

DNA Methylation as a Therapeutic Target in Cancer

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Abstract Targeting DNA methylation for cancer therapy has had a rocky history. The first reports on DNA methylation changes in cancer described global loss of methylation, which has been suggested to drive tumorigenesis through activation of oncogenic proteins or induction of chromosomal instability. In this context, reducing DNA methylation was viewed as a tumor-promoting event rather than a promising cancer therapy. The idea of inhibiting DNA methylation therapeutically emerged from subsequent studies showing that, in parallel to global decreases in methylation, several genes (including many critical to the tumor phenotype) displayed gains of methylation in their promoters during tumorigenesis, a process associated with epigenetic silencing of expression and loss of protein function. This led to revival of interest in drugs discovered decades ago to be potent inhibitors of DNA methyltransferases. These drugs have now been approved for clinical use in the United States in the treatment of myelodysplastic syndrome, thus opening the floodgate for a whole new approach to cancer therapy—epigenetic therapy.

Background

DNA methylation refers to the addition of a methyl group to one of the four bases that constitute the coding sequence of DNA. In humans, methylation is normally added only on the 5 position of the cytosine base in a post-DNA synthesis reaction catalyzed by one of several DNA methyltransferases. DNA methylation plays a key role in chromatin structure, suppression of the activity of endogenous parasitic sequences, and stable suppression of gene expression (epigenetic silencing), a process normally reserved for special situations such as the inactive X-chromosome and imprinted genes (1).

Epigenetics has emerged as an important biological process relevant to all multicellular organisms. Studies in various models have shown that epigenetic regulation is essential for proper embryogenesis and development. There are several epigenetic processes described, and all seem to be interrelated to some degree. Besides DNA methylation, posttranslational modifications of histones (referred to as a histone code; ref. 2) seem to mediate epigenetics in many organisms (including mammals), and RNA interference is a key epigenetic process in plant cells (3) and likely in mammalian cells as well.

Why target DNA methylation? The role of DNA methylation in cancer development was initially inferred from observational studies of epigenetic patterns in normal and neoplastic tissues. Early studies using global measurement of 5-methylcytosine content suggested that loss of methylation was a common

feature of carcinogenesis (4). Subsequently, this loss of methylation was also shown in individual genes in which it was suggested as a mechanism of activation of gene expression (5). Marked loss of 5-methylcytosine was also shown to be associated with chromosomal breaks (6) and to lead to an increased cancer incidence in specific animal models (7). All this, of course, does not generate much enthusiasm for treating patients with drugs that would further reduce DNA methylation levels.

The situation changed with the realization that DNA methylation patterns in cancer had two faces (8). Global hypomethylation was but one aspect of the story. In parallel, many genes were shown to have DNA methylation increases affecting their promoters, and these increases were linked to silencing of gene expression and loss of protein function (9). It is estimated that hundreds of genes are thus silenced in every cancer (10) and many of these genes play important physiologic roles in the creation or perpetuation of the neoplastic phenotype. The stable nature of epigenetic silencing has led to the hypothesis that it constitutes a viable mechanism of inactivating tumor-suppressor genes in cancer (11), a hypothesis that has been confirmed many times over the past years. Finally, reducing methylation in mouse models was also shown to prevent cancer formation under specific circumstances (12, 13), providing further evidence for a pathogenic role of increased DNA methylation in cancer.

The plethora of genes and pathways affected by DNA methylation makes this specific therapeutic target also remarkably nonspecific in its effects. For example, restoration by hypomethylation of the expression of a silenced receptor can exquisitely sensitize cells to the effects of receptor ligands (e.g., retinoic acid receptor β and all-*trans* retinoic acid responsiveness; ref. 14). This seems to be very specific. In the same cells, however, it is likely that hypomethylation also affects other silenced pathways (e.g., cell cycle control, apoptosis, angiogenesis, and invasion) and there are even examples of oncogenes silenced in cancer by DNA methylation (15) and

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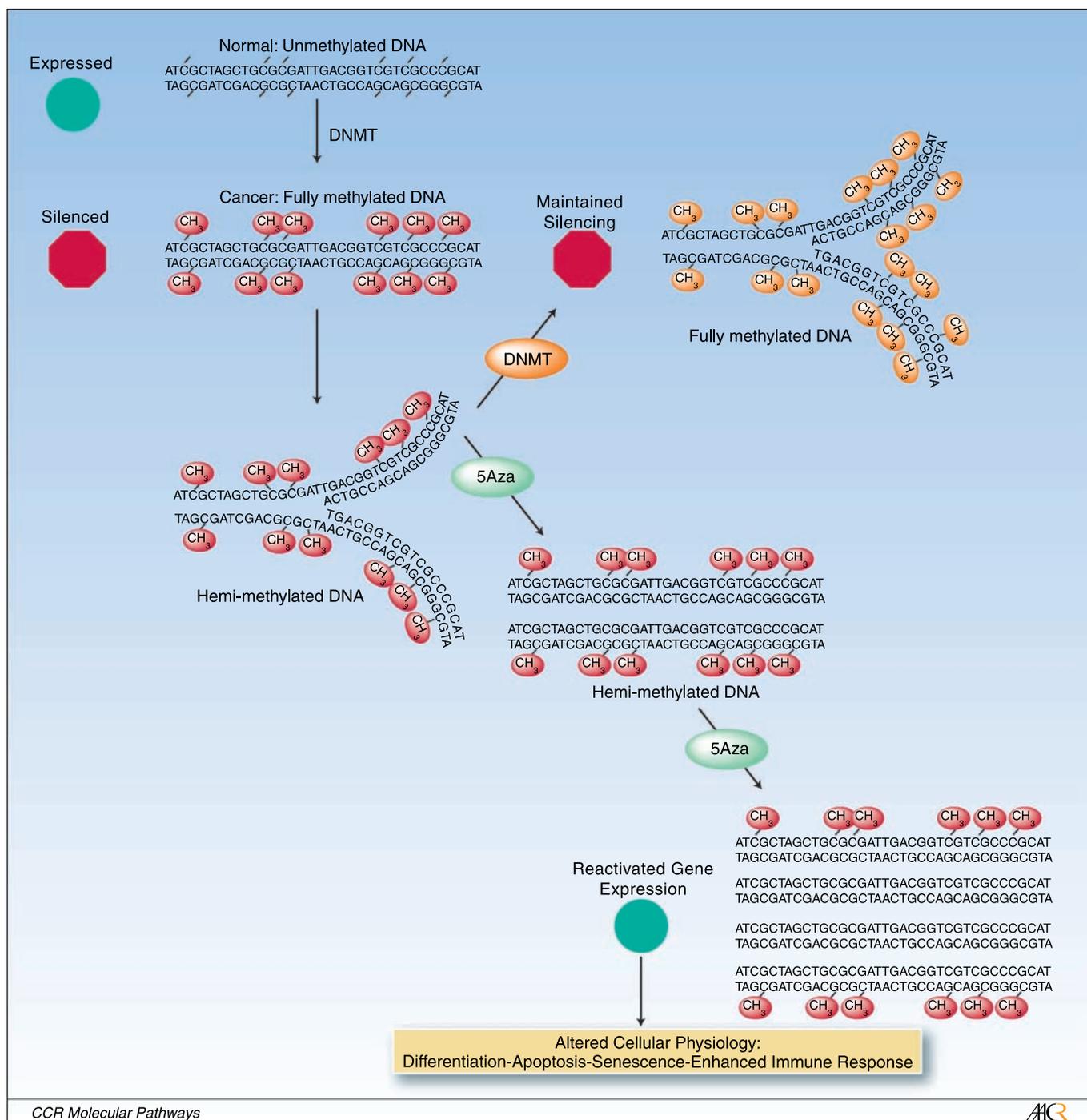


Fig. 1. Proposed mechanism of action of DNA methylation inhibitors in cancer therapy. A hypothetical tumor-suppressor gene promoter is shown switching methylation status from unmethylated and expressed in normal tissue to hypermethylated and silenced in cancer tissue. This switch requires the activity of DNA methyltransferases (*DNMT*), which are also required to maintain the hypermethylated state after each round of DNA replication. Inhibitors of DNA methyltransferases will result in failure to remethylate after DNA replication, which eventually leads to appearance of totally unmethylated alleles that reactivate gene expression. This effect on gene expression is then hypothesized to have pleiotropic effects on cancer cell biology, including induction (or facilitation) of differentiation, apoptosis, senescence, and immune response, for example.

reactivated by hypomethylation induction. Nevertheless, a therapeutic ratio for hypomethylation therapy exists and is related to the fact that tumors are much more dependent on gene silencing (e.g., of tumor suppressor genes) for their phenotype than normal adult cells. The effects of hypomethylation therapy are then the sum of multiple effects on cellular physiology, and it is likely that the net effect is favorable

therapeutically. Indeed, this nonspecificity can be viewed as advantageous (multiple defects are corrected simultaneously) while realizing its potential problems (risk of toxicity, cancer induction, etc.).

How would one target DNA methylation? Being a postsynthetic event, in proliferating cells DNA methylation is critically dependent on continued expression of DNA methyltransferases.

Inhibition of the expression of these proteins would therefore result in progressive reduction in DNA methylation in newly divided cells (Fig. 1), a phenomenon associated with reactivation of gene expression in hypomethylated cells (16). Broadly, DNA methylation inhibitors fall into three classes: (a) nucleoside inhibitors; (b) nonnucleoside weak inhibitors, often discovered serendipitously; and (c) rationally designed inhibitors.

5-Azacytidine and 5-aza-2'-deoxycytidine are cytosine analogues that trap all DNA methyltransferases and target them for degradation (16). At low doses that do not inhibit proliferation, these drugs are effective hypomethylating agents and they have shown clinical activity as anticancer agents (see further). Other nucleoside inhibitors include zebularine, which has shown promise *in vitro* (17) but is not being pursued clinically at this time, and 5-fluoro-2'-deoxycytidine (18), which has entered clinical trials. One limitation of nucleoside analogues is the requirement for DNA incorporation and active DNA synthesis, which limits the activity of the drugs in hypoproliferating cells (including potentially cancer stem cells). This has led to an interest in developing different inhibitors for the DNA methyltransferases, and this is an area of active investigation. Among the described weak inhibitors are orally available drugs such as procainamide and hydralazine (19). The mechanism of action of these drugs is not well understood and their clinical potential is likely limited by their low level of hypomethylation induction (20). Nevertheless, they could be useful as starting points for the design of other nonnucleoside inhibitors of DNA methylation. Rationally designed inhibitors of DNA methyltransferase proteins are beginning to be described (21). One limitation to this approach is the fact that three separate DNA methyltransferase genes encode for proteins with DNA methyltransferase activity. The most abundant DNA methyltransferase in cancer cells is DNA methyltransferase 1, and there is controversy about whether inhibiting this protein alone is sufficient to induce hypomethylation and a therapeutic effect in cancer cells (22, 23). Cooperation between different DNA methyltransferases (24) implies the need to inhibit several of them simultaneously for optimal clinical benefit, and drug design strategies need to take this fact into account. Finally, inhibition of DNA methyltransferase activity suffers from extreme nonspecificity, as previously discussed. There is considerable interest in devising ways of hypomethylating specific genes and there are theoretical means of doing so using unmethylated oligonucleotides (25) or other approaches (26), but none of these is of practical clinical utility at the present time.

Clinical-Translational Advances

5-Azacytidine was the first hypomethylating agent approved by the U.S. Food and Drug Administration for the treatment of a neoplasm (the myelodysplastic syndrome; ref. 27), and the deoxy analogue of 5-azacytidine was also recently approved for the same indication (28). Both drugs produce remissions or clinical improvements in more than half of the patients treated (29, 30). Features of responses include the requirement for multiple cycles of therapy, slow responses, and actual clonal elimination (based on cytogenetic changes). Optimization of

therapy has included reducing the dose to favor hypomethylation over cytotoxicity (31), prolonging administration schedules (32), and increasing dose intensity (within low doses; ref. 30). Side effects have been primarily hematologic, with no unexpected problems, chromosomal changes, or secondary malignancies (to date; ref. 33). Molecularly, hypomethylation and gene reactivation have been shown (30, 34, 35) and seem to be required for responses (30). All the data accumulated thus far are consistent with an epigenetic effect of these drugs *in vivo*, leading to clinical responses via clonal elimination, although the precise mechanism of clearance of neoplastic cells is unknown. Given the plethora of potential effects of this therapy, the mechanism may vary in different patients and likely includes a combination of induction of senescence, differentiation, apoptosis, and perhaps clearance by immune activation in some cases. Although the therapy is effective, with complete responses lasting months to years in some patients, resistance seems to develop in the majority of patients, and the mechanisms of resistance are unknown.

The data in myelodysplastic syndrome represent a proof-of-concept for epigenetic therapy for cancer. Current data suggest that myeloid malignancies (myelodysplastic syndrome, acute myelogenous leukemia, and chronic myelogenous leukemia) are the neoplasms most sensitive to inhibitors of DNA methylation. There is no known reason why this should be true, however, and why solid tumors would not respond as well. Nevertheless, older studies suggested lack of activity for these agents in various solid tumors (36). Most of these were done with high doses, a limited number of exposure days, and typically response evaluation after one cycle, which are all factors that would reduce the apparent efficacy of these drugs. There may be pharmacologic or pharmacodynamic reasons that favor hematologic malignancies in this regard (drug uptake, proportion of proliferating cells, etc.), but the activity of these agents in solid tumors deserves a second look with appropriate dosing schedules. Indeed, there is already some evidence for activity of 5-aza-2'-deoxycytidine in malignant melanoma at low doses combined with interleukin-2 (37).

With the proof-of-concept at hand, current investigations are aimed at optimizing the results obtained with epigenetic therapy. This will undoubtedly entail combination therapy. Rational design of combinations includes combining different epigenetic therapies (DNA methylation and histone acetylation inhibitors based on *in vitro* synergy; ref. 38), combining hypomethylating agents with drugs that exploit gene activation (e.g., *all-trans* retinoic acid), and attempting to integrate epigenetic therapy with more standard therapy. Two trials have recently been reported that have combined hypomethylating agents with histone deacetylase inhibitors [5-azacytidine and phenylbutyrate (32); decitabine and valproic acid (39)]. Both trials documented epigenetic effects of the combination and concluded that the therapy was promising, although it is clear that randomized trials will be required to definitely establish synergy (or even additive effects) between the two classes of drugs. Vorinostat (suberoylanilide hydroxamic acid), a highly potent histone deacetylase inhibitor, was recently approved by the Food and Drug Administration for the treatment of cutaneous T-cell lymphoma (40), and combinations of this drug with potent

DNA methylation inhibitors are eagerly awaited. Combinations of hypomethylating agents and biological agents are also ongoing. Promising results have been described for the combination of decitabine with interleukin-2 in melanoma (37), and combinations of hypomethylating agents and retinoic acid are ongoing. Finally, integration of hypomethylating agents with standard therapies is also following rational designs. For example, *in vitro* studies have shown that decitabine reverses resistance to various anticancer agents *in vitro* (41), and a clinical trial of this approach in ovarian cancer is ongoing.

Conclusions

Hypomethylation therapy is a beautiful example of translational research at work. New findings on hypermethylation in cancer led to a reevaluation of hypomethylating drugs *in vitro* and *in vivo*, resulting in Food and Drug Administration–approved drugs that are helping patients live longer with fewer side effects than conventional cytotoxic therapy. It seems likely that the field of epigenetic therapy will grow exponentially, with new drugs and new indications discovered via a continued dialogue between the laboratory and the clinic.

References

- Bird A. DNA methylation patterns and epigenetic memory. *Genes Dev* 2002;16:6–21.
- Jenuwein T, Allis CD. Translating the histone code. *Science* 2001;293:1074–80.
- Zamore PD. RNA interference: listening to the sound of silence. *Nat Struct Biol* 2001;8:746–50.
- Lapeyre JN, Becker FF. 5-Methylcytosine content of nuclear DNA during chemical hepatocarcinogenesis and in carcinomas which result. *Biochem Biophys Res Commun* 1979;87:698–705.
- Feinberg AP, Vogelstein B. Hypomethylation of ras oncogenes in primary human cancers. *Biochem Biophys Res Commun* 1983;111:47–54.
- Ehrlich M. DNA hypomethylation, cancer, the immunodeficiency, centromeric region instability, facial anomalies syndrome and chromosomal rearrangements. *J Nutr* 2002;132:2424–9S.
- Gaudet F, Hodgson JG, Eden A, et al. Induction of tumors in mice by genomic hypomethylation. *Science* 2003;300:489–92.
- Baylin SB, Herman JG, Graff JR, Vertino PM, Issa JPJ. Alterations in DNA methylation—a fundamental aspect of neoplasia. *Adv Cancer Res* 1998;72:141–96.
- Herman JG, Baylin SB. Gene silencing in cancer in association with promoter hypermethylation. *N Engl J Med* 2003;349:2042–54.
- Toyota M, Issa JP. Epigenetic changes in solid and hematopoietic tumors. *Semin Oncol* 2005;32:521–30.
- Jones PA, Laird PW. Cancer epigenetics comes of age. *Nat Genet* 1999;21:163–7.
- Laird PW, Jackson-Grusby L, Fazeli A, et al. Suppression of intestinal neoplasia by DNA hypomethylation. *Cell* 1995;81:197–205.
- Belinsky SA, Klinge DM, Stidley CA, et al. Inhibition of DNA methylation and histone deacetylation prevents murine lung cancer. *Cancer Res* 2003;63:7089–93.
- Cote S, Momparler RL. Activation of the retinoic acid receptor β gene by 5-aza-2'-deoxycytidine in human DLD-1 colon carcinoma cells. *Anticancer Drugs* 1997;8:56–61.
- Toyota M, Shen L, Ohe-Toyota M, Hamilton SR, Sinicrope FA, Issa JP. Aberrant methylation of the cyclooxygenase 2 CpG island in colorectal tumors. *Cancer Res* 2000;60:4044–8.
- Egger G, Liang G, Aparicio A, Jones PA. Epigenetics in human disease and prospects for epigenetic therapy. *Nature* 2004;429:457–63.
- Cheng JC, Matsen CB, Gonzales FA, et al. Inhibition of DNA methylation and reactivation of silenced genes by zebularine. *J Natl Cancer Inst* 2003;95:399–409.
- Gowher H, Jeltsch A. Mechanism of inhibition of DNA methyltransferases by cytidine analogs in cancer therapy. *Cancer Biol Ther* 2004;3:1062–8.
- Cornacchia E, Golbus J, Maybaum J, Strahler J, Hanash S, Richardson B. Hydralazine and procainamide inhibit T cell DNA methylation and induce autoreactivity. *J Immunol* 1988;140:2197–200.
- Chuang JC, Yoo CB, Kwan JM, et al. Comparison of biological effects of non-nucleoside DNA methylation inhibitors versus 5-aza-2'-deoxycytidine. *Mol Cancer Ther* 2005;4:1515–20.
- Siedlecki P, Boy RG, Musch T, et al. Discovery of two novel, small-molecule inhibitors of DNA methylation. *J Med Chem* 2006;49:678–83.
- Rhee I, Jair KW, Yen RW, et al. CpG methylation is maintained in human cancer cells lacking DNMT1. *Nature* 2000;404:1003–7.
- Robert MF, Morin S, Beaulieu N, et al. DNMT1 is required to maintain CpG methylation and aberrant gene silencing in human cancer cells. *Nat Genet* 2003;33:61–5.
- Rhee I, Bachman KE, Park BH, et al. DNMT1 and DNMT3b cooperate to silence genes in human cancer cells. *Nature* 2002;416:552–6.
- Yao X, Hu JF, Daniels M, et al. A methylated oligonucleotide inhibits IGF2 expression and enhances survival in a model of hepatocellular carcinoma. *J Clin Invest* 2003;111:265–73.
- Jouvenot Y, Ginja V, Zhang L, et al. Targeted regulation of imprinted genes by synthetic zinc-finger transcription factors. *Gene Ther* 2003;10:513–22.
- Issa JP, Kantarjian HM, Kirkpatrick P. Azacitidine. *Nat Rev Drug Discov* 2005;4:275–6.
- Kantarjian H, Issa JP, Rosenfeld CS, et al. Decitabine improves patient outcomes in myelodysplastic syndromes: results of a phase III randomized study. *Cancer* 2006;106:1794–803.
- Silverman LR, Demakos EP, Peterson BL, et al. Randomized controlled trial of azacitidine in patients with the myelodysplastic syndrome: a study of the cancer and leukemia group B. *J Clin Oncol* 2002;20:2429–40.
- Kantarjian H, Oki Y, Garcia-Manero G, et al. Results of a randomized study of three schedules of low-dose decitabine in higher risk myelodysplastic syndrome and chronic myelomonocytic leukemia. *Blood* 2007;109:52–7.
- Issa JP, Garcia-Manero G, Giles FJ, et al. Phase I study of low-dose prolonged exposure schedules of the hypomethylating agent 5-aza-2'-deoxycytidine (Decitabine) in hematopoietic malignancies. *Blood* 2004;103:1635–40.
- Gore SD, Baylin S, Sugar E, et al. Combined DNA methyltransferase and histone deacetylase inhibition in the treatment of myeloid neoplasms. *Cancer Res* 2006;66:6361–9.
- Yang AS, Estecio MR, Garcia-Manero G, Kantarjian HM, Issa JP. Comment on "Chromosomal instability and tumors promoted by DNA hypomethylation" and "Induction of tumors in mice by genomic hypomethylation." *Science* 2003;302:1153.
- Yang AS, Doshi KD, Choi SW, et al. DNA methylation changes after 5-aza-2'-deoxycytidine therapy in patients with leukemia. *Cancer Res* 2006;66:5495–503.
- Mund C, Hackanson B, Stresemann C, Lubbert M, Lyko F. Characterization of DNA demethylation effects induced by 5-aza-2'-deoxycytidine in patients with myelodysplastic syndrome. *Cancer Res* 2005;65:7086–90.
- Abele R, Clavel M, Dodion P, et al. The EORTC Early Clinical Trials Cooperative Group experience with 5-aza-2'-deoxycytidine (NSC127716) in patients with colo-rectal, head and neck, renal carcinomas and malignant melanomas. *Eur J Cancer Clin Oncol* 1987;23:1921–4.
- Gollob JA, Sciambi CJ, Peterson BL, et al. Phase I trial of sequential low-dose 5-aza-2'-deoxycytidine plus high-dose intravenous bolus interleukin-2 in patients with melanoma or renal cell carcinoma. *Clin Cancer Res* 2006;12:4619–27.
- Cameron EE, Bachman KE, Myohanen S, Herman JG, Baylin SB. Synergy of demethylation and histone deacetylase inhibition in the re-expression of genes silenced in cancer. *Nat Genet* 1999;21:103–7.
- Garcia-Manero G, Kantarjian HM, Sanchez-Gonzalez B, et al. Phase I/II study of the combination of 5-aza-2'-deoxycytidine with valproic acid in patients with leukemia. *Blood* 2006.
- Duvic M, Talpur R, Ni X, et al. Phase II trial of oral vorinostat (suberoylanilide hydroxamic acid, SAHA) for refractory cutaneous T-cell lymphoma (CTCL). *Blood* 2007;109:31–9.
- Plumb JA, Strathdee G, Sludden J, Kaye SB, Brown R. Reversal of drug resistance in human tumor xenografts by 2'-deoxy-5-azacytidine-induced demethylation of the hMLH1 gene promoter. *Cancer Res* 2000;60:6039–44.