

Evidence for Haemosporidian Parasite Infections in Spectacled Eiders (*Somateria fischeri*) Sampled in Alaska, USA during the Breeding Season

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ABSTRACT: We assessed hematozoa infection in Spectacled Eiders (*Somateria fischeri*) at two areas in Alaska, US. No *Haemoproteus* or *Plasmodium* species were detected. *Leucocytozoon* prevalence was 6.5% for adults across sites and 41.9% for juveniles sampled in the Arctic, providing evidence for local transmission. All *Leucocytozoon* haplotypes were previously detected in waterfowl.

Haemosporidian parasites, or hematozoa, have been associated with mortality events or population declines in songbirds, penguins, and waterfowl (Herman et al. 1975; van Riper et al. 1986; Hill et al. 2010). Hematozoa transmission is regulated by temperature and the presence of suitable vectors; it has been predicted that the prevalence and distribution of hematozoa may be affected by ecological change (Garamszegi 2011; Loiseau et al. 2012; Zamora-Vilchis et al. 2012), particularly in high-latitude areas. In Alaska, information on the distribution of hematozoa in wild birds has increased in recent years (Ramey et al. 2012; Reeves et al. 2015; Smith et al. 2016), yet the diversity of host taxa sampled remains limited. For example, molecular techniques have not been used to assess hematozoa infection in sea ducks (family: Anatidae; tribe: Mergini).

Spectacled Eiders (*Somateria fischeri*) are sea ducks inhabiting arctic and subarctic regions of eastern Siberia and Alaska and are listed as “threatened” under the Endangered Species Act by the US Fish and Wildlife Service. In Alaska, breeding populations inhabit the Yukon-Kuskokwim Delta (YKD) and Arctic Coastal Plain (ACP) on the western and northern coasts, respectively. Spectacled Eiders nest within wetlands adjacent to the Bering, Chukchi, Beaufort, and East Siberian seas and spend non-breeding seasons offshore

in the Arctic Ocean and Bering Sea (Petersen et al. 2000). Consequently, exposure of Spectacled Eiders to hematozoa via biting insects is presumably restricted to breeding areas.

We screened blood samples from Spectacled Eiders collected as part of a larger ecological study, to assess the prevalence and diversity of hematozoa infecting this species. Our objectives were to 1) estimate hematozoa prevalence in adult and juvenile Spectacled Eiders, 2) assess transmission on the ACP, and 3) characterize the genetic and morphologic diversity of parasites infecting Spectacled Eiders relative to other avian hosts. Our results provide a baseline for future assessments of health effects from parasite infections on Spectacled Eiders and changes in parasite prevalence in subarctic and arctic habitats.

We captured adult Spectacled Eiders on the YKD ($n=46$) and ACP ($n=77$) in May–June 2008 and June–August 2009–12, respectively, using methods described in Sexson et al. (2016). Additionally, we captured flightless ducklings ($n=62$) 5 wk after peak hatch on the ACP in 2010–11. Jugular blood samples (1–2 mL) were collected from each individual and preserved in either 2-mL vials with 0.75 mL Longmire Buffer, or BD Vacutainer EDTA tubes (Franklin Lakes, New Jersey, USA). We made thin blood smears on glass slides from 110 ACP samples. We conducted all capture and field sampling protocols under authorizations granted by the US Geological Survey’s Animal Care and Use Committee (2008-04) and the US Fish and Wildlife Service (TE012158-0). Molecular methods for detecting and genetically characterizing hematozoa followed Ramey et al. (2013). Sparse age- and

TABLE 1. Number and percentage of Spectacled Eider (*Somateria fischeri*) blood samples from the Yukon-Kuskokwim Delta (YKD) and Arctic Coastal Plain (ACP), Alaska, USA, detected as positive for *Leucocytozoon* infection using molecular techniques by location in Alaska, year, age, and sex.^a

| Location | Year | Age | Sex | Leucocytozoon | | | Haplotypes detected |
|----------|------|-----|-----|----------------|-------------------|---------------------------|------------------------------------|
| | | | | Total <i>n</i> | Positive <i>n</i> | % | |
| YKD | 2008 | Ad | F | 31 | 3 | 10 | SPEI 02, SPEI 03, SPEI 05 |
| | | | M | 15 | 2 | 13 | SPEI 03 |
| ACP | 2009 | Ad | F | 7 | 0 | 0 | |
| | | | M | 13 | 2 | 15 | SPEI 02, SPEI 08 |
| | 2010 | Ad | F | 21 | 1 | 5 | SPEI 06 |
| | | | M | 3 | 0 | 0 | |
| | | Juv | F | 5 | 2 | 40 | SPEI 06, SPEI 07 |
| | | | M | 14 | 11 | 79 | SPEI 01, SPEI 06, SPEI 07, SPEI 08 |
| | 2011 | Ad | F | 24 | 0 | 0 | |
| | | | Juv | F | 22 | 8 | 36 |
| | | M | 21 | 5 | 24 | SPEI 07, SPEI 08, SPEI 09 | |
| 2012 | Ad | F | 9 | 0 | 0 | | |

^a Ad = adult; Juv = juvenile; M = male; F = female.

sex-specific data, along with low infection rates, precluded incorporation of occupancy model estimates of detection probability and of potential false negatives in prevalence estimates. Blood smears were stained with Giemsa and examined at 400–1,000× magnification to identify the morphospecies of hematozoa in samples that tested positive for infection via molecular screening. All data related to this study are archived in Reed (2018).

Using molecular methods, we detected *Leucocytozoon* infections in 18.4% of samples (34/185; Table 1). Morphologic assessment of blood smears identified *Leucocytozoon simondi* as the parasite present in ACP samples (Fig. 1a; voucher specimens archived at the Smithsonian Institution, Museum of Natural History, USNM1486967–USNM1486972). *Leucocytozoon* prevalence in juveniles (41.9%; 26/62) was higher than in adults (6.5%; 8/123; Table 1), possibly because of downy ducklings having less physical protection from biting insects than fully feathered birds. Detection of *Leucocytozoon* infections in juvenile eiders provides evidence of transmission within the ACP, corroborating previous reports of *Leucocytozoon* transmission among geese in this region (Ramey et al. 2014).

We did not detect *Haemoproteus* or *Plasmodium* infections in Spectacled Eiders at either the YKD or ACP. Previous studies reported low prevalence of *Plasmodium* infections (4%) in Northern Pintails (*Anas acuta*) sampled farther inland on the YKD and *Haemoproteus* parasites in a variety of waterfowl species sampled at interior (14%) and coastal areas (1–3%) of the YKD and ACP (Ramey et al. 2012, 2014, 2015). Spectacled Eiders may be infected with *Haemoproteus* and *Plasmodium* at prevalences below the detection threshold of our methodology; alternatively, parasite development may be restricted by the relatively low temperatures along the narrow band of coastal habitat occupied by eiders sampled in this study. *Haemoproteus* and *Plasmodium* parasites previously detected in other waterfowl at the YKD and ACP may have been acquired in areas farther from the coast or other regions during non-breeding season.

We identified nine *Leucocytozoon* haplotypes (GenBank MG734969–MG734977) with nucleotide identity differing by 0.25–9.3% (1–37/400 base pairs; Fig. 1b). These nine haplotypes shared 100% identity with those previously detected in 16 avian species (predominately waterfowl) sampled at loca-

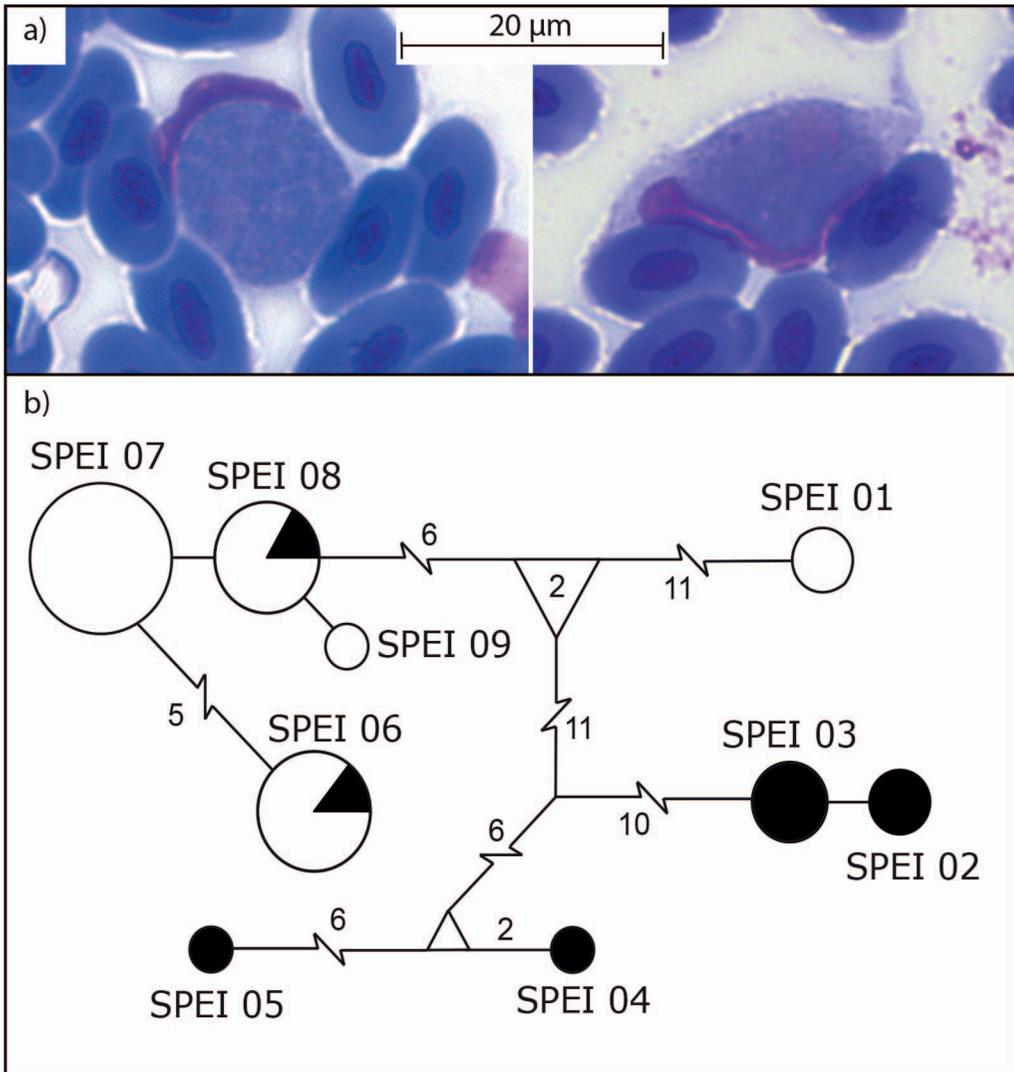


FIGURE 1. (a) *Leucocytozoon simondi* gametocytes in blood smears from Spectacled Eiders (*Somateria fischeri*) on the Arctic Coastal Plain of Alaska, USA (2010–11). (b) Minimum spanning network for *Leucocytozoon* mitochondrial DNA cytochrome *b* haplotypes identified by molecular techniques in blood sampled from Spectacled Eiders in Alaska (2008–12). Circle sizes are proportional to haplotype frequency. Host age class is represented by color (black: adult, white: juvenile). A single mutation separates nodes unless indicated by number. Lines separating nodes are drawn to scale unless indicated by a break.

tions in Alaska, elsewhere in North America, and in East Asia (see the Supplementary Table). These results are consistent with prior research, suggesting broad spatial distributions and genetic diversity for *Leucocytozoon* lineages infecting waterfowl (Reeves et al. 2015). Two *Leucocytozoon* haplotypes were identified in both juvenile and adult Spectacled Eiders (Fig. 1b), including one adult

female and her brood, indicating that parasites may be transmitted within and among family groups on the ACP during brood rearing. However, we observed most parasite haplotypes exclusively in either adult or juvenile birds (Fig. 1b). This suggests more complex spatiotemporal dynamics or the involvement of additional host species in *Leucocytozoon* transmission. Additional re-

search into transmission dynamics on the YKD and ACP, health consequences of hematozoa infections in Spectacled Eiders, and changes in parasite prevalence and distribution over time, would be helpful for refining the ecological context of our results.

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SUPPLEMENTARY MATERIAL

Supplementary material for this article is online at <http://dx.doi.org/10.7589/2018-01-012>.

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