Phylogenetic Analysis Informed by Geological History Supports Multiple, Sequential Invasions of the Mediterranean Basin by the Angiosperm Family Araceae

GUILHEM MANSION,1 GIDEON ROSENBAUM,2 NICOLA SCHOENENBERGER,3 GIANLUIGI BACCHETTA,4 JOSEP A. ROSELLÓ,5 AND ELENA CONTI1

1Institut für Systematische Botanik, Universität Zürich, Zollikerstrasse 107, 8008 Zürich, Switzerland; E-mail: guilhem.mansion@systbot.uzh.ch (G.M.)
2School of Physical Sciences–Earth Sciences, The University of Queensland Steele Building, St Lucia, Brisbane 4072, Australia
3Laboratoire de Botanique Évolutive, Université de Neuchâtel, Emile Argand 11, 2007 Neuchâtel, Switzerland
4Centro Conservazione Biodiversità (CCB), Dipartimento di Scienze Botaniche, Università degli Studi di Cagliari, Sant’Ignazio da Lconi, 13, 09123 Cagliari, Italy
5Jardí Botànic, Universitat de València, 46006 Valencia, Spain

Abstract.—Despite the remarkable species richness of the Mediterranean flora and its well-known geological history, few studies have investigated its temporal and spatial origins. Most importantly, the relative contribution of geological processes and long-distance dispersal to the composition of contemporary Mediterranean biotas remains largely unknown. We used phylogenetic analyses of sequences from six chloroplast DNA markers, Bayesian dating methods, and ancestral area reconstructions, in combination with paleogeographic, paleoclimatic, and ecological evidence, to elucidate the time frame and biogeographic events associated with the diversification of Araceae in the Mediterranean Basin. We focused on the origin of four species, Ambrosia bassii, Biaurum dispar, Helicodiceros muscivorus, Arum pictum, subendemic or endemic to Corsica, Sardinia, and the Betic Archipelago. The results support two main invasions of the Mediterranean Basin by the Araceae, one from an area connecting North America and Eurasia in the Late Cretaceous and one from the Anatolian microplate in western Asia during the Late Eocene, thus confirming the proposed heterogeneous origins of the Mediterranean flora. The subendemic Ambrosia bassii and Biaurum dispar likely diverged sympatrically from their widespread Mediterranean sister clades in the Early-Middle Eocene and Early-Middle Miocene, respectively. Combined evidence corroborates a relictual origin for the endemic Helicodiceros muscivorus and Arum pictum, the former apparently representing the first documented case of vicariance driven by the initial splitting of the Hercynian belt in the Early Oligocene. A recurrent theme emerging from our analyses is that land connections and interruptions, caused by repeated cycles of marine transgressions-regressions between the Tethys and Paratethys, favored geodispersalist expansion of biotic ranges from western Asia into the western Mediterranean Basin and subsequent allopatric speciation at different points in time from the Late Eocene to the Late Oligocene. [Araceae; biogeography; DIVA; endemism; fossil calibration; Mediterranean Basin; molecular dating; paleomap.]

A key idea in biogeography is that geological and climatic changes profoundly affect biotic distributions. For example, the separation of landmasses caused by tectonic events divides previously continuous species ranges, enabling the origin of vicariant taxa (Nelson and Platnick, 1981). Alternatively, geological dynamics can reduce or even eliminate physical barriers to range expansion, as when tectonic plates come into closer proximity and even contact, sometimes through the formation of temporary land corridors. Under these circumstances, species can extend their range via overland migration, in a process termed “geodispersal” by Lieberman (2000). Range expansion can also occur via long-distance dispersal (or LDD) involving movement beyond existing geographic barriers (Lomolino et al., 2006). Understanding the relative contribution of vicariance, geodispersal, and LDD to the composition of contemporary biotas is one of the most challenging goals of biogeography (Lomolino et al., 2006). If congruence between past geological connections and species origins can be demonstrated at the temporal and spatial levels, then vicariant and/or geodispersalist explanations for current distributions can be accepted. However, if congruence at either level is lacking, LDD over existing barriers has to be invoked.

Multiple lines of evidence are necessary to discern among the above-mentioned processes, including paleogeographic and paleoclimatic reconstructions, paleontological record, and estimation of the phylogenetic relationships, ancestral areas of distribution, and ages of selected groups. In this paper, we follow a multifaceted, integrative approach to infer the biogeographic history of Araceae in the Mediterranean Basin. This region represents an ideal case study to elucidate the role of earth-driven processes and LDD in shaping contemporary biogeographic patterns, because detailed reconstructions have been recently developed for its geological history, characterized by horizontal movements of microplates and formation of land connections at different points in time and space (Biju-Duval et al., 1977; Dercourt et al., 1986; Meulenkamp and Sissingh, 2003; Rosenbaum et al., 2002a, 2002b; Steininger and Rögl, 1984; see below). Furthermore, hypotheses on the geographic sources of the Mediterranean flora are available, although they were derived from qualitative assessments of similarities with other floras worldwide, rather than explicit estimations of areas and times of origin for the Mediterranean taxa or a detailed understanding of the paleogeography of the basin. The existing hypotheses posit that the tropical elements (e.g., Araceae, Oleaceae, Vitaceae) of the Mediterranean flora evolved from African, Asian, and American lineages, while its temperate elements (e.g., Boraginaceae, Platanaceae) evolved either from Irano-Turanian (i.e., Anatolian, Iranian, and Eurasian) lineages or diversified in situ (e.g., Arbutus, Lavatera; Quézel, 1978, 1985; Raven, 1973; Thompson, 2005). The current Mediterranean vegetation thus may have highly heterogeneous roots.
Climatically defined by hot, dry summers and cool, wet winters, and floristically distinguished by sclerophyllous vegetation (Daget, 1977; Quézel, 1985), the Mediterranean is home to 24,000 plant species (approximately 10% of known seed plants; Greuter, 1991), among which 13,000 are endemic (Quézel, 1978, 1985; Suc, 1984). The region, listed among the earth’s hotspots of biodiversity (Myers et al., 2000), contains 10 areas characterized by high species richness, including Corsica, Sardinia, and the Balearic Islands, with approximately 6000 plant species (Médail and Quézel, 1997), of which 10% to 12% are endemic (Bacchetta and Pontecorvo, 2005; Contandriopoulos, 1990; Gamisan and Jeanmonod, 1995; Mariotti, 1990; Médail and Quézel, 1997). Available biogeographic hypotheses propose that a majority of taxa endemic to the mentioned islands developed in situ or came from Europe and to a lesser extent North Africa, whereas a minority originated in the eastern Mediterranean (Cardona and Contandriopoulos, 1977, 1979; Contandriopoulos, 1990; Contandriopoulos and Cardona, 1984; Gamisan and Jeanmonod, 1995).

The Mediterranean Basin consists of two distinct domains. In the eastern part, the Ionian, Herodotus, and Levant basins represent remnants of the Early Mesozoic Neotethys Ocean (Garfunkel, 2004). The western Mediterranean Basin, including the Tyrrenian, Ligurian, Provençal, Algerian, and Alboran basins, started to form during the Oligocene (ca. 34 to 30 million years ago or mega annum [Ma]) in an overall convergence setting between the African and Eurasian plates (Dercourt et al., 1986; Dewey et al., 1973; Krijgsman, 2002). The formation of these basins has been attributed to extension in a back-arc region controlled by backward migration of subduction zones (Biju-Duval et al., 1977; Gueguen et al., 1998; Rosenbaum et al., 2002a). Similar processes also occurred in the Aegean Sea, which formed as a back-arc basin since the Middle Miocene (ca. 16 Ma; Lepichon and Angelier, 1979), whereas the other eastern Mediterranean basins have been subjected to subduction and collision processes (Robertson et al., 1991; Stampfli et al., 1991; Steininger and Rögl, 1984). During the Late Miocene (ca. 8 to 6 Ma), subduction processes in the westernmost Mediterranean led to the closure of marine gateways between the Atlantic Ocean and the Mediterranean Sea and triggered the desiccation of the Mediterranean Sea (ca. 6 Ma; Duggen et al., 2003; Hsü, 1972). This Messinian event was followed by the onset of the Mediterranean climatic regime during the Pliocene (ca. 3 Ma) and the alternation of warmer and colder conditions during the glacial cycles of the Pleistocene (1.8 to 0.01 Ma; Hugues et al., 2006; Suc, 1984). Overall, the complex geological history and paleoclimatology of the Mediterranean Basin likely drove biotas to fragment and merge as dispersal barriers appeared and disappeared through time.

Within the western Mediterranean, Corsica, Sardinia, the Balearic Islands (off the coast of Spain), the internal parts of the Betic Mountains of Spain and the Rif Mountains of Morocco, the Kabylies (in the Atlas mountains of Algeria), and Calabria (in the southern tip of the Italian peninsula) are all remnants of a Paleozoic mountain chain (the Hercynian belt) that was situated in Iberia and southern Europe during the Early Oligocene (Fig. 1a). Corsica, Sardinia, and the Balearic Islands are referred here as the “Hercynian Islands,” reflecting their geological origin, and their endemic species are designated as “Hercynian endemics.” According to tectonic reconstructions, the Balearic-Kabylies microplate and the Corsica-Sardinia-Calabria microplate rifted off from Iberia and southern Europe at ca. 30 to 28 Ma (Alvarez et al., 1974; Rosenbaum et al., 2002a). From around 25 Ma,
The primarily tropical Araceae (3300 species and 100 genera; Mayo et al., 1997) are represented in the Mediterranean Basin by *Arum*, *Biarum*, *Dracunculus*, *Eminentia*, *Helicodiceros* (*Areat*)*, Ambrosina* (*Ambrosineae*), and *Arisarum* (*Arisareae*), with four species either endemic or subendemic to the Hercynian Islands: *Helicodiceros muscivorus* and *Arum pictum* restricted to Corsica, Sardinia, and the Balearic archipelago, *Biarum dispar* in southern Sardinia, southeastern Spain, and northwestern Africa, and *Ambrosina bassii* in southern Corsica, Sardinia, Sicily, southern Italy, and northwestern Africa. According to available hypotheses, both *Helicodiceros muscivorus* and *Arum pictum* would represent relictual elements having possible affinities with central or eastern Mediterranean Araceae (Boyce, 1994; Contandriopoulos, 1981; Contandriopoulos and Cardona, 1984; Hruby, 1912); *Biarum dispar* shows karyological similarities with extant Spanish species (Talavera, 1976); and *Ambrosina bassii* is closely related to the Mediterranean *Arisarum* and the North American *Peltandra*, as demonstrated by recent molecular and morphological studies (Barabé et al., 2004a, 2004b). The occurrence of reliable fossils (Bogner et al., 2005; Friis et al., 2004; Smith and Stockey, 2003; Wilde et al., 2005; Zetter et al., 2001), fundamental to generate an absolute time frame for the origin of the Mediterranean endemics through molecular dating analysis, and the lack of obvious adaptations to wind dispersal make Araceae especially suitable for an integrative biogeographic study.

Using techniques of molecular dating and reconstruction of ancestral areas of distribution, it is now possible to assign specific age intervals to the phylogenetic separation of the selected Hercynian endemics from their sister taxa and infer the regions where the divergence occurred. If the estimated times and areas are congruent with known paleogeological connections of the Hercynian Islands in the Mediterranean Basin (see Table 1), it is parsimonious to conclude that earth-driven processes possibly played an important role in the origin of the studied endemics. Obviously, the availability of land corridors for biotic migration does not preclude the possibility of LDD, but the latter modality of range expansion cannot be tested directly, because no specific time frame can be proposed for it. The hypothesis of colonization of the Hercynian islands via LDD must be invoked when either the time or place estimated for the origin of the selected endemics is incompatible with geologic reconstructions. In the study presented here, we asked the following specific questions: (1) Did the Araceae colonize the Mediterranean Basin from Asia, Africa, or America (Quézel, 1978, 1985; Raven, 1973; Thompson, 2005)? (2) When and where did the species endemic to Corsica, Sardinia, and the Balearic Islands diverge from their sister clades? (3) Are the inferred temporal and spatial origins of the investigated species congruent with the reconstructed geological connections of the Hercynian Islands with Iberia until the Early to Middle Oligocene, Apulia during the Late Oligocene to Early Miocene, and North Africa and Eurasia during the Late Miocene? (4) Are our results congruent with previous hypotheses on

<table>
<thead>
<tr>
<th>Epoch</th>
<th>Time frame (Ma)</th>
<th>Geologic event</th>
</tr>
</thead>
<tbody>
<tr>
<td>Early Oligocene</td>
<td>Until 30</td>
<td>Western Mediterranean terranes are connected to the Hercynian basement in the Iberian microplate.</td>
</tr>
<tr>
<td>Early to Middle Oligocene</td>
<td>30 to 28</td>
<td>The Corsica-Sardinia-Calabria and Balearic-Kabylies microplates break off from the Iberian microplate.</td>
</tr>
<tr>
<td>Late Oligocene/Early Miocene</td>
<td>20 to 16</td>
<td>Corsica breaks off from Sardinia. The Sardinia-Calabria block is connected to Eurasia via central Apulia to the east.</td>
</tr>
<tr>
<td>Late Oligocene/Early Miocene</td>
<td>25 to 21</td>
<td>The Balearic-Kabylies microplate rotates from Iberia to Africa; the Balearic Islands split off from the Kabylies and reach their current position.</td>
</tr>
<tr>
<td>Late Miocene (Messinian)</td>
<td>5.96 to 5.36</td>
<td>Some connections exist between North Africa, Corsica, Sardinia and Eurasia, and between the Balearic Islands and Iberia during the Messinian Salinity Crisis.</td>
</tr>
<tr>
<td>Pliocene/Pleistocene</td>
<td>5 to present</td>
<td>Calabria separates from Sardinia and collides with southern Italy.</td>
</tr>
</tbody>
</table>
the origins of Hercynian endemics? (5) Can we identify repeated biogeographic patterns and processes among the selected taxa?

MATERIAL AND METHODS

Taxon and cpDNA Sampling

Sequences were generated from a total of 88 accessions, targeting the following six chloroplast DNA (cpDNA) regions: (i) the trnL intron and (ii) the trnL-trnF intergenic spacer, included in the trnALEU intron/spacer region; (iii) part of the trnK intron and (iv) the matK gene, included in the matK region; (v) the rbcL gene; and (vi) the rps16 intron. Plant samples were either collected in the field (36) or obtained from botanical gardens and herbaria (52). A list of collected accessions and species authority names is shown in online Appendix 1, available at www.systematicbiology.org.

Taxon sampling included representatives of all seven genera (Arum, Biarum, Dracunculus, Eminium, Helicodiceros, Sauroumatum, and Typhonium) ascribed to Areaceae, the single genus (Ambrosina) ascribed to the Armosinae, and the single genus (Arisarum) ascribed to the Arisareae (Boyce, 1993; Mayo et al., 1997). More intensive sampling focused on the Mediterranean genera, including 19 of the 35 species/subspecies ascribed to Arum and eight of the 26 species/subspecies ascribed to Biarum, representing all sections recognized in these two large genera (Boyce, 1993; Riedl, 1980), both species of Dracunculus, one of the seven species of Eminium, the single species of Helicodiceros, the single species of Ambrosina, and one of the three species of Arisarum (Boyce, 1990, 1994; Lobin and Boyce, 1991; Quilichini et al., 2005). When possible, the Hercynian endemics were represented by multiple accessions within their geographic range: Arum pictum (13 accessions) and Helicodiceros muscivorus (7 accessions) were sampled in Corsica, Sardinia, and the Balearic Islands; Biarum dispar (5 accessions) in Sardinia and Spain; and Ambrosina bassii (1 accession) in Sardinia (see online Appendix 1). Based on published phylogenetic analyses of Araceae (Barabé et al., 2004a; French et al., 1995; Nie et al., 2006; Renner and Zhang, 2004; Tam et al., 2004), accessions from the closely related Arisaemataceae (2), Colocasieae (5), Peltandreae (2), and Pistieae (1) were also included in the study and Calla (Calloideae), Amorphophallus (Thomsoniaceae), Caladium, and Xanthosoma (Caladieae) were used as outgroups (Fig. 2).

DNA Extraction, PCR Parameters, and Sequencing

Total DNA was extracted from silica gel–dried samples or from herbarium specimens using the DNeasy Plant Mini Kit (Qiagen, Hombrechtikon, Switzerland). Double-stranded DNA was amplified by polymerase chain reactions (PCR) performed in 20-μL volumes containing 1× buffer, 2 mM MgCl2, 200 μM dNTPs, 0.2 μM of each primer, and one unit of Taq polymerase (Sigma, Buchs, Switzerland) in a TGradient thermocycler (Biometra, Goettingen, Germany). PCR amplifications included 1 cycle of 3 to 5 min denaturation at 94°C, followed by 30 cycles of 30 s denaturation at 94°C, 1 min annealing at 52°C to 56°C, and 1 to 2 min elongation at 72°C, finishing with a 5- to 10-min elongation step at 72°C to complete primer extension. The main characteristics of the primers used for DNA amplification and sequencing are given in Table 2. The PCR products were then run on a 1% agarose gel stained with ethidium bromide to evaluate the quality (absence of contamination) and length (by comparison with a molecular weight marker) of amplicons and finally purified using the GFX PCR DNA and Gel Band purification Kit (Bioscience Amersham, Otelfingen, Switzerland). Cycle sequencing reactions for both DNA strands of each locus were performed with the BigDye Terminator Cycle Sequencing Reaction Kit (Applied Biosystems, Foster City, CA) according to the manufacturer’s instructions. Sequencing reactions were then purified on 96-well multiscreen filtration plates (Millipore, Billirica) and loaded on an ABI Prism 3100 Genetic Analyzer. New sequences were carefully checked, edited, and assembled using Sequencher 4.2 (Gene Code, Ann Arbor, MI), and submitted to GenBank (see online Appendix 1).

Phylogenetic Inference

DNA sequences were aligned manually using Se-Al (Rambaut, 1996) and combined in a matrix of 88 accessions and 4498 aligned characters (88-taxon matrix). Difficult-to-align regions (i.e., poly-AT regions ranging from 50 base pairs in Calla palustris to ca. 190 base pairs in Arum pictum), situated between positions 253 and 254 of the final alignment, were excluded from the analysis. Gaps were either treated as missing data or coded with Gapcoder (Young and Healy, 1987) and added as binary states at the end of the matrix. The final matrix was submitted to TreeBASE (study accession number SN3636).

The data set was analyzed using Bayesian inference, with a selected model of sequence evolution for each of the six partitions (i.e., trnL intron, trnL-trnF intergenic spacer, trnK intron, matK gene, rbcL gene, rps16 intron). The best respective models were chosen using the Akaike information criterion, as implemented in MrModelTest (Nylander, 2004). A binary model (Lset coding = variable) was applied to the coded gaps. Bayesian runs were performed with MrBayes 3.1 (Huelsenbeck and Ronquist, 2001), using one cold and three heated Markov chain Monte Carlo (MCMC) chains run for 5 × 106 cycles, sampling trees every 100 generations, and with a default temperature parameter value of 0.2. The MCMC runs were repeated twice, and the first 400 trees were discarded as burn-in after checking for (i) stationarity on the log-likelihood curves; (ii) similarity of the majority-rule topologies and final likelihood scores; (iii) the values of standard deviation of split frequencies (<0.001); and (iv) the value of the potential scale reduction factor (close to 1). The remaining trees were used to produce a majority-rule consensus tree and to calculate the posterior probability (pp) values.
Figure 2. MrBayes 50% majority-rule consensus tree inferred from the combined 88-taxon data set (4553 aligned positions). Posterior probabilities, ranging from 0 to 1, are reported above the branches. Tribal ranks and distribution of the Hercynian endemics (highlighted in bold) are reported to the right of the tree. Asterisks indicate the taxa selected for biogeographic and dating analyses. Area designations as in Figure 3.
For the molecular dating and biogeographic reconstruction, both requiring fully resolved trees, the initial data set of 88 accessions was reduced by pruning similar accessions and keeping representatives of all major clades recovered in the broader analysis (see Figs. 2, 3). The reduced data set, comprising 42 accessions (42-taxon data set), was analyzed using Bayesian inference with the same settings as above. A list of selected taxa with their current distribution is shown in online Appendix 2 (available at www.systematicbiology.org).

**Divergence Time Estimation**

To evaluate whether the sequences in the 42-taxon data set evolved under constant rates, a likelihood-ratio (LR) test was performed by comparing the likelihood scores of the trees resulting from Bayesian analysis with and without a clock (Felsenstein, 1981). Because rate constancy was rejected (LR = 216, df = 39, P < 0.01), we used the Bayesian approach implemented in Multidivtime (Kishino et al., 2001; Thorne and Kishino, 2002; Thorne et al., 1998) to estimate divergence times at the nodes of interest. This program assumes autocorrelation of substitution rates between ancestral and descendant nodes. Branch lengths and their variance-covariance matrix were first estimated on the 42-taxon cladogram using the BASEML and ESTBRANCHES programs in PAML version 3.14 (Thorne et al., 1998; Yang, 2000) under the F84+G model of substitution. Output branch lengths were then used as priors in MCMC searches in Multidivtime (Thorne and Kishino, 2002), to find the most likely model of rate change given the sequence data, the chosen topology, node constraints, and a Brownian motion parameter nu that determines the permitted rate change between an ancestral and a descendant node. Mean values for four prior distributions and their respective standard deviation (SD) were specified in units of 10 Ma and determined following Thorne’s protocol. (1) The prior of the age at root node (rate) was set to 0.0023 unit (SD = 0.0023), this value being calculated by dividing the median distance between the ingroup root and the tips by rttm. (3) The prior of the Brownian motion parameter nu (brownmean) was set to 0.1 (SD = 0.1) substitutions per site per million years, so that brownmean x rttm = 1. (4) Finally, prior of the age of a larger clade to which the group belongs (bigtime) was set to 11 units, an age corresponding to the oldest known fossil for the Araceae and, indeed, the entire Monocots (Friis et al., 2004). This fossil, represented by pollen grains of Mayoa sp. from the Barremian-Aptian zones of the western Portugal basin, was assigned to the Spatiphyllaeae (Friis et al., 2004), a tribe included in the Monstroideae-Pothoideae clade that diverged early in a global phylogenetic analysis of Araceae encompassing 45 genera (Barabé et al., 2004). The Markov chain was run for 100,000 cycles and sampled every 100 cycles, after an initial burn-in period of 10,000 cycles. The MultiDivTime analysis was performed at least twice with different initial conditions and the output files checked for convergence of the Markov chain.

To check for robustness of the results obtained with Bayesian dating, divergence times were also estimated by using penalized likelihood (PL) implemented in r8s (Sanderson, 2002, 2003). The PL method was applied to the same 42-taxon ML phylogram used for Bayesian dating analyses to estimate divergence times for nodes 1 through 10 (see Fig. 3). Application of the cross-validation procedure determined the optimal smoothing value to be 0.0025. This value was used to calculate nodal ages from (i) the ML phylogram and (ii) 1000 phylograms generated from bootstrap resampling in PAUP (Swofford, 2002). Nodal ages obtained from the 1000 phylograms were summarized (using the “profile” command in r8s) and the resulting standard deviations for nodal ages were used to derive 95% confidence intervals for the point estimates obtained using the ML phylogram.

In both dating analyses, the transformation of relative ages into absolute ages was performed using two fossil constraints on nodes F1 (Peltandreae crown node) and F2 (Colocasiae crown node; Fig. 3a; online Appendix 3.

**Table 2.** General information for primers used in this study. F = forward; R = reverse; A = amplification by PCR; S = sequencing. References: 1Taberlet et al., 1991; 2Sang et al., 1997; 3Winkworth et al., 2002; 4Johnson and Soltis, 1995; 5Asmussen and Chase, 2001; 6Oxelman et al., 1997.
FIGURE 3. (a) Chronogram inferred from Bayesian dating analysis of the 42-taxon data set. F1 and F2 indicate fossil constraints at 60 Ma and 50 Ma, respectively. Labeled nodes (1 to 10), with mean age estimates and 95% confidence intervals (grey bars), obtained with MULTIDIVTIME (Thorne and Kishino, 2002), are discussed in the text. Contemporary areas of distribution are reported to the right of the tree (Naf, North Africa; Ib, Iberian microplate; Co, Corsica; Sa, Sardinia; Ba, Balearic Islands; Ap, Apulian microplate; Ae, Aegean microplate; Ar, Arabian plate; An, Anatolian plate; Ir, Iranian plate; Eu, Eurasian plate; Ma, Macaronesian Islands; Taf, tropical Africa; Tas, tropical Asia; and Ame, American plate). Nodes with multiple optimal reconstructions are marked by an asterisk (1: AmeAp/AmeNaf/AmeSa/AmeCo/AmeNaTaf/AmeSaTaf/AmeCoTaf/AmeTaf/SaTaf/CoTaf/AnTaf; 2: Ap/Naf/Sa/Co; 5: AnCoSaBa/AnCo/AnSa/AnCo/AnAr/AnArSa/AnSrSa/AnSrCoSa/AnSrCoBa/AnSrCo/AmSrCoSa/AnSrCoBa/AnSrCoBa/AnSrCoBa/AnSrCo/AnSrCoSa/AnSrCoBa/AnSrCoBa/AnSrCoBa/AnSrCoBa; 8: SaAn/CoAn/BaAn; 9: Anlb/Anaeb; alternative reconstructions are separated by a slash). The ancestral areas listed at the nodes were selected based on congruence between the inferred nodal ages and paleogeological models. Circle = vicariance; diamond = duplication; + = dispersal; PP, Pliocene and Pleistocene; Ma, mega annum (million years ago). (b) Map showing the areas used for biogeographic reconstructions.
Biogeographic Analyses

We used the dispersal vicariance method (Ronquist, 1997), implemented in the software DIVA (Ronquist, 1996), to reconstruct ancestral areas of distribution. Unlike “pattern-based methods,” which rely on hierarchical area cladograms (van Veller and Brooks, 2001), DIVA reconstructs ancestral distributions for a given phylogeny without a priori assuming any specific area relationships (“event-based methods”; Ronquist, 1997). This strategy allows for the reconstruction of reticulate biogeographic scenarios in regions where past tectonic events have permitted repeated contacts between different microplates, as in the case of the Mediterranean Basin (Oberprieler, 2005; Sanmartin, 2003). When multiple area reconstructions were possible, the equivocal nodes were marked by an asterisk and the ancestral areas selected at these nodes were based on geographic contiguity with areas at adjacent nodes during the relevant geological periods (Figs. 3a, 4), thus assuming, when possible, continuous range expansion.

DIVA analyses were performed on a fully resolved Bayesian cladogram generated from the 42-taxon data set (Fig. 3a). Calla palustris, used as the dating outgroup in Multidivtime (Thorne and Kishino, 2002) analyses, was not included in the DIVA reconstructions. As recommended by Ronquist (1996, 1997), we restricted the number of areas inferred at internal nodes by using the “maxareas” option (e.g., Donoghue et al., 2003; Jonsson and Fjeldsa, 2006; Oberprieler, 2005). The maximum number of possible areas at each node was set to four units (“maxareas” = 4) because (i) all but two of the sampled species now occur in no more than four areas; (ii) the two species with a more widespread distribution (Arum maculatum, seven areas; Arisarum vulgare, 10 areas) are common in three to four areas and rare in the remaining ones (Boyce, 1993; Maire, 1957; Moggi, 1959). To check the sensitivity of the ancestral area reconstruction to the assumption of ancestral species inhabiting at most four areas, another optimization was also performed without constraints of unit areas for each ancestral node. Finally, areas where species occurrence was linked with human introduction were excluded from the analysis. The areas assigned to the taxa used in the dated phylogeny (Fig. 3b) are described in detail in online Appendix 4 (available at www.systematicbiology.org).

RESULTS

Phylogenetic Reconstructions

The larger cpDNA matrix consisted of 88 taxa and 4553 aligned positions from the following partitions: trnL intron = 444 bp; trnL-trnF intergenic spacer = 453 bp; trnK intron = 230 bp; matK gene = 1392 bp; rbcL gene = 1264 bp; rps16 intron = 712 bp; unambiguous coded gaps = 56 binary characters. The reduced cpDNA matrix, used for subsequent dating and biogeographic analyses, consisted of 42 taxa and 4508 aligned positions from the same partitions listed above. Independent Bayesian analyses of the two matrices, performed under specific models of nucleotide substitution assigned to each of the seven partitions (online Appendix 5; available at www.systematicbiology.org), yielded congruent topologies and similar pp values for the resulting 50% majority-rule consensus trees (Figs. 2, 3a).

Both the 88-taxon and the 42-taxon consensus trees (Figs. 2, 3a) strongly support (pp = 1.00) two independent origins for the Mediterranean Araceae. The Mediterranean clade M1, comprising the Hercynian subendemic Ambrosina and the widespread Mediterranean Arisarum, is sister to the geographically disjoint, tropical/temperate Peltandreae (pp = 1.00). The Mediterranean clade M2, comprising Arum, Dracunculus, Biarum, Helicodiceros, and Eminium, is sister to the clade formed by the tropical Typhonium and Sauromatum (pp = 1.00), all seven genera being included in Areaceae (Fig. 2). Within M2, the Hercynian endemic Helicodiceros forms a moderately supported clade (pp = 0.81) with the eastern Mediterranean Eminium. The Sardinian accessions of Helicodiceros (2, 3, and 5) form a well-supported clade (pp = 1.00), whereas those from Corsica (1, 4, and 6) and the Balearic Islands (7) remain unresolved. The Biarum clade is sister to the Dracunculus-Arum clad (pp = 1.00). Two Sardinian accessions (4 and 5) of the Hercynian subendemic Biarum dispar form a sister clade with one Spanish accession (1) from Murcia (pp = 0.98). The two remaining accessions from Spain (2, 3) are sister to the Sardinia-Murcia clade (pp = 1.00). Lastly, an unresolved clade comprising all accessions of the Hercynian endemic Arum pictum (pp = 1.00) from Corsica, Sardinia, and the Balearic Islands is sister to a clade formed by the remaining accessions of Arum (pp = 1.00).
8.1 Ma; Early Paleocene; online Appendix 6 [available at www.systematicbiology.org]). Within M2, the Hercynian endemics Helicodiceros muscivorius and Arum pictum diverged from their sister clades in the Early Oligocene (node 5 = 31.1 ± 6.7 Ma; Fig. 3a) and Middle Miocene (node 8 = 16.1 ± 5.8 Ma; Fig. 3a), respectively, whereas the sub endemic Biarum dispar diverged in the Early Miocene (node 10 = 18.7 ± 5.8 Ma; Fig. 3a). For all the mean nodal ages estimated by penalized likelihood (r8s; Sanderson, 2002, 2003) were all included within the confidence intervals calculated by Bayesian dating (see online Appendix 6), we will mainly refer to the Bayesian dating results in the Discussion, unless otherwise stated.

**Biogeographic Reconstructions**

The constrained DIVA (Ronquist, 1996) analysis produced 22,784 equally parsimonious reconstructions, each requiring a total cost of 56 dispersal, 13 vicariance, and 27 duplication events. In the optimization shown on the chronogram of Figure 3a, the nodes numbered 1 to 10 correspond to the origin and diversification of the Mediterranean clades M1 and M2 and to the divergence of the four Hercynian endemic or sub endemic Ambrosina bassii, Helicodiceros muscivorius, Arum pictum, and Biarum dispar from their respective sister clades. In this preferred reconstruction, compatible with the assumption that contiguous range expansion is favored when possible, duplication events were inferred for nodes 2, 4, 6, 7, 10, whereas vicariance events were inferred for nodes 1, 3, 5, 8, 9, described below (Fig. 3a).

Among the 12 possible reconstructions for the equivalent node 1, dated at 82.7 ± 6.8 Ma (Fig. 3a), the America-Apulia ancestral area was selected, based on paleomaps showing the continuity of these ancestral ranges and, thus, possible expansion of plant populations in the respective areas (see Discussion; Fig. 4a). Of the four reconstructions inferred by DIVA (Ronquist, 1996) for node 2, dated at 46.1 ± 9.0 Ma (Fig. 3a), Apulia was selected as the most likely ancestral area based on paleobotanical and ecological evidence (see Discussion; Fig. 4b). All 22 reconstructions proposed for node 5, dated at 31.1 ± 6.7, involved vicariance between Anatolia and at least one of the Hercynian Islands (Fig. 3a). Vicariance events between Anatolia and the Hercynian microplates were also inferred for node 8, dated at 16.1 ± 5.8 Ma, and node 9, dated at 21.6 ± 6.2 Ma (Fig. 3a). The unconstrained DIVA analysis (favoring total vicariance) gave identical results (not shown), except for the deepest nodes 1 (one widespread ancestral range encompassing 12 areas), 2 (one widespread ancestral range encompassing 10 areas), 4 (one widespread ancestral range encompassing 11 areas), and 5 (13 alternative solutions, also involving the Anatolian and Hercynian microplates).

**DISCUSSION**

Overall, the study presented here underlines that an integrated approach, including phylogenetic hypotheses, lineage age estimates, ancestral area reconstructions, paleogeological models, and, where available, paleontological, paleoclimatic, and ecological evidence, is indispensable to improving our understanding of the relative contribution of earth-driven processes and LDD to current biogeographic patterns. Keeping in mind that the ages estimated in this study are minimum ages with sometimes large confidence intervals, we cautiously provide in the following discussion tentative hypotheses concerning the origin of the Mediterranean clades M1 and M2 and their respective endemics.

**Origin of the Mediterranean Clade M1**

The Mediterranean clade M1, formed by Ambrosina and Arisarum, likely diverged from its temperate-subtropical sister clade, comprising the American Peltandra and the African Typhonodorum (both assigned to Peltandreae), around 82.7 Ma (node 1; Fig. 3a), in an area that connected North America and Eurasia, including the Apulian microplate, during the Late Cretaceous (Dercourt et al., 1993; Fig. 4a). The presence of Peltandreae fossils, specifically leaf fragments and seeds, in the Late Cretaceous/Early Tertiary formations of North Bohemia and Kazakhstan (Wilde et al., 2005) corroborate the choice of this ancestral area for node 1. The inferred allopatric divergence between the Peltandreae and Ambrosineae-Arisaraceae clades might have been associated with range fragmentation caused by the progressive closure of the Alpine Tethys—an ancient ocean separating the Eurasian plate and the Apulian microplate throughout the Early Cretaceous—and the subsequent Alpine orogeny (Stampfli et al., 2002). Alternative processes of geodispersal, associated with episodes of marine transgression-regression, or short distance dispersal by island hopping between the respective landmasses, cannot be excluded.

Our analyses support sympatric divergence between the Hercynian sub endemic Ambrosina bassii and Arisarum in the Middle Eocene (node 2 = 46.1 ± 9.0 Ma; Fig. 3a), or in the Early Paleocene (node 2 = 62.7 ± 8.1; r8s estimation in online Appendix 6), either in Apulia, Corsica, North Africa, or Sardinia (Fig. 3a). Ecological and paleobotanical factors are instrumental in choosing among these putative ancestral areas. Ambrosina and Arisarum, which colonize stony grounds in the under-story of the maquis, form starchy dormant tubers during the dry season and resume their vegetative growth with the onset of the rainy season (Mayo et al., 1997), a life cycle commonly viewed as an adaptation to the marked seasonality typical of the Mediterranean climate (Thompson, 2005). Considering these ecological traits, the fact that the sclerophyllous vegetation typical of the Mediterranean Basin started to develop north of the Tethys during the Early Eocene (Palamarev, 1987), and the retrieval of Oligo-Miocene fossils of sclerophyllous vegetation in Italy, our combined results suggest that the initial, sympatric diversification of clade M1 likely occurred in Apulia, rather than in Corsica, North Africa, or Sardinia (Fig. 4b).

Subsequently, the Arisarum lineage expanded throughout the Mediterranean, whereas the Ambrosina
lineage dispersed into North Africa, Sardinia, and Corsica. Migration via land from Apulia into North Africa was possible until the Middle Eocene (Figs. 4a, b), when the former was connected to the latter (Dercourt et al., 1986), whereas Corsica and Sardinia were still connected to the Hercynian basement of the Iberian microplate (Alvarez, 1972; Bellon et al., 1977). Direct expansion from Apulia into Sardinia and Corsica became possible at different times during the Miocene, according to tectonic models (Table 1; Alvarez, 1972; Alvarez et al., 1974; Rosenbaum et al., 2002a), but our current sampling does not allow us to make any specific hypothesis about the most likely migration route of *Ambrosina* towards these two islands.

**Origin of the Mediterranean Clade M2**

Combined evidence from DIVA (Ronquist, 1996) and dating analyses indicates that the stem lineage of the Mediterranean clade M2 (Fig. 3a), comprising *Eminium*, *Helicodiceros*, *Biarum*, *Dracunculus*, and *Arum* and included in Areae (Fig. 2), diverged allopatrically from a tropical, primarily Asian sister clade, formed by *Saurornatum guttatum* and *Typhonium giganteum* (the latter genus being paraphyletic; Renner and Zhang, 2004), in a region encompassing Tropical Asia and Anatolia during the Late Eocene (node 3 = 37.8 ± 7.2 Ma; Fig. 3a). The vicariant hypothesis for the origin of M2 and its estimated age seem congruent with paleoreconstructions suggesting that geological continuity between Anatolia and the current tropical Asian region was disrupted by the uplift of the Tibetan plateau following plate collision between the Indian margin and Eurasia during the Middle to the Late Eocene (44 to 36 Ma; Patriat and Achacha, 1984; Ruddiman and Kutzbach, 1991).

M2 then diversified sympatrically on the Anatolian microplate at the Eocene-Oligocene boundary (node 4 = 33.8 ± 6.9 Ma; Fig. 3a), giving origin to a clade formed by the eastern Mediterranean/western Asian *Eminium* and the Hercynian endemic *Helicodiceros muscivorus*, and a clade formed by the disjunct eastern-western Mediterranean *Biarum*, the peri-Mediterranean *Arum*, and the disjunct eastern Mediterranean-Macaronesian *Dracunculus* (Mayo et al., 1997). The repeated cycles of marine transgression-regression between Tethys and Paratethys during the Oligocene (Jolivet et al., 2006; Rögl, 1999) likely enabled speciation and range expansion by both vicariant and geodispersalist processes on terrains that were part of the current Anatolian microplate (Fig. 4c).

The east-west migratory route inferred for the origin of clade M2 has also been suggested for other Mediterranean taxa, including (i) species of the steppic flora of Spain that are related to taxa found only in the Middle East and Central Asia (Polunin and Smithies, 1973); (ii) some Fabaceae with an east-west disjunct distribution; e.g., *Anthyllis hermanniae*, occurring in Anatolia and Aegean Islands, Balkan Peninsula, South Italy, Sicily, Malta, Corsica, and Sardinia (Brullo and Giussu del Galdo, 2006), and *Genista* section *Acanthospartum* occurring in Greece, Crete, Sardinia, and Majorca (Contandriopoulos and Cardona, 1984); and (iii) several groups of animals; e.g., insects (Sanmartin, 2003), scorpions, frogs, and newts (Oosterbroek and Arntzen, 1992).

**Origin of the Hercynian endemic Helicodiceros muscivorus (L. fil.) Engler**

Ancestral area reconstructions suggest that the common ancestor of *Eminium* and *Helicodiceros* originated in the Anatolian plate (node 4; Fig. 3a), then dispersed to the western Mediterranean and split allopatrically into an eastern Mediterranean lineage and a Hercynian lineage, respectively (node 5; Fig. 3a). The vicariant hypothesis for the divergence between *Eminium* and *Helicodiceros muscivorus* and the inferred age of the event (node 5 = 31.1 ± 6.7 Ma; Fig. 3a) are compatible with geological reconstructions, suggesting that (i) during the Early Oligocene, the Anatolian microplate was alternatively connected and disconnected from the Iberian microplate via intermediate microplates in the forming Mediterranean Basin, due to cycles of marine transgression-regression between the Tethys Sea to the south and the Paratethys Sea to the north (Jolivet et al., 2006; Rögl, 1999); and (ii) Corsica, Sardinia, and the Balearic Islands were part of the Iberian microplate until they started to separate from the Hercynian basement around 30 to 28 Ma (Table 1; Alvarez, 1972; Alvarez et al., 1974; Rosenbaum et al., 2002a).

The inferred origin of *H. muscivorus* by vicariance, rather than over-water dispersal, is also congruent with the low dispersability of its fruits, which consist in orange, ovoid berries, generally containing two seeds (Mayo et al., 1997). The only available evidence on dispersal vectors concerns the lizard *Podarcis lilfordii*, endemic to the Balearic islets, which has been reported to consume the fruits of *H. muscivorus* and disperse its seeds (Pérez-Mellado et al., 2000; Valentin Perez-Mellado, University of La Rioja, Spain, March 2007, personal communication). Seed dispersal by lizards, also reported in...
other island systems for different plants, e.g., *Coprosma propinququa* (Rubiaceae; Wotton, 2002) or *Withania aristata* (Solanaeae; Valido and Nogales, 1994), can only reach small distances, apparently contradicting a hypothetical origin of *H. muscivorus* by LDD.

Previous studies suggested that approximately 3% of the Corsican endemics have an eastern Mediterranean origin (Contandriopoulos, 1981; Gamisan and Jeanmonod, 1995) and that *H. muscivorus* might represent a paleoendemic (Contandriopoulos and Cardona, 1984). Our phylogeny-based results provide the first explicit evidence supporting the ancient origin of a Hercynian endemic from the eastern Mediterranean. Available investigations on other proposed Hercynian paleoendemics (Contandriopoulos and Cardona, 1984) were unable to conclusively demonstrate their relictual nature. For example, the putative paleoendemic *Nanthea perpusilla* (Asteraceae) was found to diverge from the main eastern Mediterranean *Tripleurospermum* in the Late Miocene (ca. 8.5 Ma; Oberprieler, 2005), thus not conforming to the Early Oligocene origin that would be congruent with the geological splitting of the Hercynian basement. A vicariant origin was also proposed to explain the current disjunct distribution of *Cyclamen balearicum* (Myrsinaceae) in the Balearic Islands and southern France, although no time frame or biogeographic reconstructions were generated to support the claim (Gielly et al., 2001). Similarly, no explicit temporal and biogeographic analyses have been generated to test the paleoendemic hypothesis for the origin of *Helichrysum frigidum* (Asteraceae; Galbany-Casals et al., 2004), *Hypericum balearicum* (Hypericaceae; Crockett et al., 2004), and *Naufraga barariaca* (Apiaceae; Fridlender and Boisselier-Dubayle, 2000). In animals, the split of the Corso-Sardinian block from the Hercynian basement of the Iberian microplate was assumed, rather than tested, to estimate rates of substitution in mitochondrial DNA and infer biogeographic scenarios in newts (Caccione et al., 1994, 1997), snails (Ketmaier et al., 2003), lizards (Oliverio et al., 2000), and insects (Caccione and Sordoni, 2001; Fochetti et al., 2004).

In summary, the Early Oligocene origin of *H. muscivorus* from a widespread eastern-western Mediterranean ancestor (Fig. 3a), the low levels of infraspecific morphological (Fridlender, 2000; Rosselló and Saez, 1997) and genetic variation (ranging from zero to one nucleotide differences among the eight accessions from the three island systems; Fig. 2), and its taxonomic isolation in a monospecific genus (Boyce, 1994) are congruent with the criteria commonly used to define paleoendemics, which emphasize their ancient age; the absence of their closest relatives from the same or adjacent geographical areas; their relictual nature, as the only survivors of a once more widely distributed ancestor; the lack of morphological and genetic variation, interpreted as the product of a long process of adaptation in isolation to the conditions of the refugial area; and their taxonomic/phylogenetic isolation, due to the extinction of the closest relatives (Bramwell, 1972; Favarger and Contandriopoulos, 1961; Greuter, 1972; Lomolino et al., 2006; Major, 1988).

**Origin of the Hercynian Endemic Arum pictum L. fil.**

DIVA (Ronquist, 1996) and molecular dating estimates suggest that the common ancestor of *Biurum*, *Dracunculus*, and *Arum* originated in Anatolia during the Early Oligocene (node $d = 30.2 \pm 6.7$ Ma; Fig. 3a), subsequently generating the *Arum-Dracunculus* lineage on the same microplate (node $d = 27.3 \pm 6.5$ Ma; Fig. 3a). As already remarked above for M2 and *Helicodiceros* (nodes 3 and 5; Fig. 3a), in the Early Oligocene (Jolivet et al., 2006; Rögl, 1999) extensive land connections allowed for biotic movement from Anatolia into Iberia during episodes of marine regression between the Tethys and Paratethys Seas, whereas oceanic barriers favored allopatric speciation between eastern and western Mediterranean microplates during episodes of marine transgression (Fig. 4c).

Later, the *Arum* lineage expanded its range into the western Mediterranean, until the ancestor of the Hercynian endemic *Arum pictum* diverged allopatrically from the remaining species during the Miocene (node $d = 16.1 \pm 5.8$ Ma; Fig. 3a). Migration of *A. pictum* from Anatolia into Sardinia (node 8; Fig. 3a) would have been possible between the Early and Middle Miocene via land connection with Eurasia through the Apulia microplate (Fig. 4d), with final separation of Sardinia, still attached to the Calabria block, around 16 Ma (Table 1; Alvarez, 1972; Alvarez et al., 1974; Rosenbaum et al., 2002a). However, the geological connections between Sardinia and neighboring areas are still debated (e.g., Meulen Kamp and Sissingh, 2003; Roca, 2002), and the allopatric origin of *A. pictum* might have also been achieved via LDD.

The greater or lesser ability of fruits and seeds to disperse over long distances provides another important line of evidence to discern between over-land migration and dispersal over existing barriers (Bohning-Gaese et al., 2006). The red, fleshy fruits of *A. pictum*, about 4 to 5 mm long and containing one to two spherical, reticulate, brown seeds (Boyce, 1988), are generally transported by short-distance dispersers such as ants (Boyce, 1988), snakes (Gianni Bacchetta, University of Cagliari, Italy, March 2007, personal observation) and lizards, which enhance seed germination by ingestion (Valentin Perez-Mellado, University of La Rioja, Spain, March 2007, personal communication). Furthermore, the demonstrated toxicity for birds of the calcium oxalate crystals abundant in the fruits and leaves of most Araceae (Arai et al., 1992), including *A. pictum* (Boyce, 1993), does not favor LDD by such vectors. Therefore, the low dispersibility of the propagules seems to be more congruent with a geodispersalist hypothesis for the diversification of *A. pictum* than with LDD by birds. In conclusion, as seen for *Helicodiceros muscivorus*, our phylogeny-based evidence appears consistent with published scenarios on the eastern Mediterranean origin of some Hercynian endemics (Cardona
and Contandripoulos, 1979; Contandriopoulos, 1981; Gamisan and Jeanmonod, 1995).

Origin of the Sardinian Subendemic Biarum dispar (Schott) Tavolera

Molecular dating estimates and reconstruction of ancestral areas suggest that the ancestor of Biarum diverged from its sister clade during the Early Oligocene in Anatolia (node 6 = 30.2 ± 6.7 Ma; Fig. 3a), from where it dispersed into Iberia. In the Early Miocene, the Biarum lineage split allopatrically into an eastern Mediterranean lineage, represented by B. davisii, and a widespread Mediterranean clade that includes the subendemic B. dispar (node 9 = 21.6 ± 6.2 Ma; Fig. 3a), which diverged sympatrically from its sister clade on the Iberian peninsula (node 10 = 18.7 ± 5.8 Ma; Fig. 3a), expanding into Sardinia later in the Miocene.

As seen above, terrestrial connections separating the Tethys and Paratethys Seas during the Miocene (Fig. 4d; Jolivet et al., 2006; Rögl, 1999) might have allowed for biotic exchanges between eastern and western Mediterranean. Furthermore, the formation of water barriers during periods of marine transgression might have enabled allopatric speciation. The east-west pattern of migration inferred for the expansion of Biarum is congruent with trans-Tethys biotic exchanges proposed for several groups of animals (Oosterbroek and Arntzen, 1992) and plants, including the Hercynian endemic Nananthea (Asteraceae; Oberprieler, 2005).

The ecological characteristics of Biarum seem compatible with a history of sympatric speciation and range expansion when geological connections were available, and allopatric speciation when geological connections were interrupted. Indeed, most extant Biarum species are geocarpic, i.e., bear their fruits below the ground (Mayo et al., 1997), and are generally dispersed by ants (Boyce, 1993), thus allowing for delayed seed dispersal, partitioned germination through time, and survival in inhospitable habitats (Ellner and Shmida, 1981).

One limit of our investigation is that the method used for ancestral area reconstruction (DIVA; Ronquist, 1996) does not incorporate any knowledge of past and present connections among the areas of distribution defined at the tips of the tree, thus leading to many alternative optimizations at the nodes that are not compatible with geological models. Selection of the most likely ancestral area, therefore, must be performed a posteriori, by comparing the equally optimal solutions for ancestral areas (Fig. 3a) with available geological reconstructions (Table 1; Fig. 4), introducing a subjective element in the inference of integrative biogeographic scenarios. Furthermore, by favoring a priori vicariance and range expansion against dispersal, DIVA assumes continuity in all ancestral ranges, overall making ancestral reconstruction particularly equivocal. A promising analytical development incorporates geological models into the reconstruction of ancestral areas a priori, thus helping to achieve a sounder basis for the estimation of the relative roles of long-distance and earth-driven processes (Ree et al., 2005). Ideally, further progress in this direction will allow for the widespread application of this novel approach to cases with very complex geological histories, including the Mediterranean Basin.

Despite the above-mentioned caveats and the breadth of the inferred time intervals for the divergence of the selected taxa, the phylogenetic results on Araceae allowed us to make at least some progress towards two goals: (i) the assessment of congruence between the estimated ages and geographic sources of the Mediterranean endemics with independently derived paleogeological models; (ii) the evaluation of previous, intuitive hypotheses about the origin of the Mediterranean flora in general and the Hercynian endemics in particular. Increased precision of the estimated time frames, coupled with improved methods of ancestral area inference (Ree et al., 2005) and the explicit incorporation of paleoclimatic models and ecological parameters (Jakob et al., 2007; Wiens and Donoghue, 2004; Yesson and Culham, 2006), will be necessary to further refine hypotheses about the most likely biogeographic history of the examined Mediterranean endemics.

Conclusions

Our results from phylogenetic, molecular dating, and ancestral area analyses, combined with geological, paleobotanical, paleoclimatic, and ecological evidence, support two main invasions of the Mediterranean region by the Araceae: one from an area that connected North America and Eurasia during the Late Cretaceous (clade M1) and one from the Anatolian plate in western Asia during the Late Eocene (clade M2; Figs. 3, 4). The inferred invasions are congruent with the diverse biogeographic origins postulated for the heterogeneous Mediterranean flora (Quézel, 1985). The stem lineage of the M1 clade might have been part of an ancestral subtropical flora likely widespread in the Tethyan region before the progressive opening of the North Atlantic from the Late Cretaceous to the Early Eocene (Quézel, 1985; Raven, 1973; Scotese, 2004). On the other hand, the stem lineage of M2 appears to have been an element of the tropical Irano-Turanian flora that presumably invaded the Mediterranean from the East multiple times in the Tertiary (Quézel, 1985), mainly during episodes associated with the geological evolution of the Tethys and Paratethys Seas. Indeed, a recurrent theme that emerges from our analyses is that land connections and interruptions caused by repeated cycles of marine transgression-regression between the Tethys and Paratethys (Meulenkamp and Sissingh, 2003; Oosterbroek and Arntzen, 1992; Popov et al., 2006; Quézel, 1985; Rögl, 1999; Sanmartin, 2003; Stampfl et al., 1991; Steininger and Rögl, 1984) favored geodispersalist expansion of biotic ranges from western Asia into the western Mediterranean Basin and subsequent allopatric speciation at different points in time, thus providing a geological framework for the origin of M2, Helicodiceros muscivorus, Arum pictum, and Biarum dispar from the Late Eocene to the Late Oligocene (Figs. 3, 4c, d).
It has been hypothesized that islands formed through fragmentation of continental plates (fragment islands: Gillespie and Roderick, 2002; continental islands: Lo molino et al., 2006) should be characterized by a high proportion of relictual, paleoendemic elements from the original continental biota, while oceanic islands of volcanic origin (Darwinian islands: Gillespie and Roderick, 2002) would include a large number of neo endemics derived from colonization of empty ecological niches. The integrative evidence summarized above supports a relictual origin for the Hercynian endemics Helicodiceros muscivorus and Arum pictum, a conclusion that is congruent with the suggested high representation of paleoendemics on islands that were connected with continental plates for at least part of their geological history, as in the case of Corsica, Sardinia, and the Balearic Islands (Table 1).

ACKNOWLEDGMENTS

The authors thank J. Sullivan, M. Fishbein, and S. Renner for constructive comments and reviews; F. Küpfer, D. Schererich, M. Bigazzi, F. Selvi, P. Enz, and J. A. Mejias for tissue samples; and L. Zeltner for assistance in the field; M. Chase, E. Kapinos, and L. Csiba for DNA samples; and the following herbaria, B, EDI, G, HUB, K, MA, for photographs or loans. We are also indebted to L. Hugot (Botanical Conservatory of Corsica, France) for permission to collect plants endemic to Corsica; A. Guggisberg for help and advice; I. Sanmartin for aid in the fieldwork, and the Equal Opportunity Office of the University of Zurich for consumables.

REFERENCES


First submitted 29 May 2007; reviews returned 14 September 2007; final acceptance 20 December 2007
Associate Editor: Mark Fishbein