

# Neural Control of Inflammation

## *Implications for Perioperative and Critical Care*

Benjamin E. Steinberg, M.D., Ph.D., Eva Sundman, M.D., Ph.D., Niccolo Terrando, Ph.D., Lars I. Eriksson, M.D., Ph.D., F.R.C.A., Peder S. Olofsson, M.D., Ph.D.

### ABSTRACT

Inflammation and immunity are regulated by neural reflexes. Recent basic science research has demonstrated that a neural reflex, termed the inflammatory reflex, modulates systemic and regional inflammation in a multiplicity of clinical conditions encountered in perioperative medicine and critical care. In this review, the authors describe the anatomic and physiologic basis of the inflammatory reflex and review the evidence implicating this pathway in the modulation of sepsis, ventilator-induced lung injury, postoperative cognitive dysfunction, myocardial ischemia–reperfusion injury, and traumatic hemorrhage. The authors conclude with a discussion of how these new insights might spawn novel therapeutic strategies for the treatment of inflammatory diseases in the context of perioperative and critical care medicine. (**ANESTHESIOLOGY 2016; 124:1174–89**)

**N**EURAL reflex circuits are the basic organizational units of the nervous system, capable of rapid and precise responses to a myriad of physiologic challenges in both health and disease. In particular, homeostatic autonomic reflexes regulate body temperature, heart rate, blood pressure, and a wide range of other organ functions.<sup>1</sup> Patients in perioperative or critical care are to variable extents unable to maintain homeostasis and fine-tune their internal physiology due to combinations of therapeutic interventions (*e.g.*, surgery and anesthesia) and disease.<sup>1</sup> When homeostatic reflexes fail, clinicians are tasked with replacing neural reflex control with biochemical monitoring and therapeutic interventions to support normal physiology.

Autonomic reflex circuits are composed of a sensory (afferent) arc that report to the central nervous system (CNS) and a motor (efferent) arc that project regulatory signals to target tissues. CNS integration of a multitude of sensory information allows for purposeful and rapid adaptation to changing demands. For example, the baroreflex regulates heart rate and blood pressure to optimize organ perfusion and adjust exchange of oxygen, carbon dioxide, and nutrients according to need<sup>2</sup> (fig. 1A). Detailed understanding of

this cardiovascular reflex has enabled clinicians to diagnose and treat hemodynamic instability effectively.

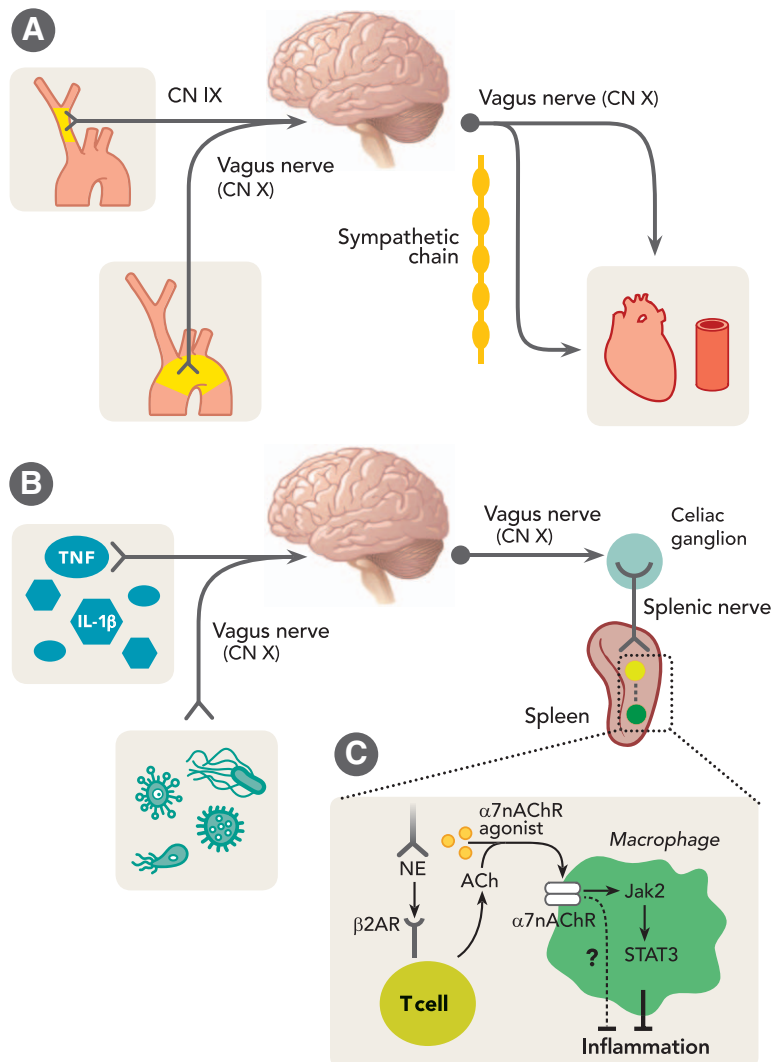
The inflammatory response is crucial for proper antimicrobial defense and healing after an aseptic injury; however, an excessive inflammatory response or failure to resolve the proinflammatory phase may lead to exaggerated tissue injury, circulatory shock, and death.<sup>3,4</sup> The available therapy for treatment of this unbalanced inflammatory reaction remains limited: steroidal and nonsteroidal antiinflammatory drugs, small-molecule compounds, and specific anticytokine drugs in clinical use are not selective to particular tissues and often produce serious undesirable side effects. For example, systemic anti-tumor necrosis factor (TNF) therapy, which has revolutionized the treatment of several chronic inflammatory conditions, may increase the risk of opportunistic bacterial, viral, and fungal infections.<sup>5,6</sup> The need for new, selective treatment options in inflammation is, therefore, pressing.<sup>7</sup>

In this context, the identification of the so-called “inflammatory reflex” provided the first description of a neural circuit capable of providing information in real time to the brain about the body’s inflammatory status to allow for rapid neural regulatory responses.<sup>8,9</sup> Yet, the neural reflexes that monitor and

This article is featured in “This Month in Anesthesiology,” page 1A. Figures 1 to 4 were enhanced by Annemarie B. Johnson, C.M.I., Medical Illustrator, Vivo Visuals, Winston-Salem, North Carolina. James C. Eisenach, M.D., served as Handling Editor for this article. Drs. Eriksson and Olofsson share senior authorship.

Submitted for publication June 4, 2015. Accepted for publication January 6, 2016. From the Department of Anesthesia, University of Toronto, Toronto, Ontario, Canada (B.E.S.); Laboratory of Biomedical Science, The Feinstein Institute for Medical Research, Manhasset, New York (B.E.S., P.S.O.); Section for Anesthesiology and Intensive Care Medicine, Department of Physiology and Pharmacology, Karolinska Institutet and Karolinska University Hospital, Stockholm, Sweden (E.S., L.I.E.); Department of Anesthesiology, Duke University, Durham, North Carolina (N.T.); Department of Anesthesia, Surgical Services and Intensive Care, Karolinska University Hospital, Stockholm, Sweden (L.I.E.); and Department of Medicine, Center for Molecular Medicine, Karolinska Institutet, Stockholm, Sweden (E.S., P.S.O.).

Copyright © 2016, the American Society of Anesthesiologists, Inc. Wolters Kluwer Health, Inc. All Rights Reserved. Anesthesiology 2016; 124:1174–89



**Fig. 1.** Reflex structure and function. (A) The baroreflex is a well-characterized reflex that maintains blood pressure. Like other reflexes, its anatomy consists of a sensory branch coupled with a motor output. The sensory component includes baroreceptors within the aortic arch and carotid sinus, which send information about blood pressure to the central nervous system *via* the glossopharyngeal (CN IX) and vagus nerves (CN X), respectively. Hypertension activates the reflex leading to cholinergic activation and adrenergic inhibition. This manifests as decreased heart rate and peripheral resistance and ultimately decreased blood pressure. Hypotension has the opposite effect and thereby increases blood pressure. (B) The inflammatory reflex similarly contains sensory and motor branches. In this case, vagus nerve sensory afferents are activated by the products of inflammatory and infectious stimuli. This information is conveyed to the brainstem. After integration by the central nervous system, the reflex is completed by sending vagus motor signals to the celiac ganglion where the splenic nerve arises. (C) The splenic nerve terminates in close proximity to a specialized acetylcholine-producing T cell in the spleen. This T cell behaves similarly to an interneuron: norepinephrine (NE) released by the splenic nerve activates  $\beta_2$  adrenergic receptors ( $\beta_2$ ARs) on the T cell, which in turn releases acetylcholine (ACh). The ACh engages the  $\alpha_7$  nicotinic acetylcholine receptor ( $\alpha_7$ nAChR) on splenic macrophages and down-regulates their production of tumor necrosis factor (TNF) resulting in an antiinflammatory effect. The intracellular mechanism for  $\alpha_7$ nAChR-mediated regulation of cytokine production in immune cells may involve Janus Kinase (Jak) 2 and signal transducer and activator of transcription (STAT) 3 signaling. Pharmacologic  $\alpha_7$ nAChR agonists (yellow circles) that activate the inflammatory reflex are being developed as potential antiinflammatory therapies. CN = cranial nerve; IL-1 $\beta$  = interleukin-1 $\beta$ .

respond to inflammatory stimuli in real time remain oftentimes overlooked. Recent research on how peripheral neural networks both sense and respond to inflammation is providing a possible framework on which to build and implement novel clinical therapies based on the neural control of inflammation.<sup>10</sup>

In this review, we elaborate the anatomic and physiologic basis of the inflammatory reflex as the prototype

of inflammation-regulating neural circuits (section “The Inflammatory Reflex”) and review the evidence implicating this reflex in modulating clinical conditions (section “Clinical Implications of the Inflammatory Reflex”). We conclude with a discussion of active areas of research into the neuroimmune interface that aim to develop new therapeutics that exploit the nervous system to control dysregulated and

nonresolving inflammation (sections “Cholinergic Anti-inflammatory Pharmacologic Intervention and Bioelectronic Medicine” and “Other Neural Reflexes that Regulate Immunity”).

### The Inflammatory Reflex

The vagus nerve (“the wandering nerve”) is the longest of the cranial nerves and innervates the majority of the visceral organs including the lungs, liver, and intestine with both sensory and motor fibers. The majority of vagus nerve fibers are sensory, detect a broad spectrum of mechanical and chemical stimuli, and send the information to the brain stem.<sup>11</sup> Notably, these same fibers monitor peripheral inflammatory responses.

The work delineating the interplay between immune mediators and the sensory vagus nerve began with studies by Watkins *et al.*, demonstrating that subdiaphragmatic vagotomies prevent the normal stress and febrile responses elicited by systemic administration of interleukin-1 $\beta$ .<sup>12–14</sup> These physiologic responses were corroborated by direct electrophysiologic recordings from the afferent fibers of the hepatic branch of the vagus nerve in rats, where intraportal injection of interleukin-1 $\beta$  lead to a dose-dependent increase in afferent fiber activity.<sup>15,16</sup> Moreover, bacterial products may also elicit reflex activity mediated by the vagus nerve. Recently, Fairchild *et al.*<sup>17</sup> observed bradycardias within minutes of administering bacteria or fungi to mice and implicated the vagus nerve by demonstrating simultaneous activation of vagus nuclei in the brain stem. Together, these data suggest that the sensory arm of the vagus nerve can detect immune and inflammatory signals within viscera and convey that information to the brain (fig. 1B). It remains unclear, however, whether the vagus nerve itself is able to directly sense cytokines and bacterial products, if intermediate players are involved, or if both direct and indirect activation pathways are at play. An intriguing possibility is that the afferent fibers of the vagus nerve convey cytokine-specific information to the brain stem, conceivably allowing the CNS to engage differential neurophysiologic responses depending on the immunologic challenge.

In line with this postulate, recent studies of the carotid body suggest that this multimodal sensory organ also serves as a peripheral monitor of inflammation in addition to oxygen, carbon dioxide, and pH, relaying information *via* the carotid sinus nerve and the glossopharyngeal nerve to the brain stem.<sup>18,19</sup> Furthermore, specific sensory nerves have a capacity to directly detect the presence of bacteria to modulate inflammation.<sup>20</sup> Primary sensory neurons in the dorsal root and trigeminal ganglia of the peripheral nervous system express functional toll-like receptors, innate immune receptors that recognize structurally conserved microbial motifs and regulate sensory function including pain and pruritus.<sup>21–24</sup> Interestingly, the selective deletion from nociceptive sensory neurons of myeloid differentiation primary response gene 88, a downstream signaling molecule in the toll-like

receptor activation pathway, results in impaired innate and adaptive immunity.<sup>25</sup> These results suggest that bacterial products could directly modulate neuronal excitability in certain sensory neuron populations.

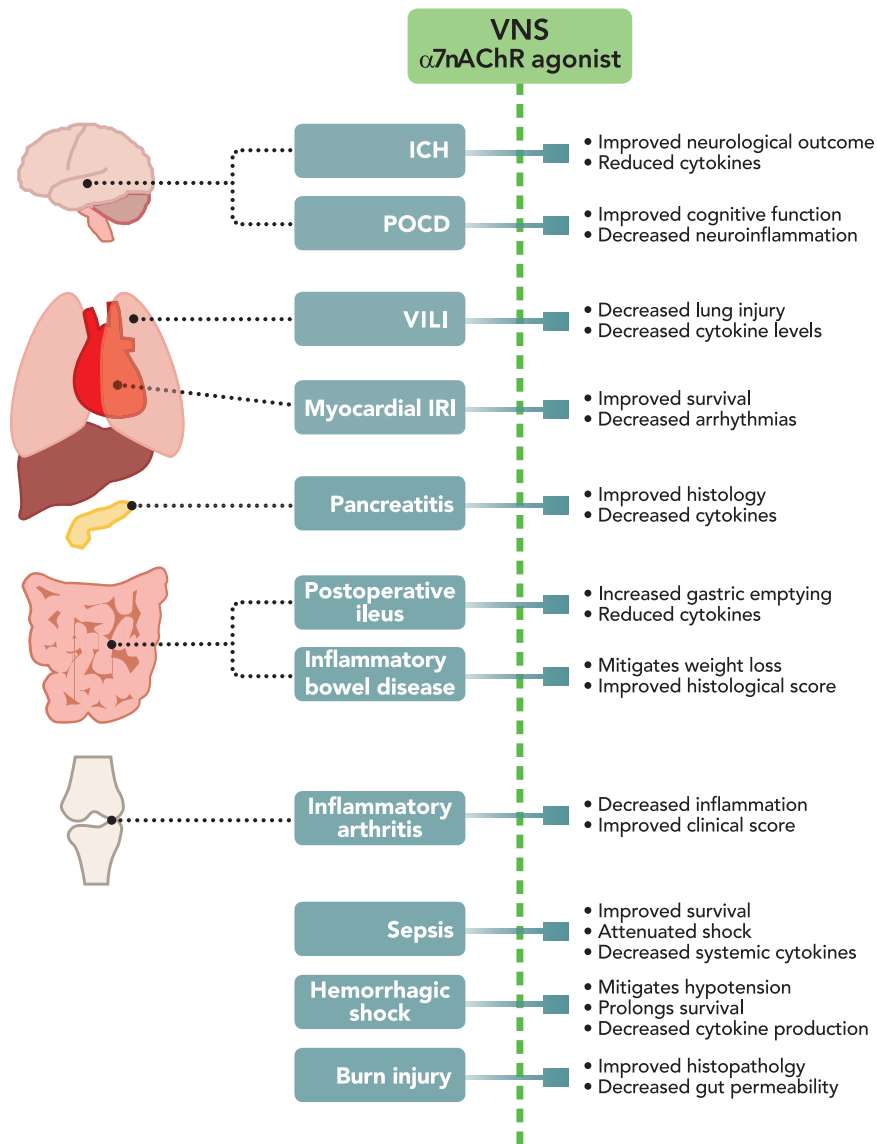
The efferent arc of the inflammatory reflex (fig. 1, B and C) was first defined by Borovikova *et al.*, who observed that electrical vagus nerve stimulation (VNS) reduced systemic levels of TNF in experimental models of severe systemic inflammation.<sup>26–28</sup> They also found that acetylcholine—the principal neurotransmitter of the vagus nerve—decreased proinflammatory cytokine production in stimulated macrophages.<sup>26</sup> Efferent fibers of the vagus nerve travel to the celiac ganglion where the splenic nerve originates. Splenic nerve axons terminate in close proximity to an acetylcholine-releasing subset of T cells.<sup>29,30</sup> Norepinephrine, the transmitter released by splenic nerve terminals, promotes acetylcholine release from these T cells. This cholinergic signal activates the homomeric neuronal subtype  $\alpha 7$  nicotinic acetylcholine receptor ( $\alpha 7$ nAChR) on immune cells, including macrophages resident within the spleen, and reduces their secretion of TNF.<sup>29,31</sup> The intracellular mechanism for  $\alpha 7$ nAChR-mediated regulation of cytokine production in immune cells is, however, not entirely clear, but it has been described to involve phosphatidylinositol-4,5-bisphosphate 3-kinase activation, Janus Kinase 2/signal transducer and activator of transcription 3 signaling, and inhibition of the assembly of the nuclear factor  $\kappa$ B complex in cells outside the CNS<sup>32–34</sup> (fig. 1C). Furthermore,  $\alpha 7$ nAChR in mitochondrial membranes may regulate interleukin-1 $\beta$  and high-mobility group box 1 in macrophages by inhibiting inflammasome activation through a mechanism involving mitochondrial DNA release.<sup>35</sup> Further studies of the intracellular mechanisms after  $\alpha 7$ nAChR activation in immune cells are clearly warranted. The physiologic effects, in cell culture and *in vivo*, are better known.<sup>9</sup> Mice devoid of  $\alpha 7$ nAChR do not respond to  $\alpha 7$  nicotinic acetylcholine ( $\alpha 7$ nACh) agonists or to electrical VNS with reduced TNF release, and higher levels of systemic TNF were observed in  $\alpha 7$ nAChR-deficient mice subjected to endotoxemia.<sup>31,36,37</sup> The  $\alpha 7$ nAChR subunit in immune cells is, therefore, a key component of the cholinergic anti-inflammatory pathway and an essential regulator of inflammation. Disruption of the integrity of the inflammatory reflex conversely inhibits resolution of inflammation.<sup>38</sup> Taken together, these data suggest that the inflammatory reflex tonically balances the production and release of inflammatory mediators and plays an important role in the resolution of inflammation.<sup>39</sup>

### Clinical Implications of the Inflammatory Reflex

Given these important effects of neural signaling on inflammation and immune system activity, it is conceivable that modulation of signals in the inflammatory reflex can be used to regulate inflammation and treat disease. In

fact, the inflammatory reflex has already been implicated as a potential therapeutic target across a variety of clinical conditions of regional and systemic inflammation<sup>26,34,39–59</sup> (fig. 2). In this context, a series of pharmacologic studies in experimental animals have demonstrated a key role for this neuronal pathway in inflammation and, in particular, the essential regulatory role of  $\alpha 7$ nAChR. Pharmacologic interventions using selective or nonselective  $\alpha 7$ nAChR agonists improve survival in experimental sepsis, reduce acute neuroinflammation, and result in improved cognitive performance after aseptic surgical injury.<sup>46,60–62</sup> Moreover, there are ongoing trials of the potential benefits from pharmacologic interventions using nicotinic agonists as well as clinical trials evaluating electrical VNS

as treatment for chronic inflammatory disorders such as rheumatoid arthritis and inflammatory bowel disease (clinicaltrials.gov: NCT01552941, NCT01569503, and NCT02311660).<sup>63</sup> These trials will evaluate the efficacy of  $\alpha 7$ nAChR agonists or VNS in these chronic conditions; the extensive preclinical and preliminary clinical data suggest promise with this approach.<sup>59,64</sup> Recent research indicates that selective pharmacologic intervention of the  $\alpha 7$ nAChR or direct electrical stimulation of the cervical vagus nerve may serve as novel treatment strategies in the management of sepsis, ventilator-induced lung injury (VILI), postoperative cognitive dysfunction (POCD), myocardial ischemia–reperfusion injury (IRI), and hemorrhage.



**Fig. 2.** Experimental disease models that respond to pharmacologic or electrical stimulation of the inflammatory reflex. A more detailed discussion of vagus nerve stimulation (VNS) in sepsis, ventilator-induced lung injury (VILI), postoperative cognitive dysfunction myocardial ischemia–reperfusion injury (IRI), and hemorrhage is provided in the main text. ICH = intracranial hemorrhage; POCD = postoperative cognitive dysfunction;  $\alpha 7$ nAChR =  $\alpha 7$  nicotinic acetylcholine receptor.

## Sepsis

Sepsis is a leading cause of worldwide morbidity and mortality and refers to a syndrome of florid systemic inflammatory response as triggered by a microbial infection. Its incidence continues to rise and it has a persistently high mortality.<sup>65</sup> At present, there are no specific therapies available targeting the inflammatory response to infection, and current treatment in sepsis is primarily based on early administration of antibiotics and mechanical removal of damaged tissues, typically combined with aggressive supportive intensive care therapy.<sup>66</sup> The paucity of therapeutic options that directly target the septic inflammatory syndrome partly reflects not only its heterogeneous etiology but also the lack of a more comprehensive understanding of its underlying pathophysiology. A dysregulated inflammatory response and impaired neuroendocrine signaling contribute to disease progression including multiorgan failure with subsequently increased morbidity and mortality.<sup>67</sup>

An association between changes in vagus nerve activity and systemic inflammatory responses has been observed. Analysis of heart rate variability, an indicator of vagus nerve activity, indicates that vagus nerve signals change before severe sepsis<sup>68</sup> and are associated with compromised cardiac function.<sup>69–71</sup> In line with these observations, early changes in heart rate variability recorded at admission in the emergency department have been associated with an increased risk of developing severe sepsis, septic shock, and death,<sup>69,72,73</sup> further implicating a potential role for the vagus nerve in septic patients.

In agreement with these clinical and experimental observations, vagotomy increases mortality in experimental models of sepsis<sup>42,60,74,75</sup> as does genetic ablation of the  $\alpha 7$ nAChR.<sup>37</sup> Reciprocally, activation of the inflammatory reflex by pharmacologic interventions using selective  $\alpha 7$ nAChR agonists or direct electrical stimulation of the vagus nerve is protective, yielding not only decreased proinflammatory cytokine burden but also increased survival,<sup>42,60,74</sup> a therapeutic benefit lost in  $\alpha 7$ nAChR-deficient mice.<sup>37</sup> Sepsis survivors commonly suffer from long-lasting organ dysfunction, including cognitive impairment, which can be reduced by antiinflammatory therapy after recovery from the acute episode in mice.<sup>76</sup> Interestingly, activation of the  $\alpha 7$ nAChR reduces microglial activation and cytokine release in CNS inflammation.<sup>77,78</sup> Hence, it is conceivable that activation of cholinergic signaling and the inflammatory reflex might be beneficial for mitigating organ dysfunction in sepsis survivors, although this remains to be studied. Notably, in a human trial, nicotine (a nonselective  $\alpha 7$ nAChR agonist) attenuated the inflammatory response in individuals exposed to intravenous endotoxin.<sup>79</sup> Hence, the  $\alpha 7$ nAChR is a key component of the inflammatory reflex, serves as a physiologic break on inflammation, and is an attractive pharmacologic target for the development of novel immunomodulatory pharmacologic therapeutics against systemic inflammatory disease and organ injury in acute inflammation due to sepsis and surgical trauma.

## Ventilator-induced Lung Injury

Mechanical ventilation in the perioperative or critical care setting renders patients susceptible to VILI through cycles of stretch and overinflation concomitant to damaging inflammatory responses, so-called biotrauma.<sup>80,81</sup> Multiple strategies to prevent VILI have focused on reducing the amount of mechanical damage to the lung tissue, while optimizing ventilation and gas exchange regimens, yet the clinical burden remains high.<sup>82</sup> Low-tidal volume as part of lung-protective ventilation strategies may still lead to increased release of inflammatory mediators<sup>83,84</sup> with subsequent risk for pulmonary edema and impaired gas exchange. This suggests that even with the development of novel lung-protective techniques, the inflammatory response needs to be addressed to fully abrogate the additional burden of VILI.

Experimental studies suggest that activation of the inflammatory reflex is beneficial in lung injury. For example, in burn-induced or hemorrhagic shock-induced acute lung injury mouse models, VNS reduced neutrophil infiltration into the lung as well as histologic lung injury.<sup>85,86</sup> Similarly, nicotine, choline, or a selective  $\alpha 7$ nAChR agonist, agents that activate target receptors in the inflammatory reflex, significantly reduce acid-induced lung injury.<sup>87</sup> Surgical vagotomy before the initiation of VILI, in contrast, lead to worsening of pulmonary inflammation and lung function, corroborating that the inflammatory reflex is a modulator of mechanical ventilation-induced biotrauma.<sup>53</sup> In reciprocal experiments, both pharmacologic and electrical stimulation of the efferent vagus nerve was protective in a model of VILI after hemorrhagic shock and resuscitation.<sup>53</sup> Similarly, stimulation of  $\alpha 7$ nAChR through the systemic administration of a partial agonist reduced TNF release and the alveolar–arterial gradient at clinically appropriate ventilation parameters.<sup>52</sup> In a recent study using a two-hit rodent lung model combining acute lung injury with barotrauma, neither pharmacologic intervention with nicotine nor VNS improved lung function, yet vagotomy lead to a worsening of the pulmonary cytokine response.<sup>88</sup> These results underscore the importance of further mechanistic studies aimed at delineating the involvement of the cholinergic system and the vagus nerve in modulating regional pulmonary and systemic inflammation. VILI offers a unique opportunity for intervention and study as the causative injury is initiated at a well-defined moment of patient care with the prescription of mechanical ventilation.

## Postoperative Cognitive Dysfunction

Surgery and trauma impair cognitive functions and affect a considerable proportion of the surgical population worldwide.<sup>89,90</sup> In surgical care, POCD is one of the most common long-term complications involving memory, learning, and attention capacity, which, when it occurs, impair postoperative rehabilitation and quality of life.<sup>91</sup>

Postoperative cognitive dysfunction develops within the first week after surgery and may remain for several months.

POCD is distinct from acute postoperative delirium, which lasts for hours or days, and from postoperative dementia, which represents a permanent reduction in higher cognitive functions. While reversible, POCD affects up to 30% of middle-aged and elderly patients at 1 week after surgery and 10% of elderly surgical patients at 3 months or even later.<sup>90,92,93</sup> Notably, the incidence of POCD is similar after regional or general anesthesia, indicating that general anesthesia *per se* has minimal direct influence on long-term deficits in cognition after surgery.

The pathophysiology of surgery-induced memory decline remains unclear although preclinical models suggest a role for surgery-induced systemic inflammation leading to activation of immune cells in the CNS with subsequent neuroinflammation<sup>94,95</sup> and neuronal dysfunction.<sup>96</sup> Surgery and tissue damage trigger an innate immune response *via* release of damage-associated molecular patterns such as high-mobility group box 1 and canonical proinflammatory cytokines (TNF, interleukin-1 $\beta$ , interleukin-6). This systemic inflammatory milieu leads to transient endothelial dysfunction and an impairment of the blood–brain barrier (BBB) integrity, which is associated with infiltration of peripheral immune cells into the brain parenchyma and later cognitive dysfunction.<sup>97,98</sup> In this context, activated peripheral macrophages appear to play a pivotal role by orchestrating a systemic release of inflammatory biomarkers combined with short-lasting BBB disintegration, ultimately resulting in macrophage migration into the CNS and distinct hippocampal neuroinflammation with neuronal impairment and ensuing cognitive dysfunction.<sup>46,99</sup> Recent experimental studies have demonstrated that prophylactic administration of  $\alpha 7$ nAChR agonists before surgery prevents trauma-induced neuroinflammation, BBB disruption, and subsequent cognitive decline by inhibiting TNF release and nuclear factor  $\kappa$ B activation in monocyte-derived peripheral macrophages.<sup>46</sup> Efforts to translate these findings into surgical patients by analysis of cerebrospinal fluid have revealed a timely increase in pro- and antiinflammatory molecules after major cardiac and orthopedic surgery,<sup>100–103</sup> suggesting that immune activation in the brain is present within 24 h in surgical patients. These findings demonstrate that therapeutics targeting cholinergic signaling within the inflammatory reflex pathway have the potential to provide a novel prevention and treatment strategy for POCD in humans.

### Myocardial Ischemia–Reperfusion Injury

Ischemic heart disease, common in perioperative and critical care patient populations, is typically treated with reperfusion therapies. A patient presenting emergently with an acute myocardial infarction may undergo reperfusion by thrombolytic therapy or primary percutaneous coronary intervention. Alternatively, stable ischemic heart disease may be treated by coronary artery bypass surgery. Timely intervention limits infarct size, preserves systolic function, and prevents the development of heart failure. Paradoxically,

however, the reperfusion process itself independently damages the myocardium, contributing up to 50% of the final infarct size.<sup>104,105</sup> The full extent of the cardiomyocyte death constitutes the IRI.

The pathophysiology of myocardial IRI is complex, involving oxidative stress, cardiomyocyte calcium overload, and mitochondrial dysfunction.<sup>106,107</sup> A robust inflammatory response accompanies an acute myocardial infarction, although the degree to which it is a direct contributor to the development of myocardial injury is unclear.<sup>106</sup> At minimum, the ischemic event and subsequent reperfusion lead to reactive oxygen species production and initiate a local and systemic inflammatory response and the release of proteases, TNF, and other cytokines.<sup>107–109</sup>

Advances directed at improving timely and effective reperfusion along with maintaining the patency of the diseased coronary vessel do not address the full extent of myocardial IRI.<sup>110</sup> To this end, VNS has emerged as a possible therapeutic strategy in the treatment of IRI, with multiple preclinical studies demonstrating that VNS is cardioprotective, improves ventricular function, and decreases arrhythmia in the setting of IRI.<sup>50,111–116</sup> Nevertheless, these animal studies have not been uniformly beneficial. Continuously applied right-sided VNS in a rabbit model of IRI produced increased infarct size, which was suspected to be due to increased sympathetic activation and catecholamine release.<sup>117</sup> Intermittent VNS, in contrast, decreased infarct size in an atropine-dependent fashion.<sup>118</sup> Similar results were observed in a swine IRI model, where left-sided intermittent VNS had a greater cardioprotective effect than a continuous stimulation protocol.<sup>119</sup>

The observed variability likely reflects important differences in the VNS protocols. For example, the decision to stimulate the right or left cervical vagus nerves, intermittently or continuously, may significantly influence the efficacy of the therapy. The right cervical vagus nerve has more efferent cardiac fibers than the left and is thought to have a greater influence on cardiac function.<sup>119</sup> More recently, noninvasive VNS strategies have been tried in IRI. Low-level transcutaneous stimulation of the auricular branch of the vagus nerve in a canine model of cardiac remodeling postmyocardial ischemia produced decreased infarct size and heart remodeling, improved systolic and diastolic function, and decreased systemic C-reactive protein and *N*-terminal probrain-type natriuretic peptide levels.<sup>120</sup>

Mechanistically, VNS during ischemia appears to decrease infarct size through nicotinic receptor activation<sup>112</sup>; however, the specific molecular pathways that are activated remain poorly defined. It has been postulated that VNS can induce a cardioprotective ischemic preconditioning-like state that includes activation of the phosphatidylinositol-4,5-bisphosphate 3-kinase pathway and inhibition of glycogen synthase kinase-3 $\beta$  by phosphorylation.<sup>118,121,122</sup> These signaling pathways ultimately reduce interstitial myoglobin, norepinephrine, and matrix metalloproteinase levels important in

tissue remodeling.<sup>123–125</sup> The detailed molecular mechanisms by which these VNS-activated pathways preserve cardiac function remains an active area of research and will likely continue to inform the design of future experimental VNS protocols in IRI management.

### Hemorrhage and Coagulopathy

Coagulopathy and hemorrhage are frequently encountered in high-energy blunt or penetrating trauma and in the critically ill patient populations. Traumatic hemorrhage remains one of the most common causes of preventable death,<sup>126</sup> with a notable morbidity and mortality burden in younger adults. Within the operating room, the ability to maintain hemostasis has benefitted from improvements in surgical approach; however, control of intraoperative bleeding can readily deteriorate in the setting of coagulopathy.

While surgical technique, coagulation factors, and blood products remain central to the management of hemorrhage, clinical best practices continue to evolve. Given the magnitude of this perioperative challenge, the development of so-called neural tourniquet approaches based on animal experiments has been proposed.<sup>127</sup> In particular, recent studies have mapped the effects of VNS and cholinergic agonists in experimental models of bleeding. VNS reduces bleeding time and blood loss in porcine traumatic injury by approximately 50%.<sup>47</sup> Notably, this was not a function of hemodynamic effects resulting from activation of vagus nerve cardiac fibers. In fact, the investigators found that the electrical stimulation as compared to sham controls had increased thrombin-antithrombin complex concentration at the site of injury, suggestive of a *bona fide* biochemical mechanism to the effect.<sup>47</sup> It is tempting to speculate that specific pharmacologic treatment or nerve stimulation strategies that physiologically facilitate coagulation might be used therapeutically pre-, intra-, and postoperatively to reduce blood loss and aid surgical hemostasis. We eagerly await future reports that will provide a greater mechanistic understanding of these observations in animal studies and inform possible novel clinical therapeutic interventions for the treatment of coagulopathies, traumatic injury, and intraoperative hemorrhage.

### Cholinergic Antiinflammatory Pharmacologic Intervention and Bioelectronic Medicine

With the cholinergic inflammatory reflex and the vagus nerve as a prototype, the role of neural control of inflammation is becoming increasingly recognized as a therapeutic target in clinical medicine. The animal studies discussed in the section “Clinical Implications of the Inflammatory Reflex” offer a framework for designing clinical trials that employ pharmacologic or electrical interventions to treat inflammatory conditions through cholinergic pathways.

### Pharmacologic Interventions of the Cholinergic System

Pharmacologically, the neuronal control of inflammation can be modulated by stimulation or inhibition of the cholinergic system using nicotinic  $\alpha 7$ nAChR agonists or antagonists, respectively. Since the initial demonstration of the role of the  $\alpha 7$ nACh receptor subtype for the regulation of inflammation, in particular peripheral macrophages,<sup>37</sup> a series of experimental studies on acute inflammation as triggered by either infection or trauma in animals and humans have demonstrated promising effects on outcomes.

Anesthesiologists and critical care physicians have a long-standing familiarity with medications that target the cholinergic system, such as acetylcholinesterase inhibitors for the reversal of nondepolarizing neuromuscular blockade. Yet, the approved agents used routinely in the perioperative setting have not been shown to have clinically significant antiinflammatory effects. Moreover, given the pleiotropic effects of direct and indirect cholinergic agonists, it is unlikely that these agents, used in isolation, will be at the forefront of pharmacologic interventions that target inflammation through the neuroimmune interface. Targeting the  $\alpha 7$ nAChR specifically will avoid the untoward side effects affecting locomotor activity and autonomic dysfunction that result from stimulation of other nicotinic receptors such as  $\alpha 3$ nACh,  $\alpha 4$ nACh,  $\alpha 5$ nACh, and  $\beta 2$ nAChR. To this end, investigators have been developing an array of specific small-molecule modulators of the  $\alpha 7$ nAChR activity.

GTS-21 (3-[2,4-dimethoxybenzylidene] anabaseine) was one of the first such  $\alpha 7$ nAChR agonists to be studied for its immune-modulating capacity. GTS-21 was found to suppress proinflammatory cytokine production in human macrophages stimulated by endotoxin<sup>128</sup> as well as improve survival in animal models of sepsis<sup>60</sup> and hemorrhage.<sup>129</sup> To extend these findings toward patient care, a double-blind placebo-controlled pilot human study was conducted in healthy volunteers exposed to intravenous endotoxin.<sup>130</sup> This study did not show any effect of the drug on proinflammatory cytokine serum levels compared with the placebo group; however, high plasma levels of the drug correlated with lower levels of TNF and interleukin-6.<sup>130</sup> The study was largely underpowered and limited by the considerable intersubject variability in cytokine levels and the achieved levels of GTS-21 or its active metabolite. It was further complicated by the incomplete information on the specificity of GTS-21 for human  $\alpha 7$ nAChR.<sup>131</sup>

The inflammatory reflex can likewise be activated by cholinergic agents that act at the level of the CNS. For example, galantamine, a Federal Drug Agency–approved drug for the treatment of Alzheimer disease, is a centrally acting acetylcholinesterase inhibitor that activates the efferent arm of the inflammatory reflex.<sup>132</sup> In an experimental model of endotoxemia in mice, galantamine decreased TNF levels and improved survival, an effect dependent on  $\alpha 7$ nAChR. Although galantamine acts centrally, the  $\alpha 7$ nAChR dependence presumably reflects the importance of this receptor in

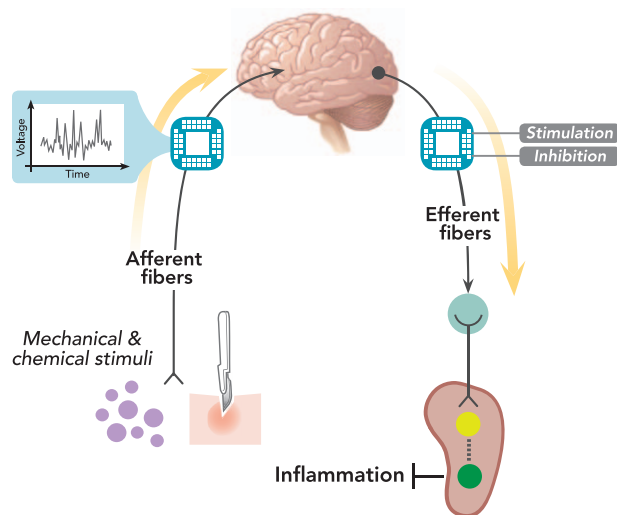
completing the antiinflammatory efferent arc at the level of the spleen.<sup>132</sup> Galantamine has been shown to be similarly beneficial in animal models of experimental colitis<sup>133</sup> and obesity-associated inflammation.<sup>134</sup> In the latter study, treatment of obese mice with galantamine decreased circulating levels of proinflammatory cytokines, such as interleukin-6, as well as decreased food intake and body weight while alleviating impaired glucose tolerance.<sup>134</sup> These encouraging preclinical data have since informed the design of a phase 4 clinical trial investigating the efficacy of galantamine in the treatment of patients with metabolic syndrome (clinicaltrials.gov: NCT02283242).

The overall strength of the preclinical findings and accumulating human data maintain the ongoing enthusiasm for pharmacologically targeting antiinflammatory cholinergic pathways. Moving forward, researchers are working toward specific neuronal nicotinic AChR agonists, including for the  $\alpha 7$ nAChR. To the best of our knowledge, as of yet, no clinical trials of these agents in perioperative or critical care have started but are greatly anticipated.

### Bioelectronic Medicine

Another approach to modulate the inflammatory reflex and the neural control of inflammation is by electrical nerve stimulation. Bioelectronic medicine<sup>63,135,136</sup> is the interdisciplinary field that brings together molecular medicine, neuroscience, engineering, and clinical medicine. The field holds great promise for targeted and specific therapy in the treatment of inflammatory diseases, for example, by modulating signals in neural reflexes (fig. 3). A current objective in the field at large is to develop electrical recording devices that interrogate the neural activity moving through the nerve and translate the neural code into an interpretable physiologic readout. While the field remains in its infancy, the confluence of improved electrical recording hardware, wireless technologies, and data analytics brings our ability to decipher an immunologic code within the realm of possibility. Ultimately, the goal is to develop either implantable or transcutaneous devices capable of recording and interpreting sensory information that inform ensuing modulatory signals in efferent nerves to support homeostasis and treat diseases.

The therapeutic use of electricity has a well-established precedence within the CNS. Deep brain stimulation is approved for treatment of Parkinson disease and depression. It was in fact within the context of illnesses of the CNS that VNS became recognized as a therapeutic modality. First used clinically nearly 30 yr ago, VNS was introduced for the treatment of epilepsy in 1988<sup>137</sup> and eventually approved in both the United States and Canada in 1997. In VNS, a bipolar helical electrode is surgically placed around the cervical vagus nerve, and an implanted stimulator similar to a conventional pacemaker delivers electrical charges that directly activate the vagus nerve. The efficacy of VNS in epilepsy is well defined across adult and pediatric populations with significant reductions in seizure frequency and duration along



**Fig. 3.** Bioelectronic medicine. Bioelectronic medicine treats disease through the use of electricity to activate or inhibit neural circuits. This therapeutic modality can interface with both the afferent and efferent branches of a neural reflex to modulate the amount of information propagating down the nerve fibers. The long-term objective of this therapeutic approach is to allow clinicians to record the electrical activity in the nerve so as to extract real-time information about patient status. This information can be incorporated into diagnosis and monitoring algorithms as well as inform therapeutic delivery of either electrical or pharmacologic treatment in a patient-specific fashion.

with improved postictal recovery.<sup>138–140</sup> The clinical indication for VNS has expanded to include chronic treatment-resistant depression,<sup>141–144</sup> and VNS is being explored in other diseases involving the CNS, such as migraine and eating disorders.<sup>145,146</sup>

More recently, clinical trials are exploiting the vagus nerve's control of chronic inflammatory responses in the context of autoimmune diseases, including inflammatory bowel disease and rheumatoid arthritis.<sup>64,147</sup> In a pilot, open-label study, eight patients with active rheumatoid arthritis were implanted with a commercially available vagus nerve stimulator and received 6 weeks of daily stimulation. According to preliminary data, the VNS-treated patients showed improved clinical symptoms, with the therapy being well tolerated by study participants.<sup>64,147</sup>

During the implantation procedure, the vagus nerve is stimulated to test device functionality. In less than 0.1% of patients, the test can elicit a bradyarrhythmia or transient asystole, although this adverse event has only been reported at the time of the intraoperative stimulation and resolves when stimulation is stopped.<sup>148</sup> Postoperative surgical site infection and vocal cord paresis are rare, occurring in approximately 3 to 6% and less than 1% of patients, respectively.<sup>148</sup> With use of the vagus nerve stimulator, the reported side effects include hoarseness, throat pain, and coughing and are largely limited to the periods of actual stimulation and commonly alleviate with time.<sup>148–150</sup> Importantly, a 1-min-long



daily electrical stimulation of the vagus nerve is sufficient to alleviate experimental inflammatory disease, with more recent findings indicating that even stimulation times of as little as 500  $\mu$ s, may be sufficient to significantly reduce an inflammatory response in animal models.<sup>151</sup> Treatment strategies with very low-stimulation duty cycles may allow for innovative, smaller stimulator designs with longer service intervals and fewer side effects in patients with chronic inflammation.

Although extensive experimental data in animal models indicate that VNS is effective for the treatment of acute inflammation, the effect on human acute inflammation has not been evaluated in clinical trials. At present, the only available data come from a few small studies in patients with VNS-treated epilepsy and demonstrate variable changes in serum cytokine levels associated with VNS.<sup>152–155</sup> These data are limited and largely inconclusive given that the patients studied had epilepsy but not inflammatory conditions *per se*. In another small study of 10 patients with treatment-resistant depression, both pro- and antiinflammatory cytokines increased in the 3 months after VNS device implantation.<sup>156</sup>

While medical device technology continues to evolve, VNS has yet to be adapted for temporary, preferably non-invasive, stimulation. Several initiatives are underway, and access to safe and reliable noninvasive methods for specific VNS would significantly facilitate clinical studies of the potential efficacy of VNS in perioperative and intensive care medicine.

### Other Neural Reflexes that Regulate Immunity

In addition to the inflammatory reflex, multiple other peripheral neural circuits that regulate inflammation have been described<sup>157,158</sup> (fig. 4). These include modulation of inflammatory pulmonary airway hyperresponsiveness and nociceptive neuron modulation of infection.

#### Airway Hyperresponsiveness

The lung contains a dense network of nociceptive neurons that respond to inhaled noxious chemical stimuli *via* transient receptor potential (TRP) channels, including TRP vanilloid 1 (TRPV1) and TRP ankyrin 1. When activated, these nociceptors initiate protective coughing reflexes and mucus secretion.<sup>159</sup> Airway hyperresponsiveness in the setting of chronic inflammation and increased mucus secretion are hallmarks of asthma. These features, including local inflammation, reflect activation of pulmonary airway afferent neural pathways. For example, in allergic and nonallergic mouse models of asthma, genetic ablation or pharmacologic inhibition of TRPV1 and TRP ankyrin 1 channels eliminates airway hyperresponsiveness,<sup>160,161</sup> which appears to be driven primarily by a subset of TRPV1-positive vagus nerve neurons.<sup>162</sup>

Stimulation of pulmonary nociceptors with capsaicin results in increased neuropeptide release and immune cell

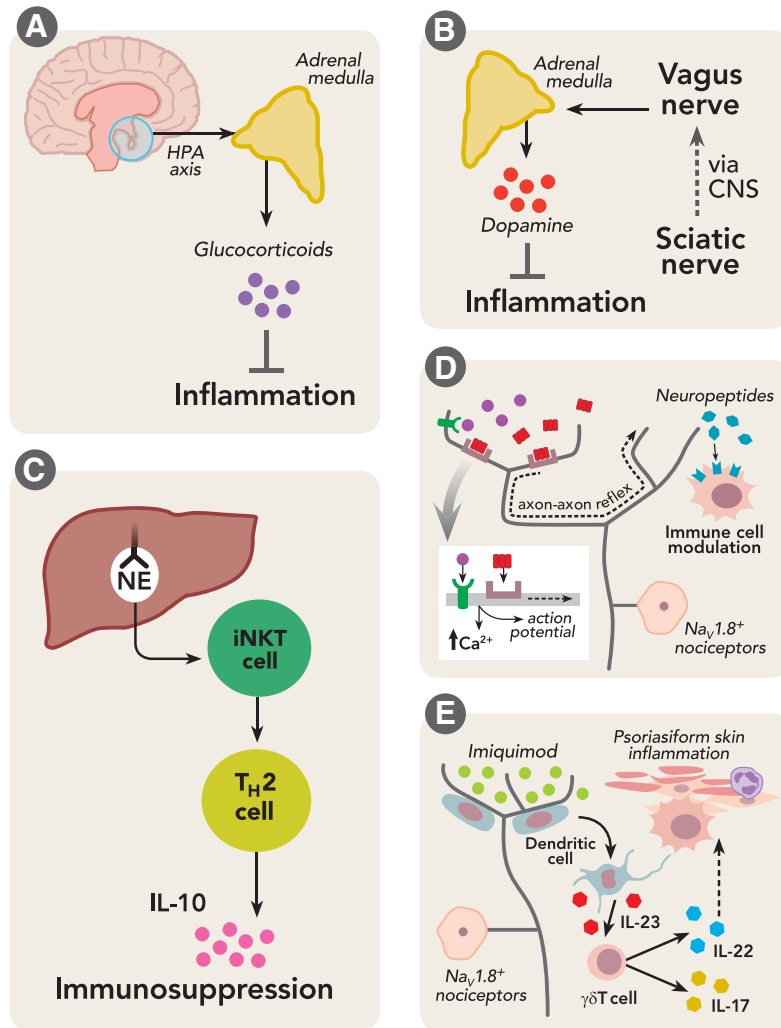
infiltration, whereas genetic ablation or pharmacologic inhibition of Na<sub>v</sub>1.8-positive neurons was protective against airway inflammation and hyperresponsiveness in murine models of allergic asthma.<sup>163</sup> Notably, interleukin-5, an important cytokine in eosinophil activation, was shown to directly stimulate nociceptors to release vasoactive intestinal peptide, which in turn activates resident immune cells and promotes the allergic response.<sup>163</sup> These data are consistent with earlier work that demonstrated that allergic asthma is diminished in mice with systemic denervation. In this case, denervation resulted in decreased interleukin-5 release and chemokine production, thereby limiting the number of eosinophils that infiltrated into the tissue.<sup>158,164,165</sup> Accordingly, current work is being directed toward whether pharmacologic inhibitors of pulmonary nociceptive pathways represent a new therapeutic target for asthma and inflammatory pulmonary disease.

#### Infection

While chemical irritants and sterile inflammatory pathways are modulated by peripheral neural networks, the same appears true for infectious inflammation. The dense network of peripheral nociceptive neurons ensures that invading microorganisms will encounter neural tissue independent of their point of entry. In turn, the peripheral nervous system interfaces with the immune system to modulate the inflammatory response to invading microorganisms. For instance, chemical ablation of sensory neurons in *Mycoplasma* infection increases tissue damage, tracheal thickness, and disease severity.<sup>166</sup> Similarly, genetic deletion of TRPV1 in mice subjected to experimental sepsis impairs bacterial clearance, worsens end-organ damage, and exacerbates a detrimental cytokine production.<sup>167</sup>

Recent work suggests that this relationship stems from direct activation of nociceptive neurons by bacterial toxins so as to initiate a rapid response to infection,<sup>20,168</sup> in part through the production of neuropeptides.<sup>169</sup> Ablation of Na<sub>v</sub>1.8-positive neurons before *Staphylococcus aureus* inoculation in mice lead to increased leukocyte infiltration to the site of infection and draining lymphadenopathy.<sup>20</sup> In this model, bacterial products, including *N*-formylated peptides and  $\alpha$ -hemolysin toxin, directly activate axon-axon reflexes *via* formyl peptide receptors and a disintegrin and metalloprotease 10, respectively, and thereby result in the release of antiinflammatory neuropeptide, such as calcitonin gene-related peptide (CGRP).<sup>20</sup> Na<sub>v</sub>1.8 neuron ablation prevents CGRP release and thus promotes the inflammatory phenotype. Conversely, administration of exogenous CGRP is protective against otherwise lethal endotoxemia, decreasing pro- and increasing antiinflammatory cytokines.<sup>170</sup>

Further examples of peripheral neural networks that modulate inflammatory processes continue to be described<sup>171–174</sup> and have been reviewed elsewhere.<sup>157,158</sup> It is likely that a large number of additional neural reflex circuits that regulate inflammation and immunity yet remain to be discovered. As our mechanistic understanding of these physiologic processes



**Fig. 4.** Examples of neural control of inflammatory processes. (A) Stimulation of the hypothalamic–pituitary–adrenal (HPA) axis initiates a neuroendocrine sequence resulting in glucocorticoid release and suppression of the inflammatory response. (B) Activation of the sciatic nerve by electroacupuncture inhibits cytokine release and improves survival in a mouse model of sepsis. The neural circuit maps from the sensory sciatic nerve to the efferent fibers of the vagus nerve. In this case, the vagus nerve signal results in the release of dopamine from the adrenal medulla. The dopamine engages dopaminergic type 1 receptors to suppress the inflammatory response. (C) After stroke, noradrenergic innervation of the liver signal to hepatic invariant natural killer T (iNKT) promotes immunosuppression. Blockade of adrenergic signaling (e.g., β-blockade with propranolol) reduces immunosuppression, protects against infection, and improves survival. (D) Infection with the bacterium *Staphylococcus aureus* results in an acute pain response caused by the direct activation of peripheral nociceptors by bacterial products. In addition to transducing a signal toward the central nervous system, receptor stimulation at the nerve terminal generates an antidromic axon–axon reflex that results in the release of neuropeptides that impair the recruitment and activation of locally infiltrating immune cells. (E) In an imiquimod-induced model of skin inflammation, a subset of nociceptors that express TRPV1 and Na<sub>v</sub>1.8 promote local inflammation through the induction of interleukin (IL)-23 production by skin-resident dendritic cells. In turn, the IL-23 activates other immune cells within the skin to secrete the IL-17 and IL-22 that ultimately propel psoriasiform skin inflammation. CNS = central nervous system; NE = norepinephrine; TH2 = T helper cell type 2.

increase, we predict that measuring and modulating activity of specific neural circuits using bioelectronic medicine will eventually become part of patient care in many diseases with an inflammatory component.

### Conclusion and Future Perspectives

The immune system, similar to other organ systems, is regulated by the CNS through neural reflexes, with the

inflammatory reflex involving α<sub>7</sub>nACh-dependent chemical neurotransmission and the vagus nerve being the most highly studied pathway. As inflammation is part of the pathogenesis of a diverse and important set of diseases commonly encountered in perioperative and critical care patients, this neural regulation may allow for new treatment possibilities. Approaching the neural control mechanisms of inflammation by novel pharmacologic and technical principles

provides a new and exciting possibility for prevention and treatment of acute and chronic inflammation.

Future research and clinical trials should focus on introducing pharmacologic interventions applying  $\alpha 7$ nAChR agonists in clinical practice and work to improve technology for recording nerve activity and delivering specific electrical signals to targeted nerves in order to ultimately mitigate the physiologic trespass of an acute traumatic or chronic inflammatory insult and improve patient health.

## Acknowledgments

This study was supported by a Clinician Investigator Program fellowship from the Ministry of Health (Ontario, Canada; to Dr. Steinberg), Svenska Läkaresällskapet (Stockholm, Sweden; to Dr. Olofsson), the Knut and Alice Wallenberg Foundation (Stockholm, Sweden; to Dr. Olofsson), and the Heart-Lung Foundation (Stockholm, Sweden; to Dr. Olofsson).

## Competing Interests

The authors declare no competing interests.

## Correspondence

Address correspondence to Dr. Olofsson: Center for Molecular Medicine, L8:03, Karolinska Institutet, Karolinska University Hospital, Stockholm, Sweden. peder.olofsson@ki.se. Information on purchasing reprints may be found at [www.anesthesiology.org](http://www.anesthesiology.org) or on the masthead page at the beginning of this issue. ANESTHESIOLOGY's articles are made freely accessible to all readers, for personal use only, 6 months from the cover date of the issue.

## References

- Glick DB: The autonomic nervous system, Miller's Anesthesia, 8th edition. Edited by Miller RD, Cohen NH, Eriksson LI, Fleisher LA, Wiener-Kronish JP, Young WL. Philadelphia, Elsevier Saunders, 2014, pp 346–86
- Thomas GD: Neural control of the circulation. *Adv Physiol Educ* 2011; 35:28–32
- Hotchkiss RS, Monneret G, Payen D: Immunosuppression in sepsis: A novel understanding of the disorder and a new therapeutic approach. *Lancet Infect Dis* 2013; 13:260–8
- Tracey KJ, Beutler B, Lowry SF, Merryweather J, Wolpe S, Milsark IW, Hariri RJ, Fahey TJ III, Zentella A, Albert JD: Shock and tissue injury induced by recombinant human cachectin. *Science* 1986; 234:470–4
- Ramiro S, Gaujoux-Viala C, Nam JL, Smolen JS, Buch M, Gossec L, van der Heijde D, Winthrop K, Landewé R: Safety of synthetic and biological DMARDs: A systematic literature review informing the 2013 update of the EULAR recommendations for management of rheumatoid arthritis. *Ann Rheum Dis* 2014; 73:529–35
- Ali T, Kaitha S, Mahmood S, Ftesi A, Stone J, Bronze MS: Clinical use of anti-TNF therapy and increased risk of infections. *Drug Healthc Patient Saf* 2013; 5:79–99
- Nathan C, Ding A: Nonresolving inflammation. *Cell* 2010; 140:871–82
- Tracey KJ: The inflammatory reflex. *Nature* 2002; 420:853–9
- Olofsson PS, Rosas-Ballina M, Levine YA, Tracey KJ: Rethinking inflammation: Neural circuits in the regulation of immunity. *Immunol Rev* 2012; 248:188–204
- Steinberg BE, Tracey KJ, Slutsky AS: Bacteria and the neural code. *N Engl J Med* 2014; 371:2131–3
- Berthoud HR, Neuhuber WL: Functional and chemical anatomy of the afferent vagal system. *Auton Neurosci* 2000; 85:1–17
- Watkins LR, Goehler LE, Relton JK, Tartaglia N, Silbert L, Martin D, Maier SF: Blockade of interleukin-1 induced hyperthermia by subdiaphragmatic vagotomy: Evidence for vagal mediation of immune-brain communication. *Neurosci Lett* 1995; 183:27–31
- Watkins LR, Wiertelak EP, Goehler LE, Smith KP, Martin D, Maier SF: Characterization of cytokine-induced hyperalgesia. *Brain Res* 1994; 654:15–26
- Hansen MK, O'Connor KA, Goehler LE, Watkins LR, Maier SF: The contribution of the vagus nerve in interleukin-1 $\beta$ -induced fever is dependent on dose. *Am J Physiol Regul Integr Comp Physiol* 2001; 280:R929–34
- Nijijima A, Hori T, Katafuchi T, Ichijo T: The effect of interleukin-1  $\beta$  on the efferent activity of the vagus nerve to the thymus. *J Auton Nerv Syst* 1995; 54:137–44
- Nijijima A: The afferent discharges from sensors for interleukin 1  $\beta$  in the hepatoportal system in the anesthetized rat. *J Auton Nerv Syst* 1996; 61:287–91
- Fairchild KD, Srinivasan V, Moorman JR, Gaykema RP, Goehler LE: Pathogen-induced heart rate changes associated with cholinergic nervous system activation. *Am J Physiol Regul Integr Comp Physiol* 2011; 300:R330–9
- Zapata P, Larraín C, Reyes P, Fernández R: Immunosensory signalling by carotid body chemoreceptors. *Respir Physiol Neurobiol* 2011; 178:370–4
- Shu HF, Wang BR, Wang SR, Yao W, Huang HP, Zhou Z, Wang X, Fan J, Wang T, Ju G: IL-1 $\beta$  inhibits IK and increases [Ca<sup>2+</sup>]<sub>i</sub> in the carotid body glomus cells and increases carotid sinus nerve firings in the rat. *Eur J Neurosci* 2007; 25:3638–47
- Chiu IM, Heesters BA, Ghasemlou N, Von Hehn CA, Zhao F, Tran J, Wainger B, Strominger A, Muralidharan S, Horswill AR, Bubeck-Wardenburg J, Hwang SW, Carroll MC, Woolf CJ: Bacteria activate sensory neurons that modulate pain and inflammation. *Nature* 2013; 501:52–7
- Liu T, Gao YJ, Ji RR: Emerging role of Toll-like receptors in the control of pain and itch. *Neurosci Bull* 2012; 28:131–44
- Liu T, Berta T, Xu ZZ, Park CK, Zhang L, Lü N, Liu Q, Liu Y, Gao YJ, Liu YC, Ma Q, Dong X, Ji RR: TLR3 deficiency impairs spinal cord synaptic transmission, central sensitization, and pruritus in mice. *J Clin Invest* 2012; 122:2195–207
- Liu T, Xu ZZ, Park CK, Berta T, Ji RR: Toll-like receptor 7 mediates pruritus. *Nat Neurosci* 2010; 13:1460–2
- Li Y, Zhang H, Zhang H, Kosturakis AK, Jawad AB, Dougherty PM: Toll-like receptor 4 signaling contributes to paclitaxel-induced peripheral neuropathy. *J Pain* 2014; 15:712–25
- Liu XJ, Zhang Y, Liu T, Xu ZZ, Park CK, Berta T, Jiang D, Ji RR: Nociceptive neurons regulate innate and adaptive immunity and neuropathic pain through MyD88 adapter. *Cell Res* 2014; 24:1374–7
- Borovikova LV, Ivanova S, Zhang M, Yang H, Botchkina GI, Watkins LR, Wang H, Abumrad N, Eaton JW, Tracey KJ: Vagus nerve stimulation attenuates the systemic inflammatory response to endotoxin. *Nature* 2000; 405:458–62
- Borovikova LV, Ivanova S, Nardi D, Zhang M, Yang H, Ombrellino M, Tracey KJ: Role of vagus nerve signaling in CNI-1493-mediated suppression of acute inflammation. *Auton Neurosci* 2000; 85:141–7
- Bernik TR, Friedman SG, Ochani M, DiRaimo R, Susarla S, Czura CJ, Tracey KJ: Cholinergic antiinflammatory pathway inhibition of tumor necrosis factor during ischemia reperfusion. *J Vasc Surg* 2002; 36:1231–6
- Rosas-Ballina M, Olofsson PS, Ochani M, Valdés-Ferrer SI, Levine YA, Reardon C, Tusche MW, Pavlov VA, Andersson U, Chavan S, Mak TW, Tracey KJ: Acetylcholine-synthesizing T cells relay neural signals in a vagus nerve circuit. *Science* 2011; 334:98–101

30. Rosas-Ballina M, Ochani M, Parrish WR, Ochani K, Harris YT, Huston JM, Chavan S, Tracey KJ: Splenic nerve is required for cholinergic antiinflammatory pathway control of TNF in endotoxemia. *Proc Natl Acad Sci USA* 2008; 105:11008–13
31. Olofsson PS, Katz DA, Rosas-Ballina M, Levine YA, Ochani M, Valdés-Ferrer SI, Pavlov VA, Tracey KJ, Chavan SS:  $\alpha 7$  nicotinic acetylcholine receptor ( $\alpha 7$ nAChR) expression in bone marrow-derived non-T cells is required for the inflammatory reflex. *Mol Med* 2012; 18:539–43
32. Arredondo J, Chernyavsky AI, Jolkovsky DL, Pinkerton KE, Grando SA: Receptor-mediated tobacco toxicity: Cooperation of the Ras/Raf-1/MEK1/ERK and JAK-2/STAT-3 pathways downstream of alpha7 nicotinic receptor in oral keratinocytes. *FASEB J* 2006; 20:2093–101
33. Sugano N, Shimada K, Ito K, Murai S: Nicotine inhibits the production of inflammatory mediators in U937 cells through modulation of nuclear factor-kappaB activation. *Biochem Biophys Res Commun* 1998; 252:25–8
34. de Jonge WJ, van der Zanden EP, The FO, Bijlsma MF, van Westerloo DJ, Bennink RJ, Berthoud HR, Uematsu S, Akira S, van den Wijngaard RM, Boeckxstaens GE: Stimulation of the vagus nerve attenuates macrophage activation by activating the Jak2-STAT3 signaling pathway. *Nat Immunol* 2005; 6:844–51
35. Lu B, Kwan K, Levine YA, Olofsson PS, Yang H, Li J, Joshi S, Wang H, Andersson U, Chavan SS, Tracey KJ:  $\alpha 7$  nicotinic acetylcholine receptor signaling inhibits inflammasome activation by preventing mitochondrial DNA release. *Mol Med* 2014; 20:350–8
36. Parrish WR, Rosas-Ballina M, Gallowitsch-Puerta M, Ochani M, Ochani K, Yang LH, Hudson L, Lin X, Patel N, Johnson SM, Chavan S, Goldstein RS, Czura CJ, Miller EJ, Al-Abed Y, Tracey KJ, Pavlov VA: Modulation of TNF release by choline requires alpha7 subunit nicotinic acetylcholine receptor-mediated signaling. *Mol Med* 2008; 14:567–74
37. Wang H, Yu M, Ochani M, Amella CA, Tanovic M, Susarla S, Li JH, Wang H, Yang H, Ulloa L, Al-Abed Y, Czura CJ, Tracey KJ: Nicotinic acetylcholine receptor alpha7 subunit is an essential regulator of inflammation. *Nature* 2003; 421:384–8
38. Mirakaj V, Dalli J, Granja T, Rosenberger P, Serhan CN: Vagus nerve controls resolution and pro-resolving mediators of inflammation. *J Exp Med* 2014; 211:1037–48
39. Andersson U, Tracey KJ: Reflex principles of immunological homeostasis. *Annu Rev Immunol* 2012; 30:313–35
40. Huston JM, Ochani M, Rosas-Ballina M, Liao H, Ochani K, Pavlov VA, Gallowitsch-Puerta M, Ashok M, Czura CJ, Foxwell B, Tracey KJ, Ulloa L: Splenectomy inactivates the cholinergic antiinflammatory pathway during lethal endotoxemia and polymicrobial sepsis. *J Exp Med* 2006; 203:1623–8
41. Hofer S, Eisenbach C, Lukic IK, Schneider L, Bode K, Brueckmann M, Mautner S, Wente MN, Encke J, Werner J, Dalpke AH, Stremmel W, Nawroth PP, Martin E, Krammer PH, Bierhaus A, Weigand MA: Pharmacologic cholinesterase inhibition improves survival in experimental sepsis. *Crit Care Med* 2008; 36:404–8
42. Huston JM, Gallowitsch-Puerta M, Ochani M, Ochani K, Yuan R, Rosas-Ballina M, Ashok M, Goldstein RS, Chavan S, Pavlov VA, Metz CN, Yang H, Czura CJ, Wang H, Tracey KJ: Transcutaneous vagus nerve stimulation reduces serum high mobility group box 1 levels and improves survival in murine sepsis. *Crit Care Med* 2007; 35:2762–8
43. Boland C, Collet V, Laterre E, Lecuivre C, Wittebole X, Laterre PF: Electrical vagus nerve stimulation and nicotine effects in peritonitis-induced acute lung injury in rats. *Inflammation* 2011; 34:29–35
44. Lee ST, Chu K, Jung KH, Kang KM, Kim JH, Bahn JJ, Jeon D, Kim M, Lee SK, Roh JK: Cholinergic anti-inflammatory pathway in intracerebral hemorrhage. *Brain Res* 2010; 1309:164–71
45. Su X, Feng X, Terrando N, Yan Y, Chawla A, Koch LG, Britton SL, Matthay MA, Maze M: Dysfunction of inflammation-resolving pathways is associated with exaggerated postoperative cognitive decline in a rat model of the metabolic syndrome. *Mol Med* 2012; 18:1481–90
46. Terrando N, Eriksson LI, Ryu JK, Yang T, Monaco C, Feldmann M, Jonsson Fagerlund M, Charo IF, Akassoglou K, Maze M: Resolving postoperative neuroinflammation and cognitive decline. *Ann Neurol* 2011; 70:986–95
47. Czura CJ, Schultz A, Kaipel M, Khadem A, Huston JM, Pavlov VA, Redl H, Tracey KJ: Vagus nerve stimulation regulates hemostasis in swine. *Shock* 2010; 33:608–13
48. Guarini S, Altavilla D, Cainazzo MM, Giuliani D, Bigiani A, Marini H, Squadrito G, Minutoli L, Bertolini A, Marini R, Adamo EB, Venuti FS, Squadrito F: Efferent vagal fibre stimulation blunts nuclear factor-kappaB activation and protects against hypovolemic hemorrhagic shock. *Circulation* 2003; 107:1189–94
49. Xiong J, Yuan YJ, Xue FS, Wang Q, Cheng Y, Li RP, Liao X, Liu JH: Postconditioning with  $\alpha 7$ nAChR agonist attenuates systemic inflammatory response to myocardial ischemia-reperfusion injury in rats. *Inflammation* 2012; 35:1357–64
50. Mioni C, Bazzani C, Giuliani D, Altavilla D, Leone S, Ferrari A, Minutoli L, Bitto A, Marini H, Zaffe D, Botticelli AR, Iannone A, Tomasi A, Bigiani A, Bertolini A, Squadrito F, Guarini S: Activation of an efferent cholinergic pathway produces strong protection against myocardial ischemia/reperfusion injury in rats. *Crit Care Med* 2005; 33:2621–8
51. Brégeon F, Xeridat F, Andreotti N, Lepidi H, Delpierre S, Roch A, Ravailhe S, Jammes Y, Steinberg JG: Activation of nicotinic cholinergic receptors prevents ventilator-induced lung injury in rats. *PLoS One* 2011; 6:e22386
52. Kox M, Pompe JC, Peters E, Vaneker M, van der Laak JW, van der Hoeven JG, Scheffer GJ, Hoedemaekers CW, Pickkers P:  $\alpha 7$  nicotinic acetylcholine receptor agonist GTS-21 attenuates ventilator-induced tumour necrosis factor- $\alpha$  production and lung injury. *Br J Anaesth* 2011; 107:559–66
53. dos Santos CC, Shan Y, Akram A, Slutsky AS, Haitsma JJ: Neuroimmune regulation of ventilator-induced lung injury. *Am J Respir Crit Care Med* 2011; 183:471–82
54. Song XM, Li JG, Wang YL, Liang H, Huang Y, Yuan X, Zhou Q, Zhang ZZ: Effect of vagus nerve stimulation on thermal injury in rats. *Burns* 2010; 36:75–81
55. Costantini TW, Bansal V, Peterson CY, Loomis WH, Putnam JG, Rankin F, Wolf P, Eliceiri BP, Baird A, Coimbra R: Efferent vagal nerve stimulation attenuates gut barrier injury after burn: Modulation of intestinal occludin expression. *J Trauma* 2010; 68:1349–54; discussion 1354–6
56. Meregiani J, Clarençon D, Vivier M, Peinnequin A, Mouret C, Sinniger V, Picq C, Job A, Canini F, Jacquier-Sarlin M, Bonaz B: Anti-inflammatory effect of vagus nerve stimulation in a rat model of inflammatory bowel disease. *Auton Neurosci* 2011; 160:82–9
57. van Westerloo DJ, Giebelen IA, Florquin S, Bruno MJ, Larosa GJ, Ulloa L, Tracey KJ, van der Poll T: The vagus nerve and nicotinic receptors modulate experimental pancreatitis severity in mice. *Gastroenterology* 2006; 130:1822–30
58. The FO, Boeckxstaens GE, Snoek SA, Cash JL, Bennink R, Larosa GJ, van den Wijngaard RM, Greaves DR, de Jonge WJ: Activation of the cholinergic anti-inflammatory pathway ameliorates postoperative ileus in mice. *Gastroenterology* 2007; 133:1219–28
59. Levine YA, Koopman FA, Faltys M, Caravaca A, Bendele A, Zitnik R, Vervoordeldonk MJ, Tak PP: Neurostimulation of the cholinergic anti-inflammatory pathway ameliorates disease in rat collagen-induced arthritis. *PLoS One* 2014; 9:e104530
60. Pavlov VA, Ochani M, Yang LH, Gallowitsch-Puerta M, Ochani K, Lin X, Levi J, Parrish WR, Rosas-Ballina M, Czura CJ, Larosa GJ, Miller EJ, Tracey KJ, Al-Abed Y: Selective alpha7-nicotinic

- acetylcholine receptor agonist GTS-21 improves survival in murine endotoxemia and severe sepsis. *Crit Care Med* 2007; 35:1139–44
61. Han Z, Li L, Wang L, Degos V, Maze M, Su H: Alpha-7 nicotinic acetylcholine receptor agonist treatment reduces neuroinflammation, oxidative stress, and brain injury in mice with ischemic stroke and bone fracture. *J Neurochem* 2014; 131:498–508
  62. Han Z, Shen F, He Y, Degos V, Camus M, Maze M, Young WL, Su H: Activation of  $\alpha$ -7 nicotinic acetylcholine receptor reduces ischemic stroke injury through reduction of pro-inflammatory macrophages and oxidative stress. *PLoS One* 2014; 9:e105711
  63. Olofsson PS: A stimulating concept: bioelectronic medicine in inflammatory disease. *Bioelectron Med* 2014; 2014:30–3
  64. Koopman FA, Schuurman PR, Vervoordeldonk MJ, Tak PP: Vagus nerve stimulation: A new bioelectronics approach to treat rheumatoid arthritis? *Best Pract Res Clin Rheumatol* 2014; 28:625–35
  65. Dombrovskiy VY, Martin AA, Sunderram J, Paz HL: Rapid increase in hospitalization and mortality rates for severe sepsis in the United States: A trend analysis from 1993 to 2003. *Crit Care Med* 2007; 35:1244–50
  66. Angus DC, van der Poll T: Severe sepsis and septic shock. *N Engl J Med* 2013; 369:840–51
  67. Deutschman CS, Tracey KJ: Sepsis: Current dogma and new perspectives. *Immunity* 2014; 40:463–75
  68. Thayer JF, Lane RD: Claude Bernard and the heart-brain connection: Further elaboration of a model of neurovisceral integration. *Neurosci Biobehav Rev* 2009; 33:81–8
  69. Buchan CA, Bravi A, Seely AJ: Variability analysis and the diagnosis, management, and treatment of sepsis. *Curr Infect Dis Rep* 2012; 14:512–21
  70. Frasure-Smith N, Lespérance F, Irwin MR, Talajic M, Pollock BG: The relationships among heart rate variability, inflammatory markers and depression in coronary heart disease patients. *Brain Behav Immun* 2009; 23:1140–7
  71. Werdan K, Schmidt H, Ebelt H, Zorn-Pauly K, Koidl B, Hoke RS, Heinroth K, Müller-Werdan U: Impaired regulation of cardiac function in sepsis, SIRS, and MODS. *Can J Physiol Pharmacol* 2009; 87:266–74
  72. Barnaby D, Ferrick K, Kaplan DT, Shah S, Bijur P, Gallagher EJ: Heart rate variability in emergency department patients with sepsis. *Acad Emerg Med* 2002; 9:661–70
  73. Chen WL, Kuo CD: Characteristics of heart rate variability can predict impending septic shock in emergency department patients with sepsis. *Acad Emerg Med* 2007; 14:392–7
  74. Wang H, Liao H, Ochani M, Justiniani M, Lin X, Yang L, Al-Abed Y, Wang H, Metz C, Miller EJ, Tracey KJ, Ulloa L: Cholinergic agonists inhibit HMGB1 release and improve survival in experimental sepsis. *Nat Med* 2004; 10:1216–21
  75. Rosas-Ballina M, Valdés-Ferrer SI, Dancho ME, Ochani M, Katz D, Cheng KF, Olofsson PS, Chavan SS, Al-Abed Y, Tracey KJ, Pavlov VA: Xanomeline suppresses excessive pro-inflammatory cytokine responses through neural signal-mediated pathways and improves survival in lethal inflammation. *Brain Behav Immun* 2015; 44:19–27
  76. Chavan SS, Huerta PT, Robbiati S, Valdes-Ferrer SI, Ochani M, Dancho M, Frankfurt M, Volpe BT, Tracey KJ, Diamond B: HMGB1 mediates cognitive impairment in sepsis survivors. *Mol Med* 2012; 18:930–7
  77. Shytle RD, Mori T, Townsend K, Vendrame M, Sun N, Zeng J, Ehrhart J, Silver AA, Sanberg PR, Tan J: Cholinergic modulation of microglial activation by alpha 7 nicotinic receptors. *J Neurochem* 2004; 89:337–43
  78. Thomsen MS, Mikkelsen JD: The  $\alpha$ 7 nicotinic acetylcholine receptor ligands methyllycaconitine, NS6740 and GTS-21 reduce lipopolysaccharide-induced TNF- $\alpha$  release from microglia. *J Neuroimmunol* 2012; 251:65–72
  79. Wittebole X, Hahm S, Coyle SM, Kumar A, Calvano SE, Lowry SF: Nicotine exposure alters *in vivo* human responses to endotoxin. *Clin Exp Immunol* 2007; 147:28–34
  80. Tremblay LN, Slutsky AS: Ventilator-induced injury: From barotrauma to biotrauma. *Proc Assoc Am Physicians* 1998; 110:482–8
  81. Santos CC, Zhang H, Liu M, Slutsky AS: Bench-to-bedside review: Biotrauma and modulation of the innate immune response. *Crit Care* 2005; 9:280–6
  82. Ventilation with lower tidal volumes as compared with traditional tidal volumes for acute lung injury and the acute respiratory distress syndrome. The Acute Respiratory Distress Syndrome Network. *N Engl J Med* 2000; 342:1301–8
  83. Vaneker M, Halbertsma FJ, van Egmond J, Netea MG, Dijkman HB, Snijdelaar DG, Joosten LA, van der Hoeven JG, Scheffer GJ: Mechanical ventilation in healthy mice induces reversible pulmonary and systemic cytokine elevation with preserved alveolar integrity: An *in vivo* model using clinical relevant ventilation settings. *ANESTHESIOLOGY* 2007; 107:419–26
  84. Wolthuis EK, Vlaar AP, Choi G, Roelofs JJ, Juffermans NP, Schultz MJ: Mechanical ventilation using non-injurious ventilation settings causes lung injury in the absence of pre-existing lung injury in healthy mice. *Crit Care* 2009; 13:R1
  85. Krzyzaniak MJ, Peterson CY, Cheadle G, Loomis W, Wolf P, Kennedy V, Putnam JG, Bansal V, Eliceiri B, Baird A, Coimbra R: Efferent vagal nerve stimulation attenuates acute lung injury following burn: The importance of the gut-lung axis. *Surgery* 2011; 150:379–89
  86. Reys LG, Ortiz-Pomales YT, Lopez N, Cheadle G, de Oliveira PG, Eliceiri B, Bansal V, Costantini TW, Coimbra R: Uncovering the neuroenteric-pulmonary axis: Vagal nerve stimulation prevents acute lung injury following hemorrhagic shock. *Life Sci* 2013; 92:783–92
  87. Su X, Lee JW, Matthay ZA, Mednick G, Uchida T, Fang X, Gupta N, Matthay MA: Activation of the alpha7 nAChR reduces acid-induced acute lung injury in mice and rats. *Am J Respir Cell Mol Biol* 2007; 37:186–92
  88. Kox M, Vaneker M, van der Hoeven JG, Scheffer GJ, Hoedemaekers CW, Pickkers P: Effects of vagus nerve stimulation and vagotomy on systemic and pulmonary inflammation in a two-hit model in rats. *PLoS One* 2012; 7:e34431
  89. Abildstrom H, Rasmussen LS, Rentowl P, Hanning CD, Rasmussen H, Kristensen PA, Moller JT: Cognitive dysfunction 1-2 years after non-cardiac surgery in the elderly. ISPOCD group. *International Study of Post-Operative Cognitive Dysfunction. Acta Anaesthesiol Scand* 2000; 44:1246–51
  90. Moller JT, Cluitmans P, Rasmussen LS, Houx P, Rasmussen H, Canet J, Rabbitt P, Jolles J, Larsen K, Hanning CD, Langeron O, Johnson T, Lauen PM, Kristensen PA, Biedler A, van Beem H, Fraidakis O, Silverstein JH, Beneken JE, Gravenstein JS: Long-term postoperative cognitive dysfunction in the elderly ISPOCD1 study. ISPOCD investigators. *International Study of Post-Operative Cognitive Dysfunction. Lancet* 1998; 351:857–61
  91. Steinmetz J, Christensen KB, Lund T, Lohse N, Rasmussen LS; ISPOCD Group: Long-term consequences of postoperative cognitive dysfunction. *ANESTHESIOLOGY* 2009; 110:548–55
  92. Johnson T, Monk T, Rasmussen LS, Abildstrom H, Houx P, Korttila K, Kuipers HM, Hanning CD, Siersma VD, Kristensen D, Canet J, Ibañez MT, Moller JT; ISPOCD2 Investigators: Postoperative cognitive dysfunction in middle-aged patients. *ANESTHESIOLOGY* 2002; 96:1351–7
  93. Price CC, Garvan CW, Monk TG: Type and severity of cognitive decline in older adults after noncardiac surgery. *ANESTHESIOLOGY* 2008; 108:8–17
  94. Terrando N, Monaco C, Ma D, Foxwell BM, Feldmann M, Maze M: Tumor necrosis factor- $\alpha$  triggers a cytokine cascade yielding postoperative cognitive decline. *Proc Natl Acad Sci USA* 2010; 107:20518–22

95. Cibelli M, Fidalgo AR, Terrando N, Ma D, Monaco C, Feldmann M, Takata M, Lever IJ, Nanchahal J, Fanselow MS, Maze M: Role of interleukin-1beta in postoperative cognitive dysfunction. *Ann Neurol* 2010; 68:360–8
96. Terrando N, Gómez-Galán M, Yang T, Carlström M, Gustavsson D, Harding RE, Lindskog M, Eriksson LI: Aspirin-triggered resolvin D1 prevents surgery-induced cognitive decline. *FASEB J* 2013; 27:3564–71
97. Li RL, Zhang ZZ, Peng M, Wu Y, Zhang JJ, Wang CY, Wang YL: Postoperative impairment of cognitive function in old mice: A possible role for neuroinflammation mediated by HMGB1, S100B, and RAGE. *J Surg Res* 2013; 185:815–24
98. Vacas S, Degos V, Tracey KJ, Maze M: High-mobility group box 1 protein initiates postoperative cognitive decline by engaging bone marrow-derived macrophages. *ANESTHESIOLOGY* 2014; 120:1160–7
99. Degos V, Vacas S, Han Z, van Rooijen N, Gressens P, Su H, Young WL, Maze M: Depletion of bone marrow-derived macrophages perturbs the innate immune response to surgery and reduces postoperative memory dysfunction. *ANESTHESIOLOGY* 2013; 118:527–36
100. Reinsfelt B, Ricksten SE, Zetterberg H, Blennow K, Fredén-Lindqvist J, Westerlind A: Cerebrospinal fluid markers of brain injury, inflammation, and blood-brain barrier dysfunction in cardiac surgery. *Ann Thorac Surg* 2012; 94:549–55
101. Bromander S, Anckarsäter R, Kristiansson M, Blennow K, Zetterberg H, Anckarsäter H, Wass CE: Changes in serum and cerebrospinal fluid cytokines in response to non-neurological surgery: An observational study. *J Neuroinflammation* 2012; 9:242
102. Yeager MP, Lunt P, Arruda J, Whalen K, Rose R, DeLeo JA: Cerebrospinal fluid cytokine levels after surgery with spinal or general anesthesia. *Reg Anesth Pain Med* 1999; 24:557–62
103. Buvanendran A, Kroin JS, Berger RA, Hallab NJ, Saha C, Negrescu C, Moric M, Caicedo MS, Tuman KJ: Upregulation of prostaglandin E2 and interleukins in the central nervous system and peripheral tissue during and after surgery in humans. *ANESTHESIOLOGY* 2006; 104:403–10
104. Braunwald E, Kloner RA: Myocardial reperfusion: A double-edged sword? *J Clin Invest* 1985; 76:1713–9
105. Yellon DM, Hausenloy DJ: Myocardial reperfusion injury. *N Engl J Med* 2007; 357:1121–35
106. Hausenloy DJ, Yellon DM: Myocardial ischemia-reperfusion injury: A neglected therapeutic target. *J Clin Invest* 2013; 123:92–100
107. Álvarez P, Tapia L, Mardones LA, Pedemonte JC, Fariás JG, Castillo RL: Cellular mechanisms against ischemia reperfusion injury induced by the use of anesthetic pharmacological agents. *Chem Biol Interact* 2014; 218:89–98
108. Ahn J, Kim J: Mechanisms and consequences of inflammatory signaling in the myocardium. *Curr Hypertens Rep* 2012; 14:510–6
109. Zhang M, Chen L: Status of cytokines in ischemia reperfusion induced heart injury. *Cardiovasc Hematol Disord Drug Targets* 2008; 8:161–72
110. Fröhlich GM, Meier P, White SK, Yellon DM, Hausenloy DJ: Myocardial reperfusion injury: Looking beyond primary PCI. *Eur Heart J* 2013; 34:1714–22
111. Katare RG, Ando M, Kakinuma Y, Arikawa M, Handa T, Yamasaki F, Sato T: Vagal nerve stimulation prevents reperfusion injury through inhibition of opening of mitochondrial permeability transition pore independent of the bradycardiac effect. *J Thorac Cardiovasc Surg* 2009; 137:223–31
112. Calvillo L, Vanoli E, Andreoli E, Besana A, Omodeo E, Gneccchi M, Zerbi P, Vago G, Busca G, Schwartz PJ: Vagal stimulation, through its nicotinic action, limits infarct size and the inflammatory response to myocardial ischemia and reperfusion. *J Cardiovasc Pharmacol* 2011; 58:500–7
113. Katare RG, Ando M, Kakinuma Y, Arikawa M, Yamasaki F, Sato T: Differential regulation of TNF receptors by vagal nerve stimulation protects heart against acute ischemic injury. *J Mol Cell Cardiol* 2010; 49:234–44
114. Li M, Zheng C, Sato T, Kawada T, Sugimachi M, Sunagawa K: Vagal nerve stimulation markedly improves long-term survival after chronic heart failure in rats. *Circulation* 2004; 109:120–4
115. Uemura K, Zheng C, Li M, Kawada T, Sugimachi M: Early short-term vagal nerve stimulation attenuates cardiac remodeling after reperfusion myocardial infarction. *J Card Fail* 2010; 16:689–99
116. Mastitskaya S, Marina N, Gourine A, Gilbey MP, Spyer KM, Teschemacher AG, Kasparov S, Trapp S, Ackland GL, Gourine AV: Cardioprotection evoked by remote ischaemic preconditioning is critically dependent on the activity of vagal preganglionic neurones. *Cardiovasc Res* 2012; 95:487–94
117. Buchholz B, Donato M, Perez V, Ivalde FC, Höcht C, Buitrago E, Rodríguez M, Gelpi RJ: Preischemic efferent vagal stimulation increases the size of myocardial infarction in rabbits. Role of the sympathetic nervous system. *Int J Cardiol* 2012; 155:490–1
118. Buchholz B, Donato M, Perez V, Deutsch AC, Höcht C, Del Mauro JS, Rodríguez M, Gelpi RJ: Changes in the loading conditions induced by vagal stimulation modify the myocardial infarct size through sympathetic-parasympathetic interactions. *Pflugers Arch* 2015; 467:1509–22
119. Shinlapawittayatorn K, Chinda K, Palee S, Surinkaew S, Thunsiri K, Weerateerangkul P, Chattipakorn S, KenKnight BH, Chattipakorn N: Low-amplitude, left vagus nerve stimulation significantly attenuates ventricular dysfunction and infarct size through prevention of mitochondrial dysfunction during acute ischemia-reperfusion injury. *Heart Rhythm* 2013; 10:1700–7
120. Wang Z, Yu L, Wang S, Huang B, Liao K, Saren G, Tan T, Jiang H: Chronic intermittent low-level transcutaneous electrical stimulation of auricular branch of vagus nerve improves left ventricular remodeling in conscious dogs with healed myocardial infarction. *Circ Heart Fail* 2014; 7:1014–21
121. Tong H, Imahashi K, Steenbergen C, Murphy E: Phosphorylation of glycogen synthase kinase-3beta during preconditioning through a phosphatidylinositol-3-kinase-dependent pathway is cardioprotective. *Circ Res* 2002; 90:377–9
122. Terashima Y, Sato T, Yano T, Maas O, Itoh T, Miki T, Tanno M, Kuno A, Shimamoto K, Miura T: Roles of phospho-GSK-3β in myocardial protection afforded by activation of the mitochondrial K ATP channel. *J Mol Cell Cardiol* 2010; 49:762–70
123. Kawada T, Yamazaki T, Akiyama T, Kitagawa H, Shimizu S, Mizuno M, Li M, Sugimachi M: Vagal stimulation suppresses ischemia-induced myocardial interstitial myoglobin release. *Life Sci* 2008; 83:490–5
124. Kawada T, Yamazaki T, Akiyama T, Li M, Ariumi H, Mori H, Sunagawa K, Sugimachi M: Vagal stimulation suppresses ischemia-induced myocardial interstitial norepinephrine release. *Life Sci* 2006; 78:882–7
125. Uemura K, Li M, Tsutsumi T, Yamazaki T, Kawada T, Kamiya A, Inagaki M, Sunagawa K, Sugimachi M: Efferent vagal nerve stimulation induces tissue inhibitor of metalloproteinase-1 in myocardial ischemia-reperfusion injury in rabbit. *Am J Physiol Heart Circ Physiol* 2007; 293:H2254–61
126. Kauvar DS, Lefering R, Wade CE: Impact of hemorrhage on trauma outcome: An overview of epidemiology, clinical presentations, and therapeutic considerations. *J Trauma* 2006; 60(6 suppl):S3–11
127. Fritz JR, Huston JM: The neural tourniquet. *Bioelectron Med* 2014; 2014:25–9
128. Rosas-Ballina M, Goldstein RS, Gallowitsch-Puerta M, Yang L, Valdés-Ferrer SI, Patel NB, Chavan S, Al-Abed Y, Yang H,

- Tracey KJ: The selective alpha7 agonist GTS-21 attenuates cytokine production in human whole blood and human monocytes activated by ligands for TLR2, TLR3, TLR4, TLR9, and RAGE. *Mol Med* 2009; 15:195–202
129. Cai B, Chen F, Ji Y, Kiss L, de Jonge WJ, Conejero-Goldberg C, Szabo C, Deitch EA, Ulloa L: Alpha7 cholinergic-agonist prevents systemic inflammation and improves survival during resuscitation. *J Cell Mol Med* 2009; 13:3774–85
  130. Kox M, Pompe JC, Gordinou de Gouberville MC, van der Hoeven JG, Hoedemaekers CW, Pickkers P: Effects of the  $\alpha 7$  nicotinic acetylcholine receptor agonist GTS-21 on the innate immune response in humans. *Shock* 2011; 36:5–11
  131. Stokes C, Papke JK, Horenstein NA, Kem WR, McCormack TJ, Papke RL: The structural basis for GTS-21 selectivity between human and rat nicotinic alpha7 receptors. *Mol Pharmacol* 2004; 66:14–24
  132. Pavlov VA, Parrish WR, Rosas-Ballina M, Ochani M, Puerta M, Ochani K, Chavan S, Al-Abed Y, Tracey KJ: Brain acetylcholinesterase activity controls systemic cytokine levels through the cholinergic anti-inflammatory pathway. *Brain Behav Immun* 2009; 23:41–5
  133. Ji H, Rabbi MF, Labis B, Pavlov VA, Tracey KJ, Ghia JE: Central cholinergic activation of a vagus nerve-to-spleen circuit alleviates experimental colitis. *Mucosal Immunol* 2014; 7:335–47
  134. Satapathy SK, Ochani M, Dancho M, Hudson LK, Rosas-Ballina M, Valdes-Ferrer SI, Olofsson PS, Harris YT, Roth J, Chavan S, Tracey KJ, Pavlov VA: Galantamine alleviates inflammation and other obesity-associated complications in high-fat diet-fed mice. *Mol Med* 2011; 17:599–606
  135. Birmingham K, Gradinaru V, Anikeeva P, Grill WM, Pikov V, McLaughlin B, Pasricha P, Weber D, Ludwig K, Famm K: Bioelectronic medicines: A research roadmap. *Nat Rev Drug Discov* 2014; 13:399–400
  136. Famm K, Litt B, Tracey KJ, Boyden ES, Slaoui M: Drug discovery: A jump-start for electroceuticals. *Nature* 2013; 496:159–61
  137. Penry JK, Dean JC: Prevention of intractable partial seizures by intermittent vagal stimulation in humans: Preliminary results. *Epilepsia* 1990; 31(suppl 2):S40–3
  138. Handforth A, DeGiorgio CM, Schachter SC, Uthman BM, Naritoku DK, Tecoma ES, Henry TR, Collins SD, Vaughn BV, Gilmartin RC, Labar DR, Morris GL III, Salinsky MC, Osorio I, Ristanovic RK, Labiner DM, Jones JC, Murphy JV, Ney GC, Wheless JW: Vagus nerve stimulation therapy for partial-onset seizures: A randomized active-control trial. *Neurology* 1998; 51:48–55
  139. Tatum WO, Johnson KD, Goff S, Ferreira JA, Vale FL: Vagus nerve stimulation and drug reduction. *Neurology* 2001; 56:561–3
  140. Helters SL, Wheless JW, Frost M, Gates J, Levisohn P, Tardo C, Conry JA, Yalnizoglu D, Madsen JR: Vagus nerve stimulation therapy in pediatric patients with refractory epilepsy: Retrospective study. *J Child Neurol* 2001; 16:843–8
  141. Kennedy SH, Giacobbe P, Rizvi SJ, Placenza FM, Nishikawa Y, Mayberg HS, Lozano AM: Deep brain stimulation for treatment-resistant depression: Follow-up after 3 to 6 years. *Am J Psychiatry* 2011; 168:502–10
  142. Berry SM, Broglio K, Bunker M, Jayewardene A, Olin B, Rush AJ: A patient-level meta-analysis of studies evaluating vagus nerve stimulation therapy for treatment-resistant depression. *Med Devices (Auckl)* 2013; 6:17–35
  143. Cristancho P, Cristancho MA, Baltuch GH, Thase ME, O'Reardon JP: Effectiveness and safety of vagus nerve stimulation for severe treatment-resistant major depression in clinical practice after FDA approval: Outcomes at 1 year. *J Clin Psychiatry* 2011; 72:1376–82
  144. Christmas D, Steele JD, Tolomeo S, Eljamel MS, Matthews K: Vagus nerve stimulation for chronic major depressive disorder: 12-month outcomes in highly treatment-refractory patients. *J Affect Disord* 2013; 150:1221–5
  145. McClelland J, Bozhilova N, Campbell I, Schmidt U: A systematic review of the effects of neuromodulation on eating and body weight: Evidence from human and animal studies. *Eur Eat Disord Rev* 2013; 21:436–55
  146. Martelletti P, Jensen RH, Antal A, Arcioni R, Brighina F, de Tommaso M, Franzini A, Fontaine D, Heiland M, Jürgens TP, Leone M, Magis D, Paemeleire K, Palmisani S, Paulus W, May A; European Headache Federation: Neuromodulation of chronic headaches: Position statement from the European Headache Federation. *J Headache Pain* 2013; 14:86
  147. Zitnik RJ: Treatment of chronic inflammatory diseases with implantable medical devices. *Cleve Clin J Med* 2011; 78(suppl 1):S30–4
  148. Ben-Menachem E: Vagus nerve stimulation, side effects, and long-term safety. *J Clin Neurophysiol* 2001; 18:415–8
  149. Morris GL III, Mueller WM: Long-term treatment with vagus nerve stimulation in patients with refractory epilepsy. The Vagus Nerve Stimulation Study Group E01-E05. *Neurology* 1999; 53:1731–5
  150. Ramsay RE, Uthman BM, Augustinsson LE, Upton AR, Naritoku D, Willis J, Treig T, Barolat G, Wernicke JF: Vagus nerve stimulation for treatment of partial seizures: 2. Safety, side effects, and tolerability. First International Vagus Nerve Stimulation Study Group. *Epilepsia* 1994; 35:627–36
  151. Olofsson PS, Levine YA, Caravaca A, Chavan SS, Pavlov VA, Faltsy M, Tracey KJ: Single-pulse and unidirectional electrical activation of the cervical vagus nerve reduces TNF in endotoxemia. *Bioelectron Med* 2015; 2015:37–42
  152. Aalbers MW, Klinkenberg S, Rijkers K, Verschuure P, Kessels A, Aldenkamp A, Vles J, Majoie M: The effects of vagus nerve stimulation on pro- and anti-inflammatory cytokines in children with refractory epilepsy: An exploratory study. *Neuroimmunomodulation* 2012; 19:352–8
  153. Barone L, Colicchio G, Policicchio D, Di Clemente F, Di Monaco A, Meglio M, Lanza GA, Crea F: Effect of vagal nerve stimulation on systemic inflammation and cardiac autonomic function in patients with refractory epilepsy. *Neuroimmunomodulation* 2007; 14:331–6
  154. Majoie HJ, Rijkers K, Berfelo MW, Hulsman JA, Myint A, Schwarz M, Vles JS: Vagus nerve stimulation in refractory epilepsy: Effects on pro- and anti-inflammatory cytokines in peripheral blood. *Neuroimmunomodulation* 2011; 18:52–6
  155. De Herdt V, Bogaert S, Bracke KR, Raedt R, De Vos M, Vonck K, Boon P: Effects of vagus nerve stimulation on pro- and anti-inflammatory cytokine induction in patients with refractory epilepsy. *J Neuroimmunol* 2009; 214:104–8
  156. Corcoran C, Connor TJ, O'Keane V, Garland MR: The effects of vagus nerve stimulation on pro- and anti-inflammatory cytokines in humans: A preliminary report. *Neuroimmunomodulation* 2005; 12:307–9
  157. Sundman E, Olofsson PS: Neural control of the immune system. *Adv Physiol Educ* 2014; 38:135–9
  158. McMahon SB, La Russa F, Bennett DL: Crosstalk between the nociceptive and immune systems in host defence and disease. *Nat Rev Neurosci* 2015; 16:389–402
  159. Undem BJ, Carr MJ: The role of nerves in asthma. *Curr Allergy Asthma Rep* 2002; 2:159–65
  160. Wu Y, You H, Ma P, Li L, Yuan Y, Li J, Ye X, Liu X, Yao H, Chen R, Lai K, Yang X: Role of transient receptor potential ion channels and evoked levels of neuropeptides in a formaldehyde-induced model of asthma in BALB/c mice. *PLoS One* 2013; 8:e62827
  161. Hox V, Vanoirbeek JA, Alpizar YA, Voedisch S, Callebaut I, Bobic S, Sharify A, De Vooght V, Van Gerven L, Devos F, Liston A, Voets T, Vennekens R, Bullens DM, De Vries A, Hoet P, Braun A, Ceuppens JL, Talavera K, Nemery B, Hellings PW: Crucial role of transient receptor potential ankyrin 1 and mast cells in induction of nonallergic airway hyperreactivity in mice. *Am J Respir Crit Care Med* 2013; 187:486–93

162. Tränkner D, Hahne N, Sugino K, Hoon MA, Zuker C: Population of sensory neurons essential for asthmatic hyperreactivity of inflamed airways. *Proc Natl Acad Sci USA* 2014; 111:11515–20
163. Talbot S, Abdulnour RE, Burkett PR, Lee S, Cronin SJ, Pascal MA, Laedermann C, Foster SL, Tran JV, Lai N, Chiu IM, Ghasemlou N, DiBiase M, Roberson D, Von Hehn C, Agac B, Haworth O, Seki H, Penninger JM, Kuchroo VK, Bean BP, Levy BD, Woolf CJ: Silencing nociceptor neurons reduces allergic airway inflammation. *Neuron* 2015; 87:341–54
164. Rogerio AP, Andrade EL, Calixto JB: C-fibers, but not the transient potential receptor vanilloid 1 (TRPV1), play a role in experimental allergic airway inflammation. *Eur J Pharmacol* 2011; 662:55–62
165. Alessandri AL, Pinho V, Souza DG, Castro MS, Klein A, Teixeira MM: Mechanisms underlying the inhibitory effects of tachykinin receptor antagonists on eosinophil recruitment in an allergic pleurisy model in mice. *Br J Pharmacol* 2003; 140:847–54
166. Bowden JJ, Baluk P, Lefevre PM, Schoeb TR, Lindsey JR, McDonald DM: Sensory denervation by neonatal capsaicin treatment exacerbates *Mycoplasma pulmonis* infection in rat airways. *Am J Physiol* 1996; 270(3 pt 1):L393–403
167. Fernandes ES, Liang L, Smillie SJ, Kaiser F, Purcell R, Rivett DW, Alam S, Howat S, Collins H, Thompson SJ, Keeble JE, Riffo-Vasquez Y, Bruce KD, Brain SD: TRPV1 deletion enhances local inflammation and accelerates the onset of systemic inflammatory response syndrome. *J Immunol* 2012; 188:5741–51
168. Meseguer V, Alpizar YA, Luis E, Tajada S, Denlinger B, Fajardo O, Manenschijn JA, Fernández-Peña C, Talavera A, Kichko T, Navia B, Sánchez A, Señarís R, Reeh P, Pérez-García MT, López-López JR, Voets T, Belmonte C, Talavera K, Viana F: TRPA1 channels mediate acute neurogenic inflammation and pain produced by bacterial endotoxins. *Nat Commun* 2014; 5:3125
169. Brogden KA, Guthmiller JM, Salzet M, Zasloff M: The nervous system and innate immunity: The neuropeptide connection. *Nat Immunol* 2005; 6:558–64
170. Gomes RN, Castro-Faria-Neto HC, Bozza PT, Soares MB, Shoemaker CB, David JR, Bozza MT: Calcitonin gene-related peptide inhibits local acute inflammation and protects mice against lethal endotoxemia. *Shock* 2005; 24:590–4
171. Torres-Rosas R, Yehia G, Peña G, Mishra P, del Rocio Thompson-Bonilla M, Moreno-Eutimio MA, Arriaga-Pizano LA, Isibasi A, Ulloa L: Dopamine mediates vagal modulation of the immune system by electroacupuncture. *Nat Med* 2014; 20:291–5
172. Wong CH, Jenne CN, Lee WY, Léger C, Kuberski P: Functional innervation of hepatic iNKT cells is immunosuppressive following stroke. *Science* 2011; 334:101–5
173. Arima Y, Harada M, Kamimura D, Park JH, Kawano F, Yull FE, Kawamoto T, Iwakura Y, Betz UA, Márquez G, Blackwell TS, Ohira Y, Hirano T, Murakami M: Regional neural activation defines a gateway for autoreactive T cells to cross the blood-brain barrier. *Cell* 2012; 148:447–57
174. Riol-Blanco L, Ordovas-Montanes J, Perro M, Naval E, Thiriou A, Alvarez D, Paust S, Wood JN, von Andrian UH: Nociceptive sensory neurons drive interleukin-23-mediated psoriasiform skin inflammation. *Nature* 2014; 510:157–61