

A 62-yr-old man presented with a clinical and biochemical picture of an insulinoma had a single 1.5-cm diam mass in the tail of the pancreas at surgery, which was resected and consisted of a β -cell tumor on histology. Symptoms resolved after surgery. Blood samples for this study were obtained in the fasting state at different time points over a 3-mo period before surgery.

SIZE-EXCLUSION

CHROMATOGRAPHY— Two milliliters of plasma was concentrated two-fold in a Savant speedvac concentrator and then eluted through a Sephadex G50 column (1 x 50 cm) with a 3 M acetic acid buffer or Biogel P30 column with a phosphate buffer. Recovery of insulin and proinsulin after chromatography with this methodology was 73–102%. The entire plasma sample was used so that when the specific RIA for proinsulin became available at a later date, identical samples could not be processed by both methods.

RIAS— After separation by size-exclusion chromatography, the eluate was subjected to a previously described RIA for insulin (10). Detection limit was 0.02 pmol/ml. Intra- and interassay coefficients of variation were 6.5 and 8.0%, respectively. Proinsulin had a 60% cross-reactivity in the insulin RIA. RIA for C-peptide was carried out with a previously described method (10), with anti-serum generously provided by Bruce Frank (Lilly). The detection limit in this assay was 0.025 pmol/ml. Proinsulin had 50% cross-reactivity in the C-peptide assay. Intra- and interassay coefficients of variation were 6.9 and 8.2%, respectively. Proinsulin was measured with a highly specific RIA (11), with <0.01% cross-reactivity with insulin or C-peptide. The lower limit of sensitivity of this assay was 5 fmol/ml. Intra- and interas-

say coefficients of variation were 8.0 and 9.0%, respectively.

In this patient, the diagnosis of an insulinoma was based on the finding of hyperinsulinemia and elevated plasma C-peptide concentrations in the presence of low plasma glucose concentrations (Table 1). However, plasma proinsulin levels were not consistently elevated. Size-exclusion chromatography of plasma demonstrated a variable relationship of insulin to proinsulin in three fasting plasma samples, taken at different times (Table 1). In sample 1, almost all of the insulin immunoreactivity eluted as a peak coinciding with the proinsulin marker. In marked contrast, sample 2 consisted predominantly of a peak co-eluting with insulin. Sample 3 had an intermediate, but elevated percent of proinsulin. However, with the proinsulin RIA in four other fasting samples obtained consecutively over a 1-h period (samples 4–7), elevated concentrations were uniformly observed (Table 1).

The apparent variability in proinsulin concentrations is particularly obvious when expressed as a percentage of total insulin immunoreactivity. The likely reason for this is the fluctuation observed in plasma insulin levels, which varied dramatically, despite similar glucose concentrations in the different sam-

ples. Sample 2, evaluated by chromatography, had elevated serum insulin concentrations, despite the fact that the patient was fasting. Sample 7 also demonstrated an increase of insulin concentrations without apparent explanation because the patient was under observation and known to be fasting and resting in bed at the time. Plasma C-peptide measured in the same blood samples was elevated in each case, confirming an increase in β -cell secretion. However, proinsulin was not correspondingly elevated in sample 7 (and presumably also in sample 2, although not measured by RIA). Therefore, when expressed as a percentage of total insulin immunoreactivity, based on its known cross-reactivity in the insulin assay, proinsulin was within the normal range in sample 7.

This dissociation between fasting serum insulin and proinsulin has not, to our knowledge, been previously described. It is unlikely to be related to methodological problems because insulin levels were well correlated with elevated C-peptide concentrations ($r = 0.96, P < 0.001, n = 7$). The mechanism for the sudden increase in fasting insulin concentrations that occurred in samples 2 and 7 is unclear but may reflect erratic (12) or episodic (13) secre-

Table 1—Variable relationship of insulin to proinsulin

SAMPLE	GLUCOSE (MM)	INSULIN (PMOL/ML)	C-PEPTIDE (PMOL/ML)	PROINSULIN (FMOL/ML)	%
1	2.4	0.14	1.2		95
2	2.4	1.21	4.0		5
3	3.6	0.42	2.3		32
4	4.1	0.10	0.44	105	67
5	3.5	0.09	0.36	103	68
6	3.6	0.11	0.44	94	54
7	3.4	0.62	2.80	128	13
NORMAL FASTED	3.9–6.1	0.04–0.11	0.54 ± 0.05*	9.7 ± 1.2*	<25

Samples 1–3 were processed by size-exclusion chromatography. Samples 4–7 were obtained consecutively in the fasting state within a period of 1 h and measured in the proinsulin radioimmunoassay. %, Percentage of total insulin immunoreactivity. In samples 4–7, this was estimated with known cross-reactivity of proinsulin in the insulin radioimmunoassay.

*Means ± SE.

tions that have been previously demonstrated in these tumors (12,13).

This case is instructive in that it demonstrates that insulin concentrations may fluctuate independently of proinsulin in a patient with a proven insulinoma. Thus, caution should be used in interpreting results of the chromatographic approach; proinsulin levels may appear to be normal if expressed as percentage of total immunoreactivity. We suggest that specific RIA for proinsulin be the preferred approach for screening of insulinomas. Multiple samples in the fasting state will further elucidate whether proinsulin, like insulin, is also secreted in fluctuant fashion.

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Do You Really Get a Diet Soft Drink When You Order One?

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Our patients with diabetes mellitus periodically complain that the “diet” soft drinks they receive at restaurants, particularly fast-food chains, may actually contain sugar. The calories in a typical “regular” cola soft drink,

~160 kcal for a 12 oz drink, are derived exclusively from simple sugars (1). The consumption of a regular soft drink can thus have a substantial impact on the blood glucose of an individual with diabetes. For many years, we have used a simple and reliable method to assess the presence of sugar in soft drinks. Tes-tape (Lilly, Indianapolis, IN), a glucose oxidase- and peroxidase-impregnated urine test strip, changes color from yellow to green in the presence of very small

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