

extensively studied. To claim that the effects reported in the current study occur at a “clinically relevant concentration” misrepresents this body of research and can easily lead to erroneous conclusions. We hope that this letter can serve as a reminder of the fundamental importance of knowing where the field has been to avoid the pitfalls of the past.

Competing Interests

Dr. Hemmings serves as an editor of *ANESTHESIOLOGY* and the *British Journal of Anaesthesia*, he has received research funding from the National Institutes of Health (Bethesda, Maryland) and TEM International (München, Germany), and he has served as a consultant for Elsevier (Amsterdam, The Netherlands). Dr. Goldstein receives funding from the U.S. Department of Defense (Washington, DC) and Mallinckrodt Pharmaceuticals (Dublin, Ireland). Dr. Riegelhaupt declares no competing interests.

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References

- Han L, Fuqua S, Li Q, Zhu L, Hao X, Li A, Gupta S, Sandhu R, Lonart G, Sugita S: Propofol-induced inhibition of catecholamine release is reversed by maintaining calcium influx. *ANESTHESIOLOGY* 2016; 124:878–84
- Herring BE, McMillan K, Pike CM, Marks J, Fox AP, Xie Z: Etomidate and propofol inhibit the neurotransmitter release machinery at different sites. *J Physiol* 2011; 589(pt 5):1103–15
- Xie Z, McMillan K, Pike CM, Cahill AL, Herring BE, Wang Q, Fox AP: Interaction of anesthetics with neurotransmitter release machinery proteins. *J Neurophysiol* 2013; 109:758–67
- Kazama T, Ikeda K, Morita K: Reduction by fentanyl of the Cp50 values of propofol and hemodynamic responses to various noxious stimuli. *ANESTHESIOLOGY* 1997; 87:213–27
- Servin F, Desmots JM, Haberer JP, Cockshott ID, Plummer GF, Farinotti R: Pharmacokinetics and protein binding of propofol in patients with cirrhosis. *ANESTHESIOLOGY* 1988; 69:887–91
- Tibbs GR, Rowley TJ, Sanford RL, Herold KF, Proekt A, Hemmings HC Jr, Andersen OS, Goldstein PA, Flood PD: HCN1 channels as targets for anesthetic and nonanesthetic propofol analogs in the amelioration of mechanical and thermal hyperalgesia in a mouse model of neuropathic pain. *J Pharmacol Exp Ther* 2013; 345:363–73
- Franks NP, Lieb WR: Molecular and cellular mechanisms of general anaesthesia. *Nature* 1994; 367:607–14
- Hemmings HC Jr, Akabas MH, Goldstein PA, Trudell JR, Orser BA, Harrison NL: Emerging molecular mechanisms of general anesthetic action. *Trends Pharmacol Sci* 2005; 26:503–10

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In Reply:

We thank Dr. Riegelhaupt and colleagues for their thoughtful comments on our article.¹ We stated that our concentrations (10 to 100 μM) of propofol are “clinically relevant,” because the total concentration of propofol in the blood of anesthetized humans is typically in the range of approximately 10 to 40 μM . Dr. Riegelhaupt and colleagues correctly point out

that “free” propofol concentration in blood of anesthetized humans is estimated to be lower (approximately 0.4 μM),² because more than 97% of total propofol is bound to blood constituents, such as albumin, and thus is pharmacologically sequestered.³ They expressed concerns that our statement of clinical relevance is misleading.

There has been a long-standing debate about the concentrations necessary to cause effects *in vitro* and lower concentrations of free propofol in the blood of anesthetized humans. For instance, the best-studied effects of propofol are its actions on γ -aminobutyric acid receptor type A (GABA_A) receptors. Here, the concentration of propofol to potentiate GABA -induced chloride currents of freshly dissociated neurons is in the range of 1 to 100 μM .^{4–6} This is significantly higher than the concentration of estimated “free” propofol. Therefore, it had been argued that the effect of propofol on GABA_A receptors of neurons may not be clinically relevant.⁷ However, this concern was put to rest by the finding that a point mutation in the β_3 subunit of the receptor in a knock-in mouse was sufficient to abolish the actions of propofol in preventing a response to a painful stimulus *in vivo*.⁸ It is now almost unequivocally believed that a major target for propofol is the GABA_A receptor, and using 1 to 100 μM of propofol *in vitro* has been validated.

Similarly, the critical mechanisms that explain propofol-induced hypotension have been investigated using high concentrations of propofol applied to *in vitro* preparations of cardiac cells or arterial rings. These studies have led to important insights into the propofol actions, which include direct myocardial depression by inhibition of Ca^{2+} -induced excitation–contraction coupling, reduced Ca^{2+} influx,^{9,10} and acetylcholine-induced pulmonary vasodilation.¹¹ It is probably unproductive to dismiss these results as clinically irrelevant simply because the nominal propofol concentration used *in vitro* was higher than the estimated free propofol concentration in patients.

Why is the nominal concentration of propofol used *in vitro* higher than the estimated “free” propofol in the blood? First, we would like to point out that even in the case of *in vitro* studies, the nominal concentration is higher than the concentration available for biologic action. Drugs bind non-specifically biologic preparations and to surfaces of the dish in tissue cultures, or to superfusion lines, filter material, and chambers in synaptosomal experiments, reducing free drug concentrations. Furthermore, propofol is an oil and is insoluble in physiologic buffers. A liquid emulsion formulation is used clinically, or an aqueous formulation, fospropofol, which itself is inactive. In our experiments, we made propofol stocks in dimethyl sulfoxide (DMSO) and limited the final concentration of DMSO to 0.1% to avoid potential side effects of DMSO. In our synaptosome experiments, we applied DMSO-based stock solution in small increments to agitated physiologic buffer to make final solutions of propofol. This prevented precipitation of propofol. Occasionally we observed some precipitation once we stopped agitation of the

buffer with oxygen/carbon dioxide at the end of the experiment. These factors and observations suggest that the effective *in vitro* concentrations may be lower than the calculated/nominal concentration in our experiments, as is the case, for partially overlapping reasons, in clinical applications.

In general, it is extremely difficult to accurately reproduce the clinical setting by *in vitro* experiments. For instance, the binding affinity and kinetics between propofol and albumin *versus* propofol and GABA_A receptors or other ion channels remain unknown. As such, the direct comparison of free concentration of propofol in the blood and the nominal concentration of propofol used *in vitro* may be impossible. It would be important to test how or whether our findings are affected by increasing concentrations of albumin, which sequester propofol. Such an attempt has been made just recently.¹²

In summary, we agree that the concentration we used is significantly higher than the concentration of “free” propofol estimated to be present in anesthetized humans. At the same time, “free” concentration of propofol is likely to be lower than the nominal concentration, even in our experiments. More pertinent for potential clinical relevance is that the concentrations we used are in line with numerous previous *in vitro* studies, which led to the discovery of the clinically relevant, important actions of propofol, including its actions on GABA_A receptors.

Competing Interests

The authors declare no competing interests.

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References

- Han L, Fuqua S, Li Q, Zhu L, Hao X, Li A, Gupta S, Sandhu R, Lonart G, Sugita S: Propofol-induced inhibition of catecholamine release is reversed by maintaining calcium influx. *ANESTHESIOLOGY* 2016; 124:878–84
- Franks NP, Lieb WR: Molecular and cellular mechanisms of general anaesthesia. *Nature* 1994; 367:607–14
- Gin T: Pharmacodynamics of propofol and free drug concentrations. *ANESTHESIOLOGY* 1993; 78:604–5
- Hara M, Kai Y, Ikemoto Y: Enhancement by propofol of the gamma-aminobutyric acidA response in dissociated hippocampal pyramidal neurons of the rat. *ANESTHESIOLOGY* 1994; 81:988–94
- Orser BA, Wang LY, Pennefather PS, MacDonald JF: Propofol modulates activation and desensitization of GABAA receptors in cultured murine hippocampal neurons. *J Neurosci* 1994; 14:7747–60
- Eghbali M, Gage PW, Birnir B: Effects of propofol on GABAA channel conductance in rat-cultured hippocampal neurons. *Eur J Pharmacol* 2003; 468:75–82
- Engdahl O, Abrahams M, Björnsson A, Vegfors M, Norlander B, Ahlner J, Eintrei C: Cerebrospinal fluid concentrations of propofol during anaesthesia in humans. *Br J Anaesth* 1998; 81:957–9
- Jurd R, Arras M, Lambert S, Drexler B, Siegwart R, Crestani F, Zaugg M, Vogt KE, Ledermann B, Antkowiak B, Rudolph U: General anesthetic actions *in vivo* strongly attenuated by a point mutation in the GABA(A) receptor beta3 subunit. *FASEB J* 2003; 17:250–2
- Shigemura T, Hatakeyama N, Shibuya N, Yamazaki M, Masuda A, Chen FS, Momose Y, Ito Y: Effects of propofol on contractile response and electrophysiological properties in single guinea-pig ventricular myocyte. *Pharmacol Toxicol* 1999; 85:111–4
- Hatakeyama N, Sakuraya F, Matsuda N, Kimura J, Kinoshita H, Kemmotsu O, Yamazaki M, Hattori Y: Pharmacological significance of the blocking action of the intravenous general anesthetic propofol on the slow component of cardiac delayed rectifier K⁺ current. *J Pharmacol Sci* 2009; 110:334–43
- Horibe M, Ogawa K, Sohn JT, Murray PA: Propofol attenuates acetylcholine-induced pulmonary vasorelaxation: Role of nitric oxide and endothelium-derived hyperpolarizing factors. *ANESTHESIOLOGY* 2000; 93:447–55
- Kojima A, Bai JY, Ito Y, Ding WG, Kitagawa H, Matsuura H: Serum albumin attenuates the open-channel blocking effects of propofol on the human Kv1.5 channel. *Eur J Pharmacol* 2016; 783:117–26

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