

These PKCs are highly important to the phosphorylation of the phagocyte NADPH oxidase components, p47^{phox} and p40^{phox}. Phosphorylation of p47^{phox} is of particular interest because this posttranslational modification changes the conformation of p47^{phox}, leading to its membrane translocation and binding to p22^{phox}, a subunit of the core NADPH oxidase termed flavocytochrome b₅₅₈.⁵ In STIM1-deficient neutrophils, PKC-dependent phosphorylation of these cytosolic factors of the NADPH oxidase is markedly reduced.

The work by Zhang et al provides molecular details for STIM1-dependent Ca²⁺ influx leading to phagocyte NADPH oxidase activation and also shows several interesting aspects of Ca²⁺ signaling in relation to other neutrophil functions. It is particularly interesting that neutrophil chemotaxis, known to rely on Ca²⁺ influx,⁶ is not affected by STIM1 deficiency, suggesting that a sustained rise in cytosolic Ca²⁺ may not be required. It is possible that other forms of Ca²⁺ signaling, including transient Ca²⁺ mobilization from intracellular stores,⁷ or localized calcium flickers as reported in other types of cells,⁸ may be sufficient for neutrophil migration. Chemotaxis mediated by different receptors may also have different requirement for Ca²⁺ influx.⁹

Production of oxygen radicals is a major bactericidal function of phagocytes. However, it is a double-edged sword because of its tissue-damaging property. Zhang et al show that the *Stim1*^{-/-} chimeras are resistant to ischemia/reperfusion injury to the liver, suggesting that neutrophil production of oxygen radicals might contribute to the pathological changes in wild-type subjects. Future work using granulocyte-specific deletion of *Stim1* will be helpful to ascertain the respective contribution of different blood cells to the observed changes. These studies create new opportunities for therapeutic intervention that targets STIM1.

Conflict-of-interest disclosure: The author declares no competing financial interests. ■

REFERENCES

- Zhang H, Clemens RA, Liu F, et al. STIM1 calcium sensor is required for activation of the phagocyte oxidase during inflammation and host defense. *Blood*. 2014; 123(14):2238-2249.
- Bootman MD. Calcium signaling. *Cold Spring Harb Perspect Biol*. 2012;4(7):a011171.
- Baba Y, Kurosaki T. Physiological function and molecular basis of STIM1-mediated calcium entry in immune cells. *Immunol Rev*. 2009;231(1):174-188.

- Brécard S, Plançon S, Melchior C, Tschirhart EJ. STIM1 but not STIM2 is an essential regulator of Ca²⁺ influx-mediated NADPH oxidase activity in neutrophil-like HL-60 cells. *Biochem Pharmacol*. 2009; 78(5):504-513.
- El-Benna J, Dang PM, Gougerot-Pocidallo MA, Marie JC, Braut-Boucher F. p47^{phox}, the phagocyte NADPH oxidase/NOX2 organizer: structure, phosphorylation and implication in diseases. *Exp Mol Med*. 2009;41(4):217-225.
- Boucek MM, Snyderman R. Calcium influx requirement for human neutrophil chemotaxis: inhibition by lanthanum chloride. *Science*. 1976;193(4256):905-907.
- Fay FS, Gilbert SH, Brundage RA. Calcium signalling during chemotaxis. *Ciba Found Symp*. 1995;188:121-135, discussion 136-140.
- Wei C, Wang X, Chen M, Ouyang K, Song LS, Cheng H. Calcium flickers steer cell migration. *Nature*. 2009;457(7231):901-905.
- Partida-Sánchez S, Iribarren P, Moreno-García ME, et al. Chemotaxis and calcium responses of phagocytes to formyl peptide receptor ligands is differentially regulated by cyclic ADP ribose. *J Immunol*. 2004;172(3):1896-1906.

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● ● ● **LYMPHOID NEOPLASIA**

Comment on Rossi et al, page 2139

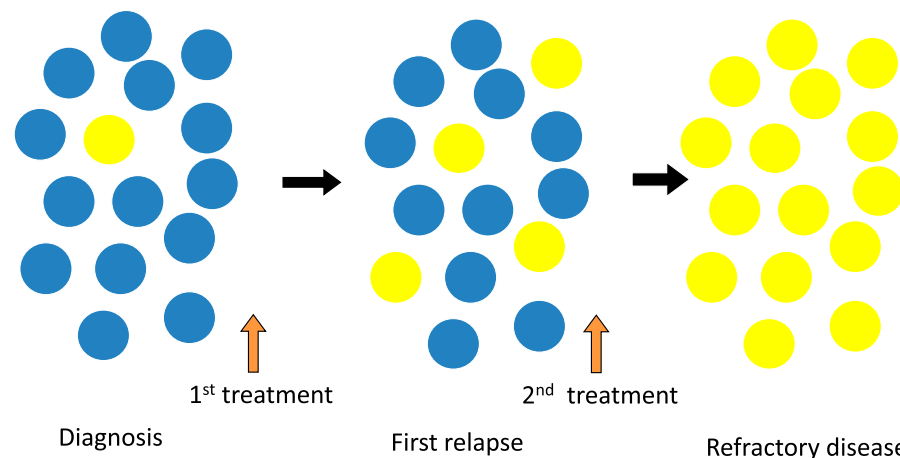
CLLonal selection: survival of the fittest?

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In this issue of *Blood*, Rossi et al provide further evidence for clonal evolution in chronic lymphocytic leukemia (CLL) and demonstrate the clinical importance of small *TP53*-deleted subclones detected at diagnosis in determining the natural history of the disease.¹

The basic principle of Darwinian evolutionary theory is the natural selection of the fittest variants. Fitness is defined by the ability to survive and reproduce and the “fittest” as those best adapted to achieve this. This concept of subclonal selection of the “fittest” variants was first applied to cancer by Nowell in 1976² and has subsequently been supported by modern genomics.³ Knowledge of the clonal diversity and clonal selection operating in any specific cancer is critical to the understanding of disease progression, response

to treatment, and development of resistance. The clonal architecture of any cancer is in a constant state of evolutionary change, which can take place over prolonged periods. Clinically important mutations may be present at an early stage of the disease but only become evident over time through selective pressure. The clinical importance of chromosomal abnormalities in CLL has been recognized since the late 1990s.⁴ Since then, more refined techniques such as fluorescence in situ hybridization (FISH) and Sanger sequencing



Clonal selection and expansion of *TP53*-mutated subclones during the clinical course of CLL. Yellow circles indicate the *TP53*-mutated CLL cell which expands through the disease course from below the level of conventional detection methods at diagnosis to become the dominant population in refractory disease. Blue circles indicate the non-*TP53*-mutated CLL cell which is the dominant clone at diagnosis but is subsequently replaced by the *TP53*-mutated subclone.

have confirmed that the presence of certain genetic alterations is associated with overall prognosis and response to treatment. CLL is characterized by relatively few recurrent somatic mutations, of which those in the *TP53* gene are the strongest predictors of chemoresistance and poor survival.⁵

In this study, Rossi et al¹ have applied highly sensitive ultra-deep next-generation sequencing to examine a large cohort of patients (309) with newly diagnosed CLL for the presence of very small *TP53*-mutated subclones (sensitivity down to 0.3% allele frequency), which would not have been detected by Sanger sequencing (which detects >20% frequency).⁶ The 5.8% of patients with such subclones had the same adverse survival as those 9% in whom *TP53* mutations were detected by conventional methods, and accounted for a third of all cases with *TP53* abnormalities.

The current recommendation is that all patients should be tested for abnormalities of *TP53* prior to initiating any line of treatment, in order to select *TP53*-independent therapy when appropriate. The prevailing dogma is that the size of the *TP53*-deleted clone is important and that below certain thresholds (variably reported as 10% or 20% using FISH), response to treatment is unaffected. Rossi et al overturn the view that small clones are clinically unimportant by showing that even very small subclones (<1%; present below the threshold of detection using current standard methods but with no apparent cutoff in the size of the clone) have an adverse impact on patient survival. They argue that the effect on outcome of the presence of these subclones is a yes/no determinant independent of clonal size.

Importantly, sequential samples showed that these subclones expanded over time, particularly under the selective pressure of chemotherapy, leading ultimately to chemorefractory disease (see figure). In the 2 patients who did not receive treatment, the clonal size remained unchanged. This would be consistent with other studies showing increasing frequency of *TP53* abnormalities in patients with disease progression and refractoriness.⁷

If multiple subclones coexist, what drives any to become dominant? There are many reasons, including limited potential for expansion due to “competition” from other clones as well as the independent effect of the surrounding microenvironment. If these

subclones are vying for space and resources, the reduction of some clones may unbalance the status quo. It is therefore unsurprising that failure of chemotherapy to completely eradicate CLL cells can result in expansion of minor, more resistant, and more dangerous subclones. “Selection” can thus be introduced artificially by the use of chemotherapeutic agents. Certain subclones are likely to gain a competitive advantage due to their “fitness” in relation to these selection pressures. It is therefore important to identify low-level molecular lesions that are known to predict for chemoresistance so that treatment can be tailored appropriately. In *TP53*-mutated CLL, this may involve use of novel targeted therapies (eg, Ibrutinib, ABT199)⁸ which have been shown to have promising activity in this subset.

What other strategies might be considered to improve therapeutic efficacy and prevent emergence of resistance? Cytotoxic drugs are likely to select for resistant cells by clearing the ground of more sensitive ones. On the other hand, cytostatic drugs (some small-molecule inhibitors) may cause cells to remain in the tissue space but without either expanding themselves or allowing expansion of other subclones. In addition, early intervention with effective treatment before clonal expansion may be a more effective way to deal with these more clinically adverse subclones. It may also be the case that carefully designed concurrent or sequential combinations of therapies may overcome some of the issues related to clonal diversity. It is important to note, however, that not all chemoresistant CLL is characterized by *TP53* mutation and it will be crucial to understand the biology of any other clinically important subclones which may be present in order to prevent their dominance. The

underlying principles are likely to be the same but the therapeutic strategies may be different.

Clearly, CLL is not a static disease, but has a clonal architecture that changes over time and is influenced by selection pressures, including treatment. Certain clinically adverse genomic changes appear to be present in the CLL cells from a very early stage of disease. Understanding how this population becomes dominant is crucial for the development of new therapeutic strategies, which will be effective by rendering them the least fit for survival.

Conflict-of-interest disclosure: The author declares no competing financial interests. ■

REFERENCES

1. Rossi D, Khiabanian H, Spina V, et al. Clinical impact of small *TP53* mutated subclones in chronic lymphocytic leukemia. *Blood*. 2014;123(14):2139-2147.
2. Nowell PC. The clonal evolution of tumor cell populations. *Science*. 1976;194(4260):23-28.
3. Campbell PJ, Pleasance ED, Stephens PJ, et al. Subclonal phylogenetic structures in cancer revealed by ultra-deep sequencing. *Proc Natl Acad Sci USA*. 2008; 105(35):13081-13086.
4. Döhner H, Stilgenbauer S, Benner A, et al. Genomic aberrations and survival in chronic lymphocytic leukemia. *N Engl J Med*. 2000;343(26):1910-1916.
5. Gonzalez D, Martinez P, Wade R, et al. Mutational status of the *TP53* gene as a predictor of response and survival in patients with chronic lymphocytic leukemia: results from the LRF CLL4 trial. *J Clin Oncol*. 2011; 29(16):2223-2229.
6. Pospisilova S, Gonzalez D, Malcikova J, et al; European Research Initiative on CLL (ERIC). ERIC recommendations on *TP53* mutation analysis in chronic lymphocytic leukemia. *Leukemia*. 2012;26(7):1458-1461.
7. Zenz T, Häbe S, Denzel T, Winkler D, Döhner H, Stilgenbauer S. How little is too much? p53 inactivation: from laboratory cutoff to biological basis of chemotherapy resistance. *Leukemia*. 2008;22(12):2257-2258.
8. Byrd JC, Furman RR, Coutre SE, et al. Targeting BTK with ibrutinib in relapsed chronic lymphocytic leukemia. *N Engl J Med*. 2013;369(1):32-42.

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● ● ● LYMPHOID NEOPLASIA

Comment on Dubois et al, page 2199

An E3 ubiquitin ligase-independent role of LUBAC

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In this issue of *Blood*, Dubois et al show a catalytic-independent role of the linear ubiquitin chain assembly complex (LUBAC) in lymphocyte activation and B-cell malignancy.¹ These data add a new layer of versatility to the recently established role of LUBAC in nuclear factor- κ B (NF- κ B) signaling.