

Early Life Body Fatness, Serum Anti-Müllerian Hormone, and Breast Density in Young Adult Women

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Abstract

Background: Emerging evidence suggests positive associations between serum anti-Müllerian hormone (AMH), a marker of ovarian function, and breast cancer risk. Body size at young ages may influence AMH levels, but few studies have examined this. Also, no studies have examined the relation of AMH levels with breast density, a strong predictor of breast cancer risk.

Methods: We examined associations of early life body fatness, AMH concentrations, and breast density among 172 women in the Dietary Intervention Study in Children (DISC). Height and weight were measured at baseline (ages 8–10) and throughout adolescence. Serum AMH concentrations and breast density were assessed at ages 25–29 at the DISC 2006 Follow-up visit. We used linear mixed effects models to quantify associations of AMH (dependent variable) with quartiles of age-specific youth body mass index (BMI) Z-scores (independent variable). We assessed

cross-sectional associations of breast density (dependent variable) with AMH concentration (independent variable).

Results: Neither early life BMI nor current adult BMI was associated with AMH concentrations. There were no associations between AMH and percent or absolute dense breast volume. In contrast, women with higher AMH concentrations had significantly lower absolute nondense breast volume ($P_{\text{trend}} < 0.01$).

Conclusions: We found no evidence that current or early life BMI influences AMH concentrations in later life. Women with higher concentrations of AMH had similar percent and absolute dense breast volume, but lower nondense volume.

Impact: These results suggest that AMH may be associated with lower absolute nondense breast volume; however, future prospective studies are needed to establish temporality. *Cancer Epidemiol Biomarkers Prev*; 25(7); 1151–7. ©2016 AACR.

Introduction

Childhood and adolescent adiposity is inversely related to breast cancer risk across the life course (1–8). We previously reported strong inverse associations of body fatness during childhood and adolescence with percent breast density, a strong predictor of breast cancer risk, in young adulthood (9). Breast density refers to the proportion of fibroglandular tissue (vs.

adipose tissue) in the breast (10). The biologic mechanism by which body fatness in youth influences breast density and subsequent cancer risk is not well established. As most breast development occurs during puberty, body fatness during this time period could have an important impact on breast morphology and breast density later in life, directly influencing breast cancer risk through this pathway.

Body fatness during childhood and adolescence could also decrease breast cancer risk via effects on ovarian function. Specifically, obesity suppresses ovarian function, leading to fewer ovulatory menstrual cycles and altered circulating levels of hormones in adolescent and premenopausal women (11, 12). Anti-Müllerian hormone (AMH) is an important marker of ovarian function; it is secreted by the ovaries starting in the prepubertal period and plays an important role in the recruitment and growth of follicles, and in the regulation of normal breast development and involution (13). In three recent prospective studies, strong positive associations between serum AMH concentrations and breast cancer risk were observed (14–16), supporting a role for AMH in breast cancer development.

Given the critical role of AMH in ovarian function, and previously demonstrated associations of AMH with breast cancer risk, it is plausible that body size at young ages may influence AMH levels. Inverse associations of AMH with adiposity have been reported in some studies (17–20), whereas others showed no clear associations (17, 18, 20–28). Furthermore, few studies have examined the possible role of adiposity earlier in life. In addition, no prior studies have examined the relation of AMH levels with

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breast density. We hypothesized that body fatness during childhood and adolescence would be inversely associated with AMH levels in young adult women and that AMH levels would be positively associated with high breast density. We examined these associations using prospectively-collected data from the Dietary Intervention Study in Children (DISC) and the DISC 2006 (DISC06) Follow-up Study. Recent evidence suggests that absolute dense and nondense areas may have independent effects on cancer risk (29, 30); therefore, we also evaluated these phenotypes separately.

Materials and Methods

Study population

The original DISC was a multicenter, randomized controlled trial to examine the safety and efficacy of a dietary intervention to reduce serum low-density lipoprotein cholesterol (LDL-C) in children (31–34). Briefly, between 1988 and 1990, 663 healthy, prepubertal 8- to 10-year-old children, including 301 girls, with elevated LDL-C were recruited to six clinical centers and randomized to a behavioral dietary intervention or usual care control group. Children participated in annual clinic visits until the trial was terminated in 1997, when the average age of participants was 16.7 years, due to a lack of treatment effect on LDL-C. At the time of the original DISC, assent was obtained from participants; their parents/guardians provided informed consent before randomization. All female DISC participants were invited to participate in the DISC06 Follow-up Study, and 260 (86.4%) of the 301 females originally randomized in DISC participated (35). Participants were recontacted before the DISC06 Follow-up Study, which took place in 2006–2008 when participants were 25–29 years old. The original DISC protocol was approved by an *National Heart, Lung, and Blood Institute* (NHLBI)-appointed independent data and safety monitoring committee and institutional review boards (IRB) at all participating clinical centers and the data coordinating center. The DISC06 Follow-up Study protocol was approved by IRBs at the Fox Chase Cancer Center, participating clinical centers, and the data coordinating center (please see Acknowledgments).

Participants in this analysis included 172 women who enrolled in the original DISC between 1988 and 1990, when they were ages 8–10 years, and also participated in the DISC06 Follow-up Study, when they were ages 25–29 years. Women who were pregnant or breastfeeding within 12 weeks ($n = 30$) before their clinic visit, had breast implants or reduction surgery ($n = 16$), or whose MRI was missing or of poor quality ($n = 32$) were excluded. Ten women without AMH measurements were also excluded.

Data collection

During the original DISC, height and weight were measured at baseline and annual clinic visits by trained study staff blinded to treatment assignment. Specifically, height was measured using a stadiometer and weight was measured on an electronic or beam balance scale. Each measurement was made twice. A third measurement was taken if the first two measurements were not within allowable tolerances (0.5 cm for height and 0.2 kg for weight) and the two closest values were averaged. BMI was calculated as weight (kg)/height (m^2). Because the interpretation of BMI in children and adolescents is specific to age and sex, we expressed the age-specific BMI as a Z-score relative to Centers for Disease Control (CDC) 2000 Growth Charts for girls (36). Information

on demographic and other characteristics, including medical history, reproductive factors, was ascertained on annual questionnaires, whereas diet and physical activity were assessed at baseline and years 1, 3, 5, and last childhood visits (35, 37). DISC06 follow-up visits took place at the original six DISC clinics and were scheduled during the luteal phase of the menstrual cycle when possible (>85% of visits occurred within 14 days of onset of next menses). Medical history and other information was updated at the DISC06 follow-up visit.

Blood collection and laboratory assays

Blood samples were collected at the DISC06 follow-up visit in the morning after an overnight fast by venipuncture using standard procedures. Blood was allowed to stand at room temperature for 45 minutes to allow complete clotting. Blood was then centrifuged and serum was separated and pipetted in 0.5-mL aliquots into cryovials, which were labeled and stored at -80°C .

AMH was measured in serum using a single lot of picoAMH ELISA Kit (AL-124, Ansh Labs; ref. 38). The limit of detection (LOD) of the picoAMH ELISA is 1.2 pg/mL; no participant samples were below the LOD. The manufacturer-specified inter-assay coefficients of variation (CV) are 4.5%, 2.2%, and 3.8% at 22.6, 86.5, and 373 pg/mL, respectively, for the picoAMH assays. Internal quality control samples included in the assay run resulted in CVs of 3.2% and 4.0% at mean AMH concentrations of 91.2 and 285 pg/mL, respectively. Finally, four participant samples were run in duplicate with mean CV of 4.7%.

Breast density assessment

At the DISC06 Follow-up visit, breast density was assessed following a standardized protocol by noncontrast MRI. Equipment standards at each site were consistent with American College of Radiology guidelines for breast MRI (39) and required that imaging be performed using a whole-body MRI scanner of 1.5 T or higher field strength and a dedicated breast imaging radiofrequency coil. A standard image-acquisition protocol was prescribed consisting of two pulse sequences performed in both the transaxial and coronal orientations with a 32 to 40 cm field of view for bilateral coverage: a three-dimensional fast gradient echo sequence without fat suppression, and a three-dimensional fast gradient echo sequence with fat suppression.

To ensure accuracy and uniformity of data acquisition at the different clinical centers, MRI technologists at the sites were trained (by C. Klifa) to recognize and correct failures due to incomplete fat suppression, motion artifacts, and inadequate breast coverage. In addition, acceptable image quality on three volunteers was required for site certification. Participant scans that were inaccurate due to artifacts, motion, or technique were excluded ($n = 21$).

All MRI image data were processed at the University of California, San Francisco (San Francisco, CA) by C. Klifa using customized software to identify the chest wall–breast tissue boundary and skin surface, and to separate breast fibroglandular and fatty tissue using a segmentation method based on fuzzy C-means clustering (40). Total volumes of fibroglandular and fatty tissue were computed separately for each breast and averaged for analysis. Outcomes of interest were percent dense breast tissue volume (ratio of fibroglandular volume to total volume of the breast), absolute (total) volume of dense tissue (fibroglandular volume), and absolute (total) volume of nondense tissue (fatty breast tissue volume).

Statistical analyses

We used linear mixed effects models to examine the association between body mass index (BMI) Z-scores at baseline (ages 8–10 years; predictor) and AMH at ages 25–29 years (outcome). The analysis was repeated, changing the predictor to BMI Z-score at each annual clinic visit during childhood and adolescence; for simplicity, we present results for baseline, year 3, year 5, and last childhood visits. For our predictor, body fatness during childhood, we created quartiles of age- and female-specific BMI Z-scores computed from CDC Growth Charts (36). We also assessed the cross-sectional association between BMI in kg/m² at the follow-up visit and AMH concentration at the same visit.

To improve normality of the outcome, we applied a natural log transformation to measured AMH concentration. We used the generalized extreme Studentized deviate many-outlier procedure (41) to formally identify potential outliers in AMH values; none were identified. Linear mixed effects models were fit by maximum likelihood with clinic included as a random effect and empirical (robust) SEs to allow for correlated outcomes within clinics. The associations between quartiles of childhood BMI Z-scores and adult AMH concentration were quantified by adjusted least square means and 95% confidence intervals (CI). Because of the log transformation, we applied Duan "smearing estimate" to back-transform AMH estimates to original (untransformed) units of ng/mL; this method adjusts for bias arising in retransformation of estimates from nonparametric or generalized linear models (42). Tests for trend were based on models including childhood or adolescent BMI Z-score (or current BMI) as a continuous variable. All models were adjusted for clinic as a random effect and treatment assignment as a fixed effect. Subsequent models additionally controlled for current BMI and current BMI-squared, to allow for a possible curvilinear association with BMI, (Model 2) and age at menarche, duration of hormone use, number of live births, race, education, alcohol consumption, smoking status, and family history of breast cancer (Model 3). Categorical covariates were categorized as shown in Table 1.

The same analytic approach (i.e., linear mixed effects models to estimate least square means and 95% CIs) was used to examine the cross-sectional associations of quartiles of log-transformed AMH (now as the independent variable) concentrations with measures of breast density (i.e., percent dense breast volume, absolute dense breast volume, and absolute nondense breast volume; dependent variable). For the absolute measures of breast density, we applied natural log transformation and backtransformed estimates to original units of cm³, as described above. Multivariable models were adjusted for the same covariates listed previously. Further adjustment for menstrual cycle day of blood collection or testosterone levels did not appreciably change the beta coefficients; therefore, these variables were not retained in the final model. All analyses were carried out using SAS 9.3.

Results

Women were on average 27 years old at the DISC06 Follow-up visit, with mean BMI 25.4 kg/m², mean percent dense breast volume 27.4%, and mean AMH concentration 4.2 ng/mL. The majority of women were white, educated, nulliparous, and were past or current users of hormonal contraceptives (Table 1). Participant characteristics were generally similar across quartiles of AMH levels; however, women in the lowest quartile of AMH had lower percent dense breast volume (23.5%) compared with

those in the top quartile (29.1%), primarily reflecting greater nondense breast volume in the lowest quartile group. Women in the lowest quartile of AMH were also somewhat more likely to be parous and to be current users of hormonal contraceptives compared with the other three quartiles (Table 1).

Neither early life BMI nor current adult BMI were associated with AMH concentrations in this population. In fully adjusted multivariable models, adjusted mean AMH concentrations were 2.8, 3.1, 3.9, and 3.0 ng/mL for successive quartiles of BMI Z-score at baseline clinic visit (ages 8–10; $P_{\text{trend}} = 0.62$). Similarly, null results were observed for BMI assessed at the other clinic visits during childhood and adolescence, as well as for current adult BMI at the DISC06 Follow-up visit (Table 2).

In cross-sectional analyses adjusting for clinic and treatment assignment only, there was a suggestive, but nonsignificant, positive association between AMH and percent dense breast volume. Mean percent dense breast volume was 23.5% (95% CI, 16.3–30.7) in the lowest quartile of AMH versus 29.3% (95% CI, 19.6–30.0) in the highest quartile ($P_{\text{trend}} = 0.24$). However, the apparent association was substantially attenuated upon adjustment for current BMI, a strong negative predictor of percent dense breast volume. In the fully adjusted multivariable model, the corresponding means for percent breast density were 25.8% (95% CI, 20.1–31.5) and 25.8% (95% CI, 20.0–31.6; $P_{\text{trend}} = 0.54$). Similarly, there was no apparent association of AMH concentration with absolute dense breast volume. Women with higher AMH concentrations, however, had significantly lower absolute nondense breast volume after controlling for predictors of breast density and absolute dense volume. Specifically, mean absolute nondense breast volume was 328.7 cm³ (95% CI, 275.8–391.7) among women in the lowest quartile of AMH compared with 280.6 cm³ (95% CI, 231.8–339.6) for the highest quartile ($P_{\text{trend}} < 0.01$; Table 3).

Results were similar among nulliparous women ($n = 122$) and parous women ($n = 50$) and among current/former hormonal contraceptive users ($n = 161$; data not shown). There were too few women ($n = 11$) reporting never using hormonal contraceptives for meaningful subanalysis in this group. Finally, there was no significant interaction between AMH and treatment group assignment for any of the density phenotypes evaluated ($P_{\text{interaction}} > 0.05$).

Discussion

In summary, we found no evidence that earlier life BMI influences AMH concentrations in young adulthood. Similarly, current BMI was not associated with AMH concentrations. We also found no association between AMH and percent or absolute dense breast volume measured concurrently; however, AMH was inversely associated with absolute nondense breast volume.

Early life, including childhood and adolescence, is hypothesized to be a critical time window for breast carcinogenesis (43). This is a time of rapid growth and development, with especially high rates of mammary gland cell proliferation during puberty, which could increase vulnerability to molecular damage and explain why exposures during this time period might be important for breast density and breast cancer risk later in life (43–45). An inverse association between early life adiposity and breast cancer risk is well established (1–8). Our previous analyses in this population demonstrated an inverse association between youth adiposity and dense breast volume (9), in agreement with several

Table 1. Participant characteristics at the DISCO6 follow-up visit, overall and by quartile of AMH

Descriptive characteristics	Overall		Q1	Q2	Q3	Q4
	n	Mean (SD) or %	n = 43	n = 43	n = 43	n = 43
Age, y	172	27.2 (1.0)	27.2 (1.0)	27.1 (1.1)	27.2 (1.0)	27.2 (1.0)
Percent dense breast volume (%)	172	27.4 (20.0)	23.5 (19.8)	26.7 (19.9)	30.0 (19.6)	29.1 (20.7)
Absolute dense breast volume (cm ³)	172	104.0 (70.3)	101.0 (73.0)	110.5 (69.2)	110.1 (83.0)	94.4 (51.0)
Absolute nondense breast volume (cm ³)	172	418.7 (369.3)	465.6 (348.9)	459.4 (426.6)	362.7 (327.2)	387.0 (369.3)
AMH (ng/mL)	172	4.21 (3.28)	1.2 (0.4)	2.6 (0.5)	4.3 (0.6)	8.7 (3.2)
BMI (kg/m ²)	172	25.4 (5.4)	25.8 (5.0)	26.0 (6.2)	25.1 (4.6)	24.9 (5.6)
Height (cm)	172	165.4 (6.4)	163.6 (5.8)	166.6 (6.1)	165.0 (7.0)	166.2 (6.3)
BMI Z-score at 8–10 years old	172	0.23 (0.90)	0.28 (1.0)	0.35 (0.9)	0.15 (0.8)	0.15 (0.9)
Age at menarche, y	172	12.9 (1.3)	12.6 (1.1)	13.0 (1.3)	12.9 (1.3)	13.1 (1.4)
Duration of hormonal contraceptive use (y) ^a	161	5.6 (3.5)	6.7 (3.1)	5.6 (3.9)	4.9 (3.6)	5.3 (3.1)
Menstrual cycle length (days) ^b	71	27.9 (2.6)	26.1 (3.9)	27.7 (2.1)	28.9 (1.6)	28.2 (2.6)
Race						
White	155	90.1%	95.3%	93.0%	86.0%	86.0%
Non-white	17	9.9%	4.7%	7.0%	14.0%	14.0%
Education						
High school, vocational, or technical school	17	9.9%	9.3%	9.3%	9.3%	11.6%
Some college	41	23.8%	30.2%	14.0%	30.2%	20.9%
College/Bachelor's	92	53.5%	53.5%	72.1%	39.5%	48.8%
Graduate school	22	12.8%	7.0%	4.7%	20.9%	18.6%
Number of live births						
0	122	70.9%	60.5%	76.7%	72.1%	74.4%
1	29	16.9%	23.3%	18.6%	9.3%	16.3%
2+	21	12.2%	16.3%	4.7%	18.6%	9.3%
Ever breast fed (among parous)						
Yes	38	74.5%	88.2%	60.0%	66.7%	81.8%
No	13	25.5%	11.8%	40.0%	33.3%	18.2%
Hormonal contraceptive use						
Never	11	6.4%	4.7%	4.7%	9.3%	7.0%
Former	62	36.1%	20.9%	39.5%	37.2%	46.5%
Current	99	57.6%	74.4%	55.8%	53.5%	46.5%
Family history of breast cancer						
Yes	7	4.1%	0.0%	7.1%	7.1%	2.4%
No	161	95.9%	100.0%	92.9%	92.9%	97.6%
Alcohol consumption						
Never/former	15	8.7%	11.6%	9.3%	4.7%	9.3%
Current, <3 per week	67	39.0%	32.6%	51.2%	39.5%	32.6%
Current, 3–<6 per week	32	18.6%	27.9%	20.9%	14.0%	11.6%
Current, 6–<10 per week	37	21.5%	14.0%	14.0%	32.6%	25.6%
Current, 10+ per week	21	12.2%	14.0%	4.7%	9.3%	20.9%
Smoking history						
Never	94	54.7%	51.2%	62.8%	53.5%	51.2%
Former	36	20.9%	25.6%	11.6%	23.3%	23.3%
Current	42	24.4%	23.3%	25.6%	23.3%	23.3%
Treatment assignment						
Intervention	86	50.0%	51.2%	44.2%	60.5%	44.2%
Usual care	86	50.0%	48.8%	55.8%	39.5%	55.8%

^aAmong current or former hormonal contraceptive users.

^bAmong women not using hormonal contraceptives.

other studies of this association (46–51). Given that obesity is known to influence ovarian function (11, 12), body fatness during childhood and adolescence could also decrease breast cancer risk by influencing AMH levels. However, our null results do not support this hypothesis. Similarly, in a birth cohort study of >1,300 adolescent girls (mean age 15.5), neither birth weight nor current BMI were associated with AMH levels (52, 53). Cross-sectional analyses in Chinese girls also reported no associations of BMI in childhood (ages 0–10 years) or adolescence (ages 11–18 years) with AMH measured concurrently (54). In contrast, in a study of adolescent girls who were normal ($n = 43$), oligomenorrheic ($n = 27$), or had polycystic ovarian syndrome ($n = 150$), AMH levels were significantly lower in obese compared with nonobese girls within each group (55). Another study of 10 normal weight and 10 obese ovulatory young women between ages 18 and 35, AMH levels were 34% lower in the obese group

(17). AMH levels peak in the mid-20s and subsequently decline with age, becoming nondetectable by menopause (15, 19, 56, 57). As participants in the DISCO6 Follow-up Study were ages 25–29 years and consequently had relatively high AMH concentrations, there may have been insufficient variation at low concentrations in AMH levels to observe associations with current or earlier life BMI.

Epidemiologic evidence for associations between current adult BMI and serum AMH levels is mixed. Similar to our findings, many studies have not found an association between current BMI and serum AMH levels (21–26); however, some studies have reported lower AMH levels among obese women (17–20). In light of these conflicting results, some researchers have suggested that inverse associations between BMI and AMH in some studies may reflect residual confounding by age (which is inversely related to AMH; ref. 22) or the phenomenon of

Table 2. Mean and 95% CI for AMH concentration (ng/mL) at the DISCO6 follow-up visit according to quartile of age-specific BMI Z-score

Quartile of age-specific BMI Z-score		Q1	Q2	Q3	Q4	P ^a
BMI at baseline visit (ages 8–10) <i>n</i> = 172	Model 1	3.33 (2.48–4.45)	2.93 (2.38–3.60)	3.39 (2.58–4.46)	2.90 (2.33–3.61)	0.43
	Model 2	3.14 (2.46–4.02)	2.90 (2.41–3.47)	3.48 (2.52–4.82)	3.02 (2.31–3.96)	0.90
	Model 3	2.82 (2.18–3.64)	3.06 (2.74–3.43)	3.85 (2.62–5.70)	2.98 (2.46–3.61)	0.62
BMI at year 3 visit (ages 11–13) <i>n</i> = 161	Model 1	3.57 (2.67–4.77)	3.10 (2.60–3.69)	2.76 (2.01–3.80)	2.93 (2.39–3.56)	0.52
	Model 2	3.53 (2.70–4.62)	3.13 (2.65–3.69)	2.78 (1.95–4.00)	2.90 (2.25–3.73)	0.78
	Model 3	3.32 (2.44–4.52)	3.28 (2.87–3.74)	2.88 (2.01–4.13)	2.87 (2.24–3.67)	0.80
BMI at year 5 visit (ages 13–15) <i>n</i> = 146	Model 1	3.40 (2.82–4.10)	3.21 (2.25–4.59)	2.68 (2.03–3.54)	3.13 (2.41–4.06)	0.44
	Model 2	3.22 (2.44–4.26)	3.11 (2.25–4.30)	2.81 (1.92–4.13)	3.23 (2.27–4.60)	0.80
	Model 3	2.68 (1.98–3.62)	3.28 (2.48–5.09)	3.27 (2.10–5.09)	3.22 (2.19–4.75)	0.62
BMI at last visit (ages 15–17) <i>n</i> = 149	Model 1	3.78 (2.62–4.43)	2.57 (2.02–3.26)	2.68 (2.11–3.41)	3.12 (2.23–4.37)	0.09
	Model 2	3.42 (2.43–4.81)	2.52 (2.00–3.17)	2.85 (2.17–3.74)	3.30 (2.18–4.98)	0.06
	Model 3	2.98 (2.13–4.17)	2.62 (2.07–3.31)	3.29 (2.55–4.25)	3.14 (2.03–4.85)	0.77
BMI ^b at follow-up visit (ages 25–29) <i>n</i> = 172	Model 1	3.50 (2.50–4.91)	2.84 (2.20–3.66)	3.38 (3.11–3.67)	2.85 (2.27–3.56)	0.47
	Model 2	3.53 (2.43–5.11)	3.12 (2.51–3.89)	3.48 (3.04–3.98)	2.58 (1.86–3.58)	0.48
	Model 3	3.53 (2.43–5.11)	3.12 (2.51–3.89)	3.48 (3.04–3.98)	2.58 (1.86–3.58)	0.48

NOTE: Model 1 means estimated from linear mixed effects models including clinic as a random effect and treatment group as a fixed effect. Model 2 means estimated from linear mixed effects models including clinic as a random effect and adjusted for treatment group and current BMI (continuous, kg/m² and squared BMI) as fixed effects. Model 3 means estimated from linear mixed effects models including clinic as a random effect and adjusted for treatment group, current BMI (continuous, kg/m² and squared BMI), number of live births, duration of hormone use, age at menarche, race, education, alcohol consumption, smoking status, and family history of breast cancer, as fixed effects (4 missing).

^aTest for trend.

^bQuartiles of BMI in kg/m² (untransformed); cut-points are Q1, <21.4; Q2, 21.5–24.1; Q3, 24.1–28.7; Q4, >28.7.

serum hormone dilution due to increased blood volume among larger women (21).

To date, only five studies have evaluated AMH and breast cancer risk. Of 30 women undergoing breast biopsy, 22 were determined to have cancer or precancer and these women had significantly lower AMH levels than those with benign breast disease (*P* < 0.001; ref. 58). However, in a case–control study among women ages 28–44 years (108 cases, 99 controls), there was no significant difference in AMH levels by case status (59). Neither of these studies measured AMH prior to breast cancer diagnosis, thus the influence of disease could not be ruled out. The first prospective analysis to evaluate this association (*n* = 105 cases, 204 controls) showed a strong positive association between serum AMH levels and breast cancer risk (*P*_{trend} < 0.001; ref. 14). These findings were recently supported by two case–control studies nested within the Sister Study cohort (*n* = 452 cases, 902 controls; ref. 15) and the Nurses' Health Study II (*n* = 539 cases, 471 controls; ref. 16), both of which also demonstrated a significant positive association

between AMH concentrations and breast cancer risk, with a more than 2-fold increased risk among women with the highest AMH concentrations compared with those with AMH <LOD.

To our knowledge, no previous studies have evaluated possible associations of AMH with breast density. An important limitation of this analysis is its cross-sectional design: AMH and breast density were measured concurrently, limiting inferences about a possible temporal association. Also, because the original DISCO study population excluded children whose weight-for-height was greater than the 90th percentile or lower than the 5th percentile at baseline (31), our findings may not be generalizable to very lean or very obese children. Major strengths of this study include the objective and repeated measures of childhood and adolescent weight and height. In addition, while confounding by unmeasured factors cannot be ruled out, we had detailed questionnaire information and the ability to adjust for many potential confounders. We used an ultrasensitive AMH assay with demonstrated reproducibility. Similarly, systematic measurement of breast

Table 3. Mean and 95% CI for three breast density phenotypes at the DISCO6 follow-up visit according to quartile of AMH (*n* = 172)

Quartile of AMH	Q1	Q2	Q3	Q4	P ^a
Percent dense breast volume					
Model 1	23.5 (16.3–30.7)	26.9 (23.0–30.9)	29.7 (22.8–36.6)	29.3 (19.6–39.0)	0.24
Model 2	25.5 (18.4–32.5)	27.5 (24.8–30.2)	29.8 (25.2–34.3)	26.6 (25.2–34.3)	0.36
Model 3	25.8 (20.1–31.5)	27.9 (25.4–31.3)	29.6 (25.7–33.4)	25.8 (20.0–31.6)	0.54
Absolute dense breast volume (cm ³)					
Model 1	69.1 (54.2–88.0)	88.9 (81.2–97.3)	84.6 (68.3–104.7)	76.3 (50.7–114.9)	0.46
Model 2	72.0 (56.2–92.1)	90.0 (77.6–104.5)	85.1 (69.0–104.9)	73.2 (50.2–106.7)	0.68
Model 3 ^b	70.3 (53.9–91.7)	90.0 (73.0–111.1)	89.2 (70.4–113.0)	72.0 (55.0–94.2)	0.55
Absolute non-dense breast volume (cm ³)					
Model 1	356.8 (287.4–442.9)	323.4 (256.7–407.4)	258.8 (256.7–407.4)	247.2 (188.7–323.9)	0.03
Model 2	335.2 (275.3–408.3)	307.2 (282.6–334.0)	263.2 (220.4–314.3)	272.9 (232.3–320.6)	<0.01
Model 3 ^c	328.7 (275.8–391.7)	303.7 (278.4–331.2)	267.4 (234.2–305.3)	280.6 (231.8–339.6)	<0.01

NOTE: Model 1 least square means estimated from linear mixed effects models including clinic as a random effect and treatment group as a fixed effect. Model 2 least square means estimated from linear mixed effects models including clinic as a random effect and adjusted for treatment group and current adult BMI (continuous, kg/m²) as fixed effects. Model 3 least square means estimated from linear mixed effects models including clinic as a random effect and adjusted for treatment group, current adult BMI (continuous, kg/m²), number of live births, duration of hormone use, age at menarche, race, education, alcohol consumption, smoking status, and family history of breast cancer, as fixed effects (4 missing).

^aTest for trend.

^bAdditionally adjusted for log-non-dense volume.

^cAdditionally adjusted for log-dense volume.

density via MRI afforded us the ability to consider dense versus nondense breast volume separately.

While we observed no apparent association between AMH concentrations and percent or absolute dense breast volume in cross-sectional analyses, our results suggest that higher AMH levels may be associated with lower absolute nondense breast volume. Considering recent evidence that suggests higher amounts of nondense breast tissue may be inversely associated with breast cancer risk, independent of absolute dense breast tissue amount (29, 30), our findings, if confirmed in future studies, could be consistent with the hypothesis that AMH is associated with increased breast cancer risk. However, future prospective studies are needed to establish temporality of associations of AMH with breast density.

Disclosure of Potential Conflicts of Interest

E.S. LeBlanc reports receiving commercial research grants from Amgen, AstraZeneca, Bristol-Myers Squibb, and Merck. No potential conflicts of interest were disclosed by the other authors.

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Acquisition of data (provided animals, acquired and managed patients, provided facilities, etc.): C. Klifa, N.M. Hylton, L.G. Snetselaar, L. Van Horn
Analysis and interpretation of data (e.g., statistical analysis, biostatistics, computational analysis): K.A. Bertrand, H.J. Baer, E.J. Orav, J.F. Dorgan
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References

- Ahlgren M, Melbye M, Wohlfahrt J, Sorensen TI. Growth patterns and the risk of breast cancer in women. *N Engl J Med* 2004;351:1619–26.
- Baer HJ, Colditz GA, Rosner B, Michels KB, Rich-Edwards JW, Hunter DJ, et al. Body fatness during childhood and adolescence and incidence of breast cancer in premenopausal women: a prospective cohort study. *Breast Cancer Res* 2005;7:R314–25.
- Michels KB, Terry KL, Willett WC. Longitudinal study on the role of body size in premenopausal breast cancer. *Arch Intern Med* 2006;166:2395–402.
- Ruder EH, Dorgan JF, Kranz S, Kris-Etherton PM, Hartman TJ. Examining breast cancer growth and lifestyle risk factors: early life, childhood, and adolescence. *Clin Breast Cancer* 2008;8:334–42.
- Bardia A, Vachon CM, Olson JE, Vierkant RA, Wang AH, Hartmann LC, et al. Relative weight at age 12 and risk of postmenopausal breast cancer. *Cancer Epidemiol Biomarkers Prev* 2008;17:374–8.
- Baer HJ, Tworoger SS, Hankinson SE, Willett WC. Body fatness at young ages and risk of breast cancer throughout life. *Am J Epidemiol* 2010;171:1183–94.
- Bandera EV, Chandran U, Zirpoli G, Ciupak G, Bovbjerg DH, Jandorf L, et al. Body size in early life and breast cancer risk in African American and European American women. *Cancer Causes Control* 2013;24:2231–43.
- Andersen ZJ, Baker JL, Bihmann K, Vejborg I, Sorensen T, Lyng E. Birth weight, childhood body mass index, and height in relation to mammographic density and breast cancer: a register-based cohort study. *Breast Cancer Res* 2014;16:R4.
- Bertrand KA, Baer HJ, Orav EJ, Klifa C, Shepherd JA, Van Horn L, et al. Body fatness during childhood and adolescence and breast density in young women: a prospective analysis. *Breast Cancer Res* 2015;17:95.
- Yaffe MJ. Mammographic density. Measurement of mammographic density. *Breast Cancer Res* 2008;10:209.
- Apter D, Vihko R. Endocrine determinants of fertility: serum androgen concentrations during follow-up of adolescents into the third decade of life. *J Clin Endocrinol Metab* 1990;71:970–4.
- Caprio S, Hyman LD, Limb C, McCarthy S, Lange R, Sherwin RS, et al. Central adiposity and its metabolic correlates in obese adolescent girls. *Am J Physiol* 1995;269:E118–26.
- Nakhuda GS. The role of mullerian inhibiting substance in female reproduction. *Curr Opin Obstet Gynecol* 2008;20:257–64.
- Dorgan JF, Stanczyk FZ, Egleston BL, Kahle LL, Shaw CM, Spittle CS, et al. Prospective case-control study of serum mullerian inhibiting substance and breast cancer risk. *J Natl Cancer Inst* 2009;101:1501–9.
- Nichols HB, Baird DD, Stanczyk FZ, Steiner AZ, Troester MA, Whitworth KW, et al. Anti-Müllerian hormone concentrations in premenopausal women and breast cancer risk. *Cancer Prev Res* 2015;8:528–34.
- Eliassen AH, Zeleniuch-Jacquotte A, Rosner B, Hankinson SE. Plasma anti-Müllerian hormone concentrations and risk of breast cancer among premenopausal women in the Nurses' Health Studies. *Cancer Epidemiol Biomarkers Prev*. 2016Mar 9. [Epub ahead of print].
- Steiner AZ, Stanczyk FZ, Patel S, Edelman A. Antimüllerian hormone and obesity: insights in oral contraceptive users. *Contraception* 2010;81:245–8.
- Su HI, Sammel MD, Freeman EW, Lin H, DeBlasis T, Gracia CR. Body size affects measures of ovarian reserve in late reproductive age women. *Menopause* 2008;15:857–61.
- Freeman EW, Sammel MD, Lin H, Gracia CR. Anti-müllerian hormone as a predictor of time to menopause in late reproductive age women. *J Clin Endocrinol Metab* 2012;97:1673–80.
- Moy V, Jindal S, Lieman H, Buyuk E. Obesity adversely affects serum anti-müllerian hormone (AMH) levels in Caucasian women. *J Assist Reprod Genet* 2015;32:1305–11.
- Lambert-Messerlian G, Plante B, Eklund EE, Raker C, Moore RC. Levels of antimüllerian hormone in serum during the normal menstrual cycle. *Fertil Steril* 2016;105:208–13.
- La Marca A, Sighinolfi G, Giulini S, Taglia M, Argento C, Sala C, et al. Normal serum concentrations of anti-Müllerian hormone in women with regular menstrual cycles. *Reprod Biomed Online* 2010;21:463–9.

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23. Halawaty S, Elkattan E, Azab H, ElGhamry N, Al-Inany H. Effect of obesity on parameters of ovarian reserve in premenopausal women. *J Obstet Gynaecol Can* 2010;32:687–90.
24. Dölleman M, Verschuren WM, Eijkemans MJ, Dollé ME, Jansen EH, Broekmans FJ, et al. Reproductive and lifestyle determinants of anti-Müllerian hormone in a large population-based study. *J Clin Endocrinol Metab* 2013;98:2106–15.
25. Sahmay S, Usta T, Erel CT, Imamoğlu M, Küçük M, Atakul N, et al. Is there any correlation between amh and obesity in premenopausal women? *Arch Gynecol Obstet* 2012;286:661–5.
26. Shaw CM, Stanczyk FZ, Egleston BL, Kahle LL, Spittle CS, Godwin AK, et al. Serum antimüllerian hormone in healthy premenopausal women. *Fertil Steril* 2011;95:2718–21.
27. Freeman EW, Gracia CR, Sammel MD, Lin H, Lim LC, Strauss JF. Association of anti-müllerian hormone levels with obesity in late reproductive-age women. *Fertil Steril* 2007;87:101–6.
28. Skaöba P, Cygal A, Madej P, Dąbkowska-Huó A, Sikora J, Martirosian G, et al. Is the plasma anti-Müllerian hormone (AMH) level associated with body weight and metabolic, and hormonal disturbances in women with and without polycystic ovary syndrome? *Eur J Obstet Gynecol Reprod Biol* 2011;158:254–9.
29. Bertrand KA, Scott CG, Tamimi RM, Jensen MR, Pankratz VS, Norman AD, et al. Dense and non-dense mammographic area and risk of breast cancer by age and tumor characteristics. *Cancer Epidemiol Biomarkers Prev* 2015;24:798–809.
30. Pettersson A, Graff RE, Ursin G, Santos Silva ID, McCormack V, Baglietto L, et al. Mammographic density phenotypes and risk of breast cancer: a meta-analysis. *J Natl Cancer Inst* 2014;106.
31. DISC Collaborative Research Group. Dietary intervention study in children (DISC) with elevated low-density-lipoprotein cholesterol. Design and baseline characteristics. *Ann Epidemiol* 1993;3:393–402.
32. Kwiterovich PO, Hartmuller G, Van Horn L, Christoffel KK, Gernhoffer N, Gidding S, et al. Efficacy and safety of lowering dietary intake of fat and cholesterol in children with elevated low-density lipoprotein cholesterol: The Dietary Intervention Study in Children (DISC). *JAMA* 1995;273:1429–35.
33. Obarzanek E, Hunsberger SA, Van Horn L, Hartmuller VV, Barton BA, Stevens VJ, et al. Safety of a fat-reduced diet: the Dietary Intervention Study in Children (DISC). *Pediatrics* 1997;100:51–9.
34. Obarzanek E, Kimm SY, Barton BA, Van Horn LL, Kwiterovich PO Jr, Simons-Morton DG, et al. Long-term safety and efficacy of a cholesterol-lowering diet in children with elevated low-density lipoprotein cholesterol: seven-year results of the Dietary Intervention Study in Children (DISC). *Pediatrics* 2001;107:256–64.
35. Dorgan JF, Liu L, Klifa C, Hylton N, Shepherd JA, Stanczyk FZ, et al. Adolescent diet and subsequent serum hormones, breast density, and bone mineral density in young women: results of the Dietary Intervention Study in Children follow-up study. *Cancer Epidemiol Biomarkers Prev* 2010;19:1545–56.
36. Kuczumarski RJ, Ogden CL, Grummer-Strawn LM, Flegal KM, Guo SS, Wei R, et al. CDC growth charts: United States. *Adv Data* 2000:1–27.
37. Pettee Gabriel K, Klifa C, Perez A, Kriska AM, High RR, Snelaelar L, et al. Adolescent and young adult exposure to physical activity and breast density. *Med Sci Sports Exerc* 2013;45:1515–23.
38. Kumar A, Kalra B, Patel A, Shah S, Savjani G, Themmen APN, et al. Development of stable picoanti-müllerian hormone (AMH) ELISA: sensitive, reliable and reproducible results. *Endocrine Reviews* 2014;35:SUN-0013.
39. American College of Radiology. ACR Practice Guidelines for the performance of contrast-enhanced magnetic resonance imaging (MRI) of the breast [Amended 2014 (Resolution 39)]. Available from: http://acr.org/~media/ACR/Documents/PGTS/guidelines/MRI_Breast.pdf.
40. Klifa C, Carballido-Gamio J, Wilmes L, Laprie A, Lobo C, DeMicco E, et al. Quantitation of breast tissue index from MR data using fuzzy clustering. *Conf Proc IEEE Eng Med Biol Soc* 2004;3:1667–70.
41. Rosner B. Percentage points for a generalized ESD many-outlier procedure. *Technometrics* 1983;25:165–72.
42. Duan N. Smearing estimate: a nonparametric retransformation approach. *J Am Stat Assoc* 1983;79:605–10.
43. Colditz GA, Frazier AL. Models of breast cancer show that risk is set by events of early life: prevention efforts must shift focus. *Cancer Epidemiol Biomarkers Prev* 1995;4:567–71.
44. Russo J, Gusterson BA, Rogers AE, Russo IH, Wellings SR, van Zwieten MJ. Comparative study of human and rat mammary tumorigenesis. *Lab Invest* 1990;62:244–78.
45. Russo J, Tay LK, Russo IH. Differentiation of the mammary gland and susceptibility to carcinogenesis. *Breast Cancer Res Treat* 1982;2:5–73.
46. Samimi G, Colditz GA, Baer HJ, Tamimi RM. Measures of energy balance and mammographic density in the Nurses' Health Study. *Breast Cancer Res Treat* 2008;109:113–22.
47. Sellers TA, Vachon CM, Pankratz VS, Janney CA, Fredericksen Z, Brandt KR, et al. Association of childhood and adolescent anthropometric factors, physical activity, and diet with adult mammographic breast density. *Am J Epidemiol* 2007;166:456–64.
48. Harris HR, Tamimi RM, Willett WC, Hankinson SE, Michels KB. Body size across the life course, mammographic density, and risk of breast cancer. *Am J Epidemiol* 2011;174:909–18.
49. McCormack VA, dos Santos Silva I, De Stavola BL, Perry N, Vinnicombe S, Swerdlow AJ, et al. Life-course body size and perimenopausal mammographic parenchymal patterns in the MRC 1946 British birth cohort. *Br J Cancer* 2003;89:852–9.
50. Jeffreys M, Warren R, Gunnell D, McCarron P, Smith GD. Life course breast cancer risk factors and adult breast density (United Kingdom). *Cancer Causes Control* 2004;15:947–55.
51. Lope V, Perez-Gomez B, Moreno MP, Vidal C, Salas-Trejo D, Ascunce N, et al. Childhood factors associated with mammographic density in adult women. *Breast Cancer Res Treat* 2011;130:965–74.
52. Fraser A, McNally W, Sattar N, Anderson EL, Lashen H, Fleming R, et al. Prenatal exposures and anti-Müllerian hormone in female adolescents: the Avon Longitudinal Study of Parents and Children. *Am J Epidemiol* 2013;178:1414–23.
53. Anderson EL, Fraser A, McNally W, Sattar N, Lashen H, Fleming R, et al. Anti-müllerian hormone is not associated with cardiometabolic risk factors in adolescent females. *PLoS One* 2013;8:e64510.
54. Cui L, Qin Y, Gao X, Lu J, Geng L, Ding L, et al. Antimüllerian hormone: correlation with age and androgenic and metabolic factors in women from birth to postmenopause. *Fertil Steril* 2016;105:481–5.
55. Park AS, Lawson MA, Chuan SS, Oberfield SE, Hoeger KM, Witchel SF, et al. Serum anti-müllerian hormone concentrations are elevated in oligomenorrheic girls without evidence of hyperandrogenism. *J Clin Endocrinol Metab* 2010;95:1786–92.
56. Sowers MR, Eyvazzadeh AD, McConnell D, Yosef M, Jannausch ML, Zhang D, et al. Anti-müllerian hormone and inhibin B in the definition of ovarian aging and the menopause transition. *J Clin Endocrinol Metab* 2008;93:3478–83.
57. Lie Fong S, Visser JA, Welt CK, de Rijke YB, Eijkemans MJ, Broekmans FJ, et al. Serum anti-müllerian hormone levels in healthy females: a nomogram ranging from infancy to adulthood. *J Clin Endocrinol Metab* 2012;97:4650–5.
58. McCoy AC, Kliethermes B, Zhang K, Qin W, Sticca R, Bouton M, et al. Serum Müllerian inhibiting substance levels are lower in premenopausal women with breast precancer and cancer. *BMC Res Notes* 2011;4:152.
59. Su HI, Flatt SW, Natarajan L, DeMichele A, Steiner AZ. Impact of breast cancer on anti-müllerian hormone levels in young women. *Breast Cancer Res Treat* 2013;137:571–7.