Dexmedetomidine Attenuates the Microcirculatory Derangements Evoked by Experimental Sepsis

Marcos L. Miranda, M.D., M.Sc., Michelle M. Balarini, M.D., M.Sc., Eliete Bouskela, M.D., Ph.D.

ABSTRACT

Background: Dexmedetomidine, an \( \alpha \)-2 adrenergic receptor agonist, has already been used in septic patients although few studies have examined its effects on microcirculatory dysfunction, which may play an important role in perpetuating sepsis syndrome. Therefore, the authors have designed a controlled experimental study to characterize the microcirculatory effects of dexmedetomidine in an endotoxemia model that allows \textit{in vivo} studies of microcirculation.

Methods: After skinfold chamber implantation, 49 golden Syrian hamsters were randomly allocated in five groups: (1) control animals; (2) nonendotoxemic animals treated with saline; (3) nonendotoxemic animals treated with dexmedetomidine (5.0 \( \mu \)g kg\(^{-1} \) h\(^{-1} \)); (4) endotoxemic (lipopolysaccharide 1.0 mg/kg) animals treated with saline; and (5) endotoxemic animals treated with dexmedetomidine. Intravital microscopy of skinfold chamber preparations allowed quantitative analysis of microvascular variables and venular leukocyte rolling and adhesion. Mean arterial blood pressure, heart rate, arterial blood gases, and lactate concentrations were also documented.

Results: Lipopolysaccharide administration increased leukocyte rolling and adhesion and decreased capillary perfusion. Dexmedetomidine significantly attenuated these responses: compared with endotoxemic animals treated with saline, those treated with dexmedetomidine had less leukocyte rolling (11.8 \( \pm \) 7.2\% vs. 24.3 \( \pm \) 15.0\%; \( P < 0.05 \)) and adhesion (237 \( \pm \) 185 vs. 510 \( \pm \) 363; \( P < 0.05 \)) and greater functional capillary density (57.4 \( \pm \) 11.2\% of baseline values vs. 45.9 \( \pm \) 11.2\%; \( P < 0.05 \)) and erythrocyte velocity (68.7 \( \pm \) 17.6\% of baseline values vs. 54.4 \( \pm \) 14.8\%; \( P < 0.05 \)) at the end of the experiment.

Conclusions: Dexmedetomidine decreased lipopolysaccharide-induced leukocyte–endothelial interactions in the hamster skinfold chamber microcirculation. This was accompanied by a significant attenuation of capillary perfusion deficits, suggesting that dexmedetomidine yields beneficial effects on endotoxemic animals’ microcirculation. (\textit{Anesthesiology} 2015; 122:619-30)

EPISTEMES is a pathology highlighted by its high incidence, morbidity, mortality, and cost to healthcare system.\(^1\)\(^,\)\(^2\) Depending on its severity and duration, increased venular leukocyte adhesion and aggregation, microthrombosis, and microvascular vasconstriction may occur, leading to ischemia, impaired organ perfusion and function, and death. Thus, drugs that assist in reversal of microcirculatory changes could be decisive in sepsis treatment.\(^3\)

Many unique characteristics favor dexmedetomidine use during sepsis syndrome. This potent and highly selective \( \alpha \)-2 adrenergic receptor agonist evokes sedation combined with arousability, preserves spontaneous ventilation, and has been associated with reduced incidence of delirium and increased pressor response to norepinephrine.\(^4\)\(^,\)\(^5\)\(^,\)\(^7\) Unfortunately, few studies have examined the influence of \( \alpha \)-2 adrenergic receptor agonists on microcirculation and tissue perfusion in septic patients.

Dexmedetomidine has vasodilator effects that could assist in the recruitment of microcirculation.\(^3\)^\(^,\)\(^4\) Furthermore, \( \alpha \)-2 adrenergic receptor agonists have effects on immunity, inflammation, and coagulation.\(^9\)^\(^,\)\(^10\) Together, these effects may contribute to restoration of microcirculatory function, improving tissue perfusion, and reducing organ failure.\(^11\)^\(^,\)\(^12\) Based on these findings, we hypothesized that dexmedetomidine could reverse the microcirculatory derangements evoked by sepsis. Thus, the current controlled experimental study was carried out to test this hypothesis in an endotoxemia rodent model that allows \textit{in vivo} studies of the microcirculation.
Materials and Methods

Experiments were performed on 49 male golden Syrian hamsters (Mesocricetus auratus; 95 to 105 g) housed one per cage under controlled conditions of light (12:12 h light–dark cycle) and temperature (21.0° ± 1.0°C), with free access to water and standard chow. All procedures were approved by the Rio de Janeiro State University Animal Care and Use Committee, Rio de Janeiro, RJ, Brazil (protocol number CEUA/060/2010) and are consistent with the United States National Institutes of Health Guide for the Care and Use of Laboratory Animals.13

Animal Preparation

The chamber implantation procedure has been described previously by Endrich et al.14 in details and is briefly shown in figure 1. Six days after chamber implantation (recovery period), animals were reanesthetized and the left carotid artery was catheterized (polyethylene-50 catheter) allowing continuous mean arterial blood pressure (MAP) and heart rate (HR) monitoring and blood sampling. The left jugular vein was also catheterized (polyethylene-10 catheter) for fluid infusion and drug injection. These catheters were tunneled under the skin, exteriorized at the dorsal side of the neck, filled with heparinized saline solution (40 IU/ml), and attached to the chamber frame with tape. Experiments were performed on awake animals after 24 h of catheter implantation.

MAP, HR, and Temperature Monitoring

Mean arterial blood pressure was continuously monitored during the study period through the arterial catheter and a blood pressure transducer (TSD104A; BIOPAC Systems, Goleta, CA). Analog pressure signals were digitized (MP100 Data Acquisition System; BIOPAC Systems) and processed using data acquisition software for cardiovascular experiments (AcqKnowledge Software v. 3.5.7, BIOPAC Systems). The software automatically determined HR from the pressure trace and expressed it as beats per min. Rectal and skinfold chamber temperatures were monitored with a beaded type K thermocouple probe (Wavetek 23XT, Wavetek Corporation, San Diego, CA). The body temperature of the hamsters was maintained with a heating pad placed near the animal controlled by a rectal thermistor (LB750, Uppsala Processdata AB, Uppsala, Sweden).

Intravital Microscopy

Intravital microscopy was performed as stated in figure 1. Acquired microcirculatory images were recorded as video files in digital media for later evaluation. Quantitative off-line analysis of all videos generated during the experiments was performed using Cap-Image 7.2, a computer-assisted image analysis system (Dr. Zeintl Biomedical Engineering, Heidelberg, Germany)15 by an investigator blinded to drug treatment. In each animal, 2 arterioles, 2 venules, and 10 capillary fields were chosen taking into account the absence of inflammation or bleeding in the microscopic field and the presence of histological landmarks that could facilitate the subsequent return to the same field, since the same vessels and capillary fields were studied throughout the experiment. Arteriolar and venular mean internal diameters were measured as the perpendicular distance (in micrometers) between the vessel walls. Arteriolar and venular blood flow velocities were determined by a semiquantitative score using an ordinal scale16: 0, no flow; 1, intermittent flow; 2, sluggish flow; 3, normal flow; and 4, supranormal flow. The functional capillary density (FCD) was considered to be the total length (in centimeters) of spontaneously erythrocyte-perfused capillaries per square centimeter of tissue surface area (cm²/cm²). Erythrocyte velocity in capillaries (Erythrocyte-Vel) was assessed by frame-to-frame analysis and determined as the ratio between the distance traveled by an erythrocyte and the time required for this displacement (expressed as mm/s). One capillary per quadrant of capillary field was studied in each animal during Erythrocyte-Vel assessment (40 capillaries in total). The selection of these capillaries was based on two criteria: they should be representative of the mean Erythrocyte-Vel of their quadrant and have good image quality for reliable analysis.

Evaluation of Venular Leukocyte–Endothelial Interactions

Intravital fluorescence microscopy was performed as stated in figure 1. According to their interaction with the microvascular endothelium, leukocytes were classified as passing, rolling, and adherent. Passing leukocytes were defined as leukocytes traversing an observed venular segment without sticking contact (adherence) to the endothelial lining. The number of passing leukocytes was expressed as cells per minute. Rolling leukocytes were defined as leukocytes moving along the endothelial lining at a velocity slower than that of the surrounding erythrocytes. The number of rolling leukocytes was expressed as a percentage of the number of passing leukocytes. A leukocyte was considered to be adherent to the venular endothelial lining if it remained stationary for more than 30 s. Adherent cells were counted in a 100 μm venular segment and the number of adherent leukocytes was expressed as the number of adherent cells per square millimeter of endothelial surface (cells/mm²), calculated from the venular mean internal diameter and length (100 μm), assuming cylindrical vessel geometry. One venule was studied in each animal, and a single period of 60 s was analyzed off-line for all cell counts.

Arterial Blood Gases and Lactate Analysis

Blood samples were withdrawn from the arterial catheter and immediately analyzed in a blood gas analyzer (CG4+ cartridge, i-STAT System, Abbott Laboratories, Abbott Park, IL) for pH, partial pressure of oxygen (PO₂), partial pressure of carbon dioxide (PCO₂), bicarbonate level (HCO₃⁻), base excess (BE), total carbon dioxide content (TCO₂), arterial oxyhemoglobin saturation (SatO₂), and arterial lactate concentrations.
Fig. 1. Animal preparation, intravital microscopy setup, and experimental protocol. (A) During anesthesia with sodium pentobarbital (90 mg/kg intraperitoneal) animals’ dorsal hair was depilated and the skin lifted away, creating a skinfold. (B) One layer of the skinfold was microsurgically excised in a circular area of 15 mm in diameter. (C and D) The remaining layer, consisting of epidermis, subcutaneous tissue, and thin striated skin muscle, was sandwiched between two titanium frames, one of which was covered with a circular cover glass, creating the window chamber (close-up view in E). After 7 days, unanesthetized animals were placed in a restraining plexiglass tube attached to an intravital microscope (Ortholux, Leitz, Wetzlar, Germany) equipped
Experimental Protocol

Animals were suitable for experiments if their baseline MAP, HR, and rectal temperature were within the normal range and if they showed no signs of inflammation or bleeding in the skinfold chamber. Included animals were randomly allocated in five groups: CONTROL (control animals; n = 5), SALINE–SALINE (nonendotoxic animals treated with normal saline [NaCl 0.9%; NS]; n = 10), SALINE–DEXMEDETOMIDINE (nonendotoxic animals treated with dexmedetomidine; n = 10), lipopolysaccharide–SALINE (endotoxic animals treated with NS; n = 12), and lipopolysaccharide–DEXMEDETOMIDINE (endotoxic animals treated with dexmedetomidine; n = 12).

After baseline determination of MAP, HR, and microcirculatory parameters, animals belonging to endotoxic groups (lipopolysaccharide–SALINE and lipopolysaccharide–DEXMEDETOMIDINE) received an intravenous injection of 1.0 mg/kg (0.2 ml) of *Escherichia coli* serotype 055:B5 lipopolysaccharide (Sigma-Aldrich, St. Louis, MO). Nonendotoxic groups (SALINE–SALINE and SALINE–DEXMEDETOMIDINE) received an intravenous injection of 0.2 ml of NS. One hour after lipopolysaccharide or NS administration, a continuous intravenous infusion of NS or dexmedetomidine solution (5.0 μg kg⁻¹ h⁻¹) was initiated and maintained at a 0.1 ml/h infusion rate for 3 h.

As shown in figure 1, sequential measurements of MAP, HR, and microcirculatory parameters were performed at five time points: at baseline (t0), just before the initiation of the continuous intravenous infusion of NS or dexmedetomidine (t1), and after 1 h (t2), 2 h (t3), and 3 h (t4) of infusion. After 3 h of continuous NS or dexmedetomidine infusion, all animals received an intravenous injection of 0.15 mg/kg (0.4 ml) of rhodamine 6G. After a 10-min waiting period, animals were assessed by intravital fluorescence microscopy for evaluation of venular leukocyte–endothelial interactions. Blood sampling for arterial blood gases and lactate analysis was performed at the end of the study period.

Hamsters allocated in the CONTROL group served as baseline controls for leukocyte–endothelial interactions, lactate, and arterial blood gases parameters. To measure these parameters, animals were submitted to a partial experimental protocol: after baseline determination of MAP and HR, animals bypassed NS or lipopolysaccharide administration and NS or dexmedetomidine 3-h infusion protocol, going directly to rhodamine 6G administration (evaluation of leukocyte–endothelial interactions) and blood sampling.

After arterial blood sampling, all animals were sacrificed by an intravenous overdose of sodium pentobarbital (>200 mg/kg; Hypnol 3%; Syntec, Cotia, SP, Brazil).

Statistical Analysis

Results are expressed as means ± SD for each group, unless otherwise noted. Sample size was based on previous experience with the endotoxemia and microcirculation models used. Normally, with these models it is possible to observe significant differences between groups with the inclusion of 5 to 6 animals per group. In this study, a larger number of animals (10 to 12 in each group) was used because of the lack of data regarding the use of dexmedetomidine in association with the hamster skinfold window chamber model. Endotoxic groups included a larger number of animals (10 to 12 in each group) was used because of the lack of data regarding the use of dexmedetomidine in association with the hamster skinfold window chamber model. Endotoxic groups included a larger number of animals (10 to 12 in each group) was used because of the lack of data regarding the use of dexmedetomidine in association with the hamster skinfold window chamber model. Endotoxic groups included a larger number of animals (10 to 12 in each group) was used because of the lack of data regarding the use of dexmedetomidine in association with the hamster skinfold window chamber model. Endotoxic groups included a larger number of animals (10 to 12 in each group) was used because of the lack of data regarding the use of dexmedetomidine in association with the hamster skinfold window chamber model. Endotoxic groups included a larger number of animals (10 to 12 in each group) was used because of the lack of data regarding the use of dexmedetomidine in association with the hamster skinfold window chamber model.

Statistical comparisons of normally distributed variables (assessed by Shapiro–Willk test) were performed using two-way ANOVA for repeated measures and one way ANOVA as appropriate, whereas Friedman and Kruskal–Wallis tests were used for other variables. When appropriate, an adequate test was used for post hoc analysis: Bonferroni method or Dunn multiple comparisons. Venuolar leukocyte–endothelial interactions were correlated with capillary perfusion changes during the experiment, as depicted by percentage reductions of functional capillary density and erythrocyte velocity in capillaries values. These correlations were performed using Spearman rank correlation. All statistical analyses were performed using GraphPad Prism 6.03 (GraphPad Software, La Jolla, CA) and the significance level was set as P value less than 0.05 for a two-tailed test.

Results

The average body weight of hamsters was 100.7 ± 3.2 g with no significant differences among groups. All animals...
survived the entire experimental protocol leaving no missing data for statistical analysis.

**Microcirculatory Parameters**

At baseline, there were no significant differences in arteriolar and venular mean internal diameters and blood flow velocities between study groups. In venules, lipopolysaccharide elicited vasodilation and decreased blood flow velocity. In arterioles, lipopolysaccharide induced moderate vasoconstriction and also decreased blood flow velocity. There were no significant differences in arteriolar and venular lipopolysaccharide responses between lipopolysaccharide–SALINE and lipopolysaccharide–DEXMEDETOMIDINE groups (table 1). At baseline, FCD and Erythrocyte-Vel did not significantly differ between study groups. In endotoxemic groups, FCD and Erythrocyte-Vel decreased after lipopolysaccharide administration. However, a major decrease was observed in lipopolysaccharide–SALINE group, which led to a significant difference between NS- and dexmedetomidine-treated endotoxemic animals. Thus, although significantly reduced when compared with nonendotoxemic animals, after lipopolysaccharide administration these variables exhibited significantly and maintained better temporal evolution in dexmedetomidine-treated animals than in those treated with NS (fig. 2).

**Evaluation of Venular Leukocyte–Endothelial Interactions**

The number of passing leukocytes decreased after endotoxemia induction ($P < 0.05$ vs. CONTROL or nonendotoxemic groups); statistically similar reductions were observed in lipopolysaccharide–SALINE and lipopolysaccharide–DEXMEDETOMIDINE groups (fig. 3). In endotoxemic animals, dexmedetomidine significantly reduced the number of rolling leukocytes to 49% of the value observed in lipopolysaccharide–SALINE group (fig. 3). Endotoxia significantly increased the number of adherent leukocytes in lipopolysaccharide–SALINE group, whereas there were no significant differences between CONTROL, SALINE–SALINE, SALINE–DEXMEDETOMIDINE, and lipopolysaccharide–DEXMEDETOMIDINE groups (fig. 3). Representative photomicrographs are shown in figure 4.

**Correlations between Venular Leukocyte–Endothelial Interactions and Capillary Perfusion Changes**

The number of passing leukocytes per minute was negatively correlated to the percentage reduction of FCD and Erythrocyte-Vel values during the experiment (from t0 to t4). On the other hand, the number of adherent leukocytes per square millimeters was positively correlated to the percentage reduction of these microcirculatory parameters. No other significant correlation was found (fig. 5).

**MAP, HR, and Temperature Alterations**

Mean arterial blood pressure and HR basal values were not significantly different among the experimental groups and were comparable to control values from healthy animals reported by the literature. Systemic administration of lipopolysaccharide elicited statistically similar reductions in MAP levels in lipopolysaccharide–SALINE and lipopolysaccharide–DEXMEDETOMIDINE groups (fig. 6). Lipopolysaccharide caused an initial increase (t1) followed by a significant decrease in HR (fig. 6). Dexmedetomidine induced a significant HR decrease in nonendotoxemic and endotoxemic animals, but a major decrease was observed in the endotoxemic group (lipopolysaccharide–DEXMEDETOMIDINE; fig. 6). Considering rectal and skinfold chamber temperatures, there were no significant differences between NS- and dexmedetomidine-treated animals.

### Table 1. Arteriolar and Venular Mean Internal Diameters and Blood Flow Velocities

<table>
<thead>
<tr>
<th></th>
<th>Arteriolar Mean Internal Diameter (μm)</th>
<th>Arteriolar Blood Flow Velocity</th>
<th>Venular Mean Internal Diameter (μm)</th>
<th>Venular Blood Flow Velocity</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>SALINE–SALINE</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>t0</td>
<td>66.3 ± 12.0</td>
<td>3 (3/3)</td>
<td>72.1 ± 15.6</td>
<td>3 (3/3.5)</td>
</tr>
<tr>
<td>t4</td>
<td>69.8 ± 11.7*</td>
<td>3 (3/3)*</td>
<td>70.6 ± 13.6</td>
<td>3 (3/3)</td>
</tr>
<tr>
<td><strong>SALINE–DEXMEDETOMIDINE</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>t0</td>
<td>61.9 ± 7.4</td>
<td>3 (3/3)</td>
<td>66.1 ± 11.9</td>
<td>3 (3/4)</td>
</tr>
<tr>
<td>t4</td>
<td>61.4 ± 10.7</td>
<td>3 (3/3)</td>
<td>61.9 ± 8.0*</td>
<td>3 (3/3)</td>
</tr>
<tr>
<td><strong>Lipopolysaccharide–SALINE</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>t0</td>
<td>66.7 ± 11.6</td>
<td>3 (3/3)</td>
<td>69.9 ± 10.1</td>
<td>4 (3/4)</td>
</tr>
<tr>
<td>t4</td>
<td>53.0 ± 12.8†</td>
<td>2 (2/3)</td>
<td>83.0 ± 12.†</td>
<td>3 (2/3)‡</td>
</tr>
<tr>
<td><strong>Lipopolysaccharide–DEXMEDETOMIDINE</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>t0</td>
<td>66.8 ± 13.0</td>
<td>3 (3/3)</td>
<td>75.2 ± 11.8</td>
<td>4 (3/4)</td>
</tr>
<tr>
<td>t4</td>
<td>54.8 ± 16.9†</td>
<td>2 (2/3)</td>
<td>82.1 ± 13.2‡</td>
<td>2 (2/2.5)§</td>
</tr>
</tbody>
</table>

Mean internal diameters are expressed as means ± SD for each group. Blood flow velocities are expressed as median (Q1/Q3). t0, baseline; t4, 3h after normal saline (NS) or dexmedetomidine infusion.

* $P < 0.05$ as compared with endotoxemic groups at the same time point. † $P < 0.0001$ as compared with group baseline. ‡ $P < 0.05$ as compared with group baseline. § $P < 0.05$ as compared with nondotoxemic groups at the same time point.

Lipopolysaccharide–DEXMEDETOMIDINE = endotoxemic, dexmedetomidine treated (n = 12); Lipopolysaccharide–SALINE = endotoxemic, NS treated (n = 12); SALINE–DEXMEDETOMIDINE = nondotoxemic, dexmedetomidine treated (n = 10); SALINE–SALINE = nondotoxemic, NS treated (n = 10).
A significant increase in arterial lactate concentration was observed in endotoxemic groups (lipopolysaccharide–SALINE and lipopolysaccharide–DEXMEDETOMIDINE); however, there were no statistical differences between NS- and dexmedetomidine-treated groups (table 2). By contrast, a marked reduction in arterial HCO₃, BE, and TCO₂ was observed in lipopolysaccharide–SALINE group, which led
to a significant difference between NS- and dexmedetomidine-treated endotoxemic animals (table 2). \( PO_2 \) increased after endotoxia induction. \( PCO_2 \) was not significantly different between study groups.

**Discussion**

In our study, lipopolysaccharide administration increased venular leukocyte rolling and adhesion and decreased functional capillary density and erythrocyte velocity in the hamster skinfold chamber microcirculation, and induced metabolic acidosis, whereas dexmedetomidine treatment significantly attenuated these responses. Therefore, our key result was that dexmedetomidine treatment in the setting of experimental sepsis has partially restored microcirculatory function, improving tissue perfusion.

The lipopolysaccharide dose used in our study was adjusted to affect microcirculatory parameters without the induction of severe hypotension, whereas dexmedetomidine dose (5.0 \( \mu g \text{ kg}^{-1} \text{ h}^{-1} \)) was chosen based on previously published experimental studies that showed that this dose was able to reduce cytokine levels in endotoxemic rodents, increasing survival rate.\(^\text{17-19} \) Although 5.0 \( \mu g \text{ kg}^{-1} \text{ h}^{-1} \) is a high dose compared to that recommended for human use, smaller animals tend to have higher drug needs in relation to their body mass due to their higher metabolic rates.\(^\text{20} \) With this dose, animals achieved a mild sedation state, assessed by a murine sedation scale,\(^\text{21} \) and had a decrease in HR. MAP was not significantly affected by the infusion. MAP and HR evolution observed in our study is in agreement with the literature. Despite theoretical concerns regarding a possible circulatory worsening with dexmedetomidine use in septic patients, studies have shown that in critically ill patients both cardiac output and blood pressure are satisfactorily maintained during dexmedetomidine infusion, regardless of the fall in HR.\(^\text{6,22-25} \) Indeed, \( \alpha \)-2 adrenergic receptor agonists may even restore pressor responsiveness to norepinephrine in experimental sepsis.\(^\text{7} \)

In regard to the microcirculation, \( \alpha \)-2 adrenergic receptor agonists have dual vasomotor effects: direct vasoconstriction (via “post”-synaptic \( \alpha \)-1 and “extra”-synaptic \( \alpha \)-2 adrenergic receptors when high plasma concentrations of dexmedetomidine are present, such as observed during bolus injection or fast infusion) and indirect vasodilatation (via central sympathetic inhibition, reduced peripheral release of norepinephrine, and nitric oxide dilation). Because nitric oxide dregulation and heterogeneity of production is one of the hallmarks of sepsis, it would be
expected that the vasoconstrictor effect prevails, reducing tissue and organic perfusion. However, no significant differences were observed in arteriolar and venular mean internal diameters between lipopolysaccharide–SALINE and lipopolysaccharide–DEXMEDETOMIDINE groups in our study.

Considering FCD and Erythrocyte-Vel temporal evolution, dexmedetomidine was associated with significant
attenuation of capillary perfusion deficits induced by lipopolysaccharide administration. Several factors are related to the microcirculatory impairment observed after endotoxemia induction, such as systemic hypotension and vasoconstriction. Furthermore, lipopolysaccharide evokes leukocyte–endothelial interactions, resulting in capillary obstruction by leukocyte plugs and increased presence of rolling or adherent leukocytes in venules, which may hamper adequate capillary flow. As better capillary perfusion in lipopolysaccharide injection.

Table 2. Arterial Blood Gases and Lactate

<table>
<thead>
<tr>
<th></th>
<th>Lactate (mmol/l)</th>
<th>pH</th>
<th>HCO₃ (mmol/l)</th>
<th>BE (mmol/l)</th>
<th>TCO₂ (mmol/l)</th>
<th>PCO₂ (mmHg)</th>
<th>PO₂ (mmHg)</th>
<th>SatO₂ (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>CONTROL</td>
<td>1.54±0.22</td>
<td>7.38±0.03</td>
<td>32.3±2.4</td>
<td>7.0±2.3</td>
<td>33.8±2.4</td>
<td>54.4±7.6</td>
<td>68.6±9.7</td>
<td>92.4±3.5</td>
</tr>
<tr>
<td>SALINE–SALINE</td>
<td>1.19±0.22</td>
<td>7.39±0.04</td>
<td>30.1±3.4</td>
<td>5.1±3.9</td>
<td>31.8±3.5</td>
<td>49.5±5.1</td>
<td>78.3±13.0</td>
<td>94.4±3.0</td>
</tr>
<tr>
<td>Lipopolysaccharide–DEXMEDETOMIDINE</td>
<td>1.36±0.57</td>
<td>7.40±0.04</td>
<td>30.4±1.4</td>
<td>5.6±1.5</td>
<td>31.9±1.7</td>
<td>48.6±5.5</td>
<td>94.2±20.9</td>
<td>96.6±1.4</td>
</tr>
<tr>
<td>Lipopolysaccharide–SALINE</td>
<td>2.99±0.72*</td>
<td>7.31±0.04†</td>
<td>24.4±2.1†</td>
<td>−1.7±2.7‡</td>
<td>25.9±2.2‡</td>
<td>48.0±4.1</td>
<td>106.7±31.9§</td>
<td>95.8±3.6</td>
</tr>
<tr>
<td>Lipopolysaccharide–DEXMEDETOMIDINE</td>
<td>2.65±0.38*</td>
<td>7.33±0.10</td>
<td>29.0±2.4</td>
<td>3.2±3.1</td>
<td>30.8±2.5</td>
<td>55.1±14.1</td>
<td>131.0±24.8*</td>
<td>97.7±3.4§</td>
</tr>
</tbody>
</table>

Blood sampling for arterial blood gases and lactate analysis was performed at the end of the study period. Values are expressed as means ± SD for each group.

*P < 0.05 as compared with CONTROL, SALINE–SALINE, or SALINE–DEXMEDETOMIDINE groups. †P < 0.05 as compared with any other group. §P < 0.05 as compared with CONTROL or SALINE–SALINE.
CONTROL = baseline controls (n = 5); Lipopolysaccharide–DEXMEDETOMIDINE = endotoxemic, dexmedetomidine treated (n = 12); Lipopolysaccharide–SALINE = endotoxemic, NS treated (n = 12); NS = normal saline; SALINE–DEXMEDETOMIDINE = nonendotoxemic, dexmedetomidine treated (n = 10); SALINE–SALINE = nonendotoxemic, NS treated (n = 10).

baseline conditions are reached again 3 h after lipopolysaccharide (when 20 to 30% of passing leukocytes are rolling) and an increase happens 8 h after lipopolysaccharide administration (to about 60% of the value of passing leukocytes). Lower pH, HCO₃, BE, and TCO₂ values found in lipopolysaccharide–SALINE group are indicative of metabolic acidosis, probably secondary to tissue hypoperfusion. So, the absence of metabolic acidosis in lipopolysaccharide–DEXMEDETOMIDINE group suggests better tissue perfusion in this group compared with lipopolysaccharide–SALINE group, corroborating our microcirculatory findings. In two previous studies, Taniguchi et al. have already observed an attenuation in the degree of acidosis in

Anesthesiology 2015; 122:619-30
dexmedetomidine-treated endotoxemic rats when compared with endotoxemic controls. Since there is a latency between perfusion deficit correction and normalization of hyperlactatemia in septic patients, we hypothesize that the short treatment period with dexmedetomidine has been insufficient for a decrease in lactate concentrations in lipopolysaccharide–DEXMEDETOMIDINE group. The contradictory PO2 increase after lipopolysaccharide injection has been previously observed in endotoxemic hamsters and is likely to be related to an adaptive characteristic of the species to different oxygen consumption situations.

**Perspectives and Limitations**

Propofol, benzodiazepines, and dexmedetomidine are commonly used in intensive care units. Studies addressing the effects of propofol and midazolam on microcirculation agree that they exert negative effects, even in nonseptic states, leading to microcirculatory derangements. Conversely, we have shown that dexmedetomidine preserves microcirculatory parameters in nonendotoxemic animals and attenuates the effects of lipopolysaccharide on microcirculation. If these findings are replicated in humans, the clinical implication is that dexmedetomidine may improve microcirculatory function of septic patients, which is in line with many of the beneficial effects observed experimentally and clinically with dexmedetomidine use.

Our study has some limitations. First, even though lipopolysaccharide administration may reproduce many clinical features of sepsis syndrome it is in fact a model of experimental endotoxemia, so we cannot generalize/translate our results to the much more complex clinical scenario of human sepsis. Second, the absence of fluid resuscitation and nor- epinephrine infusion in our study further limits the generalizability of our results as both treatments are commonly used in human sepsis and have significant microcirculatory effects. In our study animals were not fluid resuscitated because this study was designed to evaluate the effects of dexmedetomidine on the microcirculation independently of fluid therapy effects. For a similar reason, we did not control lipopolysaccharide-induced changes in MAP with nor- epinephrine infusion (returning MAP to baseline values) in endotoxemic animals. Third, the study of the skin and subcutaneous muscle microcirculation may not be representative of microcirculatory changes in organs of crucial importance in the pathophysiology of sepsis, such as splanchnic organs, limiting our conclusions. However, the hamster skinfold window chamber model is widely used for microvascular studies in endotoxemic animals because the first reactions after lipopolysaccharide administration seem to be comparable in different tissues and organs. Furthermore, unlike other microcirculatory models, this one permits the existence of a recovery period between surgical manipulation for chamber implantation and the actual experiments, allowing recovery of microcirculatory function affected by surgical trauma. In addition, experiments can be performed without background anesthesia, which has microcirculatory effects of its own. Fourth, we did not attempt to delineate a specific dexmedetomidine action via α-2 adrenergic receptor (effect reversal with idazoxan) or via imidazoline receptor (effect reversal with efaroxan). Finally, central sympathetic actions of dexmedetomidine could potentiate the cholinergic antiinflammatory pathway. Indeed, several studies have shown that dexmedetomidine reduces plasma concentration of proinflammatory cytokines in septic animals and humans. Since we did not quantitate these cytokines (lack of standardized laboratory tests for golden Syrian hamsters), we cannot make correlations between our leukocyte–endothelial interactions results and the inflammatory response.

**Conclusions**

In our study, dexmedetomidine administration was effective in reducing lipopolysaccharide-induced leukocyte–endothelial interactions in the hamster skinfold chamber microcirculation. This was accompanied by a significant attenuation of capillary perfusion deficits. These results suggest that dexmedetomidine yields beneficial effects on endotoxemic animals’ microcirculation. Further studies in experimental models closer to human sepsis are required to confirm this benefit.

**Acknowledgments**

The authors thank Eliane Furtado, B.Sc. (Laboratory for Clinical and Experimental Research in Vascular Biology—BioVasc, Biomedical Center, Rio de Janeiro State University, Rio de Janeiro, RJ, Brazil), for her surgical and intravital microscopy support; Maria-das-Graças Souza, Ph.D. (Laboratory for Clinical and Experimental Research in Vascular Biology—BioVasc, Biomedical Center, Rio de Janeiro State University); Flávia Sousa, B.Sc. (Laboratory for Clinical and Experimental Research in Vascular Biology—BioVasc, Biomedical Center, Rio de Janeiro State University), and Fátima Cyrino, B.Sc. (Laboratory for Clinical and Experimental Research in Vascular Biology—BioVasc, Biomedical Center, Rio de Janeiro State University), for their technical assistance, and Cláudio Ribeiro, L.A.T. (Laboratory for Clinical and Experimental Research in Vascular Biology—BioVasc, Biomedical Center, Rio de Janeiro State University), and Paulo Lopes, L.A.T. (Laboratory for Clinical and Experimental Research in Vascular Biology—BioVasc, Biomedical Center, Rio de Janeiro State University), for their help with animal care.

This study was supported by grants from CNPq (National Council for Scientific and Technological Development, Brasilia, Brazil; grant number 303836/2010–5) and FAPERJ (State of Rio de Janeiro Agency for Research Support, Rio de Janeiro, Brazil; grant number E26/110.754/2012).

**Competing Interests**

The authors declare no competing interests.

**Correspondence**

Address correspondence to Dr. Miranda: Laboratory for Clinical and Experimental Research in Vascular Biology—
BioVasc, Pavilhão Reitor Haroldo Lisboa da Cunha, Rio de Janeiro State University, Rua São Francisco Xavier 524, 20550-013 Rio de Janeiro, RJ, Brazil. marcoslmiranda@gmail.com. Information on purchasing reprints may be found at www.anesthesiology.org or on the masthead page at the beginning of this issue. ANESTHESIOLOGY’s articles are made freely accessible to all readers, for personal use only, 6 months from the cover date of the issue.

References


11. Yeh YC, Sun WZ, Ko WJ, Chan WS, Fan SZ, Tsai JC, Lin TY; Dexmedetomidine prevents alterations of intestinal microcirculation that are induced by sepsis in a novel rat model. Anesth Analg 2012; 115:46–53


32. Yeh YC, Sun WZ, Ko WJ, Chan WS, Fan SZ, Tsai JC, Lin TY; Dexmedetomidine prevents alterations of intestinal microcirculation that are induced by sepsis in a novel rat model. Anesth Analg 2012; 115:46–53


Downloaded from anesthesiology.pubs.asahq.org by guest on 04/23/2019
38. Hoffmann JN, Vollmar B, Laschke MW, Inthorn D, Schildberg FW, Menger MD: Hydroxyethyl starch (130 kD), but not crystalloid volume support, improves microcirculation during normotensive endotoxemia. Anesthesiology 2002; 97:460–70