Errata

Ouseph MM, Huang Y, Banerjee M, et al. Autophagy is induced upon platelet activation and is essential for hemostasis and thrombosis. *Blood*. 2015;126(10):1224-1233.

On page 1224 in the 3 September 2015 issue, the 11th author's name is misspelled as Massaki Komatsu. The correct spelling is Masaaki Komatsu. The error has been corrected in the online version, which now differs from the print version.

DOI 10.1182/blood-2015-09-668111

Walz S, Stickel JS, Kowalewski DJ, et al. The antigenic landscape of multiple myeloma: mass spectrometry (re)defines targets for T-cell–based immunotherapy. *Blood*. 2015;126(10): 1203-1213.

On page 1210 of the 3 September 2015 issue, there are two errors in Figure 5. In Figure 5A, row 5, column 5, "25.0%" should read "16.7%." In Figure 5B, the wrong set of ELISPOT wells is shown. The corrected Figure 5 is shown below. The errors have been corrected in the online version, which now differs from the print version.

In addition, in supplemental Figure 3B, the PDIA4$_{136\text{-}153}$ well was erroneously depicted. A corrected supplemental Figure 3 has replaced the incorrect figure in the supplemental Data file on the *Blood* Web site.

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A

Peptide	Sequence	Source protein	HLA	CD8+ T-cell response in MM	Allotype matched presentation in primary MM samples [%]
P_1	KAMEAASSL	MMSET	A*02	2/16 (12.5%)	20.0
P_2	SLLEQGLVEA	MMSET	A*02	1/8 (12.5%)	20.0
P_3	FVFPGELLL	SLC1A5	A*02	2/14 (14.3%)	20.0
P_4	FLFQLLQLL	SEMA4A	A*02	1/14 (7.1%)	20.0
P_5	LPPPPHVPL	SLX1A	B*07	1/6 (16.7%)	25.0
P_6	LAHVGPRL	SLX1A	A*02	2/14 (14.3%)	20.0

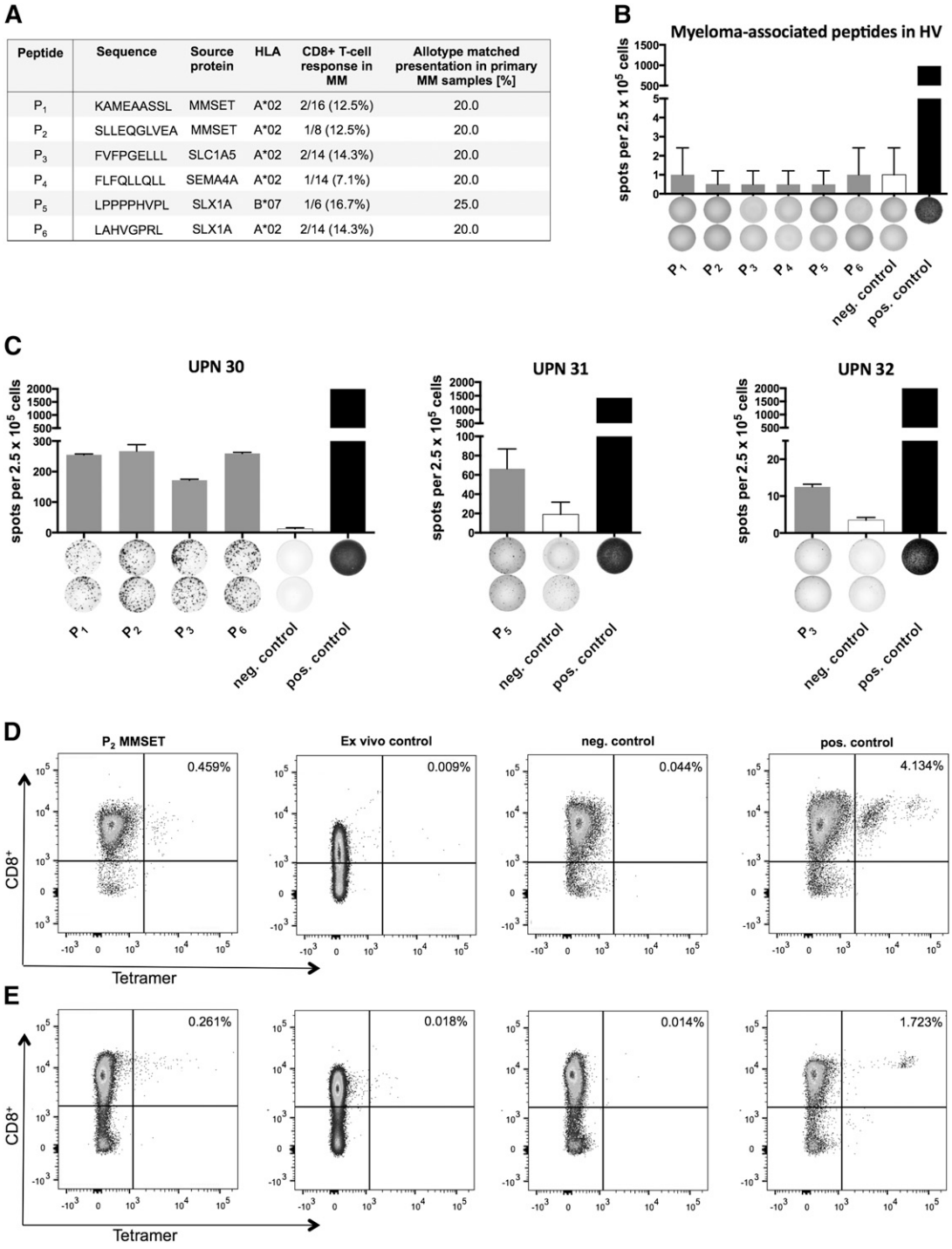

Figure 5. Functional characterization of myeloma-associated antigens. (A) Myeloma-associated T-cell epitopes with their corresponding HLA restrictions and frequencies of immune recognition by myeloma patient-derived T cells in IFNγ-ELISPOT assays. (B) Example of myeloma-associated T-cell epitopes evaluated in an IFNγ-ELISPOT using HV PBMCs. An EBV epitope mix containing the frequently recognized peptides BRLF109-117 YVLDHLIVV (A*02) and EBNA3247-255 RPPIFIRRL (B*07 served as positive control). Benign-tissue–derived peptides KLFEKVKEV (HLA-A*02) and KPSEKIQVL (B*07) served as negative control. (C) Examples of myeloma-associated T-cell epitopes evaluated in IFNγ-ELISPOTs using MM patient PBMCs (n = 3). Results are shown only for immunoreactive peptides. An EBV epitope mix containing 5 frequently recognized peptides (BRLF109-117 YVLDHLIVV [A*02], EBNA3471-479 RLRAEAQVK [A*03], EBNA3247-255 RPPIFIRRL [B*07], BZLF1190-197 RAKFKQLL [B*08], EBNA6162-171 AEGGVGWRHW [B*44]) was used as positive control. Benign-tissue–derived peptides KLFEKVKEV (HLA-A*02) and KPSEKIQVL (B*07) served as negative control. (D-E) Tetramer staining of $CD8^+$ T cells after 3 cycles of aAPC-based in vitro primings using T cells derived from (D) a healthy individual and (E) a myeloma patient: Leftmost panels, P_2-tetramer staining of $CD8^+$ T cells primed with P_2-aAPCs (SLLEQGLVEA, A*02); left middle panels, ex vivo P_2-tetramer staining of $CD8^+$ T cells; right middle panels, control staining with A*02-tetramer containing a nonrelevant A*02-restricted control peptide (KAMEAASSL, A*02) on $CD8^+$ T cells derived from the same population as T cells depicted in the left panels. Rightmost panels, Positive control: tetramer staining of $CD8^+$ T cells primed with CMV-aAPCs (NLVPMVATV, A*02). EBV, Epstein-Barr virus; neg., negative; pos., positive; UPN, uniform patient number.