

# Novel Therapeutic Options in Anaplastic Large Cell Lymphoma: Molecular Targets and Immunological Tools

Olaf Merkel<sup>1</sup>, Frank Hamacher<sup>1</sup>, Eveline Siffert<sup>1</sup>, Lukas Kenner<sup>2,3</sup>, and Richard Greil<sup>1</sup> for the European Research Initiative on Anaplastic Large Cell Lymphoma

## Abstract

Anaplastic large cell lymphoma (ALCL) is a CD30-positive, aggressive T-cell lymphoma, and about half of the patients with this disease harbor the t(2;5)(p21;q35) translocation. This chromosomal aberration leads to fusion of the NPM gene with the ALK tyrosine kinase, leading to its constitutive activation. To date, treatment options include polychemotherapy (e.g., cyclophosphamide, doxorubicin, vincristine, and prednisone), which is sometimes combined with radiation in the case of bulky disease, leading to remission rates of ~80%. However, the remaining patients do not respond to therapy, and some patients experience chemo-resistant relapses, making the identification of new and better treatments imperative. The recent discovery of deregulated ALK in common cancers such as non-small cell lung cancer and neuroblastoma has reinvigorated industry interest in the development of ALK inhibitors. Moreover, it has been shown that the ALK protein is an ideal antigen for vaccination strategies due to its low expression in normal tissue. The characterization of microRNAs that are deregulated in ALCL will yield new insights into the biology of ALCL and open new avenues for therapeutic approaches in the future. Also, CD30 antibodies that have been tested in ALCL for quite a while will probably find a place in forthcoming treatment strategies. *Mol Cancer Ther*; 10(7); 1127–36. ©2011 AACR.

## Introduction

Anaplastic large cell lymphoma (ALCL) is a rare, aggressive, mature T-cell lymphoma that affects both children and adults. ALCL accounts for 10% to 15% of all non-Hodgkin's lymphoma (NHL) cases in children and 2% to 8% of NHL cases in adults. Clinically, ALCL can be distinguished as a systemic disease or a localized primary cutaneous entity. Systemic ALCL is a very aggressive lymphoma that involves different extranodal secondary sites, including soft tissue, skin, bone, lungs, and liver (1). Primary cutaneous ALCL accounts for <2% of the disease frequency but has a 5-year disease-free survival rate of ~90% upon standard treatment (2). The primary systemic and cutaneous subtypes have a similar histologic appearance, with so-called hallmark cells

showing large cells with abundant cytoplasm and eccentric, lobulated nuclei (3). However, the clear distinction of ALCL from other disease entities was made possible by the work of Stein and colleagues (4), who showed that the surface antigen CD30 is expressed by ALCL cells. In about half of the patients, the translocation t(2;5)(p23;q35) can be found, resulting in the expression of the nucleophosmin-anaplastic lymphoma kinase (NPM-ALK) fusion protein. ALK<sup>+</sup> lymphomas represent 50% to 85% of systemic ALCL cases and occur mainly in the first 3 decades of life, predominantly in males. ALK<sup>-</sup> ALCL occurs in older patients, with a peak at 60 years of age and lower male predominance. ALK<sup>-</sup> ALCL patients have a less favorable prognosis than ALK<sup>+</sup> ALCL patients after treatment with polychemotherapy. Other fusion partners of the ALK kinase, such as tropomyosin-3, 5-aminoimidazole-4-carboxamide ribonucleotide formyltransferase/IMP cyclohydrolase, clathrin heavy chain, and TRK fused gene 1, have been described (Fig. 1). In most cases, the fusion partners contain a dimerization domain that leads to constitutive activation of the ALK kinase (5). NPM is a multifunctional 23 kDa protein that is involved in diverse processes such as ribosome biogenesis, regulation of cell division, and DNA repair. ALK is a *trans*-membrane receptor tyrosine kinase (RTK) of the insulin receptor superfamily (6). The normal function of ALK is still elusive, especially because knockout mice have only very subtle phenotypes. In normal tissue, ALK expression has mainly been found in the neonatal brain, peripheral nervous system, and spinal fluid, suggesting a function in neuronal development.

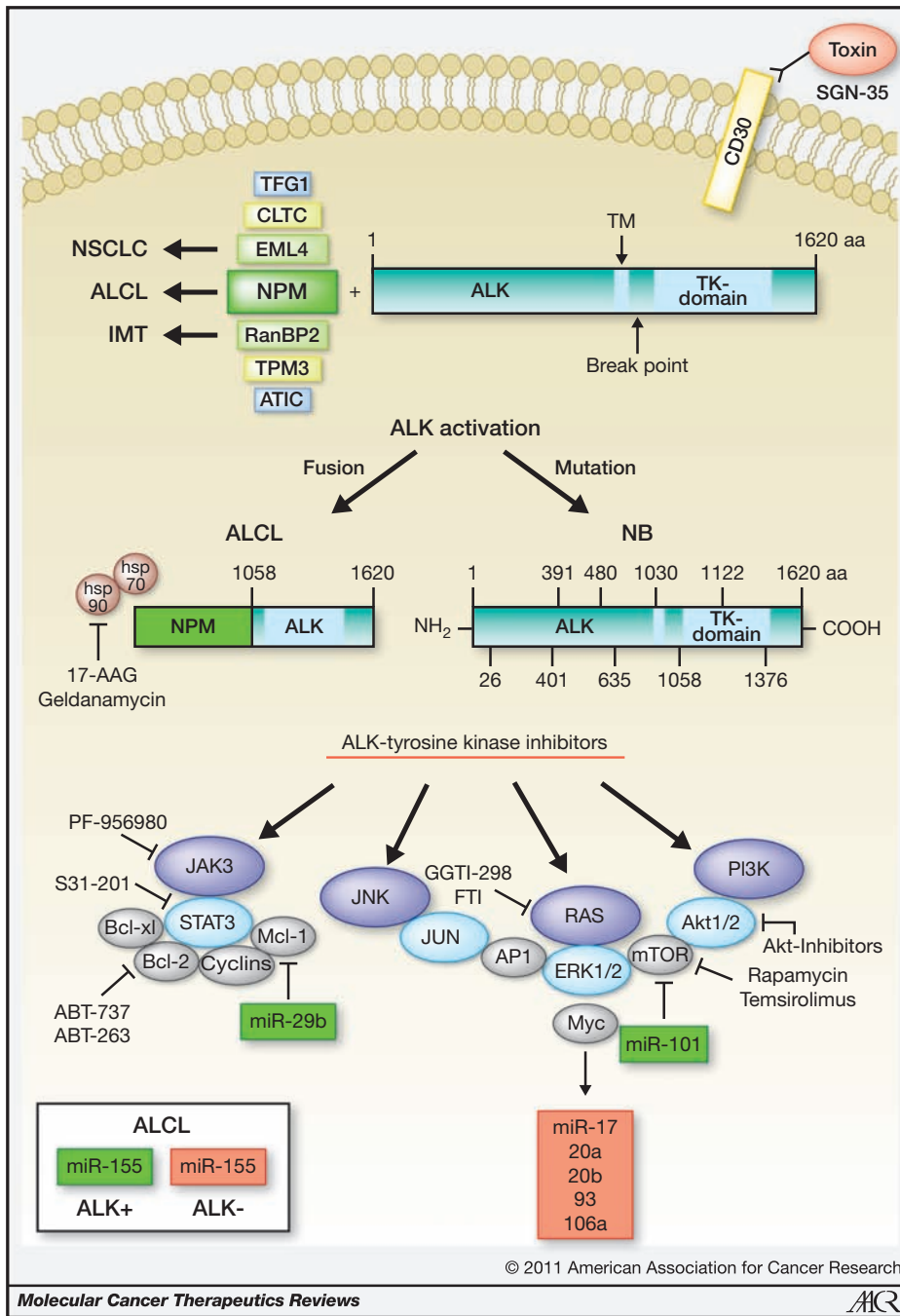
**Authors' Affiliations:** <sup>1</sup>Laboratory for Immunological and Molecular Cancer Research, Third Medical Department of Hematology, Medical Oncology, Hemostaseology, Rheumatology and Infectiology, Paracelsus Medical University Salzburg, Salzburg, Austria; <sup>2</sup>Department of Clinical Pathology, Medical University Vienna, and <sup>3</sup>Ludwig Boltzmann Institute for Cancer Research, Vienna, Austria

**Note:** The website for the European Research Initiative on Anaplastic Large Cell Lymphoma is [www.erialcl.net](http://www.erialcl.net)

**Corresponding Author:** Olaf Merkel, Laboratory for Immunological and Molecular Cancer Research, Third Medical Department, Paracelsus Medical University Salzburg, Müllner Hauptstrasse 48, 5020 Salzburg, Austria. Phone: 43-662-4482-1551; Fax: 43-662-4482-1570; E-mail: [o.merkel@salk.at](mailto:o.merkel@salk.at)

**doi:** 10.1158/1535-7163.MCT-11-0042

©2011 American Association for Cancer Research.



**Figure 1.** Sites of action of current and future ALCL therapeutics. The ALK kinase can be activated either by fusion to other proteins that lead to dimerization and thereby constitutive activation (ALCL and IMT), or by point mutations within the full-length protein (NB). Both are amenable to ALK-TK inhibition. Another approach is to inhibit one of the many ALK-activated downstream pathways, such as PI3K/Akt1/mTOR (7), JAK/Stat3 (8), and RAS/ERK (78). Several inhibitors are known to act directly on members of these pathways [e.g., PF-956980, JAK3 inhibitor; S31-201, STAT3 inhibitor; ABT-737 and ABT-263, BH-3 mimetics inhibiting Bcl-2 and Bcl-xl; GGTI-298 (geranylgeranyltransferase inhibitor-298) and FTI (farnesyltransferase inhibitor), Ras inhibitors; rapamycin and temsirolimus, mTOR inhibitors; and several Akt inhibitors]. In contrast, for ALCL patients without the ALK translocation, other treatment options would have to be considered. One such option may be the MMAE-coupled CD30 antibody SGN-35 (Seattle Genetics). Recently, we characterized oncogenic miRNAs that are specifically overexpressed in ALCL, ALK<sup>+</sup> (miR-17-92 cluster), or ALCL, ALK<sup>-</sup> (miR-155), as well as miRNAs that have low expression in both forms of ALCL (e.g., miR-29b and miR-101). These may represent novel molecular targets for future ALCL therapies.

Downloaded from <http://ascjournal.org/mct/article-pdf/10/7/1127/2320245/1127.pdf> by guest on 21 April 2024

The fusion of the N-terminal part of NPM to the kinase domain of ALK results in its constitutive activation. This leads to the activation of multiple downstream pathways, including PI3K/Akt/mTOR (7), JAK/Stat3 (8), c-myc (9), and cJun/JunB (ref. 10; Fig. 1). Mouse models that express the NPM-ALK within the hematopoietic system have shown the transforming capacity of this fusion protein (10–12). However, it remains a mystery why the ALCL variant that does not bear the translocation has the worst prognosis. The skewed age distribution, with adult ALCL

patients often lacking the NPM-ALK translocation, may influence the perception of a more negative outcome for ALK negative patients. In patients older than 40 years, a prognostic difference between the ALK<sup>+</sup> and ALK<sup>-</sup> variants of ALCL has not been found (13). In the most recent World Health Organization classification of this disease, the ALK<sup>+</sup> and ALK<sup>-</sup> subgroups were classified as 2 different disease entities, mainly due to their different prognostic properties (14). The molecular features of ALK-negative ALCL are largely unknown. Interest in

ALK kinase activation and its functional consequences increased tremendously after the recent finding that the echinoderm microtubule-associated protein-like 4-ALK kinase fusion protein (EML4-ALK) can be detected in >6% of non-small cell lung cancer (NSCLC) patients (15). Soda and colleagues (16) developed a mouse model for EML4-ALK-driven lung cancer and showed that ALK inhibitors can be effectively used for treatment in the murine system. By the end of 2009, Pfizer had launched a phase 3 clinical trial evaluating ALK inhibitors for the treatment of NSCLC bearing the EML4-ALK fusion. In patients with neuroblastoma (NB), point mutations were found that resulted in constitutive ALK activation in 6% to 12% of the patients (17–20). Recently, the EML4-ALK translocation was found with the use of exon array profiling in subsets of breast and colorectal cancer (21). ALK<sup>+</sup> patients are invariably treated with polychemotherapy—in most cases with cyclophosphamide, doxorubicin, vincristine, and prednisone (CHOP), which causes complete remission in ~80% of patients. Approximately 60% of ALK<sup>+</sup> patients only enter remission following CHOP treatment, leaving room for improvement of treatment strategies.

### ALK Kinase Inhibitors

The ALK cDNA encodes a protein of 177 kDa. The protein contains an extracellular ligand-binding domain, a transmembrane-spanning region, and a cytoplasmic kinase catalytic part (the TK domain), which has a major autophosphorylation site that regulates the conformation of the activation loop (Fig. 1). Full-length endogenous ALK supports tumor formation in NBs and glioblastomas, where it is suggested to be activated by the ligands pleiotropin and midkine (22, 23); however, the importance of the ligands is under debate (24). In ALCL and NSCLC, ALK is activated by chromosomal rearrangements leading to the fusion of the ALK gene to another gene, resulting in its dimerization and persistent activation. In NB, specific activating mutations within the kinase domain of ALK have been described. In all of these cases, ALK inhibition may be an effective therapeutic strategy; however, currently no ALK inhibitors have been approved for clinical cancer therapy. Several different classes of small molecules that have been identified as ALK inhibitors were shown to have marked antitumor efficacy in preclinical models. In contrast, attempts to abrogate NPM-ALK expression via siRNA were not successful, which may be due to the long half-life (>48 h) of the NPM-ALK protein (25). Piva and colleagues (26) found that antagonizing the NPM-ALK mRNA with small hairpin RNA through retroviral transfection was a more successful approach and established the essential function of this fusion protein in the proliferation of ALK<sup>+</sup> cell lines *in vitro* as well as in a murine ALCL engraftment model.

The ALK inhibitors staurosporine and 7-hydroxystaurosporine (UCN-01) are natural products that are isolated

from *Streptomyces staurosporeus*. UCN-01 was reported to have antitumor activity in a single patient with ALK-positive ALCL that was refractory to conventional CHOP therapy (27). Derivatives of staurosporine include the inhibitors CEP-14083 and CEP-14513 (Cephalon, Frazer, PA), whose clinical use is hampered by unfavorable physical properties such as low solubility and stability (28, 29). Second-generation inhibitors, such as compound 18 (Cephalon, Frazer, PA), showed improved properties *in vivo* with oral bioavailability and effective inhibition of tumor growth in the SCID mouse Sup-M2 tumor xenograft model (30). Another class of naturally occurring ALK modulators consists of 17-allylamino-17-demethoxygeldanamycin (17-AAG), geldanamycin, and herbimycin A. These modulators lead to indirect ALK inhibition via binding to heat shock protein 90 (HSP-90), thereby enhancing the proteasome-mediated degradation of the ALK protein (31, 32). Other inhibitor classes include pyridine, thiazole, aminopyridine, diaminopyridine, and fused ring systems. Li and colleagues (33) identified a series of novel pyridones as kinase inhibitors of ALK by stepwise *in vitro* screening followed by iterative template modification and computational ranking. One substance in this inhibitor family is the Novartis compound NVP-TAE684, which inhibits proliferation of Karpas-299 and SU-DHL-1 cell lines with an IC<sub>50</sub> range of 2 to 5 nM, hindering autophosphorylation of NPM-ALK (34). This compound is highly NPM-ALK specific, as the activity of other insulin receptor kinases is not significantly impaired. Another member of this inhibitor class is the GlaxoSmithKline compound GSK1838705, which is an ATP-competitive inhibitor of ALK, insulin-like growth factor (IGF)-1R, and insulin receptor (35). This compound is of special interest because of its good pharmacokinetic properties and excellent oral bioavailability (see Table 1 and Fig. 2).

The only compound currently undergoing clinical testing is the Pfizer compound PF-02341066 (crizotinib; Fig. 2), which was initially developed as an inhibitor of c-Met (36) but was found to have a similar IC<sub>50</sub> for ALK (24 nM). *In vitro*, it showed antiproliferative activity in ALCL cell lines (Karpas-299 and SU-DHL-1) leading to cell cycle arrest and apoptosis. The rather moderate side effects were predicted by the fact that it could be administered repeatedly in mice, dogs, and primates at concentrations up to 200 mg/kg/day for up to 30 days (36). It was tested in a monotherapy trial in heavily pretreated NSCLC patients with tumors harboring a rearrangement of EML4-ALK. The overall response rate was 64% and the disease control rate reached 90%, with a median duration of treatment of 25.5 weeks (37). Based on these promising results, at the end of 2009, Pfizer began to screen 1,500 patients with NSCLC for ALK translocations. Eighty-two (5.5%) of these patients had an ALK translocation (although in some cases not involving EML4) and were therefore eligible for PF-02341066 (crizotinib) treatment. It is interesting to note that this patient subgroup consisted mostly of relatively young nonsmokers with

**Table 1.** Selected small-molecule ALK-TK inhibitors and imatinib

Inhibitor	Company	Chemical type	Binding site	IC <sub>50</sub> (nM)	Reference	Clinical trial
UCN-01	Natural comp.	7-Hydroxystaurosporine	ATP pocket	123–150	(74)	Yes
CEP-14083	Cephalon	Staurosporine-derived	ATP pocket	2	(29)	No
CEP-14513	Cephalon	Staurosporine-derived	ATP pocket	4	(29)	No
Geldanamycin	Natural comp.	Benzoquinone	HSP90 HSP70		(31, 75)	Yes
Herbimycin	Natural comp.	Benzoquinone	HSP90 HSP70		(32)	No
NVP-TAE684	Novartis	Diaminopyrimidine	ATP pocket	<10	(34)	No
GSK1838705	GlaxoSmithKline	Diaminopyrimidine	ATP pocket	0.5	(35)	Yes
PF-02341066 (crizotinib)	Pfizer	Aminopyridine-derived	ATP pocket	11–24	(5)	Yes
Compound 18	Cephalon	Diaminopyrimidine	ATP pocket	4	(30)	No
WZ-5–126	None	Unknown	ATP pocket	3	(76)	No
Imatinib	Novartis	Diaminopyrimidine	ATP pocket	—	(77)	Yes

NOTE: Imatinib, the BCR-ABL inhibitor and a founding member of the group of TK inhibitor drugs, has been added as reference.

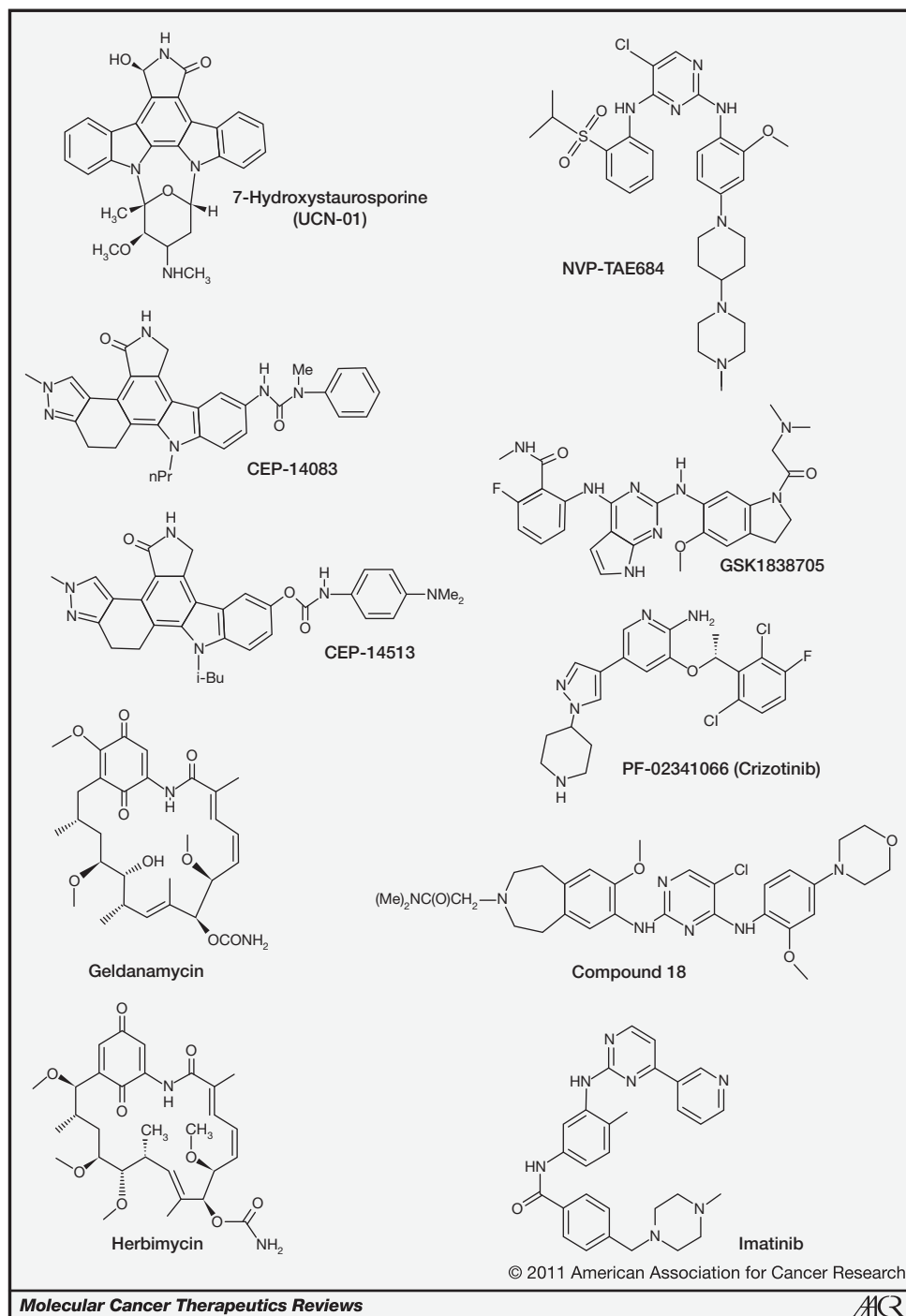
wild-type EGFR and KRAS. More than 91% of the included patients had received at least 1 previous therapy, and, remarkably, 41% had received 3 or more. Clinical activity was reported for 90% of the patients [57% responses (1 complete and 46 partial) and 33% stable disease] during the mean treatment duration of 6.4 months. Only grade 1 and 2 gastrointestinal side effects were reported (38). This result is encouraging, considering the generally poor prognosis for this class of patients. Crizotinib belongs to the TK inhibitor family, the most prominent member of which is imatinib (Gleevec; Novartis). Based on the experiences with this substance, we know that resistance mutations can eventually occur after prolonged treatment. This underscores the need for a wide range of chemically different ALK inhibitors. Indeed, such resistance mutations in the kinase domain of ALK (C1156Y and L1196M) were recently observed to develop in a patient, and their functional importance was confirmed by *in vitro* experiments (39). In addition, the first impressive clinical results regarding the use of crizotinib in patients with inflammatory myofibroblastic tumor (IMT), about half of whom carry the Ran-binding protein 2-ALK (RanBP2-ALK) translocation, were recently presented (40).

### CD30 Antibodies

CD30 is a *trans*-membrane protein that belongs to the tumor necrosis factor receptor family. It is expressed on activated (but not resting) B and T cells, and in addition to its expression on ALCL cells, it can also be found on Reed-Sternberg cells in Hodgkin's lymphoma (HL). CD30 antibodies and derivatives were tested in patients with this disease >15 years ago. In one of the first promising studies, the anti-CD30 monoclonal antibody was bound to saporin, a protein with ribosome-inactivating activity. This immunotoxin was given to 4 patients with pretreated,

refractory HL. In 3 of 4 patients, a rapid reduction of tumor mass was observed; however, the clinical responses were transient (41). In patients with HL and NHL, investigators attempted to bind CD30 to the toxin ricin or <sup>131</sup>I. However, the presence of side effects such as decreases in serum albumin, edema, and hypotension with the ricin, or severe hematotoxicity in the case of the Iodine-131-coupled antibody, dampened the enthusiasm for these types of treatments (42–44). In a seminal phase 1/2 study by Ansell and colleagues (45), it was shown that a fully human anti-CD30 immunoglobulin G1 monoclonal antibody (MDX-060) was well tolerated at concentrations up to 15 mg/kg in a mixed patient population, including patients with HL and ALCL. Despite high expectations, however, only 6 of 72 patients showed a clinical response. In a more recent study with 39 heavily pretreated HL patients (82% of whom had failed at least one prior therapy regimen) and 41 systemic ALCL patients using the anti-CD30 antibody SGN-30 (Seattle Genetics), none of the HL patients had an objective response (46). In contrast, in the ALCL arm, 2 patients achieved a complete response (4.9%) and 5 showed a partial response (12%), pointing to a limited clinical activity of this antibody. Side effects were rather mild, with fatigue and nausea being the most frequently reported symptoms. In a phase 2 study in which the same antibody was tested in CD30-positive, heavily pretreated patients with cutaneous ALCL, lymphomatoid papulosis, and transformed mycosis fungoides, SGN-30 showed clinical activity (complete or partial response) in 16 out of 23 patients. Specifically, in cutaneous ALCL, clinical activity was seen in all patients, with responses in 82% of the treated patients (47). These are encouraging results, especially given the mild side-effects profile. However, these data have to be considered in context with current treatment strategies using pegylated liposomal doxorubicin and gemcitabine, which have response rates for cutaneous ALCL ranging from 69% to 80% (48). The latest

**Figure 2.** Organic structures of selected small-molecule ALK-TK inhibitors and imatinib. The structure of the substance WZ-5-126 is not given because it was not publicly available at the time of submission.



addition to these CD-30 antibody-based treatment strategies is SGN-35 (brentuximab vedotin; Seattle Genetics). In this case, the CD-30 antibody is coupled to the antitubulin agent monomethyl auristatin E (MMAE) with an enzyme-cleavable dipeptide linker. This drug has been tested in 42 patients with CD30-positive relapsed or refractory HL ( $n = 40$ ) or ALCL ( $n = 2$ ) patients with a median of 3 previous therapy regimens. It is remarkable that in 36 of 42

patients (86%) tumor regression was observed, and among the 11 complete remissions, 2 systemic ALK<sup>+</sup> ALCL cases were found (49). The success of this coupled antibody is even more surprising in light of the limited activity of the CD30 antibody alone (SGN-30) in systemic ALCL, and stresses the role of the attached cytoskeleton toxin. Another appealing feature of the armed CD-30 antibody strategy is that, in contrast to ALK inhibitors,

it can be used in all subtypes of ALCL, including ALK<sup>-</sup> ALCL, ALK<sup>+</sup> ALCL, and cutaneous ALCL.

### Inhibitors of NPM-ALK-Activated Downstream Pathways

ALK hyperactivation potentiates a multitude of partially overlapping mitogenic cellular pathways that lead to transformation, proliferation, and cell survival. The main activated pathways include the PI3K/Akt/mTOR, ras-ERK, and JAK/STAT pathways (ref. 1; see also Fig. 1). JAK3, for example, is a lymphocyte-specific member of the Janus kinase signal transducer family that is expressed in ALCL cells but not in resting T cells. JAK3 is a signal transducer from ALK to STAT3, which in turn activates antiapoptotic proteins within the bcl2-rheostat such as Bcl2, Bcl-XL, and Mcl1, as well as cyclins, C-EBP $\beta$ , and survivin. Several JAK3 small-molecule inhibitors, including PF-956980 (Pfizer), have been described (50, 51). SHP1 is a phosphatase tumor suppressor that is constitutively deactivated through promoter methylation in 50% of ALK<sup>+</sup> ALCL, and deactivates the JAK3/STAT3 pathway. Using the methyl-transferase inhibitor 5-aza-deoxycytidine, Han and colleagues (52) showed that SHP1 could be reactivated, leading to reduced levels of JAK3, *p*-JAK3, and *p*-STAT3, and sensitized cells to doxorubicin-induced apoptosis. Also, a small-molecule (S3I-201) that directly deactivates STAT3 by inhibiting DNA binding and STAT3 complex formation, identified from the National Cancer Institute chemical library, has been described and may be of therapeutic interest (53).

For the STAT3 downstream targets Bcl2 and Bcl-X<sub>L</sub>, a potent inhibitor called ABT-737 has been developed by Abbott Laboratories, and a derivate of this inhibitor, ABT-236 (navitoclax; Abbott Laboratories), which is orally available, has also been described (54, 55). Overexpression of the related Mcl1 protein has often been described as conferring resistance to ABT-737 and ABT-263; therefore, it is tempting to speculate that the reintroduction of microRNAs (miRNAs) that are down-regulated in ALCL and target Mcl1, such as miR-101 and miR-29b, might sensitize ALCL cells to the effect of Bcl-2 inhibitors. The ras-ERK pathway, which is mainly responsible for cell proliferation, may be inhibited by farnesyltransferase or geranylgeranyltransferase inhibitors (56). The AKT/mTOR pathway is one of the most prominent pathways that are often overactivated in cancer. Therefore, multiple approaches have been described to inhibit its activity. In the case of AKT, classical ATP competitive inhibitors, phosphatidyl analogs that block the essential binding of AKT to PI(3,4,5)P<sub>3</sub>, pseudosubstrate inhibitors, and allosteric inhibitors have been described (57). The mTOR protein can be selectively suppressed by rapamycin and its orally available analog, temsirolimus. The feasibility of this strategy was demonstrated in ALCL mouse models, in which temsirolimus was shown to effectively reduce tumor growth (58).

### miRNAs and ALCL

miRNAs are small, evolutionary conserved, noncoding RNAs that have been described to regulate the expression of target genes through translational inhibition or mRNA degradation. To date, ~1,000 miRNAs have been described in humans, and it is predicted (based on computational algorithms) that they are involved in regulating ~30% of the proteins that are transcribed from the human genome (59, 60). The roles of miRNAs are as diverse as their targeted proteins, and therefore there are almost no aspects of cancer biology that are not affected by miRNAs. Investigators have described oncogenic miRNAs (miR-21 and miR-17-92 cluster), as well as miRNAs that act as tumor suppressors (miR-34a, miR-15a/16-1, and miR-29a, b, c). To complicate the issue, one miRNA can play tremendously different roles depending on its cellular context. For example, miR-34a, which is a downstream target of p53, is overexpressed in CLL (61) but is down-regulated in NSCLC, glioblastoma, and NB, raising the question as to whether it is a tumor suppressor or an oncogene. Until recently, the role of miRNAs in ALCL was undefined. We did a comprehensive screen of deregulated miRNA expression in human formalin-fixed, paraffin-embedded (FFPE) tissue samples, a murine ALCL tumor model, and 5 ALCL cell lines. miR-101 was down-regulated in primary ALCL FFPE tissues, the ALCL mouse model, and all tested ALCL cell lines. When miR-101 was reintroduced into the ALCL cell lines, we observed a reduced proliferation of ALK<sup>+</sup> but not ALK<sup>-</sup> cell lines. When we inhibited the described miR-101 target mTOR using the rapamycin analog temsirolimus, we found that proliferation of the ALK<sup>+</sup> ALCL cell lines was inhibited much more strongly than that of the ALK<sup>-</sup> cell lines, suggesting a stronger dependence of ALK<sup>+</sup> cell lines on the mTOR pathway. Moreover, we identified miRNAs that are differentially expressed in ALK<sup>+</sup> versus ALK<sup>-</sup> ALCL. Members of the oncogenic miR-17-92 cluster were more strongly expressed in ALK<sup>+</sup>, whereas the oncogenic miR-155, coded within the B-cell integration cluster RNA, was expressed >10-fold higher in ALK<sup>-</sup> ALCL (Fig. 1). Given the poor prognosis for ALCL patients who do not bear the ALK translocation (with a 5-year survival of only 30%–40%), the need for new treatment options is obvious. miRNA-155 has been linked to B-cell differentiation, oncogenesis, and immune function, making it the first molecular therapeutic target for ALK<sup>-</sup> ALCL (62–64).

### Vaccination Strategies

In the years since some groundbreaking studies introduced the concept of cancer immune surveillance through the innate and adapted immune systems of patients, many studies have followed up on this concept and dramatically increased our knowledge about the interactions between the immune system and cancer (65–67). In general, the control of cancer by the immune system, often called immune editing, consists

of 3 phases: elimination, equilibrium, and evasion. Elimination of tumor cells through the immune system involves building a balance between tumor growth and immune response, resulting in a small, undetectable amount of tumor cells. In the equilibrium phase, the tumor is held in check by antibodies and thus is unable to grow. In the evasion phase, the tumor, either by suppressing the immune response or by editing its exposed antigens, is able to escape the immune system and grow into a clinically apparent cancer (68). Ideally, an antigen that is used for vaccination would be specifically expressed in the tumor; it must have an important, causal part in the multifactorial process that leads to cancer, and it must be expressed stably even after it is attacked by the immune system. As highly specific antigens for tumor vaccination, one could envisage the protein products of mutated tumor suppressors, as well as proteins encoded by oncogenic viruses. However, these strictly tumor-specific antigens are relatively rare, and therefore researchers have widened their spectrum of target antigens to tumor-associated antigens, which in many cases are lineage markers that are also expressed on the corresponding healthy cells. Sipuleucel-T (Dendreon Corporation), a vaccine for prostate cancer that is based on *ex vivo* stimulation of patients' monocytes with a fusion protein consisting of prostatic acid phosphatase and granulocyte macrophage colony-stimulating factor, received Food and Drug Administration approval in April 2010. Furthermore, melanoma and other epithelial cancers are being investigated in phase 1/2 trials (68). It is clear that only ALCL forms that bear the ALK translocation would be amenable to this type of treatment. Given the above-mentioned preconditions for a tumor antigen, ALK is an ideal target for potential tumor vaccination. First, with the exception of the immunoprivileged central and peripheral nervous systems, the ALK kinase is not expressed, or is expressed at very low levels, in normal tissues. Second, it has been shown that the ALK kinase has a strong transforming capacity in normal T cells and can cause cancer in transgenic animal models, where the NPM-ALK fusion protein is specifically expressed in CD4 T cells or bone marrow (10–12). These facts have led many groups to test various strategies for tumor vaccination in murine ALCL model systems. The ALK-fusion proteins have immunogenic properties per se, hence Pulford and colleagues (69) showed that antibodies against ALK can be found in patients with ALK-positive ALCL. In a more recent study (70), the same group showed that these antibodies can persist for >10 years after the ALCL has been cured in a patient, and it is likely that these antibodies will remain present for the rest of that person's life. ALK-specific antibodies were observed in 87 of 95 chemo-naïve ALCL patients, and it was shown that high antibody titers correlated with a reduced incidence of relapse. Given the positive prognostic value of the ALK translocation within the

ALCL patient cohort, it has been hypothesized that this may be due to a permanent antitumor immune response toward the ALK-fusion protein in these patients.

Efforts to actively tune the immune response toward ALK have also been undertaken. In a study by Passoni and colleagues (71), 2 ALK-derived, HLA-A\*0201-binding peptides were able to elicit an ALK-specific cytotoxic T-lymphocyte response in peripheral blood mononuclear cells of HLA-matched healthy donors and in transgenic mice bearing the human HLA-A\*0201 locus. Moreover, the anti-ALK-peptide-primed cytotoxic T cells were able to lyse HLA-matched ALCL and NB cell lines that expressed the ALK protein.

In a different approach using DNA vaccination, Chiarle and colleagues (72) electroporated Balb/C mice twice with a plasmid containing the intracytoplasmic domain of ALK. The great advantage of DNA vaccination, as opposed to vaccination with HLA-specific peptides, is that it can be done independently of the HLA haplotype. One week after the last immunization, the mice were injected with  $1 \times 10^6$  cells of an ALCL, ALK<sup>+</sup> cell line. Of interest, the vaccinated animals were completely protected from ALCL engraftment, and this protection remained present for >70 days. However, when the immunization was carried out after engraftment of the ALCL cells, the attenuating effect on tumor growth was minimal. Therefore, the authors tried to debulk the tumor using 1 bolus of doxorubicin, followed by vaccination on days 7, 14, and 21 after administration of the doxorubicin bolus. The chemotherapy alone led to a remission rate of 60%, which was further enhanced by additional vaccination to 87%. This suggests the feasibility of using a vaccination strategy after a tumor is debulked by classical chemotherapy.

## Conclusions

Despite the relative success of classical chemotherapy (together with radiation) in ALCL, it is clear that novel treatments are needed for resistant or relapsing patients. Furthermore, it seems that the concept of TK inhibition, which has shown success in chronic myeloid leukemia [*i.e.*, with imatinib (Gleevec; Roche)], may also hold promise for other cancers, including ALCL. This is substantiated by recent successful clinical trials of the ALK-TK inhibitor crizotinib in solid tumor entities such as NSCLC and IMT (38, 40), which leaves us optimistic about its effectiveness in NB and ALCL. However, we have to await the results of clinical trials that are currently being done. Also, the CD30 antibody-based therapeutic strategies that have been tested for a long time with only limited success are far from dead. The recent clinical trial with the armed CD-30 antibody SGN-35 showed that this approach not only holds promise but translated into complete clinical responses in 10/40 heavily pretreated HL and 2/2 ALK<sup>+</sup> ALCL patients. It is important to note that this agent may also be active in ALK<sup>-</sup> ALCL patients,

in whom ALK kinase inhibitors are useless. It is clear that the treatment of ALK<sup>-</sup> ALCL patients remains a big challenge for the future. To date, not much is known about the specific molecular properties of ALK<sup>-</sup> ALCL that might be instrumental in developing new drugs. Recently, overexpression of the oncogenic miR-155 and a chromosomal translocation t(6;7)(p25.3;q32.3) resulting in miR-29 activation was reported in this class of patients (58, 73). The recent increase in the therapeutic armamentarium for lymphomas should give us confidence that also in the context of ALK<sup>-</sup> ALCL, which up to now has proved resistant to the identification of specific molecular targets, basic research elucidating novel genetic and immunologic features will allow the development of new and more-effective treatment options.

### Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed.

### References

- Chiarle R, Voena C, Ambrogio C, Piva R, Inghirami G. The anaplastic lymphoma kinase in the pathogenesis of cancer. *Nat Rev Cancer* 2008;8:11–23.
- Querfeld C, Khan I, Mahon B, Nelson BP, Rosen ST, Evens AM. Primary cutaneous and systemic anaplastic large cell lymphoma: clinicopathologic aspects and therapeutic options. *Oncology (Williston Park)* 2010;24:574–87.
- Fornari A, Piva R, Chiarle R, Novero D, Inghirami G. Anaplastic large cell lymphoma: one or more entities among T-cell lymphoma? *Hematol Oncol* 2009;27:161–70.
- Dürkop H, Latza U, Hummel M, Eitelbach F, Seed B, Stein H. Molecular cloning and expression of a new member of the nerve growth factor receptor family that is characteristic for Hodgkin's disease. *Cell* 1992;68:421–7.
- Webb TR, Slavish J, George RE, Look AT, Xue L, Jiang Q, et al. Anaplastic lymphoma kinase: role in cancer pathogenesis and small-molecule inhibitor development for therapy. *Expert Rev Anticancer Ther* 2009;9:331–56.
- Morris SW, Kirstein MN, Valentine MB, Dittmer KG, Shapiro DN, Saltman DL, et al. Fusion of a kinase gene, ALK, to a nucleolar protein gene, NPM, in non-Hodgkin's lymphoma. *Science* 1994;263:1281–4.
- Staber PB, Vesely P, Haq N, Ott RG, Funato K, Bambach I, et al. The oncoprotein NPM-ALK of anaplastic large-cell lymphoma induces JUNB transcription via ERK1/2 and JunB translation via mTOR signaling. *Blood* 2007;110:3374–83.
- Zamo A, Chiarle R, Piva R, Howes J, Fan Y, Chilosi M, et al. Anaplastic lymphoma kinase (ALK) activates Stat3 and protects hematopoietic cells from cell death. *Oncogene* 2002;21:1038–47.
- Wellmann A, Doseeva V, Butscher W, Raffeld M, Fukushima P, Stetler-Stevenson M, et al. The activated anaplastic lymphoma kinase increases cellular proliferation and oncogene up-regulation in rat 1a fibroblasts. *FASEB J* 1997;11:965–72.
- Turner SD, Tooze R, MacLennan K, Alexander DR. Vav-promoter regulated oncogenic fusion protein NPM-ALK in transgenic mice causes B-cell lymphomas with hyperactive Jun kinase. *Oncogene* 2003;22:7750–61.
- Chiarle R, Gong JZ, Guasparri I, Pesci A, Cai J, Liu J, et al. NPM-ALK transgenic mice spontaneously develop T-cell lymphomas and plasma cell tumors. *Blood* 2003;101:1919–27.
- Turner SD, Merz H, Yeung D, Alexander DR. CD2 promoter regulated nucleophosmin-anaplastic lymphoma kinase in transgenic mice causes B lymphoid malignancy. *Anticancer Res* 2006; 26[5A]:3275–9.
- Savage KJ, Harris NL, Vose JM, Ullrich F, Jaffe ES, Connors JM, et al. ALK<sup>-</sup> anaplastic large-cell lymphoma is clinically and immunophenotypically different from both ALK<sup>+</sup> ALCL and peripheral T-cell lymphoma, not otherwise specified: report from the International Peripheral T-Cell Lymphoma Project. *Blood* 2008;111:5496–504.
- Swerdlow SH, Campo E, Harris NL, Jaffe ES, Pileri SA, Stein H, et al. WHO classification of tumours of haematopoietic and lymphoid tissues. Lyon, France: IARC Press; 2008.
- Soda M, Choi YL, Enomoto M, Takada S, Yamashita Y, Ishikawa S, et al. Identification of the transforming EML4-ALK fusion gene in non-small-cell lung cancer. *Nature* 2007;448:561–6.
- Soda M, Takada S, Takeuchi K, Choi YL, Enomoto M, Ueno T, et al. A mouse model for EML4-ALK-positive lung cancer. *Proc Natl Acad Sci USA* 2008;105:19893–7.
- Chen Y, Takita J, Choi YL, Kato M, Ohira M, Sanada M, et al. Oncogenic mutations of ALK kinase in neuroblastoma. *Nature* 2008;455:971–4.
- George RE, Sanda T, Hanna M, Fröhling S, Luther W 2nd, Zhang J, et al. Activating mutations in ALK provide a therapeutic target in neuroblastoma. *Nature* 2008;455:975–8.
- Janoueix-Lerosey I, Lequin D, Brugières L, Ribeiro A, de Pontual L, Combaret V, et al. Somatic and germline activating mutations of the ALK kinase receptor in neuroblastoma. *Nature* 2008;455:967–70.
- Mossé YP, Laudenslager M, Longo L, Cole KA, Wood A, Attiyeh EF, et al. Identification of ALK as a major familial neuroblastoma predisposition gene. *Nature* 2008;455:930–5.
- Lin E, Li L, Guan Y, Soriano R, Rivers CS, Mohan S, et al. Exon array profiling detects EML4-ALK fusion in breast, colorectal, and non-small cell lung cancers. *Mol Cancer Res* 2009;7:1466–76.
- Stoica GE, Kuo A, Aigner A, Sunitha I, Souttou B, Malerczyk C, et al. Identification of anaplastic lymphoma kinase as a receptor for the growth factor pleiotrophin. *J Biol Chem* 2001;276:16772–9.
- Stoica GE, Kuo A, Powers C, Bowden ET, Sale EB, Riegel AT, et al. Midkine binds to anaplastic lymphoma kinase (ALK) and acts as a growth factor for different cell types. *J Biol Chem* 2002;277:35990–8.
- Mathivet T, Mazot P, Vigny M. In contrast to agonist monoclonal antibodies, both C-terminal truncated form and full length form of Pleiotrophin failed to activate vertebrate ALK (anaplastic lymphoma kinase)? *Cell Signal* 2007;19:2434–43.
- Ritter U, Damm-Welk C, Fuchs U, Bohle RM, Borkhardt A, Woessmann W. Design and evaluation of chemically synthesized siRNA targeting the NPM-ALK fusion site in anaplastic large cell lymphoma (ALCL). *Oligonucleotides* 2003;13:365–73.

### Acknowledgments

We thank Ninon Taylor for proofreading the manuscript. We also thank the many scientists who have contributed to the field and whose work is not mentioned here due to the stringent limit on the number of publications cited.

### Grant Support

Klinische Malignom und Zytokinforschung Salzburg-Innsbruck GmbH (R. Greil); Jubiläumsfond der Österreichischen Nationalbank (grant 12170 to R. Greil); Spezialforschungsprogramm P021 (R. Greil); Fonds zur Förderung der wissenschaftlichen Forschung P-18478-B12 (L. Kenner); Genome Austria Research project Inflammobiota (L. Kenner); and Novus Sanguis Consortium (L. Kenner).

The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked *advertisement* in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

Received January 17, 2011; revised April 14, 2011; accepted April 16, 2011; published OnlineFirst June 28, 2011.



26. Piva R, Chiarle R, Manazza AD, Tauli R, Simmons W, Ambrogio C, et al. Ablation of oncogenic ALK is a viable therapeutic approach for anaplastic large-cell lymphomas. *Blood* 2006;107:689–97.
27. Sausville EA, Arbusk SG, Messmann R, Headlee D, Bauer KS, Lush RM, et al. Phase I trial of 72-hour continuous infusion UCN-01 in patients with refractory neoplasms. *J Clin Oncol* 2001;19:2319–33.
28. Piva R, Pellegrino E, Mattioli M, Agnelli L, Lombardi L, Boccalatte F, et al. Functional validation of the anaplastic lymphoma kinase signature identifies CEBPB and BCL2A1 as critical target genes. *J Clin Invest* 2006;116:3171–82.
29. Wan W, Albom MS, Lu L, Quail MR, Becknell NC, Weinberg LR, et al. Anaplastic lymphoma kinase activity is essential for the proliferation and survival of anaplastic large-cell lymphoma cells. *Blood* 2006;107:1617–23.
30. Mesaros EF, Burke JP, Parrish JD, Dugan BJ, Anzalone AV, Angeles TS, et al. Novel 2,3,4,5-tetrahydro-benzo[d]azepine derivatives of 2,4-diaminopyrimidine, selective and orally bioavailable ALK inhibitors with antitumor efficacy in ALCL mouse models. *Bioorg Med Chem Lett* 2011;21:463–6.
31. Bonvini P, Gastaldi T, Falini B, Rosolen A. Nucleophosmin-anaplastic lymphoma kinase (NPM-ALK), a novel Hsp90-client tyrosine kinase: down-regulation of NPM-ALK expression and tyrosine phosphorylation in ALK(+) CD30(+) lymphoma cells by the Hsp90 antagonist 17-allyl-amino,17-demethoxygeldanamycin. *Cancer Res* 2002;62:1559–66.
32. Turturro F, Arnold MD, Frist AY, Pulford K. Model of inhibition of the NPM-ALK kinase activity by herbimycin A. *Clin Cancer Res* 2002;8:240–5.
33. Li R, Xue L, Zhu T, Jiang Q, Cui X, Yan Z, et al. Design and synthesis of 5-aryl-pyridone-carboxamides as inhibitors of anaplastic lymphoma kinase. *J Med Chem* 2006;49:1006–15.
34. Galkin AV, Melnick JS, Kim S, Hood TL, Li N, Li L, et al. Identification of NVP-TAE684, a potent, selective, and efficacious inhibitor of NPM-ALK. *Proc Natl Acad Sci USA* 2007;104:270–5.
35. Sabbatini P, Korenchuk S, Rowand JL, Groy A, Liu Q, Leperi D, et al. GSK1838705A inhibits the insulin-like growth factor-1 receptor and anaplastic lymphoma kinase and shows antitumor activity in experimental models of human cancers. *Mol Cancer Ther* 2009;8:2811–20.
36. Zou HY, Li Q, Lee JH, Arango ME, McDonnell SR, Yamazaki S, et al. An orally available small-molecule inhibitor of c-Met, PF-2341066, exhibits cytoreductive antitumor efficacy through antiproliferative and antiangiogenic mechanisms. *Cancer Res* 2007;67:4408–17.
37. Bang Y, Kwak EL, Shaw AT, Camidge DR, Iafrate AJ, Maki RG, et al. Clinical activity of the oral ALK inhibitor PF-02341066 in ALK-positive patients with non-small cell lung cancer (NSCLC). *J Clin Oncol (Meeting Abstracts)* 2010;28(suppl)3.
38. Kwak EL, Bang YJ, Camidge DR, Shaw AT, Solomon B, Maki RG, et al. Anaplastic lymphoma kinase inhibition in non-small-cell lung cancer. *N Engl J Med* 2010;363:1693–703.
39. Choi YL, Soda M, Yamashita Y, Ueno T, Takashima J, Nakajima T, et al. EML4-ALK mutations in lung cancer that confer resistance to ALK inhibitors. *N Engl J Med* 2010;363:1734–9.
40. Butrynski JE, D'Adamo DR, Hornick JL, Dal Cin P, Antonescu CR, Jhanwar SC, et al. Crizotinib in ALK-rearranged inflammatory myofibroblastic tumor. *N Engl J Med* 2010;363:1727–33.
41. Falini B, Bolognesi A, Flenghi L, Tazzari PL, Broe MK, Stein H, et al. Response of refractory Hodgkin's disease to monoclonal anti-CD30 immunotoxin. *Lancet* 1992;339:1195–6.
42. Borchmann P, Schnell R, Fuss I, Manzke O, Davis T, Lewis LD, et al. Phase 1 trial of the novel bispecific molecule H22xKi-4 in patients with refractory Hodgkin lymphoma. *Blood* 2002;100:3101–7.
43. Schnell R, Staak O, Borchmann P, Schwartz C, Matthey B, Hansen H, et al. A phase I study with an anti-CD30 ricin A-chain immunotoxin (Ki-4.dgA) in patients with refractory CD30+ Hodgkin's and non-Hodgkin's lymphoma. *Clin Cancer Res* 2002;8:1779–86.
44. Schnell R, Dietlein M, Staak JO, Borchmann P, Schomaecker K, Fischer T, et al. Treatment of refractory Hodgkin's lymphoma patients with an iodine-131-labeled murine anti-CD30 monoclonal antibody. *J Clin Oncol* 2005;23:4669–78.
45. Ansell SM, Horwitz SM, Engert A, Khan KD, Lin T, Strair R, et al. Phase I/II study of an anti-CD30 monoclonal antibody (MDX-060) in Hodgkin's lymphoma and anaplastic large-cell lymphoma. *J Clin Oncol* 2007;25:2764–9.
46. Forero-Torres A, Leonard JP, Younes A, Rosenblatt JD, Brice P, Bartlett NL, et al. A Phase II study of SGN-30 (anti-CD30 mAb) in Hodgkin lymphoma or systemic anaplastic large cell lymphoma. *Br J Haematol* 2009;146:171–9.
47. Duvic M, Reddy SA, Pinter-Brown L, Korman NJ, Zic J, Kennedy DA, et al. A phase II study of SGN-30 in cutaneous anaplastic large cell lymphoma and related lymphoproliferative disorders. *Clin Cancer Res* 2009;15:6217–24.
48. Duvic M. Improved understanding of peripheral T-cell lymphomas. *Oncology (Williston Park)* 2010;587:592–3.
49. Younes A, Bartlett NL, Leonard JP, Kennedy DA, Lynch CM, Sievers EL, et al. Brentuximab vedotin (SGN-35) for relapsed CD30-positive lymphomas. *N Engl J Med* 2010;363:1812–21.
50. Changelian PS, Flanagan ME, Ball DJ, Kent CR, Magnuson KS, Martin WH, et al. Prevention of organ allograft rejection by a specific Janus kinase 3 inhibitor. *Science* 2003;302:875–8.
51. Changelian PS, Moshinsky D, Kuhn CF, Flanagan ME, Munchhof MJ, Harris TM, et al. The specificity of JAK3 kinase inhibitors. *Blood* 2008;111:2155–7. Erratum in: *Blood* 2009;114:3132.
52. Han Y, Amin HM, Frantz C, Franko B, Lee J, Lin Q, et al. Restoration of shp1 expression by 5-AZA-2c-deoxycytidine is associated with downregulation of JAK3/STAT3 signaling in ALK-positive anaplastic large cell lymphoma. *Leukemia* 2006;20:1602–9.
53. Siddiquee K, Zhang S, Guida WC, Blaskovich MA, Greedy B, Lawrence HR, et al. Selective chemical probe inhibitor of Stat3, identified through structure-based virtual screening, induces antitumor activity. *Proc Natl Acad Sci USA* 2007;104:7391–6.
54. Oltersdorf T, Elmore SW, Shoemaker AR, Armstrong RC, Augeri DJ, Belli BA, et al. An inhibitor of Bcl-2 family proteins induces regression of solid tumours. *Nature* 2005;435:677–81.
55. Tse C, Shoemaker AR, Adickes J, Anderson MG, Chen J, Jin S, et al. ABT-263: a potent and orally bioavailable Bcl-2 family inhibitor. *Cancer Res* 2008;68:3421–8.
56. Sebt SM, Hamilton AD. Farnesyltransferase and geranylgeranyltransferase I inhibitors and cancer therapy: lessons from mechanism and bench-to-bedside translational studies. *Oncogene* 2000;19:6584–93.
57. Lindsley CW. The Akt/PKB family of protein kinases: a review of small molecule inhibitors and progress towards target validation: a 2009 update. *Curr Top Med Chem* 2010;10:458–77.
58. Merkel O, Hamacher F, Laimer D, Sifft E, Trajanoski Z, Scheideler M, et al. Identification of differential and functionally active miRNAs in both anaplastic lymphoma kinase (ALK)+ and ALK- anaplastic large-cell lymphoma. *Proc Natl Acad Sci USA* 2010;107:16228–33.
59. Bartel DP. MicroRNAs: genomics, biogenesis, mechanism, and function. *Cell* 2004;116:281–97.
60. Bartel DP. MicroRNAs: target recognition and regulatory functions. *Cell* 2009;136:215–33.
61. Asslaber D, Piñón JD, Seyfried I, Desch P, Stöcher M, Tinhofer I, et al. microRNA-34a expression correlates with MDM2 SNP309 polymorphism and treatment-free survival in chronic lymphocytic leukemia. *Blood* 2010;115:4191–7.
62. Costinean S, Zanoni N, Pekarsky Y, Tili E, Volinia S, Heerema N, et al. Pre-B cell proliferation and lymphoblastic leukemia/high-grade lymphoma in E(mu)-miR155 transgenic mice. *Proc Natl Acad Sci USA* 2006;103:7024–9.
63. Rodriguez A, Vigorito E, Clare S, Warren MV, Couttet P, Soond DR, et al. Requirement of bic/microRNA-155 for normal immune function. *Science* 2007;316:608–11.
64. Thai TH, Calado DP, Casola S, Ansel KM, Xiao C, Xue Y, et al. Regulation of the germinal center response by microRNA-155. *Science* 2007;316:604–8.
65. Finn OJ. Cancer immunology. *N Engl J Med* 2008;358:2704–2715.
66. Koebel CM, Vermi W, Swann JB, Zerafa N, Rodig SJ, Old LJ, et al. Adaptive immunity maintains occult cancer in an equilibrium state. *Nature* 2007;450:903–907.

67. Dunn GP, Bruce AT, Ikeda H, Old LJ, Schreiber RD. Cancer immunoeediting: from immunosurveillance to tumor escape. *Nat Immunol* 2002;3:991–998.
68. Rice J, Ottensmeier CH, Stevenson FK. DNA vaccines: precision tools for activating effective immunity against cancer. *Nat Rev Cancer* 2008;8:108–20.
69. Pulford K, Falini B, Banham AH, Codrington D, Robertson H, Hatton C, et al. Immune response to the ALK oncogenic tyrosine kinase in patients with anaplastic large-cell lymphoma. *Blood* 2000;96:1605–7.
70. Ait-Tahar K, Damm-Welk C, Burkhardt B, Zimmermann M, Klapper W, Reiter A, et al. Correlation of the autoantibody response to the ALK oncoantigen in pediatric anaplastic lymphoma kinase-positive anaplastic large cell lymphoma with tumor dissemination and relapse risk. *Blood* 2010;115:3314–9.
71. Passoni L, Scardino A, Bertazzoli C, Gallo B, Coluccia AM, Lemonnier FA, et al. ALK as a novel lymphoma-associated tumor antigen: identification of 2 HLA-A2.1-restricted CD8<sup>+</sup> T-cell epitopes. *Blood* 2002;99:2100–6.
72. Chiarle R, Martinengo C, Mastini C, Ambrogio C, D'Escamard V, Forni G, et al. The anaplastic lymphoma kinase is an effective oncoantigen for lymphoma vaccination. *Nat Med* 2008;14:676–80.
73. Feldman AL, Dogan A, Smith DI, Law ME, Ansell SM, Johnson SH, et al. Discovery of recurrent t(6;7)(p25.3;q32.3) translocations in ALK-negative anaplastic large cell lymphomas by massively parallel genomic sequencing. *Blood* 2011;117:915–9.
74. Gunby RH, Tartari CJ, Porchia F, Donella-Deana A, Scapozza L, Gambacorti-Passerini C. An enzyme-linked immunosorbent assay to screen for inhibitors of the oncogenic anaplastic lymphoma kinase. *Haematologica* 2005;90:988–90.
75. Chen Z, Sasaki T, Tan X, Carretero J, Shimamura T, Li D, et al. Inhibition of ALK, PI3K/MEK, and HSP90 in murine lung adenocarcinoma induced by EML4-ALK fusion oncogene. *Cancer Res* 2010;70:9827–36.
76. McDermott U, Iafrate AJ, Gray NS, Shioda T, Classon M, Maheswaran S, et al. Genomic alterations of anaplastic lymphoma kinase may sensitize tumors to anaplastic lymphoma kinase inhibitors. *Cancer Res* 2008;68:3389–95.
77. Druker BJ. Translation of the Philadelphia chromosome into therapy for CML. *Blood* 2008;112:4808–17. doi:10.1182/blood-2008-07-077958.
78. Pulford K, Morris SW, Turturro F. Anaplastic lymphoma kinase proteins in growth control and cancer. *J Cell Physiol* 2004;199:330–58.