The effect of the supplementation with a primary carbon source on the resistance to oxygen exposure of methanogenic sludge

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Abstract Anaerobic methanogenic consortia have a considerable resistance to oxygen exposure. Yet, most research has been focused on the study of the tolerance to oxygen of anaerobic immobilized biomass. Less is known on the potential of the anaerobic suspended biomass for withstanding exposure to oxygen and the effect of a primary degradable substrate on such resistance. Thus, the objective of this work was to determine the effect of the amount of a primary degradable substrate (sucrose) on the resistance of a methanogenic suspended biomass to oxygen exposure. It was found that the inhibition of disperse anaerobic sludge by oxygen exposure decreases when the concentration of the supplemented carbon source increases. This is in agreement with the fact that aerobic respiration of the added substrate by the facultative heterotrophic bacteria, always present in this type of sludge, has been found in previous studies as one of the main mechanisms protecting methanogens against O₂. From a practical point of view, this suggests that aeration of anaerobic systems should be possible without inhibiting the activity of methanogenic bacteria if an adequate ratio between oxygen and COD feeding is maintained. Such a ratio will depend however on the wastewater initial COD concentration.

Keywords Anaerobic; carbon source; methanogenic activity; oxygen exposure; resistance; sludge

Introduction Biological treatment of wastes in reactors possessing several electron acceptors has been considered with increased interest in the recent past (Estrada-Vázquez et al., 2000, 2001a). To some extent, it is a consequence of the success and wide acceptance of anaerobic digestion as a wastewater treatment option (Poggi-Varaldo and Rinderknecht-Seijas, 1996; Macarie, 2000; Monroy et al., 2000), and it is one of its logical further developmental steps (Estrada-Vázquez et al., 2000). The combined environment methanogenesis-aerobiosis shows promise for the treatment and quality-polishing of dilute non-complex wastewaters, toxic effluents and groundwater remediation.

It has been shown that anaerobic methanogenic consortia have a considerable resistance to oxygen exposure. However, most research has been focused on the study of the tolerance to oxygen of anaerobic immobilized biomass such as anaerobic granules from UASB digesters (Guiot et al., 1993; Kato et al., 1993a and b; Macarie and Guiot, 1996) and bioparticles from fluidized bed reactors (Zitomer and Shrout, 2000). Little is known on the potential of the anaerobic suspended biomass for withstanding exposure to oxygen and the effect of a primary degradable substrate on such resistance. The aim of this work was to determine the influence of the amount of a primary degradable substrate (sucrose) on the resistance of a methanogenic suspended biomass to oxygen exposure.
Materials and methods

The anaerobic sludge used as inoculum for all the study was drawn from lab scale continuous, completely mixed digesters fed with a synthetic wastewater containing 25 g COD/L (sucrose, sodium acetate, mineral salts) and operated at 35°C and 25 days hydraulic retention time (Estrada-Vázquez et al., 2001b). The sludge was characterized by the following particle size distribution (on total suspended solids basis) determined accordingly to Laguna et al. (1999): 15.10% passed the 250 µm mesh and was retained in the 97 µm mesh; 5.57% was retained in mesh size 58 µm; 79.33% was captured in the dish (actual 0 µm). The mean particle diameter was 65.0 µm (arithmetic) and 60.2 µm (geometric), assuming a minimum diameter of 29 µm for the dish. Because of its small size, and also because flocks were not apparent, the inoculum is called disperse anaerobic sludge (DAS) throughout the article.

The DAS was batch-incubated without and with sucrose (initial 0, 1, 2, and 4 g COD-sucrose/L), under a range of initial O₂ concentration in the headspace (IPOH) between 0 to 70% v/v (atmospheric pressure of 580 mm Hg in Mexico City). The assay was carried out in 160 mL serum bottles with 60 mL volume liquid and 100 mL in headspace. The final concentration of the sludge was 1,620 ± 140 mg VSS/L. Bottles were incubated at 35°C and 80 rpm (Kato et al., 1993a). The sludge resistance was assessed in terms of the acetoclastic specific methanogenic activity (SMA) recovery (R) of the cultures after the incubation under oxygen, and an oxygen inhibitory concentration 50% IC₅₀ (Kato et al., 1993a; Estrada-Vázquez et al., 2001b)

\[ R = \frac{SMA_j}{SMA_c} \times 100 \]  

where \( SMA_j \) = specific acetoclastic methanogenic activity of the culture exposed to a given IPOH, and \( SMA_c \) = specific acetoclastic methanogenic activity of the control.

Incubations under oxygen exposure lasted 3 days. After the 3-day incubation, the spent media in the bottles were replaced by a medium containing sodium acetate 30 mmol l⁻¹, the headspace was flushed and replaced with N₂:CO₂ 4:1, and the specific methanogenic activity was determined. Details on other analytical methods and procedures used in this work (serum bottle technique, SMA determination, pH, alkalinity, alpha parameter, COD, total suspended and volatile suspended solids, methane and carbon dioxide contents in biogas, etc.) can be found elsewhere (Campos-Velarde et al., 1997; Poggi-Varaldo et al., 1997; Estrada-Vázquez et al., 2000 and 2001a).

Results and discussion

The increase of initial sucrose concentration significantly improved the resistance of DAS to oxygen, as can be seen from the recoveries of the SMA of the cultures (Figure 1). The positive effect was more important between 0 to 2 g/L of initial sucrose. The recoveries of DAS incubated with 2 and 4 g/L were very similar, where remarkable high R values of up to 90% were achieved. These results confirm and extend data reported by Kato et al. (1993a and b) and Estrada-Vázquez et al. (2001a and b) for disperse and granular methanogenic sludge, respectively, and reinforce the idea of a biochemical protection of the methanogenic bacteria in the inocula by consumption of the inhibitory oxygen via aerobic respiration. The latter is probably effected by the facultative microorganisms or even sometimes strict aerobes that are usually present in anaerobic consortia (Toerien and Hattingh, 1969; Assih et al., 2002).

When the initial sucrose load is much higher than the oxygen load, the recoveries of the SMA are so high that there is no IC₅₀ in the range of IPOHs tested (R values are between 75 to 95% in the range of 2.5 to 50% IPOH for bottles with DAS incubated with 2 and 4 g/L
sucrose). If the half oxygen inhibitory percentage or concentration IC$_{50}$ is used as a measure of the toxic effect of O$_2$ exposure to the methanogenic bacteria in the DAS, the IC$_{50}$ values increase with increasing sucrose concentrations (IC$_{50}$ of 3.5, 16.9, < 50 and < 50 for the series of inocula incubated with initial concentrations of 0, 1, 2 and 4 g COD-sucrose/L, respectively, Table 1). There seems to exist an inverse relationship between the IC$_{50}$ and the specific oxygen uptake rate (SOUR) of the anaerobic cultures, see Table 1. These results are in line with the argument of the possible role of the biochemical protection outlined above, and further confirm and generalize preliminary results reported by Estrada-Vázquez et al. (2001b).

If a load ratio $\gamma$ in the experimental unit is defined as

$$\gamma = \frac{\text{Initial mass of oxygen in the bottle headspace}}{\text{Initial mass of COD available in the bottle liquid phase}} \times 100$$

(2)

It can be seen in Figure 2 that recoveries are apparently very high when $\gamma < 5\%$, either for DAS incubated with 0, 1, 2, or 4 g COD/L initial sucrose (i.e. independent of the initial sucrose concentration available in the liquid phase). Above 5\%, the recovery pattern seems to depend on both the $\gamma$ and the absolute value of initial sucrose concentration. Since the bottles with no supplementary sucrose had a basal concentration of degradable COD (200 mg/L average) coming from the digester liquor, their corresponding $\gamma$ values, although great, could be calculated. Actually, for initial concentrations of added-sucrose of 0 and 1 g/L, the recoveries (10 to 55\% and 35 to 40\%, respectively) are much lower than those for DAS incubated at 2 and 4 g/L (range 75 to 90\% for both series). This is somewhat counter-intuitive, since one might speculate that the load ratio can be the determining factor for the protection of the consortia, regardless of the absolute initial concentration of the supplemental carbon source. However, the analysis of the relationship between $\gamma$ and R values of the SMA needs further investigation.

![Figure 1](https://iwaponline.com/wst/article-pdf/48/6/119/423667/119.pdf)

**Figure 1** Effect of the initial sucrose concentration recovery of the methanogenic of activity (R) versus the initial recovery

![Figure 2](https://iwaponline.com/wst/article-pdf/48/6/119/423667/119.pdf)

**Figure 2** Effect of the loading ratio $\gamma$ (initial on the mass of O$_2$ in the headspace to initial mass acetoclastic COD available) on the activity percentage of oxygen recovery

Symbols in Figure 1 and 2: 0 g/L sucrose: ○; 1 g/L sucrose: □; 2 g/L sucrose: △; 4 g/L, ○

**Table 1** Half oxygen inhibitory concentration IC$_{50}$ of disperse methanogenic sludge exposed to oxygen

<table>
<thead>
<tr>
<th>Incubation conditions</th>
<th>IC$_{50}$ (%)</th>
<th>SOUR (mg O$_2$/gVSS.day)$^b$</th>
</tr>
</thead>
<tbody>
<tr>
<td>0 g COD-sucrose/L</td>
<td>3.5</td>
<td>259</td>
</tr>
<tr>
<td>1 g COD-sucrose/L</td>
<td>16.9</td>
<td>505</td>
</tr>
<tr>
<td>2 g COD-sucrose/L</td>
<td>&gt; 50</td>
<td>547</td>
</tr>
<tr>
<td>4 g COD-sucrose/L</td>
<td>&gt; 50</td>
<td>839</td>
</tr>
</tbody>
</table>

Notes

$^a$Inhibitory concentration 50% of oxygen; $^b$specific oxygen uptake rate; experimental values at a total equivalent oxygen concentration of 980 mg O$_2$/L liquid phase of bottle (Kato et al., 1993a)

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The time courses of oxygen and methane in the bottle headspace are depicted in Figure 3 and 4, respectively.

They further support the idea that high recoveries R are related to a lower oxygen exposure during the incubation due to oxygen consumption. For instance, bottles with 50% IPOH have nearly 2, 0.6, 0.6 and 0.4 mmol O₂/bottle at the end of the 3-day incubation, for initial supplemented sucrose 0, 1, 2, and 4 g COD/L, respectively (Figure 3).

Figure 4A shows that the effect of oxygen exposure on the cultures not supplemented with sucrose was drastically negative: methane generation for the oxygen-exposed cultures decreased almost 8-fold as compared to the strict anaerobic control. In the series supplemented with 2 and 4 g COD- sucrose/L (Figure 4C, D) the negative effect of the IPOH on the methane generation was less drastic, and the bottles with low IPOH exhibited a methanogenesis nearly of the same order of that of the anaerobic control.
Conclusions
The inhibition of disperse anaerobic sludge by oxygen exposure decreases when the concentration of the supplemented carbon source increases. This is in agreement with the fact that aerobic respiration of the added substrate by the facultative heterotrophic bacteria, always present in this type of sludge, has been found in previous studies as one of the main mechanisms protecting methanogens against O₂. From a practical point of view, this suggests that aeration of anaerobic systems should be possible without inhibiting the activity of methanogenic bacteria if an adequate ratio between oxygen and COD feeding is maintained. Such a ratio will depend however on the wastewater initial COD concentration.

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References


**Notation**

- **COD**: Chemical oxygen demand
- **DAS**: Disperse anaerobic sludge
- **IC$_{50}$**: Oxygen inhibitory concentration 50%, that is, the initial percentage of oxygen in the headspace that causes a 50% decrease in the specific methanogenic activity, with respect to the activity of the control
- **IPOH**: Initial percentage of oxygen in the headspace of the bottle
- **R**: Recovery of the specific acetoclastic methanogenic activity, given by Eq. (1)
- **SMA$_c$**: Specific acetoclastic methanogenic activity of the control culture
- **SMA$_i$**: Specific acetoclastic methanogenic activity of the culture exposed to a given IPOH
- **SOUR**: Specific oxygen uptake rate
- **γ**: Load ratio of the initial mass of oxygen in the bottle headspace to the initial mass of COD available in the bottle liquid phase, see Eq. (2)