Diet-induced change in fatty acid composition of plasma triacylglycerols is not associated with change in glucagon-like peptide 1 or insulin sensitivity in people with type 2 diabetes¹-³

Audrey E Brynes, C Mark Edwards, Arvind Jadhav, Mohammed A Ghatei, Stephen R Bloom, and Gary S Frost

ABSTRACT

Background: Polyunsaturated fatty acids (PUFAs) and monounsaturated fatty acids (MUFAs) have been shown to positively affect blood lipids; however, their comparative effects on insulin sensitivity are unclear.

Objective: Our objective was to investigate whether chronic intake of MUFAs or PUFAs improves insulin sensitivity in people with type 2 diabetes via stimulation of the endogenous gut hormone glucagon-like peptide 1 [7–36] amide (GLP-1).

Design: Nine overweight people with type 2 diabetes received isoenergetic high-MUFA (20.3 ± 3.5% of total energy) or high-PUFA (13.4 ± 1.3%) diets for 24 d in a randomized, double-blind crossover design.

Results: Weight and glycemic control remained stable throughout the study. Despite a significant change in the plasma triacylglycerol linoleic-oleic acid ratio (L:O) with both diets (MUFA: from 0.46 ± 0.03 to 0.29 ± 0.02, P < 0.005; PUFA: from 0.36 ± 0.04 to 0.56 ± 0.05, P < 0.05) and the phospholipid L:O (1.7 ± 0.1 to 2.0 ± 0.3; P = 0.04) during consumption of the PUFA diet, this change was not associated with a change in insulin sensitivity, measured by the short-insulin-tolerance test. There was a significant reduction in the ratio of total to HDL cholesterol during consumption of the PUFA diet (5.2 ± 0.4 compared with 4.7 ± 0.3; P = 0.005) but no change with the MUFA diet. There was no change in the fasting or postprandial incremental area under the curve in response to an identical standard test meal for glucose, insulin, triacylglycerol, nonesterified fatty acids, or GLP-1.

Conclusions: Over the 3-wk intervention period, diet-induced change in the triacylglycerol or phospholipid L:O was not associated with either increased stimulation of GLP-1 or a change in insulin sensitivity in people with type 2 diabetes. Am J Clin Nutr 2000;72:1111–8.

KEY WORDS MUFA, PUFA, insulin sensitivity, GLP-1, type 2 diabetes, monounsaturated fatty acids, polyunsaturated fatty acids, glucagon-like peptide 1 [7–36] amide, triacylglycerol, cholesterol

INTRODUCTION

Diet is the mainstay therapy in people with type 2 diabetes, yet the ideal dietary guidelines for people with diabetes remain controversial. Recommendations aim to promote nutritional factors that have been shown to improve outcome, such as good glycemic control (1) and maintaining ideal body weight (2), while reducing the risk of coronary heart disease through improved lipid profiles (3). UK recommendations (4) suggest that carbohydrate should make up 50–55% of daily energy intake while fat is reduced to 30–35% of energy. Of this, 10% should be saturated fat, 10% polyunsaturated fatty acids (PUFAs), and 10–15% monounsaturated fatty acids (MUFAs). However, there is concern that an intake of 50–55% of energy as carbohydrate may have an adverse effect on triacylglycerols and glycemic control (5); indeed, US American Diabetes Association guidelines (6) now recommend that 60–70% of energy be divided between carbohydrate and MUFAs, depending on patient preference and nutritional goals.

In a recent review by Garg (7), high-MUFA diets compared with high-carbohydrate diets reduced fasting triacylglycerol and VLDL cholesterol by 19% and 22%, respectively, with a modest increase in HDL and no adverse effect on LDL. It was also shown with use of the euglycemic clamp method that increasing MUFAs and decreasing carbohydrate intake improves insulin sensitivity and glycemic control while having no adverse effects on lipids (8, 9).

The lipid-lowering effects of MUFAs compared with those of PUFAs are well studied, suggesting that PUFAs may be more potent at lowering plasma LDL-cholesterol and triacylglycerol concentrations (10). However, information about the effect of specific fatty acids, especially n–6 PUFAs compared with MUFAs, on insulin sensitivity is scarce (11, 12). Many studies compare MUFAs with carbohydrate rather than with other fatty acids. The mechanism for the reported benefits of MUFAs on insulin sensitivity is unknown. New evidence suggests that gut hormones, which can be manipulated by nutrient intake, may be involved in the regulation of insulin sensitivity and insulin secretion (13).

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Glucagon-like peptide 1 [7–36] amide (GLP-1) is the most potent known endogenous gut hormone stimulant of insulin secretion in humans. In addition to its insulin-stimulating effects, GLP-1 suppresses glucagon secretion, delays gastric emptying, and is postulated to increase peripheral insulin sensitivity (14–16). It is thus being investigated as a possible treatment for type 2 diabetes (17). A preprandial subcutaneous dose of 25 nmol GLP-1 is postulated to increase peripheral insulin sensitivity (14–16). It is not known whether increasing endogenous plasma GLP-1, via dietary manipulation, could improve glycemic control and insulin sensitivity.

Clinical and metabolic measurements

Short-insulin-tolerance test

Volunteers were tested in the early morning after an overnight fast. Two intravenous cannulas were inserted: one in one arm for the sampling of arterialized blood, with the hand placed in a warm box at 55°C, and the other in the other arm to give insulin. Insulin (0.05 U Human Actrapid insulin/kg; Novo Nordisk Pharmaceuticals, Copenhagen, Denmark) was given intravenously at 0 min. Blood samples (2 mL) were taken at 15, 0, 3, 4, 6, 8, 10, 12, 14, and 15 min for the measurement of glucose. The test was terminated at 15 min with oral glucose (25 g). Subjects were then allowed to eat a light breakfast. This procedure was done as described previously by Bonora et al (27) and modified by Gelding et al (28).

The SITT is a simple, safe, accurate, and reproducible test (28). The test measures blood glucose decline over a 15-min period in response to a bolus of insulin and before the onset of counter-regulatory hormones. The SITT was adapted from the insulin tolerance test (29), which involves the measurement of blood glucose concentration over a 15-min period in response to a bolus of insulin. The SITT is thus being investigated as a possible treatment for type 2 diabetes (17). A preprandial subcutaneous dose of 25 nmol GLP-1 is thus being investigated as a possible treatment for type 2 diabetes (17).

We aimed to recruit obese subjects with type 2 diabetes treated by diet alone and who were therefore insulin resistant with residual pancreatic beta cell function. Fifteen people with type 2 diabetes were approached to take part in this study. After initial screening, 9 (5 male and 4 female) overweight people with type 2 diabetes were recruited from the Hammersmith Hospital diabetes clinic. All subjects fulfilled the World Health Organization criteria for the diagnosis of diabetes (23). They were not taking any medication or dietary supplements and had normal results on an electrocardiogram and physical examination. The mean length of time since diagnosis of diabetes was 3.0 ± 2.1 years. The mean (±SD) age was 56 ± 5.3 years, body mass index (BMI; kg/m²) was 29.8 ± 2.8, and waist-to-hip ratio was 0.97 ± 0.05 for the men and 0.93 ± 0.01 for the women. The subjects were all nonsmokers and had stable weights. Previous dietary intake was per British Diabetic Association (BDA) guidelines (4). Subjects’ habitual baseline diets are described in Table 1; the subjects’ macronutrient intakes compared favorably with the BDA guidelines. Alcohol intake and physical activity level did not differ significantly between subjects or between the 2 intervention periods. The Imperial College School of Medicine, Hammersmith Campus Research and Ethics Committee granted ethical approval. All volunteers gave written consent.

Experimental protocol

A double-blind, randomized crossover study design was undertaken (Figure 1). After a 1-wk run-in period during which subjects recorded their normal habitual diet for 7 d, participants were randomly assigned to one of two 3-wk treatments: a diet rich in MUFA or a diet rich in PUFAs. There was a minimum 2-wk washout period between the 2 diets (± 2 ± 6 d). Subjects then started the other intervention. Each dietary period included a short-insulin-tolerance test (SITT) on days 1 and 23 and a standard test meal (STM) on days 2 and 24 of each intervention. The SITT was used to measure change in insulin sensitivity. Postprandial GLP-1, insulin, glucose, and lipids were measured after STMs eaten before and after subjects had consumed the MUFA or PUFA diets. Consistent exercise and daily routines were encouraged throughout the study. No alcohol or exercise was allowed on the days before the SITT or STM. On the basis of results from previous studies, alterations in glycemia, insulinemia, and lipidemia were assumed to occur within the first 3 wk after crossover (24–26).

### Table 1

<table>
<thead>
<tr>
<th>Baseline diet</th>
<th>MUFA diet</th>
<th>PUFA diet</th>
</tr>
</thead>
<tbody>
<tr>
<td>Energy (MJ/d)</td>
<td>8.93 ± 3.58</td>
<td>9.81 ± 2.89</td>
</tr>
<tr>
<td>Polyunsaturated fatty acids (%)</td>
<td>5.2 ± 3.3</td>
<td>4.2 ± 1.1</td>
</tr>
<tr>
<td>Unknown fatty acids (%)</td>
<td>9.7 ± 3.6</td>
<td>6.6 ± 3.0</td>
</tr>
</tbody>
</table>

1 Significant difference from baseline diet: 1 P < 0.05, 2 P < 0.005.
2 Statistical analyses performed on log-transformed data.
3 Significant difference from MUFA diet, P = 0.001.

Subjects and methods

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### Clinical and Metabolic Measurements

#### Short-Insulin-Tolerance Test

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The SITT is a simple, safe, accurate, and reproducible test (28). The test measures blood glucose decline over a 15-min period in response to a bolus of insulin and before the onset of counter-regulatory hormones. The SITT was adapted from the insulin tolerance test (29), which involves the measurement of blood glucose concentration over a 15-min period in response to a bolus of insulin providing 0.1 U/kg body wt. Because this dose increased the risk of hypoglycemia, we instead used 0.05 U/kg, as used in other studies with insulin-resistant subjects (30). The results of the SITT correlate well with those of the euglycemic-hyperinsulinemic clamp method (r = 0.81, P < 0.005) (28).

### Standard Test Meals

Volunteers were tested in the early morning after an overnight fast. An intravenous cannula was inserted into the
antecubetal fossa for blood sampling. Blood samples were taken at −15, 0, 5, 15, 30, 45, 60, 90, 120, 150, and 180 min. The meal was consumed within 10 min on each occasion. A supplement drink (Liquid Ensure Plus; Abbott Laboratories Ltd, Kent, United Kingdom) was used for the STM. This was nutritionally complete (400 mL contains 20 g fat, of which the main fatty acids were 16:0, 6%; 18:0, 3%; 18:1, 59%; 18:2, 25%; and 18:3, 5%; 80 g carbohydrate; 25 g protein; and 2.5 MJ), was palatable, and produced reproducible data. We showed previously that a liquid meal stimulates a significantly greater amount of GLP-1 than does an identical solid one (21). We used identical test meals to assess the long-term background effects of ingesting MUFAs compared with PUFAs, and measured the plasma fatty acid profiles of subjects at the beginning and end of each intervention to show that the interventions had induced a dietary change in background plasma and phospholipid triacylglycerol profiles.

**Dietary interventions**

The aim of the dietary interventions was to test the effect of a chronic increase in MUFAs (mainly oleic acid, 18:1n−9) or PUFAs (mainly linoleic acid, 18:2n−6) in the diet by substituting them for high–glycemic index carbohydrates (ie, bread and potatoes) while keeping the total energy and protein intakes constant. To aid compliance, an oil high in MUFAs (olive oil) or an oil high in PUFAs (corn oil) was incorporated into a carrot cake, which was portioned to provide an additional 10%, above habitual intake, of the total daily energy intake as corn oil or olive oil, while keeping the diets isoenergetic. In practice, this corresponded to an increase from 9.1 ± 2.8% to 20.3 ± 3.5% with the MUFA diet and from 5.2 ± 3.1% to 13.4 ± 1.3% with the PUFA diet (Table 1). Total energy intake was assessed by a 7-d diary during the run-in period and validated according to estimated basal metabolic rate multiplied by an activity factor of 1.3 (2). The cakes were weighed into portions and then frozen to take home. Volunteers were instructed to include 3 slices of cake/d in their diet, ideally before the 3 main meals, and then to eat to satiety.

**Laboratory analysis**

Plasma glucose concentrations were measured by using a glucose oxidase–based autoanalyzer (Technicon; Axon Bayer

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**TABLE 2**

Biochemical test results of 9 people with diet-controlled type 2 diabetes with a high-MUFA and a high-PUFA diet

<table>
<thead>
<tr>
<th></th>
<th>MUFA diet</th>
<th></th>
<th>PUFA diet</th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Day 1</td>
<td>Day 24</td>
<td>Day 1</td>
<td>Day 24</td>
</tr>
<tr>
<td>Weight (kg)</td>
<td>87.0 ± 4.8</td>
<td>87.3 ± 4.9</td>
<td>86.7 ± 5</td>
<td>87.0 ± 4.9</td>
</tr>
<tr>
<td>Hb A1c (%)</td>
<td>6.6 ± 0.3</td>
<td>6.5 ± 0.3</td>
<td>6.7 ± 0.3</td>
<td>6.6 ± 0.3</td>
</tr>
<tr>
<td>Total cholesterol (mmol/L)</td>
<td>4.8 ± 0.3</td>
<td>4.9 ± 0.3</td>
<td>4.9 ± 0.4</td>
<td>4.6 ± 0.3</td>
</tr>
<tr>
<td>HDL cholesterol (mmol/L)</td>
<td>0.96 ± 0.1</td>
<td>0.97 ± 0.1</td>
<td>1.02 ± 0.1</td>
<td>1.00 ± 0.1</td>
</tr>
<tr>
<td>Total-HDL cholesterol ratio</td>
<td>5.2 ± 0.4</td>
<td>5.2 ± 0.3</td>
<td>5.2 ± 0.4</td>
<td>4.7 ± 0.3</td>
</tr>
<tr>
<td>LDL cholesterol (mmol/L)</td>
<td>3.05 ± 0.3</td>
<td>2.87 ± 0.3</td>
<td>3.12 ± 0.3</td>
<td>2.82 ± 0.2</td>
</tr>
<tr>
<td>Triacylglycerol (mmol/L)</td>
<td>1.51 ± 0.2</td>
<td>1.8 ± 0.3</td>
<td>1.66 ± 0.3</td>
<td>1.78 ± 0.3</td>
</tr>
<tr>
<td>Nonesterified fatty acids (mmol/L)</td>
<td>0.67 ± 0.1</td>
<td>0.59 ± 0.1</td>
<td>0.60 ± 0.1</td>
<td>0.67 ± 0.1</td>
</tr>
<tr>
<td>Insulin (pmol/L)</td>
<td>40.6 ± 6.9</td>
<td>42.2 ± 4.4</td>
<td>36.6 ± 5.1</td>
<td>36.2 ± 5.6</td>
</tr>
<tr>
<td>Glucose (mmol/L)</td>
<td>37.2 ± 6.8</td>
<td>57.6 ± 12.7</td>
<td>47.4 ± 10.7</td>
<td>56.2 ± 10.9</td>
</tr>
<tr>
<td>Nonesterified fatty acids (mmol·min−1-L−1)</td>
<td>115 ± 13</td>
<td>113 ± 17</td>
<td>117 ± 17</td>
<td>125 ± 16</td>
</tr>
<tr>
<td>GLP-1 (pmol/L)</td>
<td>385 ± 167</td>
<td>957 ± 205</td>
<td>705 ± 115</td>
<td>792 ± 139</td>
</tr>
<tr>
<td>Glucose (mmol·min−1-L−1)</td>
<td>3685 ± 699</td>
<td>3075 ± 898</td>
<td>3114 ± 930</td>
<td>4021 ± 1200</td>
</tr>
<tr>
<td>Hb A1c, glycated hemoglobin (%)</td>
<td>39.7 ± 0.3</td>
<td>39.7 ± 0.5</td>
<td>46.1 ± 1.3</td>
<td>49.1 ± 1.8</td>
</tr>
</tbody>
</table>

1 ± SEM. MUFA, monounsaturated fatty acids; PUFA, polyunsaturated fatty acids; GLP-1, glucagon-like peptide 1 [7–36] amide; Hb A1c, glycated hemoglobin.

2 Statistical analyses performed on log-transformed data.

3 Significant interaction between diet and time, *P* < 0.005.

4 Significantly different from day 1, *P* = 0.009.
Diagnostic, Newbury, United Kingdom). GLP-1 and insulin concentrations were measured in-house by using specific radioimmunoassays (19, 31). For both assays the interassay and intraassay CVs were <10%. The assays were capable of detecting 2 pmol/L with 95% confidence. All samples were included in one assay and analyzed in duplicate after the first freeze-thaw.

Triacylglycerol and nonesterified fatty acids were measured by the enzymatic colorimetric method using commercial kits (MPR2 Triacylglycerols GPO-PAP 701 kit; Roche Diagnostics Ltd, Lewes, United Kingdom; NEFA kit; W AKO Chemicals, Alpha Laboratories, Eastleigh, United Kingdom). Total and HDL-cholesterol concentrations were measured on a DAX-72 analyzer (Bayer Diagnostics, Basingstoke, United Kingdom).

**Statistical analysis**

Sample size was estimated from a previous acute study in healthy volunteers for a power of 80% and an α of 5% with an estimated difference in GLP-1 incremental areas under the curve (IAUC) of 1000 pmol·min/L and an SD of 1000 pmol·min/L (21). This suggested a minimum sample size of 10 pairs. All results are presented as means ± SEMs unless stated otherwise. For Table 2 and Table 3 a repeated-measures design was used with a $2 \times 2$ factorial structure for the repeated measures, with factors of diet (MUFA and PUFA) and day (1 and 24). Individual means have been reported only if there was a significant interaction. In Table 1, a repeated-measures design, with one repeated-measures factor, diet (baseline, MUFA, or PUFA), was used. Log transformation was used for positively skewed variables. $P < 0.05$ was taken as significant. The IAUC above baseline was calculated by using the trapezoidal rule.

**RESULTS**

Body weights did not change significantly over the study period (Table 2). Glycated hemoglobin (Hb A1c) did not change significantly over either of the 3-wk intervention periods (normal range: 4.3–5.7%). There was a significant reduction in the fasting ratio of total cholesterol to HDL cholesterol with the PUFA diet but no change with the MUFA diet. There was no change in fasting or postprandial GLP-1, insulin, or glucose concentration (Table 2, Figure 2). There was no change in fasting or postprandial triacylglycerol or nonesterified fatty acid concentrations (Table 2, Figure 3).

There was no change in insulin sensitivity measured by the SITT with either the MUFA or the PUFA diet (Table 2, Figure 4). The normal value is reported to be $\approx 175 \pm 10 \mu\text{mol} \cdot \text{L}^{-1} \cdot \text{min}^{-1}$ in young, lean nondiabetic adults (28); the lower the result, the more...
In this study, we investigated the effect of increasing the proportion of MUFAs compared with PUFAs on insulin resistance in people with type 2 diabetes. Insulin resistance is thought to explain some of the increased risk of coronary heart disease in type 2 diabetes. It is important to explore the role of dietary change in insulin resistance. Previous evidence suggests that MUFAs increase insulin sensitivity (7); however, there is inconclusive evidence of the effects of n-6 PUFAs on insulin sensitivity (12).

We did not find an increase in insulin sensitivity with either the MUFA intervention or the PUFA intervention despite showing compliance with the dietary intervention by a change in the composition of plasma triacylglycerols and phospholipids. This observation is also supported by the lack of effect of the diets on insulin or glucose IAUCs after the test meals.

We also did not see a change in GLP-1. Because there was also no effect on insulin sensitivity, we speculate that a diet-induced change in GLP-1 is still an exciting possibility—if a different nutrient were to be identified that augments the chronic release of endogenous GLP-1, that nutrient could still be a potential treatment for type 2 diabetes.

Other diet studies that suggested a positive effect on insulin sensitivity have been conducted over a similar length of time (9). In our study we were able to independently verify changes in plasma triacylglycerol that support the reported compliance. In the subset
of subjects we analyzed for plasma phospholipid content, there was already a significant increase in the L-O phospholipid ratio with the PUFA intervention; however, we suggest that the interventions may not have been of sufficient length to enable us to see a change in the skeletal muscle phospholipids, in which increasing unsaturation is associated with increasing insulin sensitivity (34).

We chose to give an identical test meal with each intervention so that we could directly compare the effects that the change in the background plasma triacylglycerols and phospholipids had on the various variables we measured. The fact that there was a significant change in the composition of plasma triacylglycerol at the endpoints in the study confirms a change in the metabolic environment. This does not tell us whether differences would have been measured if the composition of the test meals had been high in the corresponding intervention MUFA or PUFA.

As can be seen from the SITT results (Figure 4), there was no consistent change in insulin sensitivity despite high concentrations of corresponding plasma triacylglycerols and phospholipids at the time of the test. It may be that some individuals are more sensitive to dietary effects than are others; however, there was no correlation between the percentage change in the L-O ratio and the percentage change in SITT or GLP-1 release.

Total fat intake (% of energy) increased by 5% and 7% with the MUFA and PUFA interventions, respectively. Although the intake of saturated fat did not change, the increase in total fat per se may have influenced the results of the SITT (35). However, the recent American Diabetic Association guidelines suggest no clear upper limit on the amount of energy from MUFA (36), suggesting a split between MUFA and carbohydrate depending on the individual.

The typical UK MUFA intake is reported to be 12% whereas that of a typical current Mediterranean diet is 17%, although at the time of the Seven Countries Study a 24% MUFA intake was recorded in Crete (37). We achieved an adequate 11.2% increase in MUFA to 20.3% and an increase in PUFA to 8.2%, 7% above the UK average (38), while keeping the diet palatable. An intake of 20.3% of energy as MUFA is in line with 2 other prescribed dietary intervention studies in free-living subjects, which did show a benefit with a high-MUFA diet (39, 40), although neither of those studies measured insulin sensitivity directly.

Total cholesterol and triacylglycerol were surprisingly low considering that the group was insulin resistant with diet-controlled diabetes. The total cholesterol concentration at baseline was 4.8 ± 0.3 mmol/L. There was no significant change in total cholesterol with either intervention; however, there was a reduction in the ratio of total to HDL cholesterol with the PUFA diet. This is in agreement with the results of previous studies (10).

Garg’s (7) meta-analysis of previous studies on insulin sensitivity and MUFA suggests that MUFA also reduce triacylglycerol. Baseline triacylglycerol and nonesterified fatty acids were within the normal range and there was no significant change in fasting values or IAUCs for these during either intervention. It is impossible to know whether this effect would have been different had the volunteers been less well controlled metabolically.

![FIGURE 4. Short-insulin-tolerance test (SITT) of change in insulin sensitivity with 24 d of a high–monounsaturated fatty acid (MUFA) or high–polyunsaturated fatty acid (PUFA) diet in people with type 2 diabetes (n = 9).](https://academic.oup.com/ajcn/article-abstract/72/5/1111/4729805)

### TABLE 3

<table>
<thead>
<tr>
<th>Fatty acid</th>
<th>Baseline, day 1</th>
<th>MUFA, day 24</th>
<th>Baseline, day 1</th>
<th>PUFA, day 24</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>% by wt of total fatty acids</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>14:0</td>
<td>2.17 ± 0.2</td>
<td>2.61 ± 0.3</td>
<td>2.56 ± 0.4</td>
<td>2.5 ± 0.5</td>
</tr>
<tr>
<td>16:0</td>
<td>28.5 ± 1.7</td>
<td>30.7 ± 1.5</td>
<td>30.9 ± 2.1</td>
<td>29.7 ± 2.2</td>
</tr>
<tr>
<td>16:1</td>
<td>4.37 ± 0.4</td>
<td>4.70 ± 0.4</td>
<td>4.84 ± 0.5</td>
<td>3.78 ± 0.3*</td>
</tr>
<tr>
<td>18:0</td>
<td>4.36 ± 0.8</td>
<td>3.97 ± 0.3</td>
<td>3.56 ± 0.3</td>
<td>3.75 ± 0.4</td>
</tr>
<tr>
<td>18:1</td>
<td>38.1 ± 1.4</td>
<td>42.5 ± 1.6*</td>
<td>38.3 ± 1.4</td>
<td>36.0 ± 1.6</td>
</tr>
<tr>
<td>18:2</td>
<td>17.6 ± 1.4</td>
<td>12.2 ± 1.1*</td>
<td>13.8 ± 1.5</td>
<td>20.3 ± 3.5*</td>
</tr>
<tr>
<td>18:3</td>
<td>1.09 ± 0.4</td>
<td>0.39 ± 0.2</td>
<td>0.86 ± 0.5</td>
<td>0.88 ± 0.9</td>
</tr>
<tr>
<td>L:O</td>
<td>0.46 ± 0.03</td>
<td>0.29 ± 0.02*</td>
<td>0.36 ± 0.04</td>
<td>0.56 ± 0.05*</td>
</tr>
<tr>
<td>P:S</td>
<td>0.56 ± 0.1</td>
<td>0.35 ± 0.05*</td>
<td>0.43 ± 0.08</td>
<td>0.68 ± 0.2</td>
</tr>
</tbody>
</table>

* ± SEM. MUFA, monounsaturated fatty acids; PUFA, polyunsaturated fatty acids; L:O, ratio of linoleic (18:2n-6) to oleic (18:1n-9) acid; P:S, ratio of polyunsaturated to saturated fatty acids.

**Significant interaction between diet and time:** $^1 P = 0.049$, $^4 P = 0.008$, $^6 P = 0.02$, $^7 P = 0.03$.

$^1$Significantly different from day 1 within diet: $^1 P < 0.005$, $^3 P < 0.05$. 

$^{1,3}$
Note that the cake increased the sucrrose content of the diet by a mean of 68 g/d (13% of overall energy intake); however, there was no significant increase in the total sugar intake during the study. This had no effect on the glycemic indexes of the diets (baseline, 81.2 ± 9.0; MUFAs, 83.3 ± 3.8; PUFAs, 83.4 ± 3.0), which were calculated as described previously (41). We saw no effect of sucrrose (75 g) on triacylglycerol, nonesterified fatty acids, insulin, or glucose profiles in insulin-resistant men at risk of coronary heart disease or in matched control subjects (42). In a review of the sucrrose content of diabetic diets, Ha et al (43) reported that an intake of 10% of energy as sucrrose had no effect on triacylglycerol concentrations.

This is the first study to investigate the effect of MUFAs compared with PUFAs on GLP-1 and insulin sensitivity in people with type 2 diabetes. Neither MUFAs nor PUFAs had an effect on the postprandial stimulation of GLP-1. We found no advantage in recommending increases in the diet of MUFAs in preference to PUFAs in terms of increased insulin sensitivity for people with type 2 diabetes over this 3-wk intervention; however, PUFAs did significantly reduce the ratio of total to HDL cholesterol, a recognized benefit in the prevention of coronary heart disease. Over the 3-wk intervention period, diet-induced change in the plasma triacylglycerol or phospholipid L-Q ratio was not associated with either increased stimulation of GLP-1 or a change in insulin sensitivity in obese, insulin-resistant people with type 2 diabetes.

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REFERENCES