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Cerebral Autoregulation-oriented Therapy at the Bedside

A Comprehensive Review

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ABSTRACT

This comprehensive review summarizes the evidence regarding use of cerebral autoregulation-directed therapy at the bedside and provides an evaluation of its impact on optimizing cerebral perfusion and associated functional outcomes. Multiple studies in adults and several in children have shown the feasibility of individualizing mean arterial blood pressure and cerebral perfusion pressure goals by using cerebral autoregulation monitoring to calculate optimal levels. Nine of these studies examined the association between cerebral perfusion pressure or mean arterial blood pressure being above or below their optimal levels and functional outcomes. Six of these nine studies (66%) showed that patients for whom median cerebral perfusion pressure or mean arterial blood pressure differed significantly from the optimum, defined by cerebral autoregulation monitoring, were more likely to have an unfavorable outcome. The evidence indicates that monitoring of continuous cerebral autoregulation at the bedside is feasible and has the potential to be used to direct blood pressure management in acutely ill patients. (ANESTHESIOLOGY 2017; 126:1187-99)

MORE than half a century has passed since the concept of cerebral autoregulation was first described by Lassen,¹ who found optimal and constant cerebral blood flow within a cerebral perfusion pressure range of 50 to 150 mmHg. This broad, “safe cerebral perfusion pressure” range was subsequently adopted as doctrine for the management of mean arterial blood pressure (MAP) in healthy human individuals,²⁻⁶ based primarily on animal experiments.⁷⁻¹¹ Advances in technology now offer the ability to collect data through cerebral autoregulation monitoring and refine decades-old guidelines, thus potentially improving outcomes by individualizing cerebral perfusion pressure. Recently, a much narrower cerebral perfusion pressure autoregulatory plateau of 80 to 120 mmHg was reported by using bedside cerebral autoregulation monitoring in adults with acute subarachnoid hemorrhage.¹² Additionally, the lower limit of autoregulation ranged from a MAP of 43 to 90 mmHg in individuals undergoing cardiac surgery.¹³

Cerebral autoregulation can be assessed clinically at the bedside by measuring changes in cerebral blood flow, or its

surrogates, in relation to cerebral perfusion pressure.^{14,15} The newest and most innovative application of cerebral autoregulation monitoring is the determination of individualized optimal MAP and optimal cerebral perfusion pressure with the delineation of individual autoregulatory ranges. After reviewing the literature in major databases (PubMed/MEDLINE, Embase, and Google Scholar) from 1990 through 2016 using combinations of the keywords “cerebral autoregulation,” “optimal arterial pressure,” “optimal cerebral perfusion pressure,” “cerebral oximetry,” “transcranial Doppler,” and “intracranial cerebral pressure,” we found 12 observational studies over the last 6 yr that have determined the feasibility of using cerebral autoregulation monitoring to delineate optimal MAP or optimal cerebral perfusion pressure at the bedside in adults undergoing cardiopulmonary bypass; adults with acute traumatic brain injury, intracerebral hemorrhage, or subarachnoid hemorrhage; neonates with hypoxic-ischemic encephalopathy; and children with moyamoya syndrome.¹⁶⁻²⁵ Of these studies, 66% (6 of 9) showed that patients in whom

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actual MAP or cerebral perfusion pressure was widely different from optimal MAP or optimal cerebral perfusion pressure were more likely to have an unfavorable outcome.^{16–20} The strength of these data prompted the Brain Trauma Foundation to recommend cerebral autoregulation monitoring as an option to optimize cerebral perfusion pressure in patients with acute traumatic brain injury.²⁶ Nonetheless, the guidelines for arterial blood pressure management still recommend a single target blood pressure for critically ill patients and those with acute stroke: the International Guidelines for Management of Sepsis²⁷ recommend a MAP of at least 65 mmHg; the American Heart Association/American Stroke Association guidelines recommend a systolic blood pressure of less than 140 mmHg after acute intracerebral hemorrhage²⁸ and aneurysmal subarachnoid hemorrhage before aneurysm clipping or coiling²⁹ and a systolic blood pressure of less than 180 mmHg after intravenous recombinant tissue plasminogen activator for acute ischemic stroke.³⁰ Other societies now recognize that patients with a history of hypertension may have a cerebral autoregulation curve that is shifted to the right and require a higher MAP. For example, the European Society of Intensive Care Medicine³¹ recommends an initial target MAP of at least 65 mmHg (level 1 evidence; quality of experience, low) and a higher MAP in septic patients with history of hypertension and in patients who show clinical improvement with higher blood pressure (level 2 evidence; quality of experience, moderate). These guidelines do not currently recommend cerebral autoregulation-guided therapy and leave many unanswered questions: What is the optimal MAP target in patients with a history of long-standing hypertension? Do patients with acute

brain injury and elevated intracranial pressure (ICP) have different lower and upper limits of cerebral perfusion pressure than patients without intracranial injury?

The purpose of this comprehensive review is to summarize the evidence regarding use of cerebral autoregulation-directed therapy at the bedside to optimize and individualize cerebral perfusion pressure and to assess whether doing so can improve functional outcomes. We start by describing the physiology and methods used to measure cerebral autoregulation and then discuss validation of different cerebral autoregulation indices with a principal focus on evaluating the evidence behind the determination of optimal MAP/optimal cerebral perfusion pressure and its ability to accurately predict outcomes.

Physiology of Cerebral Autoregulation

Cerebral autoregulation protects the brain against hypoperfusion caused by hypotension, as well as against hypertension-induced hyperemia.³² Four mechanisms regulate cerebral blood flow, including myogenic, neurogenic, endothelial, and metabolic responses (fig. 1). Myogenic tone is generated when the smooth muscle of small arteries and arterioles contracts in response to increased pressure and relaxes in response to decreased pressure.³³ A rapid change in transmural pressure ($\Delta P = 10$ to 25 mmHg/s) triggers immediate changes in vessel diameter.³⁴ The latency between the onset of transmural stimulation and the beginning of the vessel's mechanical response is usually less than 250 ms.³⁵ The neurogenic mechanism, also called “neurovascular coupling,” is less well elucidated and involves the control of moderate- and small-diameter vessels.

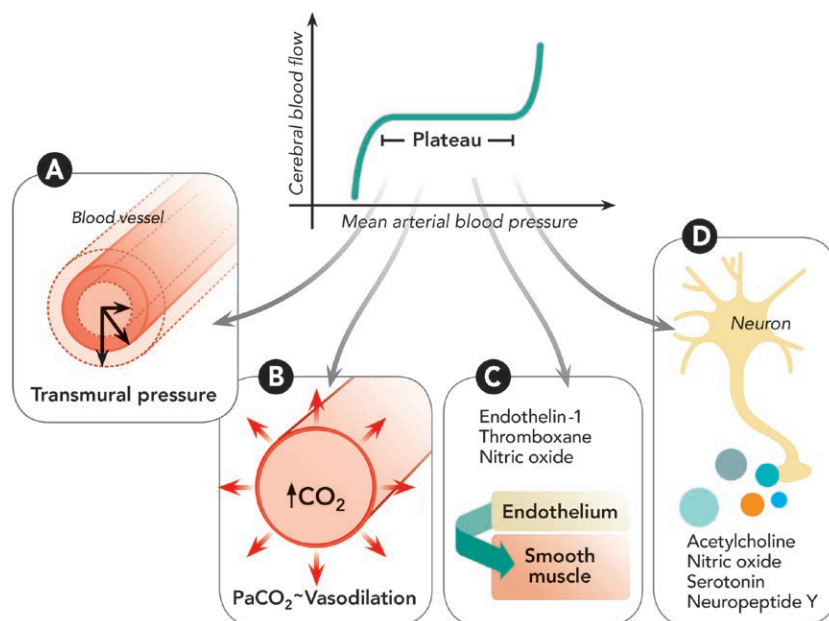


Fig. 1. Illustration of the mechanisms of cerebral autoregulation. (A) In the myogenic mechanism, changes in the transmural pressure influence changes in arterial diameter through contraction or relaxation of the smooth muscle. (B) In the metabolic mechanism, the concentration of carbon dioxide (CO_2) produced in the oxidative phosphorylation process affects small artery diameter. (C) The endothelial mechanism is based on the paracrine secretion of substances (nitric oxide and vasoconstrictors like endothelin-1 and thromboxane A₂) that stimulate the smooth muscle. (D) In the neurogenic mechanism, neuroglial cells contribute to the control of moderate- and small-diameter vessels by secreting different neurotransmitters with vasoactive properties.

Neurons secrete different neurotransmitters with vasoactive properties, such as acetylcholine or nitric oxide, which cause vasodilatation, and serotonin and neuropeptide Y, which stimulate vasoconstriction.³⁶ By using infrared videomicroscopy of interneurons and adjacent microvessels in rats, Cauli *et al.*³⁷ showed that microvessels constrict in response to interneuronal depolarization. The metabolic mechanism occurs in smaller vessels that are subject to changes in the local microenvironment that alter vasomotor responses.³⁸ For example, hypotension below the lower limit of autoregulation leads to low cerebral blood flow and a consequent accumulation of carbon dioxide. For every 1-mmHg increase in P_{aCO_2} , there is an approximately 4% increase in cerebral blood flow caused by vessel vasodilatation. Conversely, hypertension above the upper limit of autoregulation results in hyperperfusion and a drop in carbon dioxide. For every 1-mmHg decrease in P_{aCO_2} , vessel vasoconstriction will cause a 4% decrease in cerebral blood flow.³⁸ This reactivity has been attributed to the response of cerebral vessel smooth muscle to H^+ .³⁹ Last, the endothelium generates a variety of signals that influence cerebrovascular tone under normal conditions and during disease as well. The endothelium secretes vasodilators such as nitric oxide and vasoconstrictors like endothelin-1 and thromboxane A₂.⁴⁰ One of the benefits of statins is their ability to upregulate nitric-oxide synthase, causing cerebral artery dilation and increased cerebral blood flow.⁴¹

Methods to Measure Cerebral Autoregulation and Cerebrovascular Reactivity

Cerebrovascular reactivity is the ability of vascular smooth muscle to change basal tone in response to variations of physiologic parameters, such as arterial blood pressure, and metabolic factors, such as cerebral carbon dioxide and oxygen levels.¹⁵ When cerebrovascular reactivity is exhausted, cerebral blood flow becomes dependent on systemic arterial blood pressure. Cerebral autoregulation is one aspect of cerebrovascular reactivity that involves vascular tone changes in response to fluctuations in arterial blood pressure. Vessels may still demonstrate responses to further changes in carbon dioxide concentration.^{42,43} These vascular responses that continue to occur outside the MAP range of stable cerebral blood flow are also part of the cerebral autoregulatory mechanism (metabolic, endothelial, among others) that protects the brain.⁴² Therefore, the terms cerebral autoregulation and cerebrovascular reactivity should not be used synonymously, as vasodilatation reaches its maximum at arterial pressures below the lower threshold for constant cerebral blood flow.^{44,45}

Regulation of the brain vasculature's ability to maintain constant cerebral blood flow can be assessed by two modalities: static and dynamic autoregulation.⁴⁶ Static autoregulation describes the extent to which the cerebrovascular bed can constrict or dilate when cerebral perfusion pressure varies. Dynamic autoregulation also incorporates information

on the rate at which such adaptive changes in cerebrovascular resistance occur.⁴⁷ Only dynamic cerebral autoregulation allows for continuous measurement of cerebral autoregulation and therefore determination of optimal MAP and optimal cerebral perfusion pressure, the most novel application of cerebral autoregulation monitoring.

Technology for Cerebral Autoregulation Monitoring

The technology used to calculate cerebral autoregulation and cerebrovascular reactivity in the clinical setting includes transcranial Doppler, which measures cerebral blood flow velocity; near-infrared spectroscopy, which measures regional cerebral oxygen saturation; the brain tissue oxygen monitor, which measures tissue oxygen partial pressure; ICP monitors; and, more recently, ultrasound-tagged near-infrared spectroscopy, which measures cerebral blood flow velocity (table 1). All these measurements are used as surrogates for the gold standard of cerebral blood flow, which no currently available device can quantify.⁴⁸ Figure 2 shows the devices that are frequently used to measure cerebral autoregulation or cerebrovascular reactivity.

Transcranial Doppler is an accepted noninvasive tool for continuous monitoring of cerebral blood flow velocity and is a well-validated method to assess cerebral autoregulation.^{49,50} Cerebral autoregulation testing with transcranial Doppler measures cerebral blood flow velocity from the middle cerebral arteries. Because measurement of middle cerebral artery diameter is not standard, transcranial Doppler provides only a surrogate for cerebral blood flow based on the assumption that middle cerebral artery diameter changes minimally with changes in MAP.⁵¹ As cerebral blood flow velocity is a pulsatile phenomenon, it can be monitored in a time domain that relies only on spontaneous changes of MAP or cerebral perfusion pressure. A moving correlation coefficient can then be calculated between cerebral blood flow velocity and MAP or cerebral perfusion pressure; this coefficient is called the mean velocity index.⁵²

Near-infrared spectroscopy is also a noninvasive device that measures regional cerebral oxygen saturation. Near-infrared light is transmitted from a source embedded in a sensor attached to the forehead and directed toward the frontal lobe. Light in the near-infrared spectrum (700 to 950 nm) can traverse biologic tissue because of the relative transparency of tissue to light at these wavelengths. Several biologic molecules, termed chromophores, have distinct absorption spectra in the near infrared.⁵³ Oxyhemoglobin, deoxyhemoglobin, and cytochrome aa_3 (a complex protein present in the mitochondria that is involved in the oxidative phosphorylation process) are the most abundant chromophores that absorb near-infrared light between 700 and 1,000 nm.^{53,54} The amount of light detected by sensors positioned at set distances from the light source is a function of reflectance from the light-tissue angle, scattering from body tissues, and absorption by chromophores.^{53,55} This technology makes the following assumptions: cytochrome aa_3 and bilirubin are minimal, and the hemoglobin measured

Table 1. Cerebral Autoregulation Indices with Their Cutoffs to Define Impaired Autoregulation

Surrogate of CBF	Device or Monitor	Cerebral Autoregulation Index	Correlation Between	Cutoff for Impaired CA	Reference No.
Regional cerebral oxygenation	NIRS	Cerebral oximetry index	Regional cerebral oxygenation and MAP	>0.3	58
Total hemoglobin volume	NIRS	Hemoglobin volume index	Total hemoglobin volume and MAP	>0.3	81
Regional cerebral oxygenation	NIRO	Tissue oxygen index	Regional cerebral oxygenation and MAP	>0.1 >0.13	82 12
Tissue hemoglobin	NIRO	Tissue hemoglobin index	Oxygenated and deoxygenated hemoglobin and MAP	NA	—
CBF velocity	UT-NIRS	CBF velocity index	CBF velocity and MAP	NA	—
CBF velocity	TCD	Dynamic autoregulatory index	CBF velocity and MAP	<4	83
CBF velocity	TCD	Systolic flow velocity index	Systolic CBF velocity and MAP	>0.1 >0.05	82 12
CBF velocity	TCD	Mean flow velocity index	Mean CBF velocity and MAP	>0.3 >0.46	52 84
CBF velocity	TCD	Mean flow velocity index	Mean CBF velocity and cerebral perfusion pressure	>0.3	49,71,72
Tissue oxygen pressure	Brain tissue oxygen monitor	Brain tissue oxygen pressure reactivity index	Tissue oxygen pressure and cerebral perfusion pressure	>0.4	85
ICP	ICP monitor	Pressure reactivity index	5- to 10-s mean ICP and MAP	>0.3	52
ICP	ICP monitor	Diastolic coefficient index	Diastolic CBF velocity and diastolic MAP	>0.24	84
ICP	ICP monitor	Low-frequency autoregulation index	Minute-by-minute mean ICP and MAP	NA	—
ICP	ICP monitor	Low-frequency sample pressure reactivity index	20-min averages of ICP and MAP	>0.2	86

CA = cerebral autoregulation; CBF = cerebral blood flow; ICP = intracranial pressure; MAP = mean arterial blood pressure; NA = not described yet; NIRO = near-infrared oxygenation monitor; NIRS = near-infrared spectroscopy; TCD = transcranial Doppler; UT-NIRS = ultrasound-tagged near-infrared spectroscopy.

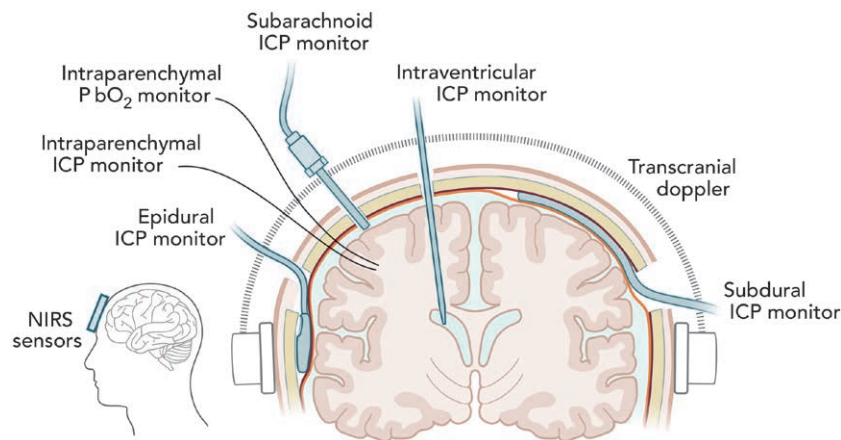


Fig. 2. Principal devices used to measure cerebral autoregulation and cerebrovascular reactivity and their positioning. ICP = intracranial pressure; NIRS = near-infrared spectroscopy.

is contained in a fixed mixture of vessels that are approximately 70 to 75% venous and 25 to 30% arterial blood volume.^{56,57} Equations used to account for variability in venous:arterial ratios are manufacturer specific; therefore, regional cerebral oxygen saturations derived from different machines are not equivalent.^{55,56} The cerebral oximetry index is derived from the correlation between regional cerebral oxygen saturation from near-infrared spectroscopy and MAP or cerebral perfusion pressure.⁵⁸

An innovative new hybrid device, CerOx (Ornim Medical Ltd., Israel), uses a single, noninvasive probe to provide a brain oximeter and blood flow monitor that utilizes a combination of near-infrared light and a localized low-power ultrasound. The ultrasound signal is a sequence of phase-modulated waves with a central frequency at 1 MHz, which is similar to the frequency (2 MHz) of transcranial Doppler.⁵⁹ This ultrasound-tagged near-infrared spectroscopy estimates changes in microcirculatory blood flow within the

interrogated volume of 1 cm^3 .^{59,60} The cerebral blood flow velocity index is derived from the correlation between cerebral blood flow velocity from ultrasound-tagged near-infrared spectroscopy and MAP or cerebral perfusion pressure.⁶¹

Other methods estimate cerebrovascular reactivity by measuring changes in ICP with ICP monitors. Normally, the cerebral blood volume and ICP vary inversely with arterial blood pressure.⁶² Therefore, if cerebrovascular reactivity is intact, a significant increase in MAP will produce vasoconstriction, a decrease in cerebral blood volume, and a decrease in ICP.^{15,39,63,64} If vessels are nonreactive, an increase in MAP would cause an increase in the cerebral blood volume and, thereby, ICP.¹⁵ The pressure reactivity index is the most commonly used index to measure cerebrovascular reactivity in patients with traumatic brain injury and is derived from the correlation between ICP and MAP.¹⁵

The disadvantages of some older cerebral autoregulation detection methods are that they require a hemodynamic stimulus to induce a change in MAP, such as thigh cuff release,⁶⁵ increase in arterial partial pressure of carbon dioxide,⁶⁶ tilt table declination,⁶⁷ application of negative body pressure,⁶⁸ carotid artery compression,⁶⁹ or vasoactive drug administration.⁷⁰ The safety of such manipulations in compromised patients prone to organ injury from alterations in MAP is of concern. Thus, newer methods of cerebral autoregulation monitoring are based on cerebral blood flow responses to spontaneous changes in cerebral perfusion pressure or MAP that may occur over time and slow-wave oscillations in cerebral blood volume and cerebral blood flow—lasting from 30 s to a few minutes—secondary to normal physiologic functions such as breathing.^{3,49,52,71–75}

The advantage of using transcranial Doppler- and near-infrared spectroscopy-derived cerebral autoregulation indices is that they are noninvasive, whereas ICP and tissue oxygen partial pressure monitors require intracranial catheters that carry risks for hemorrhage, meningitis, and ventriculitis.⁷⁶ The principal disadvantage of transcranial Doppler is the requirement for a trained technician, which restricts widespread applicability. The use of transcranial Doppler is also hampered by the 10 to 15% rate of inadequate acoustic windows prevalent in African Americans, Asians, and elderly women.⁷⁷

Limitations to these devices must be acknowledged. Near-infrared spectroscopy measures regional cerebral oxygen saturation through sensors that are placed on the forehead. Therefore, the cerebral autoregulation calculations are limited to regional cerebral oxygen saturation from the frontal lobes, with some contamination from the external carotid artery.⁷⁸ The brain tissue oxygen monitor is also a local measure and may not reflect global oxygenation and metabolism, especially in patients suffering from focal injuries. Moreover, there is an active debate on the most appropriate location to place monitoring probes.⁷⁹ ICP monitors are a global measure and may not reflect local changes. Studying

the effect of arteriolar vasoconstriction and vasodilatation through their effect on ICP will inevitably include a dampening effect.²¹ Finally, transcranial Doppler can provide only an estimation of cerebral blood flow when the diameter of the sampled artery does not change throughout the examination. Magnetic resonance imaging can be used to test this assumption.⁸⁰

Cerebral Autoregulation Indices

There are more than 21 cerebral autoregulation indices. Some measure cerebral autoregulation (cerebral oximetry index, tissue oxygen index, cerebral blood flow velocity index, systolic flow velocity index, mean flow velocity index, and brain tissue oxygen pressure reactivity index), whereas others measure cerebrovascular reactivity (pressure reactivity index, hemoglobin volume index, tissue hemoglobin index, and dynamic autoregulatory index). Table 1 provides definitions of all of the cerebral autoregulation indices and descriptions of how to measure and calculate them. Generally, when cerebral autoregulation is lost, the cerebral autoregulation indices approximate to 1, indicating pressure passivity; a negative index or one that approaches 0 indicates intact pressure reactivity. Despite this general principle, each index has a different cutoff to define impaired autoregulation, with a range from 0.069 to 0.46.^{12,49,52,58,71,72,81–86} This wide variability in cutoff values depends on the different devices used as surrogates of cerebral blood flow measurements and the population studied. The dynamic autoregulatory index is the only one that uses a different scale than the ones mentioned above, and it ranges from 0 (absent cerebral autoregulation) to 9 (most efficient cerebral autoregulation).⁸⁷

Validation of Invasive versus Noninvasive Cerebral Autoregulation Methods

Multiple new cerebral autoregulation indices have been validated against long-standing ones during the past couple of decades. This approach makes it easy to start using newer and possibly superior methods to measure cerebral autoregulation or cerebrovascular reactivity clinically. More importantly, noninvasive methods can be compared to invasive ones. A detailed description of the validation studies is presented in table 2. One of the noninvasive cerebral autoregulation indices most often used at the bedside is mean flow velocity index based on MAP. The mean flow velocity index based on MAP is derived from the correlation between cerebral blood flow velocity and MAP, and numerous studies have validated it against mean velocity index based on cerebral perfusion pressure^{52,72,88} in patients with intracranial injury ($R = 0.789$, $P < 0.001$). Mean flow velocity index based on MAP has also shown good agreement in validations against dynamic autoregulatory index and pressure reactivity index: $R = -0.38$, $P < 0.001$ ⁸⁹; $R = 0.58$, $P < 0.001$ ⁹⁰;

Table 2. Validation Studies of Cerebral Autoregulation Indices

Comparison	R Value	P Value
Studies in patients with intracranial injury		
Mean flow velocity index based on cerebral perfusion pressure vs. mean flow velocity index based on MAP ⁵²	0.566	<0.01
Mean flow velocity index based on cerebral perfusion pressure vs. mean flow velocity index based on MAP ⁷²	0.789	<0.001
Mean flow velocity index based on cerebral perfusion pressure vs. mean flow velocity index based on MAP ⁸⁸	0.755	<0.001
Mean flow velocity index based on cerebral perfusion pressure vs. dynamic autoregulatory index ⁹¹	-0.62	0.0001
Mean flow velocity index based on MAP vs. tissue oxygen index ⁹²	0.61	0.004
Mean flow velocity index based on MAP vs. systolic flow velocity index ⁹⁸	0.89	<0.001
Mean flow velocity index vs. dynamic autoregulatory index ⁸⁹	-0.38	<0.001
Mean flow velocity index vs. tissue hemoglobin index ⁹²	0.26	0.28
Pressure reactivity index vs. mean flow velocity index based on cerebral perfusion pressure ¹⁰⁰	0.58	<0.001
Pressure reactivity index vs. low-frequency sample pressure reactivity index ¹⁷	0.7	<0.00001
Pressure reactivity index vs. low-frequency sample pressure reactivity index ⁹⁷	0.846	<0.001
Pressure reactivity index vs. brain tissue oxygen pressure reactivity index ⁹⁶	0.851	<0.04354
Pressure reactivity index vs. tissue oxygen index ⁹²	0.40	0.04
Pressure reactivity index vs. tissue hemoglobin index ⁹²	0.63	<0.001
Pressure reactivity index vs. tissue hemoglobin index ¹⁰¹	0.56	0.0002
Studies in patients with no intracranial injury		
Mean flow velocity index based on MAP vs. cerebral oximetry index ⁹³	0.51	<0.001
Mean flow velocity index based on MAP vs. cerebral oximetry index ⁹⁵	0.55	<0.0001
Mean flow velocity index based on MAP vs. tissue oxygen index ⁹⁴	0.81	<0.0001
Cerebral blood flow velocity index vs. mean flow velocity index based on MAP ⁶¹	0.39	<0.001
Mean flow velocity index based on cerebral perfusion pressure vs. hemoglobin volume index ⁸¹	0.5915	<0.001

MAP = mean arterial blood pressure.

and $R = -0.62$, $P < 0.0001$.⁹¹ Other noninvasive cerebral autoregulation methods that use the near-infrared oxygenation (NIRO-200) monitor have been validated against invasive methods. For example, the tissue oximetry index and tissue hemoglobin index each showed good agreement with the pressure reactivity index ($R = 0.40$, $P = 0.04$; $R = 0.63$, $P < 0.001$, respectively).⁹² In patients without intracranial injury, cerebral oximetry index (derived from near-infrared spectroscopy, INVOS monitor [Medtronic/Covidien, Ireland]; $R = 0.51$, $P < 0.001$)⁹³ and tissue oxygen index ($R = 0.81$, $P < 0.0001$)⁹⁴ have each been validated against mean flow velocity index based on MAP. The hemoglobin volume index, derived from near-infrared spectroscopy (INVOS monitor), has also been validated against mean flow velocity index based on cerebral perfusion pressure in patients without intracranial injury ($R = 0.5915$, $P < 0.0001$).⁸¹ All of these significant correlations between the invasive and noninvasive methods and others are described in table 2.^{17,61,95-101} These results support the accuracy of noninvasive methods and their potential utility in cerebral autoregulation and cerebrovascular reactivity monitoring.

Measurement of Optimal Cerebral Perfusion Pressure and Optimal MAP in Individual Patients

Over the last decade, several advances in determining optimal cerebral perfusion pressure and optimal MAP have been made. The cerebral autoregulation indices that have been used

to determine the optimal values are validated (*i.e.*, pressure reactivity index, cerebral oximetry index, tissue hemoglobin index) and have demonstrated significant ability to predict outcomes.^{74,90,100,102} We will describe the methodology used to determine optimal cerebral perfusion pressure and optimal MAP; a summary of these study results is shown in table 3.

Second-order Polynomial Formula (U-shaped Curve)

This method has been used most frequently in studies of traumatic brain injury in which optimal cerebral perfusion pressure is calculated by fitting a U-shaped curve over 4-h periods of monitoring (table 3). That curve, also known as the U-shaped parabola, is supposed to represent the real plot of cerebral autoregulation indices *versus* cerebral perfusion pressure or MAP; as a result, the optimal cerebral perfusion pressure or optimal MAP is logically assumed to be the X-vertex of the curve modeled by the parabolic formula ($Ax^2 + Bx + C$). The estimation of the optimal cerebral perfusion pressure by this method is thought to be exact because it represents an exact MAP or cerebral perfusion pressure point that reflects the real lowest magnitude of the cerebral autoregulation index used. Despite the accuracy assumed of this method, several limitations are worth noting. First, this method does not identify optimal pressures in all monitored patients, only in up to 55% of the monitor recordings.^{16,23} Second, this formula does not take into consideration the percentage of time in each bin recorded; therefore the calculated optimal pressure can be biased by outliers. Figure 3 shows a clear typical error of the second-order polynomial

Table 3. Summary of the Reported Optimal Cerebral Perfusion Pressure and Optimal Mean Arterial Blood Pressure Sorted by the Population Studied

Patient Population	Optimal Pressure Studied	Autoregulation Index Used (Time Window)	Sample Size	Proportion of Time CPP _{OPT} or MAP _{OPT} Identified	Method	Mean ± SD or Median [IQR] of the Optimal Pressure (mmHg)	Association with Poor Outcome	Reference No.	
Traumatic brain injury	CPP _{OPT}	Pressure reactivity index (4h)	307	55%	U-shaped curve	74.7 ± 8.2	Below and above CPP _{OPT}	16	
		Low-frequency autoregulation index (1–24 h)	55	97%	DATA CAR	70.8 ± 11.4	Below and above CPP _{OPT}	21	
	MAP _{OPT}	Pressure reactivity index (4h)	30*	NA	U-shaped curve	68.87 ± 9.73† 63.6 ± 7.9‡	Only below CPP _{OPT}	18	
		Pressure reactivity index (4h)	18	NA	U-shaped curve	88 ± 7	Only below CPP _{OPT}	19	
		Low-frequency sample pressure reactivity index (4h)	307	NA	U-shaped curve	76.9 ± 10.1	No statistically significant associations with outcome	17	
Intracerebral hemorrhage	CPP _{OPT}	Pressure reactivity index (4h)	55	44%	U-shaped curve	72.5 ± 7.6	No statistically significant associations with outcome	21	
		Pressure reactivity index (4 days)	48	72%	U-shaped curve	78.6 ± 12.2	NP	22	
	MAP _{OPT}	Cerebral oximetry index (4h)	18	NA	U-shaped curve	88 ± 7	NP	19	
		Brain tissue oxygen pressure reactivity index (4h)	18	NA	U-shaped curve	85 ± 6	NP	19	
	Subarachnoid hemorrhage	CPP _{OPT}	Cerebral blood flow velocity index (4h)	18	NA	U-shaped curve	85 ± 6	NP	19
			Pressure reactivity index (48h)	38	84%	Lowest CA index	76.25 ± 9.67	NP	23
		MAP _{OPT}	Pressure reactivity index (NA)	38	57%	U-shaped curve	83 ± 7.06	No statistically significant associations with outcome	24
			Pressure reactivity index (NA)	25	NA	Lowest CA index	78 ± 2.6§ 98 ± 3.6	No statistically significant associations with mortality	25
			Cerebral oximetry index (NA)	121	NA	Lowest CA index	78 ± 12.8	Below the MAP _{OPT} (for brain injury as outcome)	20
			Cerebral blood flow velocity index (1.85h)	69	NA	Lowest CA index	71 ± 12	NP	61
Cardiac surgery	CPP _{OPT}	Mean flow velocity index based on MAP (1.85h)	69	NA	Lowest CA index	74 ± 12	NP	61	
		Mean flow velocity index based on MAP (1.48h)	109	87%	Lowest CA index	75 ± 11	NP	81	
	MAP _{OPT}	Hemoglobin volume index (1.48h)	109	100%	Lowest CA index	74 ± 13	NP	81	
		Hemoglobin volume index (6h)#	14	NA	Lowest CA index	50 [45–55]	Only below MAP _{OPT}	104	
	MAP _{OPT}	Hemoglobin volume index (6.5h)**	17	NA	Lowest CA index	45 [45–50]			
		Hemoglobin volume index (30.5h)††	NA	NA	Lowest CA index	45 [45–55]			
	MAP _{OPT}	Cerebral oximetry index (NA)	7	86%	Lowest CA index	90 [60–95]‡‡ 90 [85–90]§§	NP	103	
		Hemoglobin volume index (NA)	7	86%	Lowest CA index	70 [60–95]‡‡ 80 [75–85]§§	NP	103	

*Children between 1 and 15 yr old. †In patients with unfavorable outcome. ‡In patients with favorable outcome. §During vasospasm. ¶During normothermia. **During rewarming. ††During hypothermia. ‡‡Intraoperative data. §§Postoperative data.
 CA = cerebral autoregulation; CPP_{OPT} = optimal cerebral perfusion pressure; DATA CAR = dynamic adaptive target of active cerebral autoregulation; IQR = interquartile range; MAP = mean arterial blood pressure; MAP_{OPT} = optimal mean arterial blood pressure; NA = not available; NP = not performed.

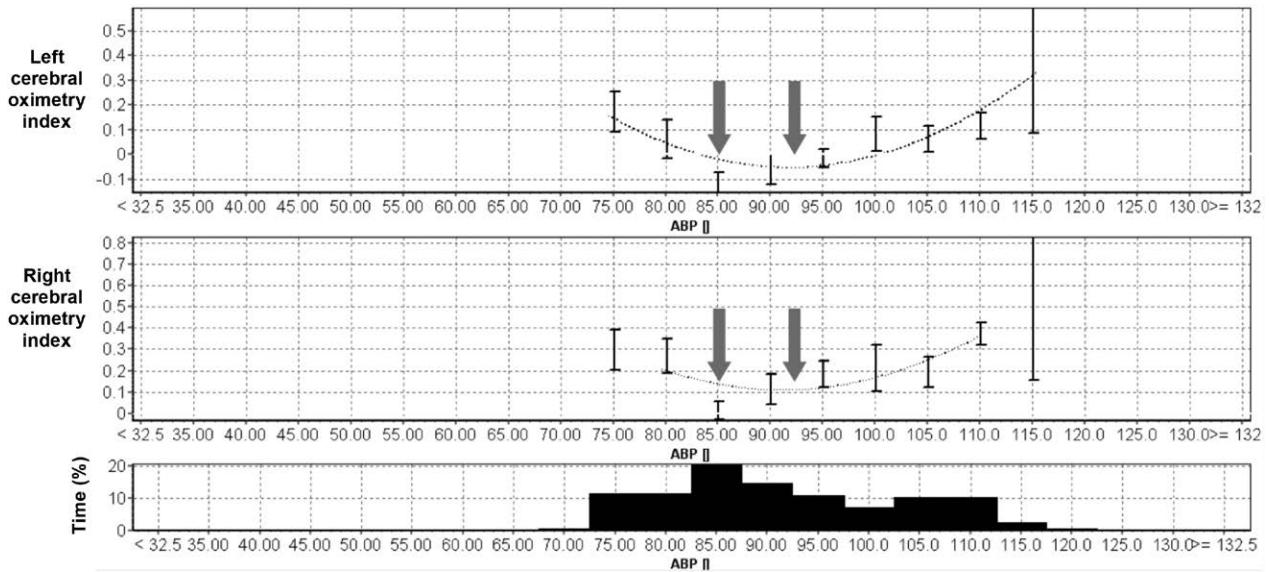


Fig. 3. A representative 4-h monitoring period shows a difference of more than 5 mmHg between the optimal mean arterial blood pressures (MAPs), defined by the U-shaped curve, and the lowest cerebral autoregulation index. This patient presented with an intracerebral hemorrhage and was continuously monitored with near-infrared spectroscopy. The *top* graph shows the left cerebral oximetry index, the *middle* graph shows the right cerebral oximetry index, and the *bottom* graph shows the histogram of monitoring time in each bin. The first arrow (at 85 mmHg) represents the optimal mean arterial blood pressure defined by the U-shaped curve method, and the second arrow (at 93 mmHg) represents the optimal mean arterial blood pressure determined by the lowest cerebral autoregulation index method.

formula that could be resolved by adjusting or weighting the curve to time or excluding bins with a monitoring time of less than 3% (fig. 3, bottom).

Recently, a study described the factors associated with such limitations in detail and concluded that the absence of slow arterial blood pressure waves, higher pressure reactivity index values, lower doses of sedative-analgesic drugs, higher vasoactive medication doses, no administration of maintenance neuromuscular blockers, and the presence of decompressive craniectomy were independently associated with the absence of a U-shaped curve.²²

Lowest Cerebral Autoregulation Index (Nadir)

Most of the studies on cardiac surgery and pediatrics and only one study on head injury have used the lowest cerebral autoregulation index as the absolute intact cerebral autoregulation. This method is based on the nadir value of the cerebral autoregulation index during the monitoring period, which could be a positive or negative value but should not be greater than the cutoff value established for the corresponding cerebral autoregulation index used. This method has the advantage of determining optimal pressures in most patients regardless of the time window for monitoring. However, it is limited by the fact that it is less objective than the polynomial derivation method and can have greater variability when more than one negative value of similar magnitude is observed at different MAPs (for example, two negative values at MAPs of 70 and 90 mmHg). It is important to recognize that no study has yet compared the last two methods discussed.

Dynamic Adaptive Target of Active Cerebral Autoregulation

Only one study has used the dynamic adaptive target of active cerebral autoregulation (DATACAR) technique, which appears to be more accurate for determining an exact and individualized optimal cerebral perfusion pressure. This method uses the same formula as the conventional U-shaped curve but additionally takes into account different time windows (*i.e.*, 1, 2, 4, 6, 8, 12, and 24 h) and assigns a weight factor to optimal cerebral perfusion pressure based on the goodness of fit of their respective U-shaped curves and the lower value of the cerebral autoregulation index of the optimal cerebral perfusion pressure. When compared with the conventional U-shaped method, this method allows optimal cerebral perfusion pressure identification in a greater number of patients and shows better accuracy for predicting outcome.²¹

Summary of the Evidence Regarding Optimal Cerebral Perfusion Pressure and Optimal MAP

Researchers have conducted multiple observational studies in adults and several in children to optimize arterial blood pressure in hospitalized patients by defining the patients' own physiologic cerebral autoregulation curve instead of using a nonindividualized target pressure recommended by guidelines. The primary objective was to provide optimum perfusion to the brain and potentially other organs (table 3). These studies calculated the optimal cerebral perfusion pressure and optimal

MAP in different populations and determined the feasibility of delineating them with cerebral autoregulation monitoring at the bedside.^{16,21–24,81,103} Four studies investigated the association of hypotension and/or hypertension based on autoregulation monitoring in adult patients with acute traumatic brain injury and functional outcomes as follows. In a large retrospective study with prospectively collected data from 327 patients in whom the pressure reactivity index was used to define optimal cerebral perfusion pressure, cerebral perfusion pressure below the optimal level increased the incidence of fatal outcome, whereas excessively high cerebral perfusion pressure levels were associated with an increased proportion of severe disability.¹⁶ Similar findings were reported in a cohort of 55 patients in whom a low-frequency autoregulation index was used to determine optimal cerebral perfusion pressure. The authors reported that having actual cerebral perfusion pressure close to the low-frequency autoregulation index—based optimal cerebral perfusion pressure was associated with increased survival.²¹ In a multivariate model, the average absolute difference between actual cerebral perfusion pressure and optimal cerebral perfusion pressure was independently associated with increased mortality. In another smaller cohort of 18 patients that used the pressure reactivity index to calculate optimal cerebral perfusion pressure, patients with a larger discrepancy (more than 10 mmHg) between actual cerebral perfusion pressure and optimal cerebral perfusion pressure were more likely to have an adverse outcome defined as a Glasgow outcome scale value equal to or greater than 3 ($P = 0.04$).¹⁹ Contrary to the aforementioned studies, one study did not find an association between optimal cerebral perfusion pressure and death or severe disability when using a new index called the low-frequency sample pressure reactivity index; however, this index has been found to have a poor predictive value for outcome by itself and also for calculation of optimal cerebral perfusion pressure.¹⁷

Only two small studies have included patients with aneurysmal subarachnoid hemorrhage ($n = 38$) and intracerebral hemorrhage ($n = 25$).^{24,25} Neither found a significant association between optimal cerebral perfusion pressure and functional outcome using the pressure reactivity index. One observational study of 121 patients undergoing cardiac surgery reported that hypotension defined with cerebral autoregulation monitoring based on the cerebral oximetry index leads to brain cellular injury characterized by elevations in serum levels of the brain-specific injury biomarker glial fibrillary acidic protein.²⁰

Several observational studies in children have calculated optimal MAP with bedside cerebral autoregulation monitoring.^{18,103,104} One study of 28 neonates with hypoxic-ischemic encephalopathy used the hemoglobin volume index to evaluate the association between blood pressure below the optimal MAP and poor outcome defined as motor and cognitive impairments at 21 to 32 months of age. The authors found that neonates with greater blood pressure deviation below optimal MAP during rewarming after therapeutic hypothermia had poor outcome.¹⁰⁴ Similar results were reported in a cohort of 30 children

with traumatic brain injury who were 6 months to 16 yr old. The authors reported that both the duration and the magnitude of negative deviations in the difference between cerebral perfusion pressure and optimal cerebral perfusion pressure were associated with unfavorable outcome defined as a Glasgow outcome scale value equal to or greater than 4.¹⁸

It is interesting to note that the mean or median calculated optimal MAP or optimal cerebral perfusion pressure differs across populations and possibly patient comorbidities. For example, patients with intracerebral hemorrhage had a higher mean optimal cerebral perfusion pressure than did patients with traumatic brain injury: 85 mmHg *versus* 75 mmHg, respectively. In addition, patients with aneurysmal subarachnoid hemorrhage and vasospasm had a higher optimal cerebral perfusion pressure than did those without vasospasm (98 mmHg *vs.* 78 mmHg, respectively). Furthermore, in some populations of patients with traumatic brain injury, the excess or deficit of cerebral perfusion pressure or MAP, based on their respective optimal values, has been associated with severe disability, whereas in patients who have undergone cardiac surgery, for example, only the deficit of MAP was associated with brain cellular injury. These differences may be explained in part by the detrimental effects of excess cerebral perfusion pressure in patients with severe acute brain injury, ICP elevation, and poor brain compliance, who may, *via* hydrostatic forces, suffer worsening cerebral edema and further rise in ICP.¹⁶ More importantly, most of the calculated mean and median optimal cerebral perfusion pressures and optimal MAPs summarized in table 3 are different from the targets recommended for blood pressure control in the current guidelines, illustrating the importance of individualizing MAP and cerebral perfusion pressure goals to achieve better outcomes.

Barriers to Adopting These Techniques into Clinical Practice

The calculation of optimal cerebral perfusion pressure and optimal MAP appears to be a useful application of cerebral autoregulation and may help clinicians individualize MAP and cerebral perfusion pressure goals to promote optimal patient management. Nevertheless, this novel technology lacks randomized controlled trial data to determine the clinical effectiveness of interventions based on optimal cerebral perfusion pressure and optimal MAP. Moreover, this technology is expensive and can be time consuming. For dynamic cerebral autoregulation monitoring and optimal cerebral perfusion pressure or optimal MAP determination, software such as ICM+ (University of Cambridge, Cambridge, United Kingdom)¹⁰⁵ is required to calculate instantaneously the correlation between the surrogate of cerebral blood flow used and MAP or cerebral perfusion pressure. Therefore, before this technology is adopted into widespread clinical practice, evidence-based data from randomized controlled trials are needed to support the premise that individualizing MAP or cerebral perfusion pressure goals based on cerebral autoregulation monitoring improves patient outcomes.

Conclusions

Monitoring of cerebral autoregulation has the potential to be used at the bedside to direct and individualize blood pressure management in the acutely ill patient. This review summarizes the evidence behind this new application of cerebral autoregulation monitoring, which has demonstrated large interindividual variability in the lower and upper limits of autoregulation, autoregulatory plateau, and optimal MAP. Cerebral autoregulation monitoring might allow clinicians to individualize management in acutely ill adults and children and thereby optimize their cerebral perfusion. Autoregulation-directed therapy should be evaluated by prospective, large-scale, randomized controlled trials in the near future.

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Competing Interests

The authors declare no competing interests.

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References

- Lassen NA: Cerebral blood flow and oxygen consumption in man. *Physiol Rev* 1959; 39:183–238
- Sheth KN, Sims JR: Neurocritical care and periprocedural blood pressure management in acute stroke. *Neurology* 2012; 79(suppl 1):S199–204
- Lang EW, Lagopoulos J, Griffith J, Yip K, Yam A, Mudaliar Y, Mehdorn HM, Dorsch NW: Cerebral vasomotor reactivity testing in head injury: The link between pressure and flow. *J Neurol Neurosurg Psychiatry* 2003; 74:1053–9
- Steven E. Lucking FAM, Robert F. Tamburro, Neal J. Thomas: *Pediatric Critical Care Study Guide: Text and Review*. London, Springer, 2012
- Powers WJ: Cerebrovascular diseases: Controversies and challenges. *Neurol Clin* 2015; 33:xiii
- Parrillo JE, Dellinger RP: *Critical Care Medicine: Principles of Diagnosis and Management in the Adult*. Philadelphia, Elsevier, 2014
- Harper AM: Autoregulation of cerebral blood flow: Influence of the arterial blood pressure on the blood flow through the cerebral cortex. *J Neurol Neurosurg Psychiatry* 1966; 29:398–403
- Ekström-Jodal B, Häggendal E, Linder LE, Nilsson NJ: Cerebral blood flow autoregulation at high arterial pressures and different levels of carbon dioxide tension in dogs. *Eur Neurol* 1971; 6:6–10
- Rapela CE, Green HD: Autoregulation of canine cerebral blood flow. *Circ Res* 1964; 15(suppl):205–12
- Strandgaard S, MacKenzie ET, Sengupta D, Rowan JO, Lassen NA, Harper AM: Upper limit of autoregulation of cerebral blood flow in the baboon. *Circ Res* 1974; 34:435–40
- Jones JV, Fitch W, MacKenzie ET, Strandgaard S, Harper AM: Lower limit of cerebral blood flow autoregulation in experimental renovascular hypertension in the baboon. *Circ Res* 1976; 39:555–7
- Budohoski KP, Czosnyka M, Smielewski P, Varsos GV, Kasprowicz M, Brady KM, Pickard JD, Kirkpatrick PJ: Cerebral autoregulation after subarachnoid hemorrhage: Comparison of three methods. *J Cereb Blood Flow Metab* 2013; 33:449–56
- Joshi B, Ono M, Brown C, Brady K, Easley RB, Yenokyan G, Gottesman RF, Hogue CW: Predicting the limits of cerebral autoregulation during cardiopulmonary bypass. *Anesth Analg* 2012; 114:503–10
- Ragauskas A, Daubaris G, Petkus V, Ragaisis V, Ursino M: Clinical study of continuous non-invasive cerebrovascular autoregulation monitoring in neurosurgical ICU. *Acta Neurochir Suppl* 2005; 95:367–70
- Czosnyka M, Smielewski P, Kirkpatrick P, Laing RJ, Menon D, Pickard JD: Continuous assessment of the cerebral vasomotor reactivity in head injury. *Neurosurgery* 1997; 41:11–9
- Aries MJ, Czosnyka M, Budohoski KP, Steiner LA, Lavinio A, Koliass AG, Hutchinson PJ, Brady KM, Menon DK, Pickard JD, Smielewski P: Continuous determination of optimal cerebral perfusion pressure in traumatic brain injury. *Crit Care Med* 2012; 40:2456–63
- Lang EW, Kasprowicz M, Smielewski P, Santos E, Pickard J, Czosnyka M: Short pressure reactivity index *versus* long pressure reactivity index in the management of traumatic brain injury. *J Neurosurg* 2015; 122:588–94
- Lewis PM, Czosnyka M, Carter BG, Rosenfeld JV, Paul E, Singhal N, Butt W: Cerebrovascular pressure reactivity in children with traumatic brain injury. *Pediatr Crit Care Med* 2015; 16:739–49
- Dias C, Silva MJ, Pereira E, Monteiro E, Maia I, Barbosa S, Silva S, Honrado T, Cerejo A, Aries MJ, Smielewski P, Paiva JA, Czosnyka M: Optimal cerebral perfusion pressure management at bedside: A single-center pilot study. *Neurocrit Care* 2015; 23:92–102
- Hori D, Ono M, Rappold TE, Conte JV, Shah AS, Cameron DE, Adachi H, Everett AD, Hogue CW: Hypotension after cardiac operations based on autoregulation monitoring leads to brain cellular injury. *Ann Thorac Surg* 2015; 100:487–93
- Depreitere B, Güiza F, Van den Berghe G, Schuhmann MU, Maier G, Piper I, Meyfroidt G: Pressure autoregulation monitoring and cerebral perfusion pressure target recommendation in patients with severe traumatic brain injury based on minute-by-minute monitoring data. *J Neurosurg* 2014; 120:1451–7
- Weersink CS, Aries MJ, Dias C, Liu MX, Koliass AG, Donnelly J, Czosnyka M, van Dijk JM, Regtien J, Menon DK, Hutchinson PJ, Smielewski P: Clinical and physiological events that contribute to the success rate of finding “optimal” cerebral perfusion pressure in severe brain trauma patients. *Crit Care Med* 2015; 43:1952–63
- Jaeger M, Dengl M, Meixensberger J, Schuhmann MU: Effects of cerebrovascular pressure reactivity-guided optimization of

- cerebral perfusion pressure on brain tissue oxygenation after traumatic brain injury. *Crit Care Med* 2010; 38:1343–7
24. Diedler J, Santos E, Poli S, Sykora M: Optimal cerebral perfusion pressure in patients with intracerebral hemorrhage: An observational case series. *Crit Care* 2014; 18:R51
 25. Bijlenga P, Czosnyka M, Budohoski KP, Soehle M, Pickard JD, Kirkpatrick PJ, Smielewski P: "Optimal cerebral perfusion pressure" in poor grade patients after subarachnoid hemorrhage. *Neurocrit Care* 2010; 13:17–23
 26. Brain Trauma Foundation, American Association of Neurological Surgeons, Congress of Neurological Surgeons, Joint Section on Neurotrauma and Critical Care AANS/CNS, Bratton SL, Chestnut RM, Ghajar J, McConnell Hammond FF, Harris OA, Hartl R, Manley GT, Nemecek A, Newell DW, Rosenthal G, Schouten J, Shutter L, Timmons SD, Ullman JS, Videtta W, Wilberger JE, Wright DW: Guidelines for the management of severe traumatic brain injury: IX. Cerebral perfusion thresholds. *J Neurotrauma* 2007; 24(suppl 1):S59–64
 27. Dellinger RP, Levy MM, Rhodes A, Annane D, Gerlach H, Opal SM, Sevransky JE, Sprung CL, Douglas IS, Jaeschke R, Osborn TM, Nunnally ME, Townsend SR, Reinhart K, Kleinpell RM, Angus DC, Deutschman CS, Machado FR, Rubenfeld GD, Webb S, Beale RJ, Vincent JL, Moreno R; Surviving Sepsis Campaign Guidelines Committee including The Pediatric Subgroup: Surviving Sepsis Campaign: International guidelines for management of severe sepsis and septic shock, 2012. *Intensive Care Med* 2013; 39:165–228
 28. Hemphill JC III, Greenberg SM, Anderson CS, Becker K, Bendok BR, Cushman M, Fung GL, Goldstein JN, Macdonald RL, Mitchell PH, Scott PA, Selim MH, Woo D; American Heart Association Stroke Council; Council on Cardiovascular and Stroke Nursing; Council on Clinical Cardiology: Guidelines for the management of spontaneous intracerebral hemorrhage: A guideline for healthcare professionals from the American Heart Association/American Stroke Association. *Stroke* 2015; 46:2032–60
 29. Connolly ES Jr, Rabinstein AA, Carhuapoma JR, Derdeyn CP, Dion J, Higashida RT, Hoh BL, Kirkness CJ, Naidech AM, Ogilvy CS, Patel AB, Thompson BG, Vespa P; American Heart Association Stroke Council; Council on Cardiovascular Radiology and Intervention; Council on Cardiovascular Nursing; Council on Cardiovascular Surgery and Anesthesia; Council on Clinical Cardiology: Guidelines for the management of aneurysmal subarachnoid hemorrhage: A guideline for healthcare professionals from the American Heart Association/American Stroke Association. *Stroke* 2012; 43:1711–37
 30. Porto GB, Spiotta AM, Chalela JA, Kellogg RT, Jauch EC: Blood pressure guideline adherence in patients with ischemic and hemorrhagic stroke in the neurointensive care unit setting. *Neurocrit Care* 2015; 23:313–20
 31. Cecconi M, De Backer D, Antonelli M, Beale R, Bakker J, Hofer C, Jaeschke R, Mebazaa A, Pinsky MR, Teboul JL, Vincent JL, Rhodes A: Consensus on circulatory shock and hemodynamic monitoring: Task force of the European Society of Intensive Care Medicine. *Intensive Care Med* 2014; 40:1795–815
 32. Czosnyka M, Smielewski P, Czosnyka Z, Piechnik S, Steiner LA, Schmidt E, Gooskens I, Soehle M, Lang EW, Matta BF, Pickard JD: Continuous assessment of cerebral autoregulation: Clinical and laboratory experience. *Acta Neurochir Suppl* 2003; 86:581–5
 33. Ibrahim J, McGee A, Graham D, McGrath JC, Dominiczak AF: Sex-specific differences in cerebral arterial myogenic tone in hypertensive and normotensive rats. *Am J Physiol Heart Circ Physiol* 2006; 290:H1081–9
 34. Osol G, Halpern W: Myogenic properties of cerebral blood vessels from normotensive and hypertensive rats. *Am J Physiol* 1985; 249:H914–21
 35. Halpern W, Osol G, Coy GS: Mechanical behavior of pressurized *in vitro* prearteriolar vessels determined with a video system. *Ann Biomed Eng* 1984; 12:463–79
 36. Hamel E: Perivascular nerves and the regulation of cerebrovascular tone. *J Appl Physiol* (1985) 2006; 100:1059–64
 37. Cauli B, Tong XK, Rancillac A, Serluca N, Lambolez B, Rossier J, Hamel E: Cortical GABA interneurons in neurovascular coupling: Relays for subcortical vasoactive pathways. *J Neurosci* 2004; 24:8940–9
 38. Rangel-Castilla L, Gasco J, Nauta HJ, Okonkwo DO, Robertson CS: Cerebral pressure autoregulation in traumatic brain injury. *Neurosurg Focus* 2008; 25:E7
 39. Yoshihara M, Bandoh K, Marmarou A: Cerebrovascular carbon dioxide reactivity assessed by intracranial pressure dynamics in severely head injured patients. *J Neurosurg* 1995; 82:386–93
 40. Golding EM, Marrelli SP, You J, Bryan RM Jr: Endothelium-derived hyperpolarizing factor in the brain: A new regulator of cerebral blood flow? *Stroke* 2002; 33:661–3
 41. Endres M, Laufs U: Effects of statins on endothelium and signaling mechanisms. *Stroke* 2004; 35(suppl 1):2708–11
 42. Enevoldsen EM, Jensen FT: Autoregulation and CO₂ responses of cerebral blood flow in patients with acute severe head injury. *J Neurosurg* 1978; 48:689–703
 43. Cortbus F, Jones PA, Miller JD, Piper IR, Tocher JL: Cause, distribution and significance of episodes of reduced cerebral perfusion pressure following head injury. *Acta Neurochir (Wien)* 1994; 130:117–24
 44. Paulson OB, Strandgaard S, Edvinsson L: Cerebral autoregulation. *Cerebrovasc Brain Metab Rev* 1990; 2:161–92
 45. Czosnyka M, Brady K, Reinhard M, Smielewski P, Steiner LA: Monitoring of cerebrovascular autoregulation: facts, myths, and missing links. *Neurocrit Care* 2009; 10:373–86
 46. Czosnyka M, Miller C; Participants in the International Multidisciplinary Consensus Conference on Multimodality Monitoring: Monitoring of cerebral autoregulation. *Neurocrit Care* 2014; 21 Suppl 2:S95–102
 47. Kontos HA, Wei EP, Navari RM, Levasseur JE, Rosenblum WI, Patterson JL Jr: Responses of cerebral arteries and arterioles to acute hypotension and hypertension. *Am J Physiol* 1978; 234:H371–83
 48. Steiner LA, Coles JP, Johnston AJ, Chatfield DA, Smielewski P, Fryer TD, Aigbirhio FI, Clark JC, Pickard JD, Menon DK, Czosnyka M: Assessment of cerebrovascular autoregulation in head-injured patients: A validation study. *Stroke* 2003; 34:2404–9
 49. Czosnyka M, Smielewski P, Kirkpatrick P, Menon DK, Pickard JD: Monitoring of cerebral autoregulation in head-injured patients. *Stroke* 1996; 27:1829–34
 50. Reinhard M, Rutsch S, Lambeck J, Wihler C, Czosnyka M, Weiller C, Hetzel A: Dynamic cerebral autoregulation associates with infarct size and outcome after ischemic stroke. *Acta Neurol Scand* 2012; 125:156–62
 51. Naqvi J, Yap KH, Ahmad G, Ghosh J: Transcranial Doppler ultrasound: A review of the physical principles and major applications in critical care. *Int J Vasc Med* 2013; 2013:629378
 52. Lang EW, Lagopoulos J, Griffith J, Yip K, Mudaliar Y, Mehdorn HM, Dorsch NW: Noninvasive cerebrovascular autoregulation assessment in traumatic brain injury: validation and utility. *J Neurotrauma* 2003; 20:69–75
 53. Jöbsis FF: Noninvasive, infrared monitoring of cerebral and myocardial oxygen sufficiency and circulatory parameters. *Science* 1977; 198:1264–7
 54. Matcher SJ, Elwell CE, Cooper CE, Cope M, Delpy DT: Performance comparison of several published tissue near-infrared spectroscopy algorithms. *Anal Biochem* 1995; 227:54–68
 55. Steppan J, Hogue CW Jr: Cerebral and tissue oximetry. *Best Pract Res Clin Anaesthesiol* 2014; 28:429–39
 56. Ghosh A, Elwell C, Smith M: Review article: cerebral near-infrared spectroscopy in adults: A work in progress. *Anesth Analg* 2012; 115:1373–83

57. Rolfe P: *In vivo* near-infrared spectroscopy. *Annu Rev Biomed Eng* 2000; 2:715–54
58. Moerman AT, Vanbiervliet VM, Van Wesemael A, Bouchez SM, Wouters PF, De Hert SG: Assessment of cerebral autoregulation patterns with near-infrared spectroscopy during pharmacological-induced pressure changes. *ANESTHESIOLOGY* 2015; 123:327–35
59. Schytz HW, Guo S, Jensen LT, Kamar M, Nini A, Gress DR, Ashina M: A new technology for detecting cerebral blood flow: A comparative study of ultrasound tagged NIRS and ¹³³Xe-SPECT. *Neurocrit Care* 2012; 17:139–45
60. Mahan GD, Engler WE, Tiemann JJ, Uzgiris E: Ultrasonic tagging of light: theory. *Proc Natl Acad Sci USA* 1998; 95:14015–9
61. Hori D, Hogue CW Jr, Shah A, Brown C, Neufeld KJ, Conte JV, Price J, Sciortino C, Max L, Laflam A, Adachi H, Cameron DE, Mandal K: Cerebral autoregulation monitoring with ultrasound-tagged near-infrared spectroscopy in cardiac surgery patients. *Anesth Analg* 2015; 121:1187–93
62. Sen AN, Gopinath SP, Robertson CS: Clinical application of near-infrared spectroscopy in patients with traumatic brain injury: A review of the progress of the field. *Neurophotonics* 2016; 3:031409
63. Bouma GJ, Muizelaar JP, Choi SC, Newlon PG, Young HF: Cerebral circulation and metabolism after severe traumatic brain injury: The elusive role of ischemia. *J Neurosurg* 1991; 75:685–93
64. Muizelaar JP, Ward JD, Marmarou A, Newlon PG, Wachi A: Cerebral blood flow and metabolism in severely head-injured children: Part 2. Autoregulation. *J Neurosurg* 1989; 71:72–6
65. Aaslid R, Lindegaard KF, Sorteberg W, Nornes H: Cerebral autoregulation dynamics in humans. *Stroke* 1989; 20:45–52
66. Cold GE, Jensen FT, Malmros R: The cerebrovascular CO₂ reactivity during the acute phase of brain injury. *Acta Anaesthesiol Scand* 1977; 21:222–31
67. Carey BJ, Manktelow BN, Panerai RB, Potter JF: Cerebral autoregulatory responses to head-up tilt in normal subjects and patients with recurrent vasovagal syncope. *Circulation* 2001; 104:898–902
68. Brown CM, Dütsch M, Hecht MJ, Neundörfer B, Hilz MJ: Assessment of cerebrovascular and cardiovascular responses to lower body negative pressure as a test of cerebral autoregulation. *J Neurol Sci* 2003; 208:71–8
69. Smielewski P, Czosnyka M, Kirkpatrick P, Pickard JD: Evaluation of the transient hyperemic response test in head-injured patients. *J Neurosurg* 1997; 86:773–8
70. Strebel S, Lam AM, Matta B, Mayberg TS, Aaslid R, Newell DW: Dynamic and static cerebral autoregulation during isoflurane, desflurane, and propofol anesthesia. *ANESTHESIOLOGY* 1995; 83:66–76
71. Lang EW, Mehdorn HM, Dorsch NW, Czosnyka M: Continuous monitoring of cerebrovascular autoregulation: A validation study. *J Neurol Neurosurg Psychiatry* 2002; 72:583–6
72. Sorrentino E, Budohoski KP, Kaszowicz M, Smielewski P, Matta B, Pickard JD, Czosnyka M: Critical thresholds for transcranial Doppler indices of cerebral autoregulation in traumatic brain injury. *Neurocrit Care* 2011; 14:188–93
73. Ono M, Joshi B, Brady K, Easley RB, Zheng Y, Brown C, Baumgartner W, Hogue CW: Risks for impaired cerebral autoregulation during cardiopulmonary bypass and postoperative stroke. *Br J Anaesth* 2012; 109:391–8
74. Ono M, Arnaoutakis GJ, Fine DM, Brady K, Easley RB, Zheng Y, Brown C, Katz NM, Grams ME, Hogue CW: Blood pressure excursions below the cerebral autoregulation threshold during cardiac surgery are associated with acute kidney injury. *Crit Care Med* 2013; 41:464–71
75. Jones SC, Williams JL, Shea M, Easley KA, Wei D: Cortical cerebral blood flow cycling: Anesthesia and arterial blood pressure. *Am J Physiol* 1995; 268:H569–75
76. Bochicchio M, Latronico N, Zappa S, Beindorf A, Candiani A: Bedside burr hole for intracranial pressure monitoring performed by intensive care physicians: A 5-year experience. *Intensive Care Med* 1996; 22:1070–4
77. Purkayastha S, Sorond F: Transcranial Doppler ultrasound: Technique and application. *Semin Neurol* 2012; 32:411–20
78. Ogoh S, Sato K, Okazaki K, Miyamoto T, Secher F, Sørensen H, Rasmussen P, Secher NH: A decrease in spatially resolved near-infrared spectroscopy-determined frontal lobe tissue oxygenation by phenylephrine reflects reduced skin blood flow. *Anesth Analg* 2014; 118:823–9
79. Lazaridis C, Andrews CM: Brain tissue oxygenation, lactate-pyruvate ratio, and cerebrovascular pressure reactivity monitoring in severe traumatic brain injury: Systematic review and viewpoint. *Neurocrit Care* 2014; 21:345–55
80. Schreiber SJ, Gottschalk S, Weih M, Villringer A, Valdueza JM: Assessment of blood flow velocity and diameter of the middle cerebral artery during the acetazolamide provocation test by use of transcranial Doppler sonography and MR imaging. *AJNR Am J Neuroradiol* 2000; 21:1207–11
81. Blaine Easley R, Kibler KK, Brady KM, Joshi B, Ono M, Brown C, Hogue CW: Continuous cerebrovascular reactivity monitoring and autoregulation monitoring identify similar lower limits of autoregulation in patients undergoing cardiopulmonary bypass. *Neurol Res* 2013; 35:344–54
82. Budohoski KP, Czosnyka M, Smielewski P, Kaszowicz M, Helmy A, Bulters D, Pickard JD, Kirkpatrick PJ: Impairment of cerebral autoregulation predicts delayed cerebral ischemia after subarachnoid hemorrhage: A prospective observational study. *Stroke* 2012; 43:3230–7
83. Severdija EE, Gommer ED, Weerwind PW, Reulen JP, Mess WH, Maessen JG: Assessment of dynamic cerebral autoregulation and cerebral carbon dioxide reactivity during normothermic cardiopulmonary bypass. *Med Biol Eng Comput* 2015; 53: 195–203
84. Reinhard M, Gerds TA, Grabiak D, Zimmermann PR, Roth M, Guschlbauer B, Timmer J, Czosnyka M, Weiller C, Hetzel A: Cerebral dysautoregulation and the risk of ischemic events in occlusive carotid artery disease. *J Neurol* 2008; 255:1182–9
85. Jaeger M, Schuhmann MU, Soehle M, Nagel C, Meixensberger J: Continuous monitoring of cerebrovascular autoregulation after subarachnoid hemorrhage by brain tissue oxygen pressure reactivity and its relation to delayed cerebral infarction. *Stroke* 2007; 38:981–6
86. Sánchez-Porras R, Santos E, Czosnyka M, Zheng Z, Unterberg AW, Sakowitz OW: “Long” pressure reactivity index (L-PRx) as a measure of autoregulation correlates with outcome in traumatic brain injury patients. *Acta Neurochir (Wien)* 2012; 154:1575–81
87. Tiecks FP, Lam AM, Aaslid R, Newell DW: Comparison of static and dynamic cerebral autoregulation measurements. *Stroke* 1995; 26:1014–9
88. Lavinio A, Schmidt EA, Haubrich C, Smielewski P, Pickard JD, Czosnyka M: Noninvasive evaluation of dynamic cerebrovascular autoregulation using Finapres plethysmograph and transcranial Doppler. *Stroke* 2007; 38:402–4
89. Liu X, Czosnyka M, Donnelly J, Budohoski KP, Varsos GV, Nasr N, Brady KM, Reinhard M, Hutchinson PJ, Smielewski P: Comparison of frequency and time domain methods of assessment of cerebral autoregulation in traumatic brain injury. *J Cereb Blood Flow Metab* 2015; 35:248–56
90. Aries MJ, Czosnyka M, Budohoski KP, Koliass AG, Radolovich DK, Lavinio A, Pickard JD, Smielewski P: Continuous monitoring of cerebrovascular reactivity using pulse waveform of intracranial pressure. *Neurocrit Care* 2012; 17:67–76
91. Czosnyka M, Smielewski P, Lavinio A, Pickard JD, Panerai R: An assessment of dynamic autoregulation from spontaneous fluctuations of cerebral blood flow velocity: A comparison of two models, index of autoregulation and mean flow index. *Anesth Analg* 2008; 106:234–9

92. Highton D, Ghosh A, Tachtsidis I, Panovska-Griffiths J, Elwell CE, Smith M: Monitoring cerebral autoregulation after brain injury: Multimodal assessment of cerebral slow-wave oscillations using near-infrared spectroscopy. *Anesth Analg* 2015; 121:198–205
93. Ono M, Zheng Y, Joshi B, Sigl JC, Hogue CW: Validation of a stand-alone near-infrared spectroscopy system for monitoring cerebral autoregulation during cardiac surgery. *Anesth Analg* 2013; 116:198–204
94. Steiner LA, Pfister D, Strebel SP, Radolovich D, Smielewski P, Czosnyka M: Near-infrared spectroscopy can monitor dynamic cerebral autoregulation in adults. *Neurocrit Care* 2009; 10:122–8
95. Brady K, Joshi B, Zweifel C, Smielewski P, Czosnyka M, Easley RB, Hogue CW Jr: Real-time continuous monitoring of cerebral blood flow autoregulation using near-infrared spectroscopy in patients undergoing cardiopulmonary bypass. *Stroke* 2010; 41:1951–6
96. Barth M, Woitzik J, Weiss C, Muench E, Diepers M, Schmiedek P, Kasuya H, Vajkoczy P: Correlation of clinical outcome with pressure-, oxygen-, and flow-related indices of cerebrovascular reactivity in patients following aneurysmal SAH. *Neurocrit Care* 2010; 12:234–43
97. Santos E, Diedler J, Sykora M, Orakcioglu B, Kentar M, Czosnyka M, Unterberg A, Sakowitz OW: Low-frequency sampling for PRx calculation does not reduce prognostication and produces similar CPPopt in intracerebral haemorrhage patients. *Acta Neurochir (Wien)* 2011; 153:2189–95
98. Soehle M, Czosnyka M, Pickard JD, Kirkpatrick PJ: Continuous assessment of cerebral autoregulation in subarachnoid hemorrhage. *Anesth Analg* 2004; 98:1133–9
99. Grözinger G, Schenk M, Morgalla MH, Thiel C, Thiel K, Schuhmann MU: The values of cerebrovascular pressure reactivity and brain tissue oxygen pressure reactivity in experimental anhepatic liver failure. *Neurocrit Care* 2012; 17:271–80
100. Budohoski KP, Czosnyka M, de Riva N, Smielewski P, Pickard JD, Menon DK, Kirkpatrick PJ, Lavinio A: The relationship between cerebral blood flow autoregulation and cerebrovascular pressure reactivity after traumatic brain injury. *Neurosurgery* 2012; 71:652–61
101. Zweifel C, Castellani G, Czosnyka M, Helmy A, Manktelow A, Carrera E, Brady KM, Hutchinson PJ, Menon DK, Pickard JD, Smielewski P: Noninvasive monitoring of cerebrovascular reactivity with near infrared spectroscopy in head-injured patients. *J Neurotrauma* 2010; 27:1951–8
102. Schmidt B, Reinhard M, Lezaic V, McLeod DD, Weinhold M, Mattes H, Klingelhöfer J: Autoregulation monitoring and outcome prediction in neurocritical care patients: Does one index fit all? *J Clin Monit Comput* 2016; 30:367–75
103. Lee JK, Williams M, Jennings JM, Jamrogowicz JL, Larson AC, Jordan LC, Heitmiller ES, Hogue CW, Ahn ES: Cerebrovascular autoregulation in pediatric moyamoya disease. *Paediatr Anaesth* 2013; 23:547–56
104. Burton VJ, Gerner G, Cristofalo E, Chung SE, Jennings JM, Parkinson C, Koehler RC, Chavez-Valdez R, Johnston MV, Northington FJ, Lee JK: A pilot cohort study of cerebral autoregulation and 2-year neurodevelopmental outcomes in neonates with hypoxic-ischemic encephalopathy who received therapeutic hypothermia. *BMC Neurol* 2015; 15:209
105. Smielewski PC, Czosnyka M: ICM+ software for brain monitoring in neurological intensive care research. Available at: <https://www.enterprise.cam.ac.uk/opportunities/icm-software-for-brain-monitoring-in-neurological-intensive-care-research/>. Accessed March 9, 2015