

Advances in HER2-Targeted Therapy: Novel Agents and Opportunities Beyond Breast and Gastric Cancer



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Abstract

The introduction of HER2-targeted therapy for breast and gastric patients with *ERBB2* (*HER2*) amplification/overexpression has led to dramatic improvements in oncologic outcomes. In the past 20 years, five HER2-targeted therapies have been FDA approved, with four approved in the past 8 years. HER2-targeted therapy similarly was found to improve outcomes in HER2-positive gastric cancer. Over the past decade, with the introduction of next-generation sequencing into clinical practice, our understanding of HER2 biology has dramatically improved. We have recognized that *HER2* amplification is not limited to breast and gastric cancer but is also

found in a variety of tumor types such as colon cancer, bladder cancer, and biliary cancer. Furthermore, HER2-targeted therapy has signal of activity in several tumor types. In addition to *HER2* amplification and overexpression, there is also increased recognition of activating *HER2* mutations and their potential therapeutic relevance. Furthermore, there is a rapidly growing number of new therapeutics targeting HER2 including small-molecule inhibitors, antibody–drug conjugates, and bispecific antibodies. Taken together, an increasing number of patients are likely to benefit from approved and emerging HER2-targeted therapies.

Introduction

HER2 (*ERBB2*) is emerging as a promising target for genomically informed therapy across a variety of tumor types. For *HER2*, gene amplification (increased copy number) is by far the most common genomic alteration and is generally, although not always, associated with protein overexpression (1–3). HER2 overexpression drives tumorigenesis through the creation of spontaneous receptor homodimers (4, 5), or heterodimers with other ERBB family members (6) resulting in activated oncogenic downstream signaling, such as PI3K/Akt/mTOR and MAPK, promoting cellular proliferation, survival, and angiogenesis (6–8). In particular, HER2–HER3 heterodimers transduce PI3K signaling via direct binding between HER3 and the p85 subunit of PI3K (9). Spontaneous formation of these heterodimers increases with amplification of the *HER2* gene (10).

Algorithms for HER2 classification have been evolving. For example, for breast cancer, 3+ HER2 protein overexpression by IHC, or *HER2* amplification assessed by ISH have been considered HER2-positive, and detailed guidelines for interpretation (11) have been developed by the American Society of Clinical Oncology and College of American Pathologists and are regularly updated.

Approved Indications for HER2-Targeted Therapy

HER2-targeted therapy has transformed outcomes for *HER2*-amplified/overexpressing (HER2-positive) breast and gastric/gastroesophageal cancer. Several therapies are approved for HER2-positive breast cancer in the adjuvant and metastatic setting: trastuzumab (metastatic and adjuvant), pertuzumab (metastatic and adjuvant), lapatinib (metastatic), ado-trastuzumab emtansine (metastatic), and neratinib (adjuvant). Trastuzumab is also approved for HER2-positive metastatic gastric/gastroesophageal junction cancers, in combination with cisplatin and a fluoropyrimidine (capecitabine or 5-fluorouracil). Furthermore, Trastuzumab-dkst (Ogivri; Mylan), a trastuzumab biosimilar, was approved for all indications included in the label of trastuzumab. FDA-approved indications are detailed in Supplementary Table S1.

Targeting *HER2* Overexpression/Amplification Beyond Breast and Gastric Cancer with Agents Approved for Breast/Gastric Cancer

With increased genomic profiling of many types of tumor, there is growing recognition that *HER2* amplification occurs in several

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Note: Supplementary data for this article are available at Clinical Cancer Research Online (<http://clincancerres.aacrjournals.org/>).

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doi: 10.1158/1078-0432.CCR-18-2275

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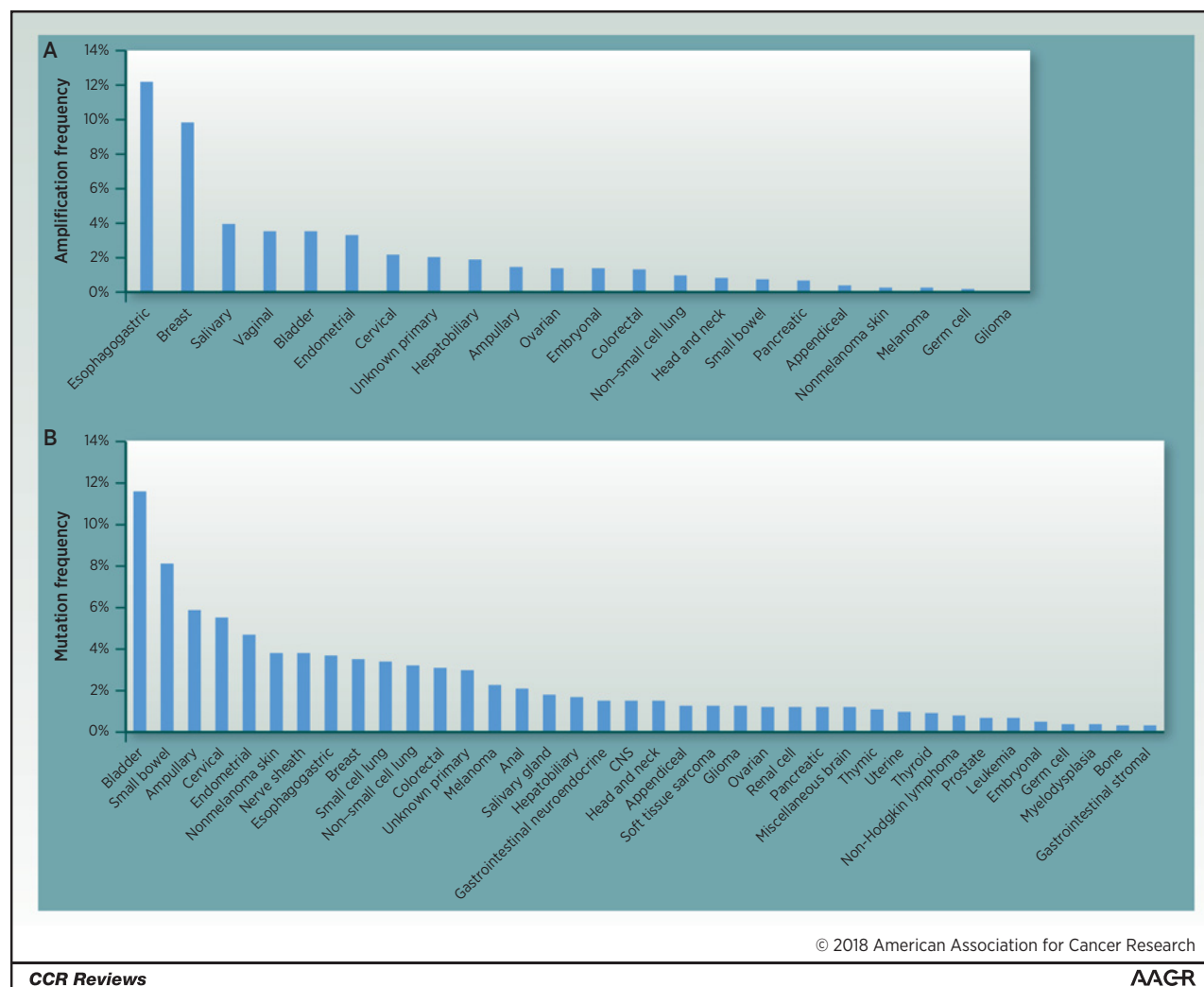


Figure 1. *HER2* alterations across tumor types. **A**, Prevalence of *HER2* amplifications across diverse cancer types in a cohort of 37,436 sequenced cases extracted from American Association for Cancer Research (AACR) Project GENIE (version 3.0.0, accessed on July 16, 2018; ref. 12). **B**, Prevalence of *HER2* mutations across diverse cancer types (version 3.0.0, accessed on July 16, 2018). CNS, central nervous system.

tumor types including salivary (3.9%), vaginal (3.6%), bladder (3.6%), endometrial (3.4%), cervical (2.2%), and colorectal cancer (1.3%; Fig. 1A; ref. 12).

The efficacy of pertuzumab and trastuzumab was tested for *HER2*-positive tumors in the MY PATHWAY basket trial (13). Thirty of 114 patients [26%; 95% confidence interval (CI), 19%–35%] with *HER2* amplification/overexpression had objective responses (OR) [two complete responses (CR), 28 partial responses (PR)]. Responses were seen in 9 tumor types: colorectal [38% (14/37) objective response rate (ORR); 95% CI, 23–55%], bladder [33% ORR (3/9); 95% CI, 8–70], biliary/gallbladder [29% ORR (2/7); 95% CI, 4–71], salivary gland [80% ORR (4/5); 95% CI, 28–99], non-small cell lung [13% ORR (2/16); 95% CI, 2–38], pancreas [22% ORR (2/9); 95% CI, 3–60], ovary [13% ORR (1/8); 95% CI, 0–53], as well as 1 patient each with prostate and skin cancer (apocrine). Furthermore, data on efficacy of pertuzumab and trastuzumab is expected from the MY PATHWAY trial as well as the ASCO TAPUR and NCI-MATCH trials; all three trials are being conducted in treatment-refractory patients.

Supporting the efficacy signal with *HER2*-targeted therapy seen in colorectal cancer in the MY PATHWAY trial, several lines of evidence point to the importance of *HER2* in colorectal cancer biology. Bertotti and colleagues used a patient-derived xenograft (PDX) platform to identify genotype–response correlations with cetuximab, and found *HER2* amplification in cetuximab-resistant, *KRAS/NRAS/BRAF/PIK3CA* wild-type PDXs (14). *HER2* amplification was also enriched in clinically nonresponsive patients with *KRAS* wild-type. Furthermore, Raghav and colleagues reported that *HER2* amplification is associated with resistance to anti-EGFR therapy (cetuximab/panitumumab) and shorter progression-free survival (15). Taken together, this data suggest that *HER2* amplification may not only be a potential target, but also a resistance marker for EGFR inhibitors.

The role of *HER2* as a target for metastatic colorectal cancer was also assessed in the HERACLES trial, which enrolled patients with *KRAS* exon 2 wild-type patients that were *HER2*-positive as defined as *HER2* 3+ overexpression in over 50% of tumor cells by IHC or 2+ IHC and a *HER2/CEP17* ratio greater than 2 in more

than 50% of cells by FISH (16). Eight (30%) of 27 patients treated with dual-targeted therapy, trastuzumab and lapatinib, achieved an OR, (one CR, seven PRs). Together, the HERACLES and MY PATHWAY studies demonstrate that HER2-targeted therapy is effective in colorectal cancer. An ongoing phase II trial, the SWOG S1613 study (NCT03365882), is comparing the efficacy of trastuzumab and pertuzumab to cetuximab and irinotecan. The trial is accruing patients with metastatic or locally advanced, unresectable colorectal cancer who have not received prior EGF or HER2 inhibitors and have *HER2*-amplified tumors that are *KRAS/NRAS/BRAF* wild-type. Furthermore, study is needed to determine the optimal therapeutic regimens and treatment sequencing.

Recently, two multihistology basket trials using adotrastuzumab emtansine (T-DM1) for *HER2*-amplified disease were also reported. Li and colleagues reported a trial conducted at Memorial Sloan Kettering Cancer Center (MSKCC, New York, NY) that demonstrated an ORR of 28%, with responses in lung cancer (43% ORR), endometrial cancer (25% ORR), salivary cancer (100% ORR), as well as in ovarian cancer and biliary cancer, but not in colorectal cancer (17). In the NCI-MATCH trial, ORR was 8%, with responses seen in 2 of 3 salivary cancers (18). This data raise some interesting points. Although the differences in ORR may simply be due to small study size, there appear to be important differences in ORR between the two trials with similar study designs, using the same drug. It will be important to review the differences in patient populations when details are available. In NCI-MATCH, 33% of patients had received >3 lines of prior therapy, thus it is possible that NCI-MATCH patients were more heavily treated, limiting therapeutic efficacy. In contrast, it is likely that in the MSKCC series, patients were offered genomic testing, and genomically matched therapy earlier in their disease course. If so, the differences in ORR would support earlier genomic testing to allow for greater benefit of genomically informed therapy. Another interesting finding was that in the MSKCC series there was efficacy in several tumor types, but there was also variability in sensitivity by tumor type, with no responses observed in colorectal cancer. This highlights that for antibody–drug conjugates, in addition to expression of the marker, sensitivity to the specific conjugate needs to be taken into consideration.

HER2 Mutations

Somatic mutations can also drive HER2 signaling. Although activating mutations have been best characterized within breast and lung cancers (19–21), mutations are reported in a variety of other tumor types (22). A query of the 214 tumor-based (noncell line) studies within the cBioPortal reveals a *HER2* mutation frequency (mutations or fusions) of 2.7% (23). Figure 1B demonstrates the frequency of mutations across tumor types in the American Association for Cancer Research (AACR) GENIE dataset: tumor types with the highest frequency of mutations in *HER2* include bladder (11.6%), small bowel (8.1%), and ampullary (5.9%; ref. 12).

However, not all *HER2* mutations are activating. Most of the known activating *HER2* mutations are shown in Fig. 2 and detailed in Supplementary Table S2. Although mutations have been reported across the entire gene, they primarily localize within two regions: the extracellular domain (ECD) and the kinase domain (KD; refs. 19, 24). Within the KD, both missense mutations and in-frame insertions lead to increased kinase activ-

ity and promote tumorigenesis (19, 25, 26). Activating mutations have also been reported in the transmembrane domain (27, 28) with enhanced protein stabilization being one proposed mechanism for gain-of-function (28).

Types of mutations vary with tumor type. In non-small cell lung cancer (NSCLC), mutations are most frequent in patients that are Asian, female, never-smokers, and in adenocarcinomas (29–31). Mutations are generally exon 20 in-frame insertions within the KD (20, 21, 29, 32). These insertions/duplications occur at the same codons identified in *EGFR*, indicative of a similar mechanism of activation (29). Indeed, functional studies have shown that *HER2*^{YVMA} insertion/duplications increase autocatalytic activity, leading to increased autophosphorylation and phosphorylation of its dimerization partners, and increased survival, proliferation, and tumorigenesis (25, 26). In NSCLC, *HER2* mutations are mutually exclusive with activating *EGFR* and *KRAS* mutations (21, 29, 30).

In breast cancer, primarily missense mutations are detected. A meta-analysis of 12,905 patients with breast cancer published across 31 articles revealed a mutation frequency of 2.7% (33). Greater than 50% of mutations were located in the ECD (S310/Y) or the KD (33). The most frequently reported mutations in the KD include L755S, V777L, D769H/Y, and L755_T759del (33, 34). Although these and other *HER2* mutations activate downstream signaling and/or increase colony formation (19), not all are tumor-promoting *in vivo*. The V777L, D769H, and G309A mutants induced tumor growth in xenograft models; however, the L755S, V842I, and R678Q mutants performed similar to wild-type, and the L755_T759del tumors grew slower than wild-type (19). Moreover, a study investigating the effects of *HER2* mutants expressed at endogenous levels found none induced tumor formation *in vivo* (35). However, *HER2* mutations may cooperate with other oncogenic events (e.g., *PIK3CA* activation).

Although rare, *HER2* gene fusions have also been reported. Within The Cancer Genome Atlas (TCGA) PanCancer Atlas study, 1.7% of esophagus, 1.4% of breast, and 1.4% of cervical samples contained a *HER2* fusion (23). Three novel fusions were reported in gastric cancer (36). Two (*NOS-HER2* and *ZNF207-HER2*) were further characterized and found to induce autophosphorylation and cellular transformation (36). Thus, fusions are another potential therapeutic target.

Targeting HER2 Mutations

Neratinib

Bose and colleagues reported that many activating *HER2* mutations are resistant to *HER2* inhibitor lapatinib, but sensitive to irreversible inhibitor neratinib (19). Ma and colleagues evaluated efficacy of neratinib in *HER2*-mutant, nonamplified breast cancer (37). The clinical benefit ratio (CBR) was 31%. Upon longitudinal circulating free DNA analysis, the mutant *HER2* allele decreased with treatment and increased with progression in 9 of 11 patients.

The efficacy of neratinib in *HER2*-mutant tumors was also tested in the SUMMIT trial (38). Neratinib demonstrated the greatest activity in patients with breast cancer (ORR at 8 weeks 32%), all with *HER2*-nonamplified tumors. Responses were observed in both ER⁺ and ER⁻ tumors and in patients with mutations in the ECD, KD as well as KD insertions. In lung cancer, only one OR (in a patient with L755S mutation) was observed among the 26 patients enrolled, but progression-free survival was 5.5 months. Responses were also observed in biliary

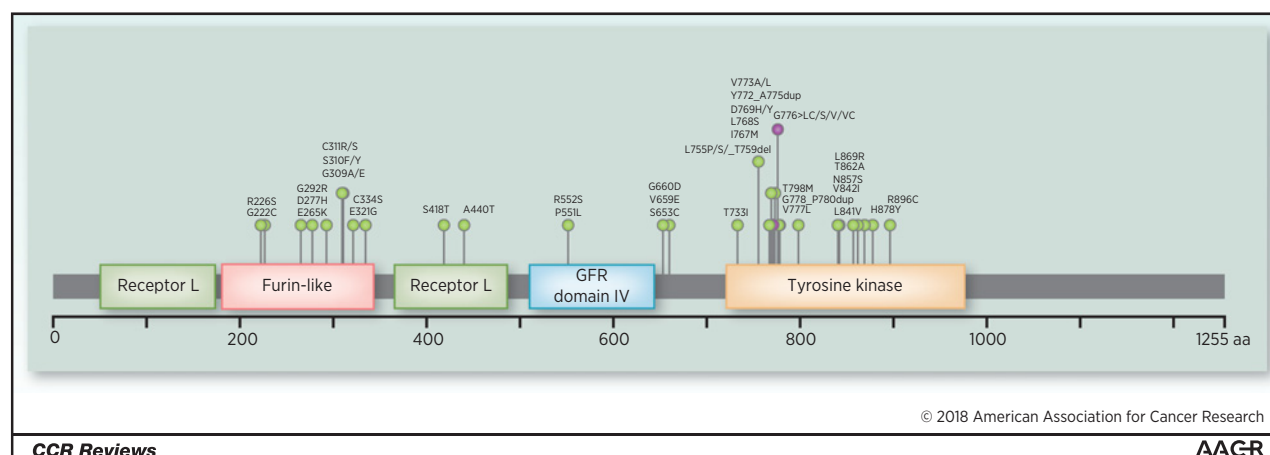


Figure 2.

Activating *HER2* mutations. Mutations defined as "activating" based on published data demonstrating that the alteration increases the activity, expression, or stability of the encoded protein. In addition, alterations shown to enhance downstream signaling or increase tumorigenic properties when expressed are shown (56). Mutation Mapper from cBioPortal was used to visualize mutations (57, 58).

tumors and cervical cancer, while none were seen among the 16 patients with bladder cancer and 12 patients with colorectal cancer enrolled, suggesting that histology may impact neratinib's efficacy.

Pertuzumab/trastuzumab

Although preclinical data have suggested that mAbs may not be as effective as irreversible kinase inhibitors for *HER2* mutations, there have been anecdotal reports of responses with trastuzumab-based therapy. In the MYPATHWAY trial, among the initial 36 patients who received trastuzumab plus pertuzumab for tumors with *HER2* mutations without amplification/overexpression; 4 patients (11%) had ORs: 3 patients of 14 with NSCLC and one with biliary cancer (13). The efficacy of pertuzumab/trastuzumab for *HER2* mutations is also being explored in the ASCO TAPUR trial.

T-DM1

Li and colleagues tested T-DM1's efficacy in *HER2*-mutant lung tumors (39). In patients with *HER2*-nonamplified lung cancer with *HER2* mutations, the ORR was 33%. Scientifically this result is somewhat surprising, but the efficacy, at least in part, is being attributed to *HER2* internalization with antibody–drug conjugate treatment.

Mechanisms of Intrinsic and Acquired Resistance to *HER2*-Targeted Therapy and Rational Combinations to Overcome Resistance

There is increasing understanding of mechanisms of intrinsic and acquired resistance to *HER2*-targeted therapy (Table 1). Much of these data have emerged from breast cancer studies, so more study is needed to determine whether they are extrapolatable to other tumor types and whether additional mechanisms of resistance are identified in other histologies. Intratumoral *HER2* heterogeneity (some cells having *HER2* overexpression/amplification and others not) as well as genomic evolution with loss of

HER2 amplification are concerns that support the use of pretreatment biopsies for confirmation of *HER2* status, and exploration of the role of liquid biopsies and functional imaging with *HER2*-PET for assessment of *HER2* status.

In spite of years of study of mechanisms of sensitivity and resistance, *HER2* aberrations remain the primary biomarker for patient selection. *HER2* copy number, including *HER2* copy number in cfDNA and *HER2*-enriched subtype by RNA-seq analysis (40) are associated with sensitivity to *HER2*-targeted therapy. However, there is evidence for intrinsic mechanisms of resistance including PI3K pathway alterations (mainly PTEN loss and *PIK3CA* mutations), and emerging data that activating receptor tyrosine kinases/RAS/RAF may limit the efficacy of anti-*HER2* therapy (38), making these survival pathways appealing targets for combination therapy. Small-molecule inhibitors of *HER2* have been associated with the emergence of gatekeeper *HER2* mutations (L755S, T862A, T798M for lapatinib; *HER2* T798I for neratinib) that may still be sensitive to alternate small-molecule inhibitors (e.g., afatinib; refs. 41, 42).

Combination therapy trials have focused on several approaches: (i) combinations of *HER2*-targeted therapies (doublets and triplets), (ii) combination with chemotherapy, (iii) overcoming ER and *HER2* cross-talk via combination with endocrine therapy, (iv) combination with CDK4/6 inhibitors, and (v) targeting intrinsic and acquired resistance mechanisms with cell signaling inhibitors such as PI3K/Akt/mTOR pathway inhibitors. A summary of targeted therapies combined with *HER2*-targeted agents are listed in Supplementary Table S3.

Novel *HER2*-Targeted Therapies

Antibody–drug conjugates

DS-8201a is a novel anti-*HER2* antibody–drug conjugate, with a derivative of DX-8951 (DXd), a topoisomerase I inhibitor (43) with a drug to antibody ratio (DAR) of approximately 8. Preclinically, DS-8201a was effective in a T-DM1-insensitive *HER2*-positive PDX model. DS-8201a, but not T-DM1, also demonstrated efficacy against *HER2*-low breast cancer models. In 2017, FDA granted DS-8201 breakthrough designation for

Table 1. Mechanisms of intrinsic and acquired resistance for HER2-targeted therapy

HER2-targeted therapy	Mechanisms of resistance	Potential strategy to overcome resistance
Trastuzumab/pertuzumab/T-DM1 (59–61)	Intrinsic/acquired: coexpression of EGFR or HER3	Dual inhibition of EGFR and HER2
	Intrinsic/acquired: overexpression of IGF1R	Treatment with mTOR/IGF1R inhibitors, inhibition of HER2 kinase activity
	Intrinsic/acquired: overexpression of MET or HGF	MET inhibitors, MET ADC
	Intrinsic/acquired: overexpression of EphA2	Treatment with an EphA2-neutralizing antibody
	Intrinsic/acquired: PI3KA activation	Inhibition of PI3K or mTOR pathways
	Intrinsic/acquired: PTEN loss	Inhibition of PI3K or mTOR pathways
	Intrinsic: high levels of catecholamines in TM	Use of β -blockers
	Intrinsic: expression of ER	Blockade of ER signaling
	Intrinsic: high p95 HER2 expression or high p95/HER2 ratio	Inhibition of HER2 kinase activity
	Intrinsic: D16 HER2 expression	Inhibition of Src kinase
	Intrinsic: <i>HER2</i> exon 20 insertion	HER2 exon 20 inhibitors
	Intrinsic: Higher expression of PDL1	Combination with a checkpoint inhibitor
	Intrinsic: Fc γ R111 deficiency/polymorphism (deficiency in NK cells and macrophages capable of binding to Fc region of trastuzumab)	Inhibition of HER2 kinase activity
	Acquired: overexpression of AXL	AXL inhibitor or AXL ADC
	Acquired: constitutive activation of Src kinase	Src kinase inhibitors
Acquired: upregulation of survivin and Mcl-1	Broad-spectrum kinase inhibitors or Mcl-1 inhibitors	
Acquired: upregulation of cyclin E	Treatment with CDK2 inhibitors	
Acquired: downregulation of p27 ^{KIP1}	Treatment with CDK2 inhibitors	
Acquired: downregulation of HER2 expression	Treatment HER2-targeted therapies with efficacy in lower HER2-expressing tumors	
T-DM1 (61)	Intrinsic: poor internalization of HER2–T-DM1 complexes/ increased recycling/defective trafficking to lysosomes	
Lapatinib/neratinib (41, 42, 59, 60)	Acquired: upregulation of ABCC1 (MRP1)	Modification of linker
	Acquired: increase in HER3 transcription and phosphorylation	Inhibition of HER3 and HER3 ADC
	Acquired: overexpression of AXL	Combination with an AXL inhibitor or AXL ADC
	Acquired: increased activation of Src family kinase activity	Combination with Src inhibitors
	Acquired: increased signaling through PI3K and AKT/ mTOR pathways	Inhibition of PI3K or mTOR pathways
	Intrinsic/acquired: low/downregulation of BIM levels	
	Acquired: upregulation of ER α , leading to FoxO3a-mediated transcription of survivin	Treatment with ER antagonists
Acquired: HER2 gatekeeper mutations (L755S, T862A, T798M for lapatinib; HER2 T798I for neratinib)	Treatment with afatinib	

Abbreviations: ADC, antibody–drug conjugate; NK, natural killer; TM, tumor microenvironment.

HER2-positive breast cancer. DS-8201a demonstrated remarkable activity in heavily pretreated patients, with a confirmed ORR of 54.5% (54/99) and disease control rate (DCR) of 93.9% (93/99) and with preliminary signal of activity in HER2-low tumors: ORR 50% (17/34) and DCR 85.3% (29/34; ref. 44). However, cases of fatal pneumonitis have also been reported (44), highlighting that greater efficacy in HER2-low cells may have a safety trade-off. Studies are ongoing to better define the efficacy as well as safety of DS-8201a.

Additional HER2 antibody–drug conjugates are in development, varying in antibody and linker payload. The use of antibodies with greater affinity for HER2 or antibody–drug conjugates with higher DAR may overcome resistance due to decreased HER2 expression. Use of toxins with greater bystander effect may help overcome resistance due to tumor heterogeneity and may prove to be effective even in HER2-low cancers, but therapeutic window will need to be assessed.

Bispecific antibodies

ZW25 is biparatropic antibody that simultaneously binds two HER2 epitopes, extracellular domain 4 (the trastuzumab-binding domain) and extracellular domain 2 (the pertuzumab-binding domain). Preclinically, its unique binding facilitates increased

tumor cell binding, ZW25-HER2 clustering, and enhanced internalization (including in the setting of lower HER2 concentrations). In a phase I trial, ZW25 led to ORRs in heavily pretreated patients with *HER2*-amplified/overexpressing breast cancer (33% ORR), gastroesophageal cancer (44% ORR), as well as other *HER2*-amplified/overexpressing tumor types (33% ORR), including colorectal cancer and gallbladder cancer (45). MCLA-128 targets both HER2 and HER3, enhancing antibody-dependent cell-mediated cytotoxicity, with a CBR of 70% in heavily pretreated patients (46). Other bispecific antibodies of special interest include antibodies that also directly engage immune mediators discussed below.

Small-molecule inhibitors

Several new HER2 tyrosine kinase inhibitors are in development, including tucatinib, poziotinib, and pyrotinib. Tyrosine kinase inhibitors are being explored in multiple tumor types and for tumors with *HER2*-amplified/overexpression as well as activating mutations. Tucatinib was granted fast track designation by the FDA in 2016 for treatment of HER2-positive metastatic breast cancer and orphan drug status in 2017 for treatment of HER2-positive central nervous system (CNS) metastasis. Tucatinib as a single agent, as well as in

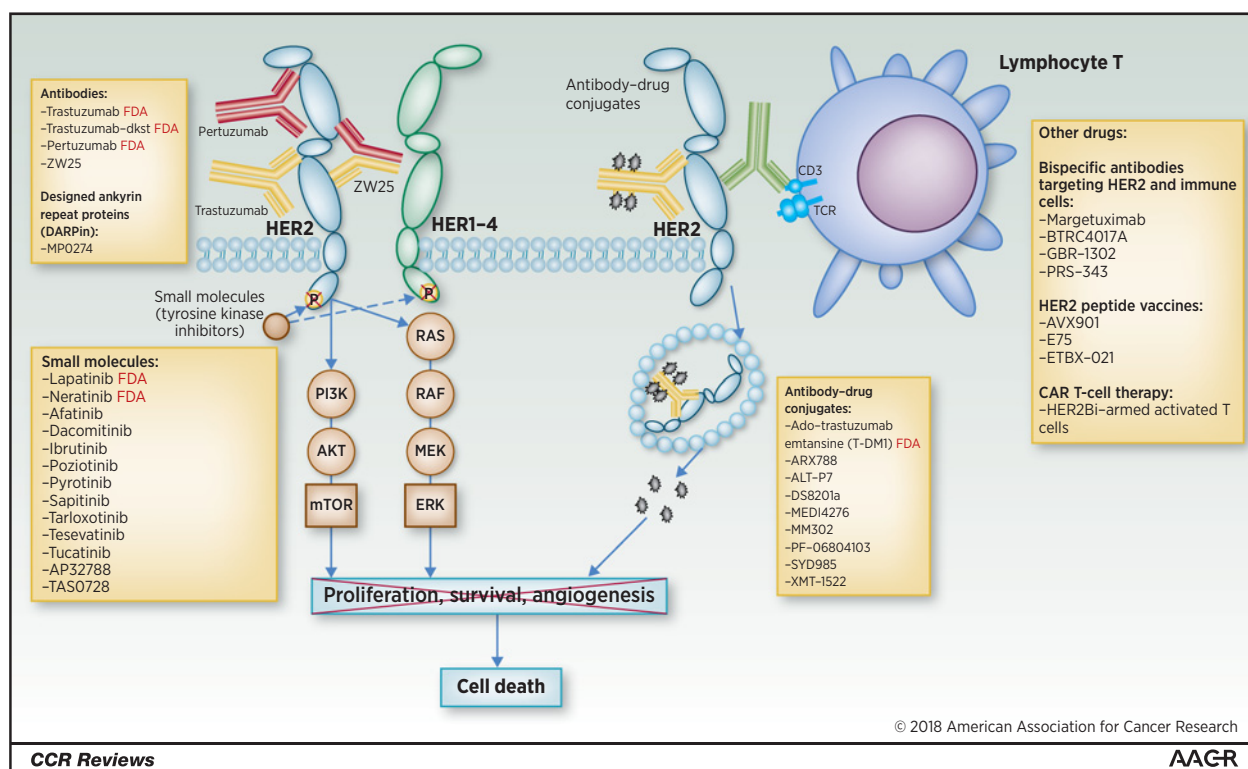


Figure 3.

Approved and emerging HER2-targeted therapies in clinical development. HER2-targeted therapies approved by the FDA for HER2-positive cancer or currently in clinical trials (from www.clinicaltrials.gov, last accessed July 11, 2018). Therapies followed by FDA in red font indicate that the drug is FDA approved. CAR, chimeric antigen receptor.

combination with standard-of-care therapies, demonstrated significant growth inhibition in HER2-positive xenografts, including breast cancer models (47, 48). Tucatinib also significantly enhanced survival in intracranial tumor xenograft models (49). In the phase I trial, tucatinib monotherapy had activity in heavily pretreated patients: ORR was 14% and the CBR (PR + stable disease \geq 24 weeks) was 27% (50). Efficacy of tucatinib has also been tested in combination studies, for example, in combination with T-DM1, 68% patients treated with the MTD had ORR of 47% and CBR was 58% (51). Furthermore, it has been explored in combination with capecitabine (ORR 83%), trastuzumab (ORR 40%), and with trastuzumab and capecitabine (ORR 61%; ref. 52). In both phase 1b combination studies (NCT01983501 and NCT02025192), clinical benefit has been noted in patients with CNS disease at similar rates compared with those without CNS disease and patients were allowed to continue on study with isolated CNS progression following locally directed therapy which allowed patients to remain on study longer (52). Validation of overall efficacy and further evaluation of the latter approach will be determined in the ongoing pivotal double-blinded randomized registration study, HER2CLIMB (NCT02614794), which is enrolling patients with and without CNS metastases. Tucatinib is also being evaluated in combination with trastuzumab in HER2-positive, RAS wild-type colorectal cancer.

Pozitotinib is a small-molecule irreversible inhibitor of EGFR, HER2, and HER4. Pozitotinib has shown clinical activity in HER2-

positive breast cancer and is being developed in combination therapy. Robichaux and colleagues reported that pozitotinib is a potent inhibitor of exon 20 mutations in EGFR and HER2, preclinically. This is notable as exon 20 mutations have been intrinsically resistant to approved targeted therapies (53). Preliminary data from a phase II trial demonstrated ORR of 64% in patients with lung cancer with exon 20 EGFR mutations. Clinical activity was also reported in a patient with lung cancer with exon 20 HER2 mutation, and this efficacy for HER2 mutations is being better defined in ongoing trials. Several other small-molecule inhibitors are in development, some with proposed greater selectivity for HER2 or for selected HER2 mutations (e.g., exon 20; Fig. 3).

DARPins

MP0274 is a proprietary designed ankyrin repeat protein (DARPin)-based agent targeting HER2. MP0274 binds to two distinct nonoverlapping epitopes on HER2, inhibiting the activity of HER2 and promoting internalization. MP0274 is now in phase I clinical trials.

HER2-targeted immunotherapy

PANACEA, the first phase Ib/II trial evaluating the antitumor efficacy of immunotherapy in combination with HER2-targeted therapy (pembrolizumab and trastuzumab), reported an ORR of 15% and DCR of 25% in PDL1-positive patients, and no responses in PDL1-negative patients (54). Although there was

modest benefit in the PDL1-positive cohort, the disease control in those who responded was durable for 1 year without chemotherapy, which is notable. Heavily pretreated metastatic breast tumors are thought to be poorly immunogenic and there is much interest in assessing the association between immune function and benefit from trastuzumab in earlier settings. Many other HER2-immunotherapy combinations are ongoing (Supplementary Table S3). There is also great interest in combinations of HER2 antibody-drug conjugates and immunotherapy, as antibody-drug conjugates elicit immune responses and have enhanced efficacy in combination with checkpoint inhibitors preclinically (44). In addition, bispecifics are being explored targeting HER2 and immune components. For example, GBR1302 is proposed to direct HER2 and CD3-redirecting cytotoxic T cells onto HER2⁺ cells, while PRS-343 increases tumor lymphocyte infiltration via bispecific targeting of 41BB (CD137) and HER2. In addition, HER2 vaccines are in clinical trials and CAR-T approaches are still being explored (55).

Conclusions

Outside of breast and gastric cancer, in what diseases, and in what clinical setting HER2 testing for amplification/overexpression should be initiated remains controversial. However, emerging data suggest efficacy of HER2-targeted therapy for HER2-amplified/overexpressing tumors across a variety of tumor types. Thus, genomic testing or specifically HER2 testing by IHC and/or ISH should be considered for advanced/metastatic disease for tumor types where HER2 is known to be amplified (Fig. 1A). HER2 also represents an important opportunity for seeking histology-agnostic approvals. It should be noted that most NGS platforms call HER2 amplification at 6 to 7 copies. Additional studies are needed to validate the efficacy of HER2-targeted therapy across tumor types and to determine whether efficacy can be extended to patients with lower levels of amplification detectable by ISH and patients with overexpression of RNA or protein in the absence of amplification.

With increasing number of HER2-targeted therapies, we will likely be able to truly personalize therapy selection, offering targeted therapies with greatest expected efficacy based on mutation type and expression status, as well as expected adverse events. Newer therapies may allow us to offer HER2-targeted therapies to patients with lower HER2 expression, leading to a redefinition of "HER2-negative." For HER2 mutations, evolving data will likely allow us to select optimal therapies individualizing therapy based on variant type.

For advanced disease, more efficacious therapy could improve outcomes. In addition, transitioning new agents to early-stage disease may allow us to offer targeted therapies alone, sparing

patients the toxicity of chemotherapy. Finally, increasing the efficacy of neoadjuvant therapy in breast cancer may increase breast-conserving surgery and spare patients axillary lymph nodal dissections, and further strengthen the evolving paradigm of avoiding surgery altogether in exceptional responders. In conclusion, greater awareness of the HER2 status of patients can enhance incorporation of HER2-targeted therapy into the multidisciplinary care across tumor types.

Disclosure of Potential Conflicts of Interest

F. Meric-Bernstam reports receiving commercial research grants from Novartis, AstraZeneca, Taiho Pharmaceuticals, Genentech, Calithera Biosciences, Debiopharm Group, Bayer, Aileron Therapeutics, PUMA Biotechnology, CytomX Therapeutics, Zymeworks, Curis, Pfizer, eFFECTOR Therapeutics, and AbbVie, speakers bureau honoraria from Sumitomo Group and Dialecta, and is a consultant/advisory board member for Aduro, Spectrum, OrigiMed, Debiopharm Group, Inflection Biosciences, Xencor, Genentech, Samsung Bioepis, Pieris Pharmaceuticals, and Darwin Health. K. Balaji is Medical Science Liaison for Lexicon Pharmaceuticals. R.K. Murthy reports receiving commercial research grants from Oncocyte, EMD Serono, Daiichi Sankyo, Genentech, and Pfizer (all funding paid directly to Dr. Murthy's institution), and is a consultant/advisory board member for PUMA and Daiichi Sankyo. J. Rodon reports receiving commercial research grants from Bayer and Novartis, and is a consultant/advisory board member for Novartis, Eli Lilly, Orion, Servier, Peptomyc, Merck Sharp & Dohme, and Kelun Pharmaceutical/Klus Pharma. S.A. Piha-Paul reports receiving commercial research grants from Taiho, Novartis, Pieris, Puma, GlaxoSmithKline, Merck Sharp & Dohme, Blue Link, Incyte, Bristol-Myers Squibb, Curis, Medivation, Pfizer, Helix, Bayer, Xuanzhu, AbbVie, Principia, FivePrime, MedImmune, Seattle Genetics, Tesaro, and Genmab. No potential conflicts of interest were disclosed by the other authors.

Acknowledgments

This work was supported by the Cancer Prevention & Research Institute of Texas (grant number RP150535; to F. Meric-Bernstam, A.M. Johnson, and K. Balaji), the Sheikh Khalifa Al Nahyan Ben Zayed Institute for Personalized Cancer Therapy (to F. Meric-Bernstam), the Nellie B. Connally Breast Cancer Research Endowment (to F. Meric-Bernstam), and The University of Texas MD Anderson Cancer Center Support Grant (NIH/NCI grant number P30 CA016672; to F. Meric-Bernstam). The authors would also like to acknowledge the AACR and its financial and material support in the development of the AACR Project GENIE registry, as well as members of the consortium for their commitment to data sharing. Interpretations are the responsibility of study authors.

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Received July 17, 2018; revised September 10, 2018; accepted November 12, 2018; published first November 15, 2018.

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