

# Cytogenetic and Molecular Tumor Profiling for Type 1 and Type 2 Papillary Renal Cell Carcinoma

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**Abstract Purpose:** The goal of this study was to evaluate immunohistochemical and cytogenetic features and their prognostic value in papillary renal cell carcinoma (PRCC) subtypes. **Experimental Design:** One hundred fifty-eight cases of PRCC were identified and reclassified by subtype. Tumoral expression of 29 molecular markers was determined by immunohistochemistry. Cytogenetic analyses were done on a prospective series of 65 patients. Associations with clinicopathologic information and disease-specific survival were assessed. **Results:** Fifty-one patients (32%) had type 1 and 107 (68%) type 2 PRCC. Type 2 patients had worse Eastern Cooperative Oncology Group performance status, higher T stages, nodal and distant metastases, higher grades, and a higher frequency of necrosis, collecting system invasion and sarcomatoid features. Type 2 showed greater expression of vascular endothelial growth factor (VEGF)-R2 in the tumor epithelium, and of VEGF-R3 in both tumor epithelium and endothelium. Loss of chromosome 1p, loss of 3p, and gain of 5q were exclusively observed in type 2, whereas type 1 more frequently had trisomy 17. Type 2 PRCC was associated with worse survival than type 1, but type was not retained as an independent prognostic factor. Lower PTEN, lower EpCAM, lower gelsolin, higher CAIX, and higher VEGF-R2 and VEGF-R3 expression, loss of 1p, 3p, or 9p, and absence trisomy 17 were all associated with poorer prognosis. **Conclusions:** Type 2 PRCC is associated with more aggressive clinicopathologic features and worse outcome. Molecular and chromosomal alterations can distinguish between PRCC subtypes and influence their prognosis. The effect of 3p loss on survival in PRCC is opposite to the relationship seen in clear cell RCC.

Renal cell carcinoma (RCC) has historically been viewed as a single entity; however, distinct subtypes have been delineated in the past several decades. The Heidelberg classification describes five subtypes, including clear cell, papillary, chromophobe, collecting duct, and unclassified RCC (1). Papillary RCC (PRCC) is the second most common with an incidence of 10% to 15% (2, 3).

The vast majority of PRCC are sporadic; however, two hereditary forms of PRCC have been described including hereditary PRCC and hereditary leiomyomatosis renal cell

cancer (4). Hereditary PRCC is a rare syndrome associated with type 1 PRCC, caused by a "gain-of-function" mutation of the *MET proto-oncogene* on chromosome 7q. This gene encodes a transmembrane receptor (c-Met) that interacts with hepatocyte growth factor (5, 6). Hereditary leiomyomatosis renal cell cancer is caused by a mutation of the *fumarate hydratase gene*, leading to cutaneous and uterine leiomyomas and an aggressive type 2 PRCC (7, 8).

Based on cytologic and histologic criteria, Delahunt and Eble (9, 10) divided PRCC into two morphologic groups, type 1 and type 2, differing in stage, grade, and prognosis. Several studies have suggested that patients with type 2 PRCC present with higher stage and nuclear grade (9–12), which leads to worse survival (12–14). Although molecular characteristics of PRCC subtypes have been investigated, many studies are hindered by small sample size, the retrospective nature, and lack of follow-up. Therefore, the goals of this study were (a) to evaluate clinicopathologic features and their prognostic value in PRCC subtypes, (b) to define an immunohistochemical profile of type 1 from type 2 PRCC, (c) to analyze the relevance of protein expression in predicting prognosis, and (d) to describe cytogenetic aberrations and their effect on survival.

## Experimental Design

**Patient selection and clinical data.** The University of California at Los Angeles Kidney Cancer Program Database

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### Translational Relevance

Papillary renal cell carcinoma (PRCC) is the second most common histologic subtype. Two distinct types of PRCC have been described yet the molecular characterization and clinical behavior has not been extensively studied. We have done a thorough assessment of each type of PRCC and determined unique pathologic, immunohistochemical, and cytogenetic differences, and their relevance in predicting prognosis. In the era of targeted cancer therapy, these differences such as VEGF receptor expression may play a role determining response to current agents such as tyrosine kinase inhibitors. Cytogenetic findings unique to each type such as trisomy 17 in papillary type 1 and the loss of 1p and 3p in type 2, may ultimately lead to future type-specific therapy.

comprises 1,825 patients treated from 1985 to 2007. After approval by the Institutional Review Board, chart and slide review of all cases were done. Patients with hereditary renal tumors, those who were not surgical candidates and those with incomplete data sets were not included. PRCC was defined as RCC with papillary or tubulo-papillary architecture in at least 75% of the microscopic fields (15). With this definition, 158 PRCCs were identified and rereviewed. Cases were subtyped into type 1 and 2 PRCC by a single expert genito-urinary pathologist (J.W.S) in accordance with Delahunt and Eble's original description (9). Because nucleolar grade but not Fuhrman grade is applicable for PRCC, all tumors were regraded according to the scheme described by Sika-Paotonu (16). Additional information included sarcomatoid features, collecting system invasion, necrosis, and multifocality. Tumor necrosis was defined as the presence of any microscopic coagulative necrosis. Multifocality was defined as separate renal cell tumors in the same kidney, diagnosed intraoperatively or by histologic examination. Radiographic, operative, and pathology reports were used to assess the tumor-node-metastasis stage (17). An Eastern Cooperative Oncology Group (ECOG) performance status was prospectively assigned at diagnosis (18).

**Tissue array construction and immunohistochemistry.** Out of the 158 tumors with type 1 or 2 PRCC, 40 specimens were randomly obtained from the Department of Pathology at the University of California at Los Angeles Medical Center. Three core tissue biopsies, 0.6 mm in diameter, were taken from morphologically representative regions of each paraffin-embedded PRCC and precisely arrayed as described previously (19).

Immunohistochemical staining was done with a panel of 29 tumor markers with a Dako Envision (Dako) or Vectastain Elite ABC (Vector) staining system (20, 21). The primary antibodies used targeted gelsolin (Sigma Co; concentration, 3.8 µg/mL), HIF-1α (Novus Biologicals; 6 µg/mL), Ki-67 (Dako; 0.5 µg/mL), vimentin (Dako; dilution, 1:1,000), CAIX (gift from Dr. Eric Stanbridge, Irvine, CA; 1:25,000 dilution), CAXII (gift from Dr. Michael Lerman, Laboratory of Immunobiology, National Cancer Institute, Frederick, MD; 1:450 dilution), EpCAM (BD Pharmingen; 20 µg/mL), p21 (Calbiochem; dilution, 1:100), p27 (Dako; 8 µg/mL), p53 (Dako; 1:100 dilution), CXCR3 (R&D Systems; 0.1 µg/mL),

pS6 (Cell Signaling; 0.125 µg/mL), pAkt (Cell Signaling; 1.5 µg/mL), PTEN (Zymed; 2 µg/mL), vascular endothelial growth factor (VEGF)-A (Santa Cruz Biotechnology; 4 µg/mL), VEGF-C (Zymed; 3 µg/mL), VEGF-D (R&D Systems; 3 µg/mL), VEGF-R1 (Santa Cruz; 1 µg/mL), VEGF-R2 (Santa Cruz; 2 µg/mL), and VEGF-R3 (gift from Dr. Kari Alitalo, Haartman Institute, University of Helsinki, Helsinki, Finland; 2 µg/mL).

Semiquantitative assessment of staining was done by a single pathologist (DBS) blinded to clinicopathologic variables and outcome. The extent of staining was recorded as percentage of the entire tumor sample that stained positive. The overall score used for subsequent statistical analysis was the pooled mean from the three spots of the same tumor.

**Cytogenetic analysis.** Tumor samples were collected immediately postoperatively in 65 consecutive patients. After short-term culture, chromosomes were banded using the GPG (G-Bands by Pancreatin using Giemsa) technique. Twenty metaphases were investigated and analyzed in accordance with the International Standing Committee on Human Cytogenetic Nomenclature by one cytogeneticist (PNR; ref. 20). Ploidy levels were classified as pseudodiploid (modal number, 46), hypodiploid (<46), hyperdiploid (47-57), and polyploid (≥58). Of the 65 tumors, 57 (88%) showed an abnormal karyotype, which form the principal study cohort.

**Statistical analysis.** A *P* value of <0.05 was considered statistically significant and the statistical software R<sup>4</sup> was used for all analyses. Categorical data were compared using the  $\chi^2$  test or Fisher's Exact test, whereas the Kruskal-Wallis test and Student's *t* test were used to assess continuous data. The end point of this study was disease-specific survival (DSS), defined from the date of nephrectomy to the date of RCC death. Cause of death was determined from the death certificate, physician correspondence, or clinical history. The Kaplan-Meier method was used to estimate survival curves, and the log-rank test applied to compare curves. Univariate and backward stepwise multivariate Cox proportional hazards regression models were fit. For multivariate Cox proportional hazards regression models, variables were removed backward as defined by the likelihood ratio statistic ( $P_{in} = 0.05$ ,  $P_{out} = 0.10$ ). The rank of elimination was given when a variable was removed from the equation, and the hazard ratio, 95% confidence interval (CI), and *P* value for the removed variables were obtained on the step of removal. The proportional hazard assumption was tested by the Schoenfeld test. For identification of high-risk patients for disease-specific death according to immunohistochemical protein expression, we used dichotomized protein expression values (high/low), identified by univariate recursive partitioning based survival tree analysis. Recursive partitioning is a method that can be used for univariate and multivariate analysis. It constructs a decision tree that classifies patients based on dichotomized, dependent variables.

### Results

**Clinicopathologic features and prognosis of type 1 and 2 PRCC.** Fifty-one patients (32%) had type 1 and 107 (68%) had type 2 PRCC. Type 2 was associated with worse ECOG performance status, higher T stage, lymph node and distant

<sup>4</sup> <http://cran.r-project.org/>

**Table 1.** Clinicopathologic features and DSS of type 1 and type 2 PRCC

	Type 1 (n = 51)	Type 2 (n = 107)	P
	No (%)	No (%)	
Age (mean ± SD)	62.1 ± 11.9 y	61.8 ± 12.7 y	0.889
ECOG			0.043
0	38 (75%)	62 (58%)	
≥1	13 (25%)	45 (42%)	
Incidental diagnosis	26 (51%)	49 (46%)	0.542
T stage			0.013
pT1	38 (75%)	53 (50%)	
pT2	4 (8%)	8 (7%)	
pT3	9 (18%)	41 (38%)	
pT4	0	5 (5%)	
Lymph node metastases (pN1/2)	3 (6%)	23 (21%)	0.013
Distant metastases (M1)	4 (8%)	29 (27%)	0.005
Tumor size (mean ± SD)	4.9 ± 4.4 cm	6.6 ± 4.4 cm	0.031
Nucleolar grade			<0.001
G1	14 (27%)	1 (1%)	
G2	35 (69%)	48 (49%)	
G3	2 (4%)	58 (54%)	
Multifocality	8 (16%)	18 (17%)	0.857
Necrosis	12 (24%)	58 (54%)	<0.001
Vascular invasion (V1/2)	5 (10%)	37 (35%)	0.001
Sarcomatoid features	0	7 (7%)	0.062
Collecting system invasion	5 (10%)	24 (22%)	0.055
5-y survival rate (± SE)	90 ± 5%	61 ± 6%	0.005

metastases, greater size, higher grade, necrosis, collecting system invasion, and sarcomatoid features (Table 1).

Median follow-up was 38 months (range, 1-199). At the time of analysis, 39 patients (25%) had died from the disease. Ninety percent of the patients who died had type 2 PRCC, corresponding to a 3.9-fold increased risk for death from type 2 compared with type 1 (95% CI, 1.39-11.1; log-rank  $P = 0.005$ ). Five-year survival rates were 61% ± 6% for all type 2 and 90% ± 5% for all type 1 patients (Table 1; Fig. 1). This survival difference, however, lost its statistical significance in multivariate analysis. Poor ECOG performance status, RCC-related symptoms, higher T stages, metastatic disease, and necrosis, but not papillary type, were independent predictors of poor prognosis (Table 2).

Survival analyses were conducted separately for localized and metastatic patients, and type 1 and type 2 cohorts. Patients presenting with localized PRCC had a 5-year survival of 95% ± 3%, similar between type 1 and 2 (97% ± 3% versus 93% ± 4%,  $P = 0.447$ ). Forty-nine patients had metastatic PRCC (44 type 2 and 5 type 1 PRCC), of which 41 presented with synchronous metastatic disease and 8 developed metastases after surgery for N0M0 PRCC. Median survival for all patients with metastatic PRCC was 11 ± 2 months. Twenty patients with metastatic PRCC received systemic therapy. Among 16 patients who received immunotherapy, there were no responders. Although administration of systemic therapy yielded a longer median survival (14 ± 4 versus 9 ± 4 months), this difference did not reach statistical significance ( $P = 0.571$ ). Median survival for patients with metastatic type 1 PRCC (5 ± 3 months,  $n = 5$ ) was worse than for type 2 (13 ± 2 months,  $n = 44$ ;  $P = 0.040$ ).

**Immunohistochemical profile.** Forty PRCC tumor samples, 13 with type 1 and 27 with type 2, were immunohistochemically evaluated. Type 2 had greater expression of VEGF-R2 in the tumor epithelium (57% versus 30%;  $P = 0.007$ ), and VEGF-R3 in both tumor epithelium (19% versus 6%;  $P = 0.028$ ) and endothelium of associated vessels (11% versus 0.2%;  $P = 0.009$ ) than type 1. All other evaluated molecular markers were not differentially expressed between type 1 and type 2.

Associations of protein expression with clinicopathologic variables are summarized in Fig. 2. Patients with higher T stages showed lower gelsolin expression, and higher endothelial VEGF-R1 and VEGF-R3 expression. Spread to distant sites was associated with lower gelsolin, lower EpCAM, and higher cytoplasmic p27, endothelial VEGF-A, VEGF-R1, and VEGF-R3 expression. Higher grades were observed in tumors with higher cytoplasmic p27, epithelial VEGF-R2, epithelial VEGF-R3, and endothelial VEGF-R3 expression. Tumors with necrosis expressed epithelial VEGF2 to a greater degree than tumors without. Finally, higher epithelial VEGF-A, epithelial VEGF-R1, and endothelial VEGF-R1 and endothelial VEGF-R3 expression were all associated with collecting system invasion.

Fourteen of the 40 patients (35%) with immunohistochemical analysis died from PRCC. We first fit univariate Cox proportional hazards models with continuous marker expressions and identified lower PTEN ( $P = 0.027$ ), lower gelsolin ( $P < 0.001$ ), lower EpCAM ( $P = 0.044$ ), higher endothelial VEGF-R2 ( $P = 0.001$ ), and higher endothelial VEGF-R3 ( $P = 0.014$ ) as predictors of diminished DSS for both types. Additionally, higher CAIX expression was associated with poorer prognosis in type 2 ( $P = 0.0370$ ). Subsequently, we identified expression level cutoff points (low/high) with recursive partitioning based survival tree analysis. Survival curves of these dichotomized marker expressions are presented in Fig. 3. A multivariate Cox model was fitted with tumor-node-metastasis stage, grade, necrosis, and significant markers. Tumor-node-metastasis stage (hazard ratio, 4.48; 95% CI, 2.03-9.88;  $P < 0.001$ ) and endothelial expression of VEGF-R2 (hazard ratio, 1.17; 95% CI 1.01-1.36;  $P = 0.039$ ) were independent prognostic factors.

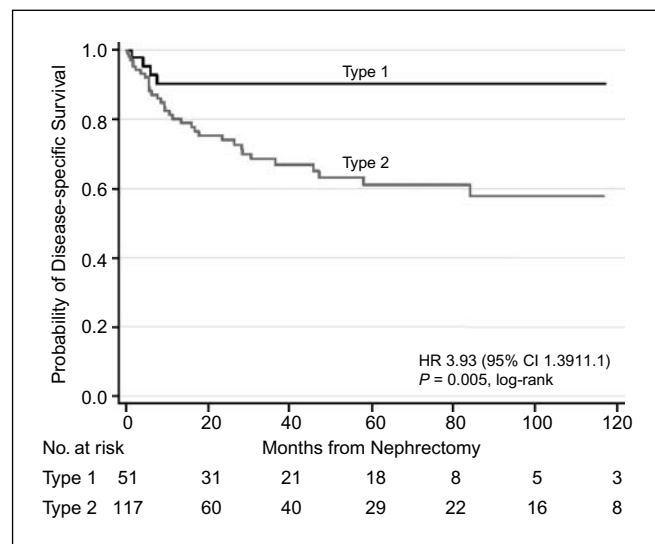


Fig. 1. DSS after nephrectomy for type 1 and type 2 PRCC. HR, hazard ratio.

**Table 2.** Univariate and multivariate models of DSS for both type 1 and type 2 PRCC

Covariate	Univariate		Rank	Multivariate	
	HR (95% CI)	P		HR (95% CI)	P
ECOG performance status	7 (3.28-14.9)	<0.0001	—	2.56 (1.06-6.21)	0.0369
Symptomatic presentation	5.75 (2.41-13.8)	<0.0001	—	2.9 (1.10-8.34)	0.0485
T stage	3.55 (2.46-5.12)	<0.0001	—	2.83 (1.70-4.72)	0.0001
Lymph node metastases	8.63 (4.42-16.9)	<0.0001	3	0.83 (0.35-1.97)	0.6767
Distant metastases	40 (17.7-90.7)	<0.0001	—	16.9 (6.49-43.9)	<0.0001
Nucleolar grade	3.8 (2.00-7.21)	<0.0001	7	1.37 (0.63-2.95)	0.4262
Multifocality	0.83 (0.35-1.98)	0.6733	2	1.08 (0.40-2.88)	0.8824
Necrosis	2.49 (1.30-4.78)	0.006	—	2.58 (1.23-5.39)	0.0119
Vascular invasion	4.92 (2.3-10.5)	<0.0001	6	1.26 (0.56-2.81)	0.5744
Sarcomatoid features	8.91 (3.54-22.4)	<0.0001	1	0.92 (0.29-2.98)	0.8938
Collecting system invasion	5.49 (2.83-10.6)	<0.0001	4	1.17 (0.55-2.46)	0.6825
Papillary subtype	3.93 (1.39-11.1)	0.01	5	0.7 (0.21-2.30)	0.5525

**Cytogenetic profile.** Of the 57 tumors with an abnormal karyotype, the following complex patterns were observed: hypodiploid in 12 cases (21%), pseudodiploid in 6 (11%), hyperdiploid (47-57) in 34 (60%), and polyploid ( $\geq 58$ ) in 5 (9%). The most frequently observed cytogenetic abnormalities were trisomy 7 ( $n = 35$ , 61%), trisomy 17 ( $n = 31$ , 54%), trisomy 12 ( $n = 28$ , 49%), trisomy 16 ( $n = 22$ , 39%), trisomy 3 ( $n = 13$ , 23%), trisomy 20 ( $n = 17$ , 30%), monosomy 21 ( $n = 8$ , 14%), loss of 3p ( $n = 8$ , 14%), gain of 5q ( $n = 7$ , 12%), loss of 1p ( $n = 6$ , 11%), loss of 9p ( $n = 6$ , 11%), trisomy 10 ( $n = 6$ , 11%), and loss of chromosome Y in men ( $n = 32$ , 71%). Trisomy 7, trisomy 10, trisomy 12, trisomy 16, trisomy 20, monosomy 21, and loss of chromosome Y were all not linked with papillary type, pathologic variables and survival.

We sought to identify chromosomal signatures of type 1 ( $n = 22$ ) and 2 ( $n = 35$ ). Type 2 PRCC had a greater number of chromosomal aberrations than type 1 (median 8 versus 6,  $P = 0.018$ ). Loss of 1p (17% versus 0%,  $P = 0.040$ ), loss of 3p (23% versus 0%,  $P = 0.016$ ), and gain of 5q (20% versus 0%,  $P = 0.025$ ) were exclusively observed in type 2 tumors, whereas type 1 tumors more frequently had trisomy 17 (73% versus 43%,  $P = 0.028$ ).

Among both subtypes, trisomy 17 was associated with lower T stages (81% versus 19%,  $P = 0.003$ ), less frequent nodal (6% versus 31%,  $P = 0.016$ ) and distant metastases (10% versus 31%,  $P = 0.044$ ), and longer survival ( $P = 0.034$ ; Fig. 4). Aberrations leading to loss of 1p material were observed in 6 tumors (11%) and included three derivative chromosomes 1 (der(1)t(1;12)(p13;p11.2), der(1)t(1;2;p13;p11.2), der(1)t(1;6)(p32;q21)), one add(1)(p36.2), one terminal deletion del(1)(p35), and one loss of the entire chromosome 1. Patients with loss of 1p had more frequently T stages 3 to 4 (83% versus 31%,  $P = 0.013$ ), lymph node metastases (67% versus 11%,  $P = 0.001$ ), and grade 3 tumors (83% versus 26%,  $P = 0.004$ ). DSS was worse for patients with loss of 1p compared with those without ( $P = 0.045$ ; Fig. 4).

Loss of chromosome 3p material was noted in 8 cases (14%). Three tumors had numerical loss of chromosome 3, three were terminal deletions with breakpoints at 3p12, 3p14, and 3p25, one tumor showed interstitial loss of 3p12-p21 and one had unbalanced translocation der(3)t(3;10)(p11.2;q11.2). Loss of 3p was associated with higher T stage (all had T3-4,  $P < 0.001$ ), lymph node involvement (63% versus 10%,  $P < 0.001$ ), distant

metastasis (63% versus 12%,  $P = 0.001$ ), higher grades (G3: 63% versus 27%,  $P = 0.042$ ), and larger tumor sizes ( $8.8 \pm 2.8$  cm versus  $5.9 \pm 5.5$  cm,  $P = 0.031$ ). In terms of outcome, patients with loss of 3p had worse survival ( $P < 0.001$ ), representing a 13.4-fold increased risk of death from PRCC (95% CI 3.35-54.3; Fig. 4).

Loss of 9p occurred in six cases, of which five had type 2 PRCC. Three were terminal deletions of the short arm of chromosome 9 with the breakpoints identified at 9p13, 9p21, 9p22, two were numerical losses of the entire chromosome, and one tumor had unbalanced translocation der(9)t(2;9)(p11.2;p13). Loss of 9p material was associated with higher T stage (all were T3-4;  $P = 0.001$ ), lymph node involvement (50% versus 14%;  $P = 0.027$ ), distant metastases (67% versus 14%;  $P = 0.002$ ), and larger tumor sizes ( $14.5 \pm 9.1$  cm versus  $5.3 \pm 3.8$  cm;  $P = 0.001$ ). Loss of 9p carried a 5.1-fold increased risk of death from PRCC (95% CI, 1.27-20.7; Fig. 4).

## Discussion

The main findings of this study are as follows: (a) type 1 and type 2 PRCC possess unique clinico pathologic and molecular profiles; (b) type 2 PRCC is associated with worse survival; however, metastatic type 1 has poorer survival than metastatic type 2 PRCC; (c) VEGF-R2 and VEGF-R3 can assist in subdividing type 1 and type 2 PRCC because they are differently expressed; (d) PTEN, EpCAM, gelsolin, CAIX, and proteins of the VEGF family are associated with survival in PRCC; (e) trisomy 17 predicts improved survival, whereas loss of 1p, 3p, or 9p material lead to worse prognosis.

Stratification of PRCC into two subtypes was first proposed by Delahunt and Eble (9). The value of subdividing has been evaluated throughout multiple studies with the consensus that type 2 tumors are usually of higher stage and grade (9, 11, 12) and are associated with poorer prognosis (12–14). However, as in our series, type does not seem to be an independent prognostic factor on multivariate analysis (13).

Multifocality and necrosis are both pathologic landmarks of PRCC. In our series, the incidence of multifocality was ~15%, which is greater than the incidence in other RCC subtypes (21, 22). We found that multifocality is equally prevalent in type 1 and 2 and is not a prognostic factor, which confirms data

from a recent report (22). Sengupta et al. (23) determined that necrosis leads to worse survival in clear cell and chromophobe RCC but not PRCC. In our study, the incidence of necrosis was higher in type 2 tumors and presence of tumor necrosis was retained as an independent prognostic factor of poor survival. An interesting finding was that prognosis of metastatic type 1 PRCC was poorer than for type 2. We speculate that unique genetic and molecular aberrations in type 1 and type 2 tumors lead to activation of different molecular pathways, which may account for this finding.

Immunohistochemical differences in PRCC subtypes have been previously reported. Ki-67, AgNOR, and topoisomerase II $\alpha$  are more highly expressed in type 2 (10, 24), whereas type 1 expresses CK7 and MUC1 to a greater degree (14, 24). In our study, we compared the immunohistochemical expression of tumor markers that are involved in the hypoxia induced pathway, the mammalian target of rapamycin pathway, the cell cycle, cell adhesion, proliferation, and angiogenesis. We found that only VEGF-R2 and VEGF-R3 are differential diagnostic markers between both subtypes. However several VEGF

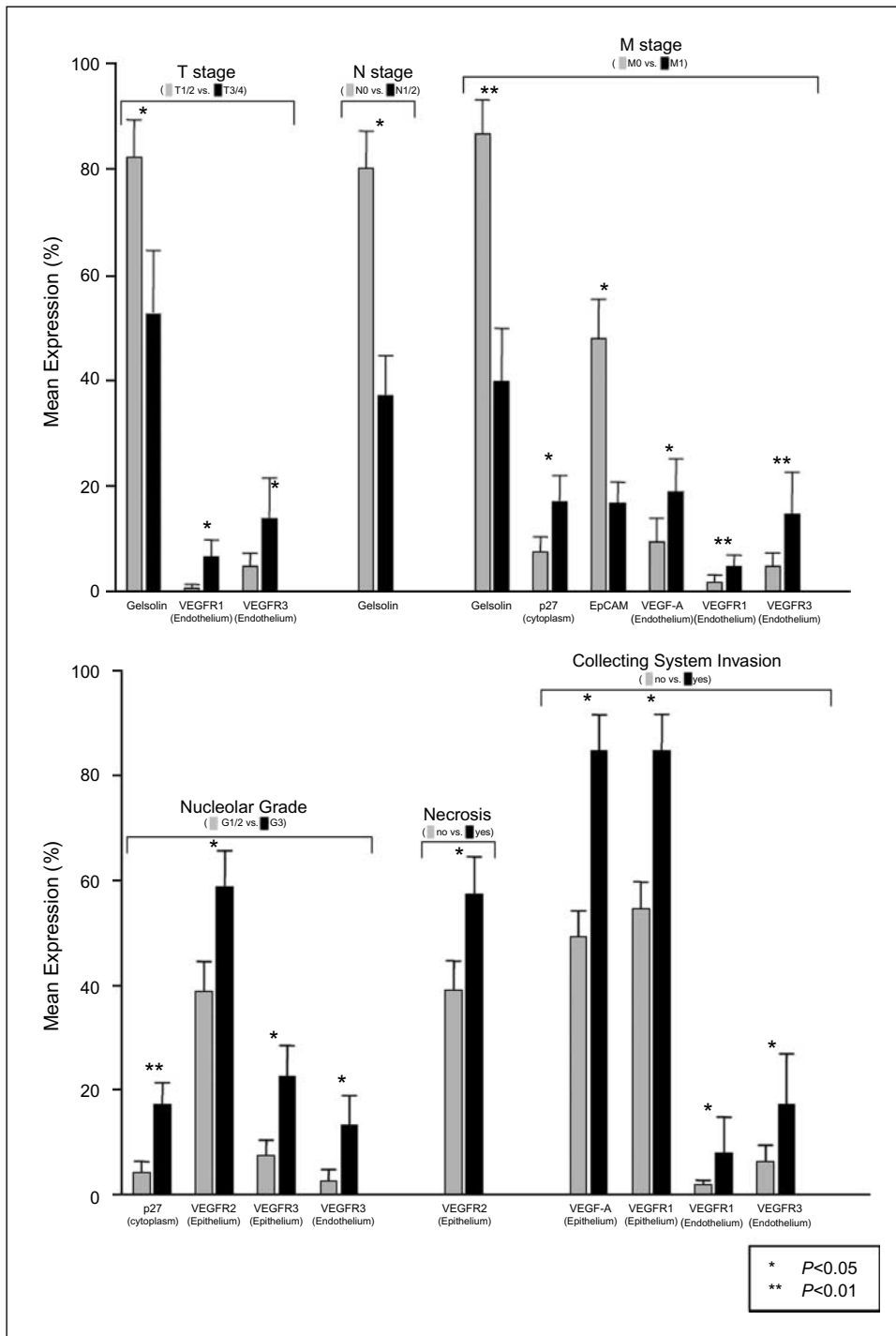


Fig. 2. Protein expression for type 1 and type 2 PRCC and association with clinical and pathologic variables.

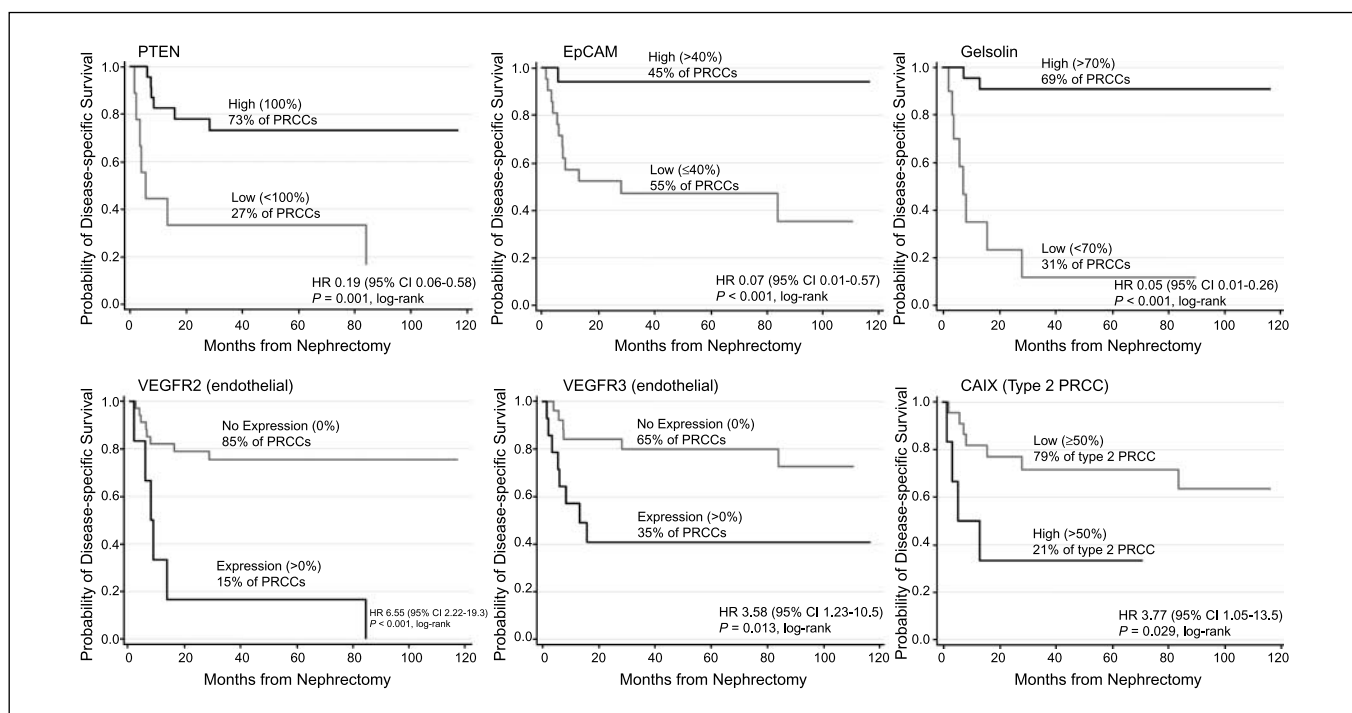


Fig. 3. Dichotomized marker expression and association with DSS.

receptors are highly expressed in both subtypes of PRCC, particularly in the tumor epithelium. This finding supports the application of VEGF-tyrosine kinase-inhibitors in PRCC. In fact, responses in PRCC were recently shown (25).

A wide variety of proteins seemed to influence DSS in PRCC including endothelial VEGF-R2 and VEGF-R3, EpCAM, gelsolin, and PTEN expression. EpCAM is an adhesion molecule that has been previously identified as a favorable prognostic factor in clear cell RCC (26). We observed higher expression of EpCAM in PRCC than in clear cell RCC and noted significantly better survival in patients with higher EpCAM expression. Higher expression of gelsolin, a member of the actin-binding protein family, is associated with worse cancer-specific survival in clear cell RCC (27). In contrast, PRCC with higher gelsolin exhibited better prognosis.

PRCC is characterized by a cytogenetic pattern distinct from other types of renal cancer. Trisomies of chromosomes 7, 12, 16, 17, and 20 are the most frequently noted karyotype aberrations (1, 11, 15, 28). Additionally, loss of chromosome Y and loss of 9p material have been reported (11, 28, 29). It has been further shown that gains of chromosome 7 and 17 are associated with type 1 (10, 28), whereas loss of 9p material is linked with type 2 cancers (11, 29). Hierarchical cluster analysis, however, has not shown distinct cytogenetic profiles for the two subtypes (11). We examined the chromosomal patterns of type 1 and 2 PRCC and their prognostic relevance. Trisomy of chromosome 17 was more frequent in type 1, whereas aberrations of chromosome 1p, 3p, 5q, and monosomy 21 are restricted to type 2. These observed cytogenetic aberrations are in accordance with previous studies (11, 28–30). Our data does not support that gains of chromosome 7p are more frequent in type 1 PRCC as suggested by Jiang and colleagues (28).

Previous studies showed that loss of 9p material correlates with of higher stages grades (11, 29). Our analyses yield similar findings in addition to poorer survival with loss of 9p material. We further showed that occurrence of trisomy 17 favors better prognosis. Indeed, the majority of grade 1 tumors exhibit trisomy 17 (30), whereas type 2 tumors have a lower incidence of trisomy 17 (11, 28). Loss of chromosome 3p in PRCC implied a more aggressive phenotype and was associated with worse survival. This is the opposite of what is seen in clear cell RCC, where loss of the *VHL* gene on chromosome 3p has been linked with a more favorable outcome (31). It is possible that other tumor-suppressor genes on the short arm of chromosome 3p may be more important in PRCC tumorigenesis.

A new strategy for treatment of PRCC is to target the c-Met receptor or its ligand, hepatocyte growth factor. Mutation of the *MET* proto-oncogene has been frequently observed in hereditary PRCC and in a subset of sporadic PRCC (32–34). Additionally, trisomy of chromosome 7, which contains the *MET* and *HGF* genes, is a frequent aberration in sporadic PRCC. Hepatocyte growth factor and MET protein expression are frequently observed in clear cell RCC and may be associated with improved survival (35, 36). In our analysis, however, trisomy 7 was not correlated with survival in PRCC.

Our results have some limitations that must be addressed including the retrospective nature. The cytogenetic analysis, however, was prospectively collected at time of nephrectomy. Further prospective studies on a larger cohort should be undertaken to confirm our findings.

## Conclusions

Unique clinicopathologic and molecular profiles of PRCC were identified. Type 2 PRCC is associated with poorer

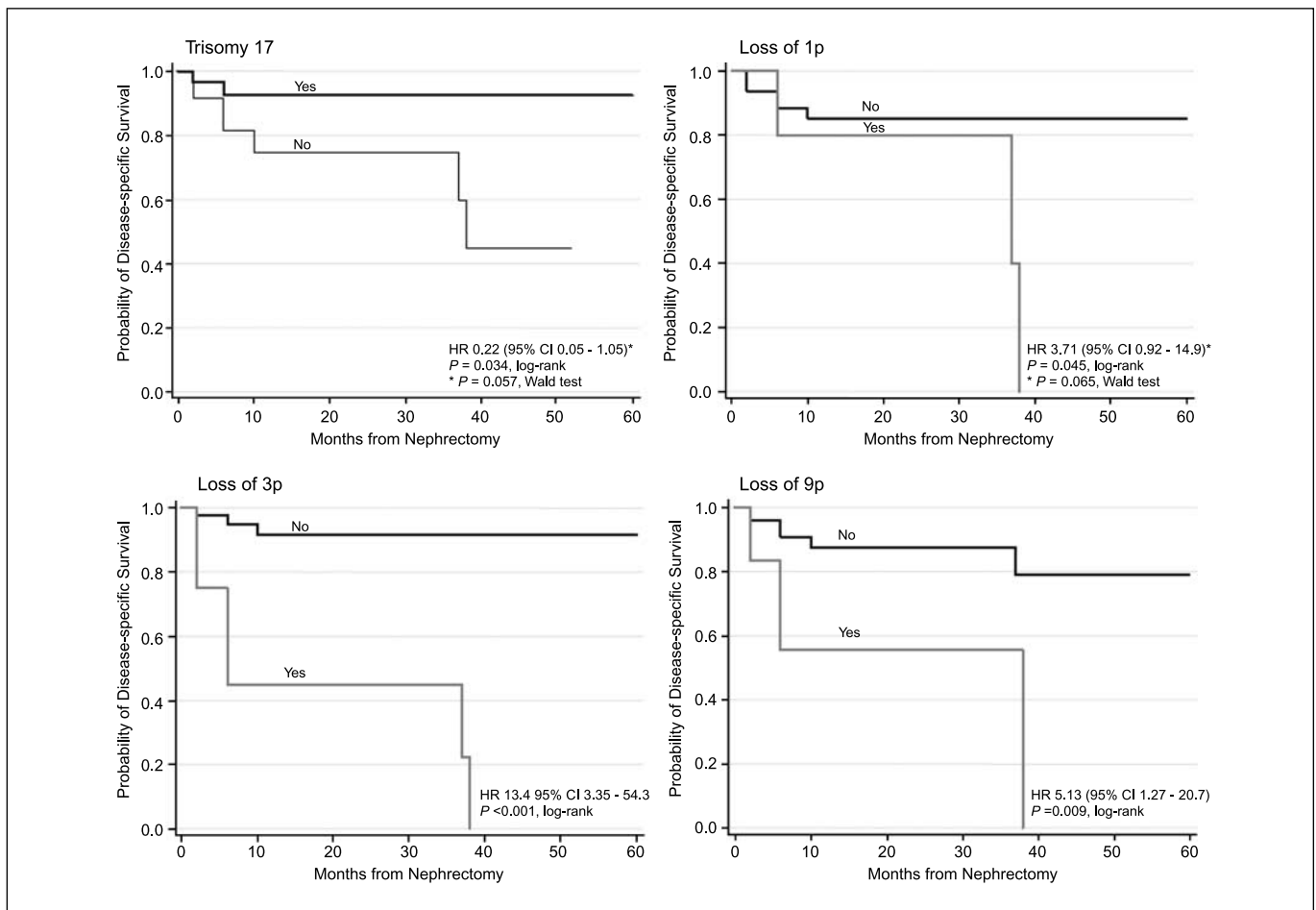


Fig. 4. Chromosomal aberrations and association with DSS.

ECOG performance status, higher stage and grade, and necrosis, leading to worse prognosis compared with type 1 PRCC. VEGFR-2 and VEGFR-3 are differentially expressed between PRCC subtypes and PTEN, EpCAM, gelsolin, CAIX, and proteins of the VEGF family are further important prognostic factors. Trisomy 17 predicts improved survival,

whereas loss of 1p, 3p, or 9p material leads to worse prognosis.

#### Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed.

#### References

- Kovacs G, Akhtar M, Beckwith BJ, et al. The Heidelberg classification of renal cell tumours. *J Pathol* 1997; 183:131–3.
- Patard JJ, Leray E, Rioux-Leclercq N, et al. Prognostic value of histologic subtypes in renal cell carcinoma: a multicenter experience. *J Clin Oncol* 2005;23:2763–71.
- Cheville JC, Lohse CM, Zincke H, Weaver AL, Blute ML. Comparisons of outcome and prognostic features among histologic subtypes of renal cell carcinoma. *Am J Surg Pathol* 2003;27:612–24.
- Linehan WM, Pinto PA, Srinivasan R, et al. Identification of the genes for kidney cancer: opportunity for disease-specific targeted therapeutics. *Clin Cancer Res* 2007;13:671–9s.
- Zhuang Z, Park WS, Pack S, et al. Trisomy 7 harbouring non-random duplication of the mutant MET allele in hereditary papillary renal carcinomas. *Nat Genet* 1998;20:66–9.
- Fischer J, Palmado G, von Knobloch R, et al. Duplication and overexpression of the mutant allele of the MET proto-oncogene in multiple hereditary papillary renal cell tumours. *Oncogene* 1998;17:733–9.
- Launonen V, Vierimaa O, Kiuru M, et al. Inherited susceptibility to uterine leiomyomas and renal cell cancer. *Proc Natl Acad Sci U S A* 2001;98:3387–92.
- Tomlinson IP, Alam NA, Rowan AJ, et al. Germline mutations in FH predispose to dominantly inherited uterine fibroids, skin leiomyomata and papillary renal cell cancer. *Nat Genet* 2002;30:406–10.
- Delahunt B, Eble JN. Papillary renal cell carcinoma: a clinicopathologic and immunohistochemical study of 105 tumors. *Mod Pathol* 1997;10:537–44.
- Delahunt B, Eble JN, McCredie MR, Bethwaite PB, Stewart JH, Bilous AM. Morphologic typing of papillary renal cell carcinoma: comparison of growth kinetics and patient survival in 66 cases. *Hum Pathol* 2001; 32:590–5.
- Gunawan B, von HA, Fritsch T, et al. Cytogenetic and morphologic typing of 58 papillary renal cell carcinomas: evidence for a cytogenetic evolution of type 2 from type 1 tumors. *Cancer Res* 2003;63:6200–5.
- Pignot G, Elie C, Conquy S, et al. Survival analysis of 130 patients with papillary renal cell carcinoma: prognostic utility of type 1 and type 2 subclassification. *Urology* 2007;69:230–5.
- Méjean A, Hopirtean V, Bazin JP, et al. Prognostic factors for the survival of patients with papillary renal cell carcinoma: meaning of histological typing and multifocality. *J Urol* 2003;170:764–7.
- Leroy X, Zini L, Leteurtre E, et al. Morphologic subtyping of papillary renal cell carcinoma: correlation with prognosis and differential expression of MUC1 between the two subtypes. *Mod Pathol* 2002;15:1126–30.

15. Kovacs G. Papillary renal cell carcinoma. A morphologic and cytogenetic study of 11 cases. *Am J Pathol* 1989;134:27–34.
16. Sika-Paotonu D, Bethwaite PB, McCredie MR, William Jordan T, Delahunt B. Nucleolar grade but not Fuhrman grade is applicable to papillary renal cell carcinoma. *Am J Surg Pathol* 2006;30:1091–6.
17. American Joint Committee on Cancer. *Kidney. AJCC Cancer Staging Manual*. 6th ed. New York: Springer; 2002. p. 323–5.
18. Oken MM, Creech RH, Tormey DC, et al. Toxicity and response criteria of the Eastern Cooperative Oncology Group. *Am J Clin Oncol* 1982;5:649–55.
19. Kononen J, Bubendorf L, Kallioniemi A, et al. Tissue microarrays for high-throughput molecular profiling of tumor specimens. *Nat Med* 1998;4:844–7.
20. International Standing Committee on Human Cytogenetic Nomenclature. *ISCN 2005: an international system for human cytogenetic nomenclature. Recommendations of the International Standing Committee on Human Cytogenetic*. Basel; Farmington (CT): Karger; 2005.
21. Blute ML, Itano NB, Cheville JC, Weaver AL, Lohse CM, Zincke H. The effect of bilaterality, pathological features and surgical outcome in nonhereditary renal cell carcinoma. *J Urol* 2003;169:1276–81.
22. Dimarco DS, Lohse CM, Zincke H, Cheville JC, Blute ML. Long-term survival of patients with unilateral sporadic multifocal renal cell carcinoma according to histologic subtype compared with patients with solitary tumors after radical nephrectomy. *Urology* 2004;64:462–7.
23. Sengupta S, Lohse CM, Leibovich BC, et al. Histologic coagulative tumor necrosis as a prognostic indicator of renal cell carcinoma aggressiveness. *Cancer* 2005;104:511–20.
24. Yang XJ, Tan MH, Kim HL, et al. A molecular classification of papillary renal cell carcinoma. *Cancer Res* 2005;65:5628–37.
25. Stadler WM, Figlin RA, Ernstoff MS, et al. The Advanced Renal Cell Carcinoma Sorafenib (ARCCS) expanded access trial: Safety and efficacy in patients (pts) with non-clear cell (NCC) renal cell carcinoma (RCC). [abstr]. *J Clin Oncol Annu Meet Proc Part 1* 2007;25:5036.
26. Seligson DB, Pantuck AJ, Liu X, et al. Epithelial cell adhesion molecule (KSA) expression: pathobiology and its role as an independent predictor of survival in renal cell carcinoma. *Clin Cancer Res* 2004;10:2659–69.
27. Visapää H, Bui M, Huang Y, et al. Correlation of Ki-67 and gelsolin expression to clinical outcome in renal clear cell carcinoma. *Urology* 2003;61:845–50.
28. Jiang F, Richter J, Schraml P, et al. Chromosomal imbalances in papillary renal cell carcinoma: genetic differences between histological subtypes. *Am J Pathol* 1998;153:1467–73.
29. Sanders ME, Mick R, Tomaszewski JE, Barr FG. Unique patterns of allelic imbalance distinguish type 1 from type 2 sporadic papillary renal cell carcinoma. *Am J Pathol* 2002;161:997–1005.
30. Kovacs G, Fuzesi L, Emanuel A, Kung HF. Cytogenetics of papillary renal cell tumors. *Genes Chromosomes Cancer* 1991;3:249–55.
31. Yao M, Yoshida M, Kishida T, et al. VHL tumor suppressor gene alterations associated with good prognosis in sporadic clear-cell renal carcinoma. *J Natl Cancer Inst* 2002;94:1569–75.
32. Schmidt L, Junker K, Nakaigawa N, et al. Novel mutations of the MET proto-oncogene in papillary renal carcinomas. *Oncogene* 1999;18:2343–50.
33. Schmidt L, Junker K, Weirich G, et al. Two North American families with hereditary papillary renal carcinoma and identical novel mutations in the MET proto-oncogene. *Cancer Res* 1998;58:1719–22.
34. Schmidt L, Duh FM, Chen F, et al. Germline and somatic mutations in the tyrosine kinase domain of the MET proto-oncogene in papillary renal carcinomas. *Nat Genet* 1997;16:68–73.
35. Miyata Y, Kanetake H, Kanda S. Presence of phosphorylated hepatocyte growth factor receptor/c-Met is associated with tumor progression and survival in patients with conventional renal cell carcinoma. *Clin Cancer Res* 2006;12:4876–81.
36. Natali PG, Prat M, Nicotra MR, et al. Overexpression of the met/HGF receptor in renal cell carcinomas. *Int J Cancer* 1996;69:212–7.