		EGFR (+)	KRAS (+)	ALK (+)	Triple (–)	Group A	Group B
No. of cases	226	127	20	11	68	36	32
Age							
Mean	60	60	60	54	61	61	60
Range	30–76	35–72	46–75	30–68	46–76	46–71	47–76
Sex							
Male	105	50	12	2	41	25	16
Female	121	77	8	9	27	11	16
Smoking habit							
Never-smoker	115	67	10	7	31	10	21
Ever-smoker	111	60	10	4	37	26	11
pStage							
IA	114	77	6	3	28	10	18
IB	54	26	8	0	20	12	8
II	58	24	6	8	20	14	6

Table 2. Genes upregulated in *ALK*-positive lung adenocarcinomas

Gene symbol ^a	Gene name	Probe ID	Fold difference
<i>ALK</i>	Anaplastic lymphoma receptor tyrosine kinase	208212_s_at	55.2
<i>EST</i>	Transcribed locus	242964_at	26.8
<i>ALK</i>	Anaplastic lymphoma receptor tyrosine kinase	208211_s_at	17.2
<i>GRIN2A</i>	Glutamate receptor, ionotropic, <i>N</i> -methyl D-aspartate 2A	242286_at	13.0
<i>GRIN2A</i>	Glutamate receptor, ionotropic, <i>N</i> -methyl D-aspartate 2A	231384_at	12.4
<i>MUC5AC</i> /// <i>MUC5B</i>	Mucin 5AC, oligomeric mucus/gel-forming /// mucin 5B, oligomeric mucus/gel-forming	222268_x_at	9.2
<i>EST</i>	Transcribed locus	1570291_at	8.1
<i>LOC100292909</i>	Hypothetical protein LOC100292909	241535_at	7.7
<i>BLID</i>	BH3-like motif containing, cell death inducer	1555675_at	7.4
<i>LOC100130894</i>	Hypothetical LOC100130894	1564158_a_at	6.1
<i>CLDN10</i>	Claudin 10	1556687_a_at	6.0
<i>KRT16</i>	Keratin 16	209800_at	5.9
<i>PROM2</i>	Prominin 2	1562378_s_at	5.6
<i>GJB5</i>	Gap junction protein, beta 5, 31.1 kDa	206156_at	5.0
<i>KIAA1644</i>	KIAA1644	221901_at	4.8
<i>EPHB1</i>	EPH receptor B1	210753_s_at	4.5
<i>LRRC4</i>	Leucine rich repeat containing 4	223552_at	4.2
<i>EST</i>	Transcribed locus	235373_at	3.4
<i>tcag7.1188</i>	Hypothetical LOC340340	1561254_at	3.3
<i>SBNO2</i>	Strawberry notch homolog 2 (<i>Drosophila</i>)	204166_at	3.3
<i>EST</i>	Transcribed locus	241083_at	3.1
<i>SLC25A37</i>	Solute carrier family 25, member 37	222528_s_at	3.1
<i>NDP</i>	Norrie disease (pseudoglioma)	206022_at	3.1
<i>EST</i>	Transcribed locus	243478_at	3.0
<i>EST</i>	Transcribed locus	239136_at	2.9
<i>RHOV</i>	ras homolog gene family, member V	241990_at	2.9
<i>YIF1B</i>	Yip1 interacting factor homolog B (<i>S. cerevisiae</i>)	231211_s_at	2.9
<i>RPRM</i>	Reprimo, TP53 dependent G2 arrest mediator candidate	219370_at	2.5
<i>SYT12</i>	Synaptotagmin XII	228072_at	2.5
<i>HES2</i>	Hairy and enhancer of split 2 (<i>Drosophila</i>)	231928_at	2.4
<i>CDH11</i>	Cadherin 11, type 2, OB-cadherin (osteoblast)	239769_at	2.2
<i>IRAK3</i>	Interleukin-1 receptor-associated kinase 3	220034_at	2.1

^aGenes with fold difference >2.0 and *P* < 0.05 between *ALK*-positive and *ALK*-negative adenocarcinomas are shown.

hierarchical clustering using these 190 probes was done on 11 *ALK*-positive adenocarcinomas and 68 triple-negative adenocarcinomas (Supplementary Figs. S1 and S2). There were 3 distinct sets of genes/probes, as indicated by red, yellow, and blue bars on the left of the heat map. Two probes for the *ALK* gene were present in the gene/probe set with a yellow bar, and 11 cases with extremely high levels of *ALK* expression comprised a small subcluster in the right side of cluster 1. All the 11 cases corresponded to the ones with *EML4-ALK* fusion gene expression.

The results strongly indicated that *ALK*-positive adenocarcinomas have distinct expression profiles in comparison with *ALK*-negative adenocarcinomas, including not only triple-negative adenocarcinomas but also *EGFR*-positive and *KRAS*-positive adenocarcinomas. Therefore, genes with fold-differences more than 2.0 and *P* values less than 0.05 in their expression between *ALK*-positive adenocarcinomas and

ALK-negative adenocarcinomas were further selected from the 190 probes. Thirty genes with 32 probes were then selected (Table 2). The *ALK* gene showed the highest level of fold difference in *ALK*-positive adenocarcinomas. Therefore, as previously reported (29–31), *ALK*-positive adenocarcinomas express high levels of *ALK* gene products, supporting that upregulation of the *ALK* gene is a biological consequence of *ALK* rearrangements in lung adenocarcinoma cells. Expression profiling further revealed that various other genes are distinctly upregulated in *ALK*-positive adenocarcinomas. In particular, fold differences of *GRIN2A* (glutamate receptor, ionotropic, *N*-methyl D-aspartate 2A) expression were more than 10, as with *ALK* expression. Moreover, *GRIN2A* was branched most closely to *ALK* in the heat map (Supplementary Fig. S2). Therefore, high levels of *GRIN2A* expression can be a characteristic unique to *ALK*-positive adenocarcinomas, in addition to upregulation of the *ALK* gene itself. The levels of *GRIN2A* expression in *ALK*-

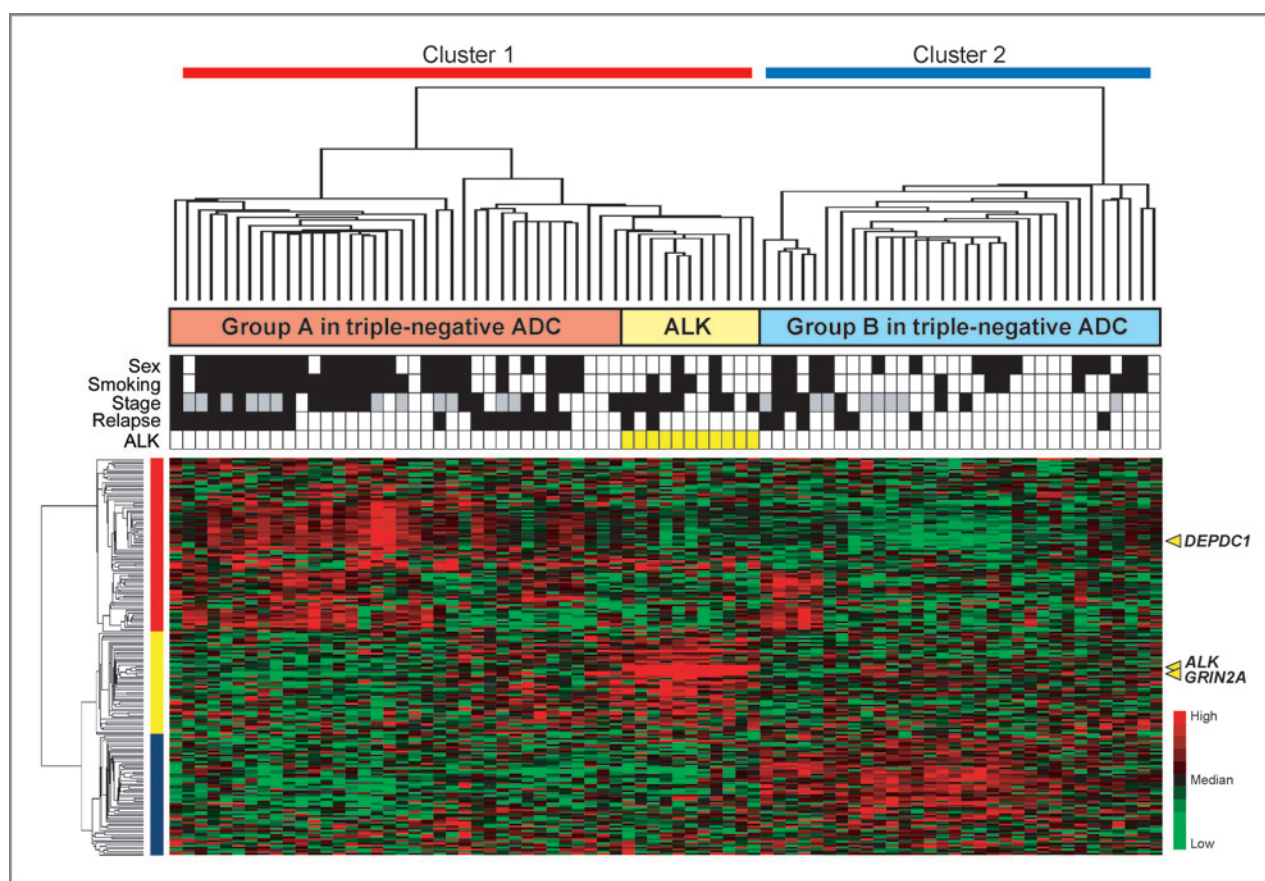


Figure 1. Unsupervised hierarchical clustering of 11 *ALK*-positive adenocarcinomas and 68 triple-negative adenocarcinomas. Triple-negative adenocarcinomas were separated into 36 group A cases and 32 group B cases, and group A cases construct cluster 1 with 11 *ALK*-positive adenocarcinoma cases. Clinical and genetic features are shown below the tree; sex (black, male; white, female); smoking status (black, ever-smoker; white, never-smoker); pathologic stage (black, stage II; gray, stage IB; white, stage IA); relapse (black, evidence of relapse; white, no evidence of relapse); *ALK* (yellow, *ALK*-fusion gene expression positive; white, negative). Three colored bars according to the main branches of probes/genes are shown on the left. Positions of probes for *ALK*, *GRIN2A*, and *DEPDC1* are shown on the right. ADC, adenocarcinoma.

positive adenocarcinomas were significantly higher than those in *ALK*-negative adenocarcinomas by quantitative RT-PCR analysis (Supplementary Fig. S3).

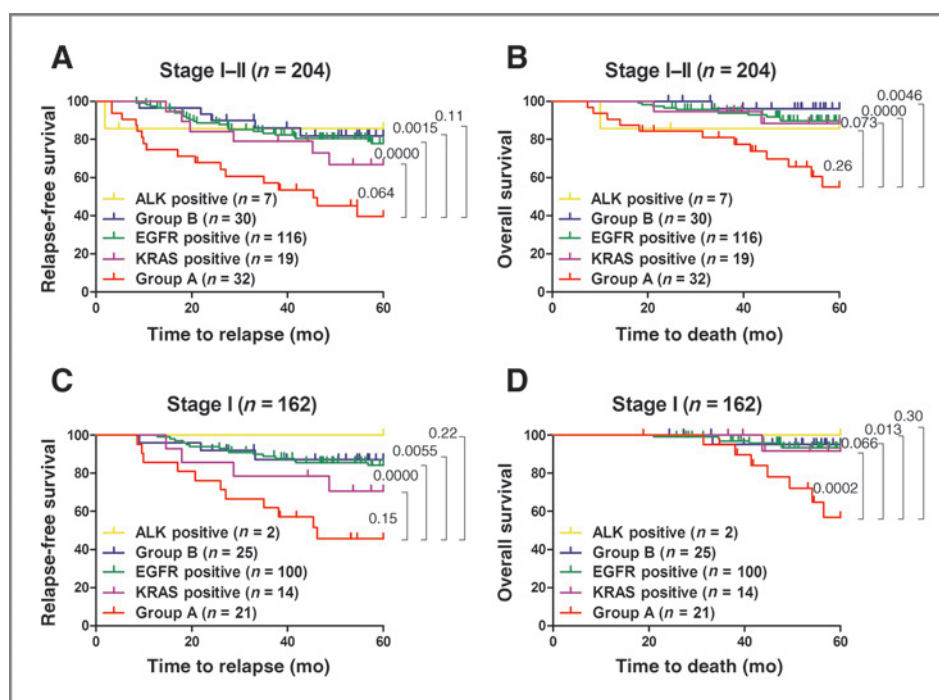
Triple-negative lung adenocarcinomas with poor prognosis identified by gene expression profiling

By the unsupervised hierarchical clustering, 68 triple-negative adenocarcinomas were separated into 2 major groups, one containing 36 cases and the other 32 cases, designated as groups A and B, respectively (Fig. 1). Group A comprised cluster 1 with 11 *ALK*-positive adenocarcinomas. Group A cases were dominant in males, ever-smokers, and advanced stages, whereas group B cases were dominant in never-smokers and early stages (Table 1), indicating that group A cases comprise an aggressive type in triple-negative adenocarcinomas. Therefore, we next compared RFS and OS among the 5 groups of patients; groups A and B, *EGFR*-positive cases, *KRAS*-positive cases, and *ALK*-positive cases (Fig. 2). Among the 226 cases, 204 cases that received complete resection and did not receive postoperative chemotherapy and/or radiotherapy were subjected to survival analysis. Group A cases ($n = 32$) showed the worst prognosis

for both RFS and OS among the 5 groups (Fig. 2A and B). In particular, group A cases showed significantly worse prognosis ($P < 0.05$) for both RFS and OS than group B cases ($n = 30$) and *EGFR*-positive cases ($n = 116$) by the log-rank test. Such differences were marginally significant between group A cases and *KRAS*-positive cases ($n = 19$) and not significant between group A cases and *ALK*-positive cases ($n = 7$), probably because the numbers of *KRAS*-positive and *ALK*-positive cases were smaller than those of group B and *EGFR*-positive cases.

Similar results were obtained from the analysis of 162 patients with stage I adenocarcinomas (Fig. 2C and D), indicating the independency of these associations with staging. Therefore, we next carried out multivariate analyses on RFS and OS of these 5 groups (Table 3). In the analysis of 204 stages I and II patients, RFS and OS of group A cases were significantly worse than those of *EGFR*-positive and group B cases, and the differences were independent of staging. HRs of *ALK*-positive and *KRAS*-positive cases were also as high as *EGFR*-positive and group B cases, although only the difference in RFS was statistically significant between group A cases and *KRAS*-positive cases. This could be also due to the small numbers

Figure 2. Kaplan–Meier survival curves for RFS and OS of 204 lung adenocarcinoma cases according to *EGFR*-positive, *KRAS*-positive, *ALK*-positive, group A, and group B. RFS and OS of stage I–II (A, B) and stage I (C, D) cases are shown.



of *KRAS*-positive and *ALK*-positive cases. Accordingly, multivariate analyses of 162 stage I patients further showed significant differences in RFS and OS between group A cases and *EGFR*-positive cases, and also between group A cases and group B cases. Because numbers of *KRAS*-positive cases and *ALK*-positive cases were small, we next compared RFS and OS between group A patients and patients in all 4 other groups combined ("Others" in Table 3). Differences in RFS as well as those in OS were highly significant and independent of staging. These results strongly indicated that group A patients comprise a distinct subclass of *EGFR*/*KRAS*/*ALK*-negative lung adenocarcinomas, and the prognoses of group A patients were the worst among the 5 groups of patients.

Clustering of lung adenocarcinomas with poor prognosis by gene expression profiling

We next carried out unsupervised hierarchical clustering of all the 226 adenocarcinoma cases, including 127 *EGFR*-positive cases and 20 *KRAS*-positive cases, to investigate whether expression profiling with a set of 174 genes with 190 probes could extract group A cases as a unique subset among all adenocarcinomas, and whether the profiling could be useful for prognosis prediction of patients with any genotypes of adenocarcinomas in general. As shown in Supplementary Fig. S4, clustering patterns of all the 226 patients were very similar to those of the 79 patients consisting of 11 *ALK*-positive cases and 68 triple-negative cases. In particular, the 11 *ALK*-positive cases comprised a small cluster in the right side of Cluster 1 (Cluster 1b), supporting that *ALK*-positive adenocarcinomas show unique expression profiles among all adenocarcinomas. Group A and group B cases also have a tendency to accumulate in Clusters 1a and Cluster 2, respectively. However, group A cases often comprise clusters with the *KRAS*-positive cases,

whereas group B cases were distributed with the *EGFR*-positive cases. Therefore, group A and group B triple-negative adenocarcinomas were not exclusive with the *EGFR*-positive and *KRAS*-positive adenocarcinomas by expression profiling of these 174 genes. Therefore, expression profiling with a set of the 174 genes was concluded to be useful to distinguish *ALK*-positive adenocarcinomas among all lung adenocarcinomas.

However, RFS of 119 patients in Cluster 1 was significantly worse than RFS of 85 patients in Cluster 2 (HR = 3.73, $P = 0.00016$). When Cluster 1 was further divided into 2 subclasses 1a and 1b of the right and left sides, respectively, Cluster 1a containing most of group A patients showed the worst prognosis among the 3 subclasses (Supplementary Fig. S4). Therefore, the expression signature of these 174 genes was indicated to be useful for prognostic prediction of adenocarcinoma patients, in particular of triple-negative adenocarcinoma patients.

Minimum set of genes characterizing triple-negative lung adenocarcinomas with poor prognosis

The above results implied that triple-negative adenocarcinomas can be classified into 2 distinct subgroups by expression profiling and prognoses of these 2 groups are significantly different from each other. Accordingly, expression of several genes among the 174 genes was expected to be independently associated with prognosis of triple-negative adenocarcinoma patients. Therefore, we next selected genes whose expression was associated with prognosis from the 174 genes evaluated by the 190 probes. To evaluate the prognostic value of each probe and to make a comparative study for association of gene expression with prognosis in other cohorts possible, we took a minimum P value approach for grouping the patients for survival analysis because of the following reason. A database

Table 3. Hazard ratios for relapse-free and overall survivals in lung adenocarcinomas

Survival	Case (n)	Variable	Univariate		Multivariate	
			HR (95% CI)	P	HR (95% CI)	P
Relapse free	Stage I-II (204)	Age	1.03 (0.99–1.07)	0.12	1.04 (0.99–1.08)	0.092
		Sex (male/female)	1.39 (0.82–2.38)	0.22	1.00 (0.49–2.05)	0.99
		Smoking habit (ever/never)	1.43 (0.84–2.44)	0.19	1.10 (0.54–2.24)	0.80
		pStage (II/I)	1.86 (1.41–2.45)	1.3E-05	3.50 (1.93–6.34)	3.6E-05
		Subgroup				
		Group A/ALK (+)	4.78 (0.63–35.99)	0.13	6.01 (0.76–47.82)	0.09
		Group A/KRAS (+)	2.43 (0.96–6.17)	0.062	2.85 (1.10–7.35)	0.031
		Group A/EGFR (+)	3.58 (1.93–6.64)	5.3E-05	2.76 (1.44–5.29)	0.0022
		Group A/Group B	4.58 (1.69–12.42)	0.0028	4.10 (1.50–11.24)	0.0061
	Group A/Others	3.56 (2.00–6.34)	1.6E-05	3.04 (1.68–5.53)	2.5E-04	
	Stage I (162)	Age	1.01 (0.96–1.06)	0.69	1.00 (0.95–1.05)	0.97
		Sex (male/female)	0.99 (0.50–1.96)	0.98	0.83 (0.33–2.07)	0.69
		Smoking habit (ever/never)	1.06 (0.54–2.08)	0.87	0.97 (0.39–2.45)	0.95
		Subgroup				
		Group A/ALK (+)	—	—	—	—
		Group A/KRAS (+)	2.31 (0.73–7.28)	0.15	2.36 (0.73–7.62)	0.15
		Group A/EGFR (+)	4.33 (2.00–9.35)	2.0E-04	4.51 (2.05–9.91)	1.7E-04
		Group A/Group B	5.36 (1.49–19.24)	0.010	5.52 (1.50–20.37)	0.010
		Group A/Others	4.18 (2.03–8.60)	1.0E-04	4.32 (2.06–9.09)	1.1E-04
Overall	Stage I-II (204)	Age	1.03 (0.98–1.08)	0.33	1.03 (0.98–1.09)	0.21
		Sex (male/female)	1.69 (0.82–3.48)	0.16	0.89 (0.33–2.41)	0.82
		Smoking habit (ever/never)	1.91 (0.92–3.97)	0.084	1.46 (0.54–3.92)	0.45
		pStage (II/I)	2.07 (1.45–2.97)	7.2E-05	3.93 (1.83–8.44)	4.6E-04
		Subgroup				
		Group A/ALK (+)	2.95 (0.38–22.78)	0.30	3.50 (0.41–29.85)	0.25
		Group A/KRAS (+)	3.12 (0.88–11.09)	0.079	3.31 (0.91–12.03)	0.069
		Group A/EGFR (+)	4.59 (2.06–10.23)	2.0E-04	3.35 (1.44–7.81)	0.005
		Group A/Group B	6.83 (1.53–30.54)	0.012	5.68 (1.24–25.95)	0.025
	Group A/Others	4.50 (2.17–9.36)	5.7E-05	3.61 (1.68–7.78)	0.0010	
	Stage I (162)	Age	0.99 (0.93–1.06)	0.73	0.98 (0.91–1.04)	0.45
		Sex (male/female)	1.15 (0.43–3.08)	0.79	0.77 (0.20–3.00)	0.70
		Smoking habit (ever/never)	1.47 (0.55–3.91)	0.45	1.26 (0.32–4.89)	0.74
		Subgroup				
		Group A/ALK (+)	—	—	—	—
		Group A/KRAS (+)	5.79 (0.71–47.3)	0.10	5.61 (0.67–46.84)	0.11
		Group A/EGFR (+)	5.83 (2.04–16.71)	0.0010	6.06 (2.08–17.71)	9.8E-04
		Group A/Group B	9.13 (1.12–74.34)	0.039	9.32 (1.10–78.61)	0.040
		Group A/Others	6.30 (2.34–16.99)	2.8E-04	6.47 (2.33–17.98)	3.4E-04

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named PrognScan was recently developed by coauthors of this study (26). In the PrognScan database, minimum *P* values for the association of gene expression with prognosis of all probes in a platform are available for a number of cohorts that have been published. Therefore, it was possible to validate the present findings using data from various other cohorts by the same criteria. According to the method described previously (26), corrected minimum *P* values were calculated for each probe to control the error rate for the evaluation of the association with RFS and OS. Expression of 11 genes evaluated with 12 probes (2 probes for the *DEPDC1* gene) showed

significant associations with both RFS and OS in 62 triple-negative adenocarcinomas and also in 46 stage I triple-negative adenocarcinomas (Table 4). Among the 11 genes, expression of 10 genes was positively correlated with poor prognosis, whereas that of the remaining 1 gene, *KIF19*, expression was negatively correlated with poor prognosis.

We first selected 174 genes as being preferentially expressed in either *ALK*-positive adenocarcinomas or triple-negative adenocarcinomas by the criteria of "probes whose expression levels in any adenocarcinomas with *EGFR* or *KRAS* mutations were lower than the mean expression

Table 4. List of genes whose expression is associated with relapse free survival and overall survival of patients with lung adenocarcinoma

Gene symbol	Probe ID (for NCC)	NCC												CAN/DF		HLM		MSK		UM		Nagoya		Duke		Seoul				
		TN, Stage I-II			TN, Stage I			All Stage I-II			All Stage I			Stage I-III		Stage I-III		Stage I-III		Stage I-III		Stage I-III		Stage I-III		Stage I-III				
		RFS n = 62	OS n = 62	P	RFS n = 46	OS n = 46	P	RFS n = 204	OS n = 204	P	RFS n = 162	OS n = 162	P	OS n = 82	OS n = 79	P	OS n = 104	OS n = 178	P	OS n = 117	OS n = 111	P	OS n = 138	OS n = 111	P	OS n = 138	OS n = 111	P		
DEPDC1	222958_s_at	0.00	2.3	0.00	3.0	0.00	3.0	0.02	2.7	0.00	2.1	0.00	1.8	0.00	2.0	0.00	2.1	—	—	0.03	1.1	—	—	0.00	1.6	0.04	1.0	0.01	0.9	
DEPDC1	235545_at	0.00	1.8	0.01	2.3	0.00	2.4	0.04	2.6	0.00	1.4	0.00	1.6	0.00	1.3	0.00	2.2	—	—	—	—	—	—	—	—	—	—	—	—	
FOSL2	218881_s_at	0.01	1.7	0.03	1.7	0.02	1.8	0.00	3.3	0.00	1.2	0.00	1.7	0.00	1.4	0.00	2.4	—	—	0.00	1.7	—	—	0.02	0.7	—	—	—	0.01	1.0
MCM4	222037_at	0.00	1.8	0.00	3.0	0.01	2.0	0.04	2.6	0.00	1.4	0.00	1.8	0.00	1.5	0.00	2.1	—	—	—	—	—	—	—	0.00	1.7	—	—	—	
UBE2S	202779_s_at	0.00	3.0	0.02	16.0	0.01	2.8	0.02	16.6	0.00	1.6	0.02	1.4	0.00	1.6	0.00	2.1	—	—	—	—	—	—	—	—	—	—	—	—	—
CD300A	217078_s_at	0.01	1.7	0.00	2.1	0.04	1.7	0.00	2.8	0.00	1.1	0.00	1.5	0.01	1.2	0.01	1.7	—	—	—	—	—	—	—	—	—	—	—	—	—
SLITRK4	232636_at	0.02	1.7	0.03	1.7	0.00	2.9	0.00	2.5	0.01	1.1	0.00	1.4	0.04	1.1	0.00	2.0	—	—	—	—	—	—	—	—	—	—	—	—	—
KRT16	209800_at	0.00	2.0	0.00	2.5	0.00	2.4	0.00	2.7	0.00	1.2	0.00	1.4	0.01	1.2	0.01	1.7	—	—	—	—	—	—	—	—	—	—	—	—	—
SIGLEC9	210569_s_at	0.00	1.9	0.01	2.0	0.00	2.1	0.04	2.1	0.00	1.6	0.00	1.4	0.00	1.7	0.00	2.2	—	—	—	—	—	—	—	—	—	—	—	—	—
D/APH3	232596_at	0.02	1.5	0.00	3.0	0.05	1.6	0.03	2.6	0.00	1.2	0.00	2.1	0.00	1.5	0.00	2.0	—	—	—	—	—	—	—	—	—	—	—	—	—
LOC152225	1562048_at	0.01	1.5	0.00	2.3	0.02	1.7	0.00	2.7	0.00	1.3	0.00	1.9	—	—	0.00	1.9	—	—	—	—	—	—	—	—	—	—	—	—	—
KIF19	1553314_a_at	0.01	-1.5	0.05	-1.6	0.00	-3.0	0.00	-2.5	—	—	0.03	-1.4	—	—	—	—	—	—	—	—	—	—	—	0.00	-1.4	—	—	—	—

Abbreviations: NCC, National Cancer Center; TN, Triple-negative; HRs (log2 ratio) with corrected P value < 0.05 are shown.

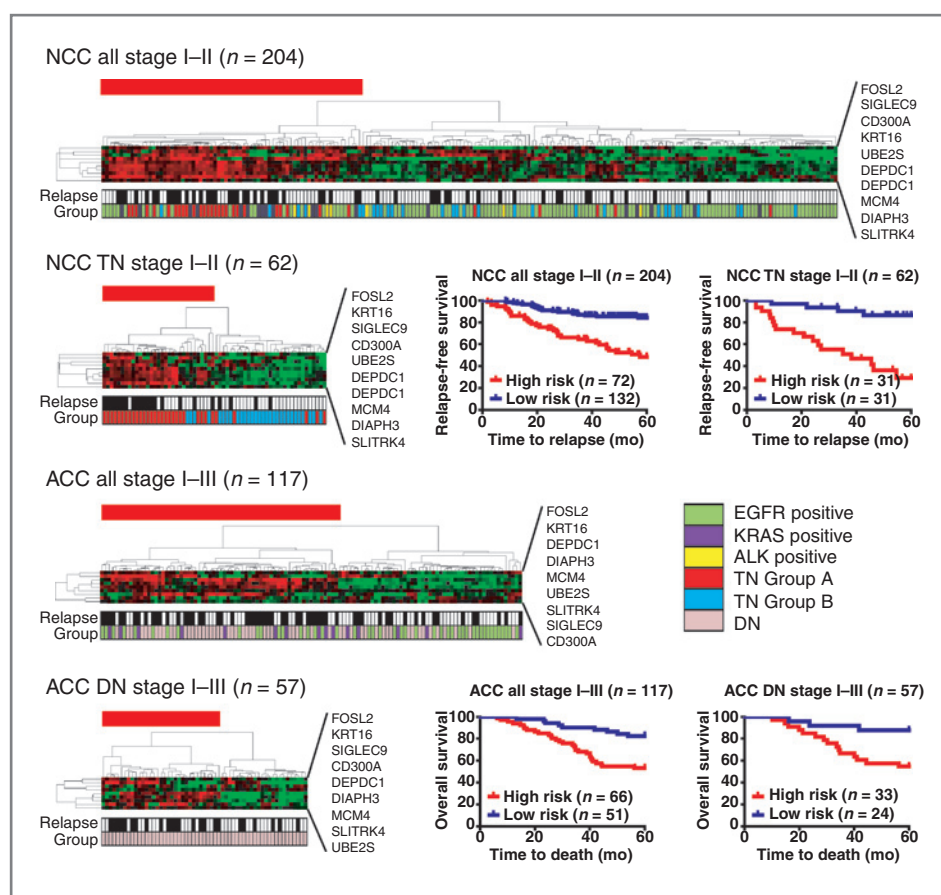


Figure 3. Unsupervised hierarchical clustering based on the expression of a set of 9 genes. All 204 stage I-II adenocarcinomas and 62 triple-negative (TN) stage I-II adenocarcinomas of the National Cancer Center (NCC) data set subjected to survival analysis were analyzed, and a cluster with higher expression of these genes than the other cluster was recognized as a high-risk group (red bar). Results of 117 adenocarcinomas, including 57 double-negative (DN) adenocarcinomas, of the Aichi Cancer Center (ACC) data set are shown below.

level of a total of 54,675 probes." Then, 11 of the 174 genes were further selected as being associated with prognosis of patients with triple-negative adenocarcinomas. Therefore, higher expression of several genes among the 11 genes was predicted to be associated with poorer prognosis, even when all adenocarcinoma cases, including *EGFR*-positive, *KRAS*-positive, and *ALK*-positive adenocarcinomas were analyzed together. Furthermore, triple-negative adenocarcinomas with poor prognosis would be separated into a high-risk group classified with this procedure. For this reason, we next analyzed all 204 adenocarcinoma cases. Among the 11 genes with 12 probes, 9 genes with 10 probes showed significant associations with both RFS and OS in all 204 adenocarcinoma cases and also in 162 stage I adenocarcinoma cases. *LOC152225* and *KIF19* were excluded because of no significant associations in stage I adenocarcinoma cases. As predicted, higher expression of the 9 genes was correlated with poorer prognosis in the analysis of RFS and OS among 204 stages I and II cases and also among 162 stage I cases.

The result strongly indicated that unsupervised hierarchical clustering using this 10 probe set (9 genes) would separate the patients into high-risk and low-risk groups for prognosis and that all group A triple-negative adenocarcinoma patients with poor prognosis would be classified into the high-risk group (Fig. 3 and Supplementary Table S3). As expected, expression profiling of these 9 genes successfully separated the 204

patients into high-risk and low-risk groups with significantly different RFS (HR = 3.79, 95% CI = 2.19–6.55, $P = 1.9E-06$) as well as OS (HR = 5.72, 95% CI = 2.53–12.87, $P = 2.5E-05$). Furthermore, if 62 triple-negative cases only were separated with these 9 genes, HRs for both RFS and OS were much higher than those with separation of all the 204 cases. All the relapsed cases in group A were separated into the high-risk group in the analyses of both cases (all the 204 cases and the 62 triple-negative cases only), supporting that triple-negative adenocarcinomas cases with poor prognosis can be selected as a high-risk group from all the adenocarcinoma cases by expression profiling of these 9 genes (Fig. 3). This profiling further separated 162 stage I cases as well as 46 stage I triple-negative adenocarcinoma cases into high-risk and low-risk groups with significantly different RFS as well as OS (Supplementary Fig. S5 and Supplementary Table S3). Again, HRs for both RFS and OS were much higher in triple-negative adenocarcinoma cases than in all adenocarcinoma cases. Accordingly, high levels of expression in these 9 genes were concluded to be distinct characteristics of triple-negative adenocarcinomas with poor prognosis.

Validation of associations using independent expression profiling data

To validate the present findings using the data of other cohorts, we searched for expression profiling data with

mutation data of the *EGFR*, *KRAS*, and *ALK* genes in various databases. However, there has been no cohort in which expression profiles specifically in triple-negative adenocarcinomas were analyzed. Therefore, unsupervised hierarchical clustering using these 9 genes was done on a cohort of 117 Japanese lung adenocarcinoma cases because expression profile data as well as *EGFR/KRAS* mutation data were available only in this cohort (32). This study included 57 adenocarcinoma cases without *EGFR* and *KRAS* mutations. Although a different array platform was used, the data for all the 9 genes were available for clustering. These cases were separated into 2 groups of 33 cases and 24 cases (Fig. 3). OS of the 33 cases was significantly shorter than that of the 24 cases (HR = 3.17, 95% CI = 1.17–8.63, $P = 2.4E-02$; Supplementary Table S3). As with our cohort, the high-risk group showed a significantly higher HR of 2.73, even when all the 117 cases were analyzed together. Although *ALK* mutation data were not available for this cohort, the results strongly supported that expression profiling of the 9 genes would be highly informative for prediction of prognosis of lung adenocarcinoma patients, in particular patients with *EGFR*- and *KRAS*-negative adenocarcinomas.

Associations of *DEPDC1* expression with prognosis of NSCLC patients

Associations of gene expression with prognosis in various cancers are available from the PrognosScan database (22). Therefore, associations of expression of these 9 genes with prognosis of NSCLC patients were examined in 7 other cohorts (Table 4). Notably, *DEPDC1* expression was positively associated with poor prognosis in 4 of the 7 cohorts; MSK, Nagoya, Duke, and Seoul. The results strongly indicated that *DEPDC1* expression can be a novel prognostic marker for patients with NSCLC. Representative data showing the association of *DEPDC1* expression with prognosis in 204 adenocarcinoma patients obtained from the minimum P value approach are shown in Supplementary Fig. S6. Associations of *DEPDC1* expression with RFS and OS were validated by quantitative RT-PCR analysis of 204 stages I and II cases and also of 162 stage I cases (Supplementary Fig. S3).

FOSL2 expression was associated with prognosis in 3 of the 7 cohorts, whereas *MCM4*, *CD300A*, and *UBE2S* expression was associated in 1 cohort, respectively (Table 4).

Discussion

In this study, we attempted to characterize *ALK*-positive adenocarcinomas and triple-negative adenocarcinomas by genome-wide expression profiling. For this purpose, we selected a set of genes that are not transcriptionally activated in any *EGFR*-positive and *KRAS*-positive adenocarcinomas, and obtained 2 pieces of unique evidence. One is that *ALK*-positive adenocarcinomas show unique expression profiles in comparison with any other types of adenocarcinomas. The other is that there is a group of patients with extremely poor prognosis among triple-negative adenocarcinomas. This group, herein designated as group A, of patients showed much worse prognoses than patients with *EGFR*, *KRAS*, or *ALK* mutations and

also than the other group, group B, of patients with triple-negative adenocarcinomas.

ALK-positive adenocarcinomas are sensitive to *ALK* TKIs with an overall response rate of 55% (8). Therefore, for the clinical application of *ALK*-targeted therapy, it is indispensable to develop a simple and reliable method for detection of *ALK* rearrangements in lung adenocarcinomas. Here, we showed that *ALK* expression is exclusively high only in *ALK*-positive adenocarcinomas and that several other genes, including *GRIN2A*, are overexpressed together with *ALK* specifically in *ALK*-positive adenocarcinomas. Therefore, *GRIN2A* can be a biomarker for detection of *ALK*-positive adenocarcinomas. *GRIN2A* encodes an *N*-methyl-D-aspartate (NMDA) receptor, which is a neurotransmitter-gated ion channel involved in regulation of synaptic function in the central nervous system (33). It was noted that the *GRIN2A* gene was recently reported to be frequently mutated in melanoma (34). Therefore, although the biological significance of *GRIN2A* upregulation in *ALK*-positive adenocarcinomas remains unclear, *GRIN2A* expression may play some important role in the phenotype unique to *ALK*-positive adenocarcinomas. Expression profiles unique to *ALK*-positive adenocarcinomas, shown here, will be also informative to improve clinical detection of *ALK* rearrangements.

Group A cases were discriminated by expression profiling of 9 genes among stage I–II cases who received complete surgical resection of tumors. Therefore, this gene set will be applicable as biomarkers to select lung adenocarcinoma patients who will benefit from adjuvant therapy after surgery, in particular to select them among patients with triple-negative adenocarcinomas. For this reason, combined analyses of this expression profiling with mutational analyses of the *EGFR*, *KRAS*, and *ALK* genes will be appropriate to pick out triple-negative adenocarcinoma patients with poor prognosis from all the adenocarcinoma patients. Molecular targeting drugs against triple-negative adenocarcinomas are not available at present; therefore, genes upregulated in group A cases will also be applicable as targets for therapy. *DEPDC1* was previously identified as being upregulated in bladder cancer and breast cancer (35–37). Because *DEPDC1* expression was hardly detectable in any normal tissues except testis, it has been considered as a cancer/testis antigen and also as a promising target of therapeutic drugs (35, 36). This study showed that *DEPDC1* is preferentially expressed in triple-negative adenocarcinomas with poor prognosis. In the PrognosScan database, *DEPDC1* expression is shown to be positively associated with poor prognosis in bladder cancer, multiple myeloma, breast cancer, glioma, and melanoma. Therefore, *DEPDC1* could be a novel target for diagnosis as well as therapy in various cancers, including lung adenocarcinoma.

Identification of genetic alterations that occur specifically in group A cases will be also of great importance for the development of target therapy for stages I and II lung adenocarcinoma patients with poor outcomes. Group A cases include males and ever-smokers as a majority (Table 1); therefore, group A cases were likely to carry several genetic alterations induced by tobacco carcinogens leading to poor outcomes. Identification of genetic alterations in

group A adenocarcinomas will further facilitate the development of targeted therapies for lung adenocarcinomas with poor prognosis.

Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed.

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References

1. Ferlay J, Shin HR, Bray F, Forman D, Mathers C, Parkin DM. Estimates of worldwide burden of cancer in 2008: GLOBOCAN 2008. *Int J Cancer* 2010;127:2893–917.
2. Parkin DM, Bray F, Ferlay J, Pisani P. Global cancer statistics, 2002. *CA Cancer J Clin* 2005;55:74–108.
3. Colvy T, Noguchi M, Henschke C, Vazquez M, Geisinger K, Yokose T, et al. Adenocarcinoma. In: Travis WD, Brambilla E, Muller-Hermelink HK, Harris CC, editors. World Health Organization classification of tumors: pathology and genetics; tumours of the lung, pleura, thymus and heart. IARC Press: Lyon, France; 2004. p. 35–44.
4. Pao W, Girard N. New driver mutations in non-small-cell lung cancer. *Lancet Oncol* 2011;12:175–80.
5. Herbst RS, Heymach JV, Lippman SM: Lung cancer. *N Engl J Med* 2008;359:1367–80.
6. Janku F, Stewart DJ, Kurzrock R. Targeted therapy in non-small-cell lung cancer—is it becoming a reality? *Nat Rev Clin Oncol* 2010;7:401–14.
7. Bronte G, Rizzo S, La Paglia L, Adamo V, Siragusa S, Ficorella C, et al. 2288; Driver mutations and differential sensitivity to targeted therapies: a new approach to the treatment of lung adenocarcinoma. *Cancer Treat Rev* 2010;36 Suppl 3:S21–9.
8. Gerber DE, Minna JD. ALK inhibition for non-small cell lung cancer: from discovery to therapy in record time. *Cancer Cell* 2010;18:548–51.
9. Sun S, Schiller JH, Gazdar AF. Lung cancer in never smokers—a different disease. *Nat Rev Cancer* 2007;7:778–90.
10. Rosell R, Moran T, Queralt C, Porta R, Cardenal F, Camps C, et al. Screening for epidermal growth factor receptor mutations in lung cancer. *N Engl J Med* 2009;361:958–67.
11. Mok TS, Wu YL, Thongprasert S, Yang CH, Chu DT, Saijo N, et al.: Gefitinib or carboplatin-paclitaxel in pulmonary adenocarcinoma. *N Engl J Med* 2009;361:947–57.
12. Konstantinopoulos PA, Karamouzis MV, Papavassiliou AG. Post-translational modifications and regulation of the RAS superfamily of GTPases as anticancer targets. *Nat Rev Drug Discov* 2007;6:541–55.
13. Soda M, Choi YL, Enomoto M, Takada S, Yamashita Y, Ishikawa S, et al. Identification of the transforming EML4-ALK fusion gene in non-small-cell lung cancer. *Nature* 2007;448:561–6.
14. Takeuchi T, Tomida S, Yatabe Y, Kosaka T, Osada H, Yanagisawa K, et al. Expression profile-defined classification of lung adenocarcinoma shows close relationship with underlying major genetic changes and clinicopathologic behaviors. *J Clin Oncol* 2006;24:1679–88.
15. Angulo B, Suarez-Gauthier A, Lopez-Rios F, Medina PP, Conde E, Tang M, et al. Expression signatures in lung cancer reveal a profile for EGFR-mutant tumours and identify selective PIK3CA overexpression by gene amplification. *J Pathol* 2008;214:347–56.
16. Motoi N, Szoke J, Riely GJ, Seshan VE, Kris MG, Rusch VW, et al. Lung adenocarcinoma: modification of the 2004 WHO mixed subtype to include the major histologic subtype suggests correlations between papillary and micropapillary adenocarcinoma subtypes, EGFR mutations and gene expression analysis. *Am J Surg Pathol* 2008;32:810–27.
17. Meyerson M, Carbone D. Genomic and proteomic profiling of lung cancers: lung cancer classification in the age of targeted therapy. *J Clin Oncol* 2005;23:3219–26.
18. Travis WD, Brambilla E, Muller-Hermelink HK, Harris CC, editors. World Health Organization classification of tumors: pathology and genetics; tumours of the lung, pleura, thymus and heart. IARC Press: Lyon, France; 2004. p. 1–344.
19. Saito M, Schetter AJ, Mollerup S, Kohno T, Skaug V, Bowman ED, et al. The association of microRNA expression with prognosis and progression in early-stage, non-small cell lung adenocarcinoma: a retrospective analysis of three cohorts. *Clin Cancer Res* 2011;17:1875–82.
20. Robins JM, Gail MH, Lubin JH. More on "Biased selection of controls for case-control analyses of cohort studies". *Biometrics* 1986;42:293–9.
21. Richardson DB. An incidence density sampling program for nested case-control analyses. *Occup Environ Med* 2004;61:e59.
22. Eisen MB, Spellman PT, Brown PO, Botstein D. Cluster analysis and display of genome-wide expression patterns. *Proc Natl Acad Sci USA* 1998;95:14863–8. Available from: <http://rana.lbl.gov/eisen/>.
23. Takano T, Ohe Y, Sakamoto H, Tsuta K, Matsuno Y, Tateishi U, et al. Epidermal growth factor receptor gene mutations and increased copy numbers predict gefitinib sensitivity in patients with recurrent non-small-cell lung cancer. *J Clin Oncol* 2005;23:6829–37.
24. Takano T, Ohe Y, Tsuta K, Fukui T, Sakamoto H, Yoshida T, et al. Epidermal growth factor receptor mutation detection using high-resolution melting analysis predicts outcomes in patients with advanced non small cell lung cancer treated with gefitinib. *Clin Cancer Res* 2007;13:5385–90.
25. Takeuchi K, Choi YL, Soda M, Inamura K, Togashi Y, Hatano S, et al. Multiplex reverse transcription-PCR screening for EML4-ALK fusion transcripts. *Clin Cancer Res* 2008;14:6618–24.
26. Mizuno H, Kitada K, Nakai K, Sarai A. 2288; PrognoScan: a new database for meta-analysis of the prognostic value of genes. 2288; *BMC Med Genomics* 2009;2:18. Available from: <http://gibk21.bio.kyutech.ac.jp/PrognoScan/index.html>.
27. Mitsudomi T. Advances in target therapy for lung cancer. *Jpn J Clin Oncol* 2010;40:101–6.
28. Suda K, Tomizawa K, Mitsudomi T. Biological and clinical significance of KRAS mutations in lung cancer: an oncogenic driver that contrasts with EGFR mutation. *Cancer Metastasis Rev* 2010;29:49–60.
29. Zhang X, Zhang S, Yang X, Yang J, Zhou Q, Yin L, et al. Fusion of EML4 and ALK is associated with development of lung adenocarcinomas lacking EGFR and KRAS mutations and is correlated with ALK expression. *Mol Cancer* 2010;9:188.
30. Boland JM, Erdogan S, Vasmatazis G, Yang P, Tillmans LS, Johnson MR, et al. Anaplastic lymphoma kinase immunoreactivity correlates with ALK gene rearrangement and transcriptional up-regulation in non-small cell lung carcinomas. *Hum Pathol* 2009;40:1152–8.
31. Mino-Kenudson M, Chirieac LR, Law K, Hornick JL, Lindeman N, Mark EJ, et al. A novel, highly sensitive antibody allows for the routine detection of ALK-rearranged lung adenocarcinomas by standard immunohistochemistry. *Clin Cancer Res* 2010;16:1561–71.

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32. Tomida S, Takeuchi T, Shimada Y, Arima C, Matsuo K, Mitsudomi T, et al. Relapse-related molecular signature in lung adenocarcinomas identifies patients with dismal prognosis. *J Clin Oncol* 2009;27:2793–9.
33. Endeley S, Rosenberger G, Geider K, Popp B, Tamer C, Stefanova I, et al. Mutations in *GRIN2A* and *GRIN2B* encoding regulatory subunits of NMDA receptors cause variable neurodevelopmental phenotypes. *Nat Genet* 2010;42:1021–6.
34. Wei X, Walia V, Lin JC, Teer JK, Prickett TD, Gartner J, et al. Exome sequencing identifies *GRIN2A* as frequently mutated in melanoma. *Nat Genet* 2011;43:442–6.
35. Kanehira M, Harada Y, Takata R, Shuin T, Miki T, Fujioka T, et al. Involvement of upregulation of DEPDC1 (DEP domain containing 1) in bladder carcinogenesis. *Oncogene* 2007;26:6448–55.
36. Harada Y, Kanehira M, Fujisawa Y, Takata R, Shuin T, Miki T, et al. Cell-permeable peptide DEPDC1-ZNF224 interferes with transcriptional repression and oncogenicity in bladder cancer cells. *Cancer Res* 2010;70:5829–39.
37. Kretschmer C, Sterner-Kock A, Siedentopf F, Schoenegg W, Schlag PM, Kimmner W. Identification of early molecular markers for breast cancer. *Mol Cancer* 2011;10:15.