

## T-Regulatory Cells: Key Players in Tumor Immune Escape and Angiogenesis

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### Abstract

T-regulatory cells (Tregs) are found infiltrating tumors in a vast array of tumor types, and tumor-infiltrating Tregs are often associated with a poor clinical outcome. Tregs are potent immunosuppressive cells of the immune system that promote progression of cancer through their ability to limit antitumor immunity and promote angiogenesis. Here, we discuss the ways in which Tregs suppress the antitumor immune response and elaborate on our recent discovery that Tregs make significant direct contributions to tumor angiogenesis. Further, we highlight several current therapies aimed at eliminating Tregs in cancer patients. Given the multifaceted role of Tregs in cancer, a greater understanding of their functions will ultimately strengthen future therapies. *Cancer Res*; 72(9); 2162–71. ©2012 AACR.

### Introduction

The tumor microenvironment is characterized by a multitude of mechanisms that support angiogenesis and immune suppression (1). Many of the immune suppressive regulatory circuits that operate in tumors are part of the physiologic regulatory mechanisms used by the immune system to maintain homeostasis to prevent autoimmunity and temper inflammation after infection or injury (1). T-regulatory cells (Tregs) are considered to be pivotal mediators of peripheral tolerance and immune suppression. Tregs are comprised of natural Tregs, which are thymically derived cells of FoxP3 lineage, and inducible Tregs, which upregulate FoxP3 expression and are derived in the periphery from naïve CD4<sup>+</sup> T-cell precursors under tolerogenic conditions (2). Tregs are highly enriched in the tumor microenvironment and are well known for their roles in tumor progression. They are considered to be important for limiting antitumor immune responses and promoting immunologic ignorance (peripheral tolerance) of cancer cells. Recently, we expanded on the roles of Tregs beyond immune suppression in tumors and showed that Tregs are directly involved in promoting angiogenic reprogramming of the tumor microenvironment (3), highlighting a multifaceted role for Tregs in promoting cancer through tumor immune escape and angiogenesis. Thus, we assert that future cancer therapy strategies must take into consideration either the elimination or the functional suppression of Tregs because these cells play an important role in the establishment of aggressive tumor phenotypes.

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**doi:** 10.1158/0008-5472.CAN-11-3687

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### Tregs Are Increased in Tumors and Are Correlated with a Poor Prognosis

Woo and colleagues (4) were the first to report an increased number of Tregs in cancer patients. They showed that regulatory CD4<sup>+</sup>CD25<sup>+</sup> T cells were increased in tumor sites in non-small cell lung and ovarian cancers, and that these cells (now appreciated as Tregs) secreted large amounts of TGF- $\beta$  that inhibited CD8<sup>+</sup> effector T-cell functions *in vitro* (4). An increased number of Tregs has been shown in a multitude of cancers, including melanoma and ovarian, breast, colorectal, lung, and pancreatic cancers [see Zou (5) and references therein]. In a study of ovarian cancer patients, Tregs that were isolated from the tumor site, ascites, or peripheral blood were equally able to suppress tumor-antigen-specific immune responses, suggesting that Tregs contribute to the promotion of ovarian cancer, likely due to their enhanced recruitment or local expansion rather than an enhanced suppressive capacity acquired in the tumor microenvironment (6).

Increased numbers of Tregs in tumors have been associated with poor survival in many solid tumors, including breast cancer (7), gastric cancer (8), and ovarian cancer (6, 9). In ovarian cancer, a low abundance of tumor-infiltrating Tregs can translate into years of added survival, highlighting the importance of these cells for tumor progression (6). However, some groups have identified Treg infiltration to be a biomarker of good clinical outcome [e.g., in colon (10) and ovarian (11) carcinomas], highlighting the complexity of Tregs as a biomarker. We have observed that Treg infiltration increases in proportion to effector T cells in cancer. Thus, Tregs could be associated with improved outcome, if considered as an isolated parameter, possibly reflecting the overall T-cell infiltration, which also predicts improved outcome in colon cancer (12, 13) and ovarian cancer (14). Therefore, of particular importance is the ratio of Tregs to CD8<sup>+</sup> effector cells, with a high CD8:Treg ratio representing the best indicator of prolonged survival (9). Mouse models further support the role of Tregs in tumor progression. Depletion of Tregs was shown to facilitate tumor

rejection and induction of antitumor immunity (15, 16), which is associated with a fundamental shift in the tumor microenvironment cytokine milieu (17). Of importance, whereas the transfer of tumor-reactive CD8<sup>+</sup> T cells is known to result in tumor elimination experimentally, the cotransfer of Tregs with CD8<sup>+</sup> cells abrogates their efficacy in both ovarian cancer and melanoma models (6, 18). Furthermore, Treg depletion *in vitro* allowed for the expansion of NYESO-1-reactive Th1 cells derived from cancer patients (19). Thus, Tregs suppress tumor-specific immunity and significantly affect the course of tumor progression across multiple tumor types.

### Mechanisms of Immune Suppression by Tregs

Much of what is known about Tregs in tumor progression is related to their ability to limit antitumor immune responses, resulting in immunologic tolerance and ignorance of the tumor. The 4 best-known mechanisms of immune regulation by Tregs are (i) secretion of soluble or membrane-tethered immunosuppressive molecules, (ii) direct cytolytic activity, (iii) metabolic disruption, and (iv) suppression of dendritic cells (DC) [for an extensive review, see Vignali and colleagues (20)].

#### Suppressive cytokines and secreted molecules

Chief among the mechanisms of T-cell suppression is the secretion of soluble or membrane-tethered mediators that inhibit effector T-cell functions through cell-contact-dependent and -independent mechanisms. The primary established Treg-derived cytokines that are responsible for this are interleukin (IL)-10, TGF- $\beta$ , and IL-35, which function by inhibiting the activities of effector T cells (20). Of importance for tumor development, both TGF- $\beta$  and IL-10 derived from Tregs have been shown to be key mediators that contribute to tumor progression by limiting antitumor immunity (21, 22). These cytokines prevent the expansion, cytokine elaboration (e.g., IFN- $\gamma$  and TNF- $\alpha$ ), and effector functions (cytolysis) of effector cells that are critically important for the control of tumor growth but also polarize DCs toward tolerogenic phenotypes. Our recent discovery that Tregs secrete VEGF (3), a known immunosuppressive molecule, adds to the panel of paracrine mechanisms through which Treg can exert suppression and affect the differentiation and function of DCs.

#### Cytolysis

An additional mechanism of regulation is the killing of effector T cells or possibly tumor antigen-presenting DCs. Tregs have been shown to exert cytolytic functions by using a variety of mediators, such as granzyme B (23, 24), the TRAIL pathway (25), and galectin-1 (26). The activation of these pathways by Tregs induces apoptosis on target effector cells. Of importance, Cao and colleagues (27) were able to show that Treg-derived granzyme-B and perforin are responsible for the suppression of natural killer (NK) cells and the ability of cytotoxic CD8<sup>+</sup> cells to eliminate tumors in multiple models.

#### Metabolic disruption

Investigators have proposed several mechanisms by which Tregs may be able to inhibit the functions of effector T cells

metabolically. Although the idea is controversial, Pandiyan and colleagues (28) suggested that Tregs can essentially "starve" effector cells by depleting local resources of IL-2, leading to effector cell apoptosis. Additionally, it was shown that Tregs catalyze ATP to adenosine through expression of CD39 and CD73, and in turn adenosine suppresses effector T-cell functions (29). Finally, Tregs were suggested to inhibit effector T-cell function by the physical transfer of cAMP through membrane gap junctions (30). The contribution of these mechanisms to tumor immune escape is unknown.

#### Dendritic cell interactions

Some evidence suggests that Tregs may mediate immune suppression through secondary cell types, with the largest body of evidence supporting deleterious interactions with DCs. Tregs induce DCs through cell-cell-mediated reverse signaling by cytotoxic T-lymphocyte antigen 4 (CTLA-4), expressed on Tregs, and CD80 and/or CD86, expressed on DCs, to upregulate indoleamine 2,3-dioxygenase (IDO) in DCs (31). IDO expression is responsible for the catabolism of tryptophan, which suppresses effector T-cell function by simultaneously depleting essential tryptophan and generating immunosuppressive tryptophan metabolites. Further, Tregs have been shown to reduce the capacity for DCs to activate effector T cells through inhibition of costimulatory molecules, suppression of DC maturation via IL-10/TGF- $\beta$  signaling, or Treg-DC interactions mediated by lymphocyte-activation gene 3 (32, 33).

#### Recruitment of Tregs to the Tumor Microenvironment

The increased number of Tregs in tumor sites is probably due to a number of factors. Tumor environments such as ovarian cancer (6) and Hodgkin lymphoma (34) contain large amounts of CC-chemokine ligand 22 (CCL22), which is likely derived from both tumor cells and tumor macrophages. CCL22 can recruit Tregs through CCR4, and Treg migration can be abrogated through CCR4 blockade *in vitro*. Recently, we identified a novel immunosuppressive and angiogenic circuit that establishes a direct role for tumor hypoxia in the recruitment of Tregs in ovarian cancer (3). Hypoxia is a key promoter of tumor angiogenesis and has been previously linked to the infiltration of Tregs in breast cancer (35). We have recently shown that hypoxia upregulates CCL28 in ovarian cancer cells, and CCL28 expression was responsible for the recruitment of CD4<sup>+</sup>CD25<sup>+</sup>Foxp3<sup>+</sup> Treg cells through ligation of the cognate receptor CCR10 expressed on Tregs (3). In ovarian cancer patients, CCL28 expression was correlated with the expression of hypoxia-inducible factor 1 $\alpha$  (HIF-1 $\alpha$ ), which is a poor-prognosis biomarker, and high CCL28 expression in patient tumors was also shown to be an indicator of poor survival. Artificial overexpression of CCL28 in mouse ovarian cancer cells led to enhanced growth of intraperitoneal tumors, as characterized by increased Treg infiltration and increased IL-10 production in the peritoneal ascites (3). Of importance, in the CCL28-overexpressing mouse tumor model, depletion of CD25<sup>+</sup> or CCR10<sup>+</sup> cells eliminated Tregs from the tumor and

abrogated the tumor growth advantage conferred by CCL28 overexpression. It is possible that numerous additional chemokines regulate Treg recruitment in cancer [see Campbell and Koch (36) for a list of potentially important chemokine receptors], and they may have nonredundant roles in recruiting as-yet-unidentified Treg subsets. In the case of CCL28–CCR10 interactions, recruitment of Tregs to the specific hypoxic environment may serve to enhance their immunosuppressive capacity as part of a biologic program (1, 37), because it has been shown that hypoxia increases the potency of Tregs, and hypoxia-exposed Tregs are more effective at suppressing the proliferation of effector cells (1, 37).

### Expansion of Tregs in the Tumor Microenvironment

For the most part, Treg cells can be divided into natural Tregs (nTreg), which are derived from the thymus and maintained peripherally by TGF- $\beta$ , or inducible Tregs (iTreg), which are induced from naïve CD4<sup>+</sup> T-cell precursors and exert suppressive characteristics similar to those observed for nTregs. Both of these Treg subtypes express FoxP3 [a more detailed discussion of this concept can be found in Curotto de Lafaille and Lafaille (2)]. Beyond recruitment of nTregs through chemokines, the tumor microenvironment promotes the continued expansion of nTregs (38) and the generation of iTregs (39) due to a tumor microenvironment that is rich in cytokines such as IL-10 (40), TGF- $\beta$  (41), and adenosine (42) derived from either the tumor cells or tumor-resident immunosuppressive DCs (43) and TIE-2<sup>+</sup> monocytes (39, 44). These circuits are a reflection of physiologic homeostatic mechanisms that tumors co-opt in tissue-specific and anatomic-compartment-restricted ways. For example, naïve CD4<sup>+</sup> cells are converted into iTregs by CD103<sup>+</sup> DCs in the mesenteric lymph nodes, a mechanism that helps to maintain gut homeostasis in a Toll-like receptor agonist-rich environment (45).

### Tregs in Tumor Angiogenesis

#### Tumor angiogenesis

Angiogenesis is defined as the sprouting of new blood vessels from preexisting ones. Under physiologic conditions, such as development, angiogenesis occurs in a stepwise manner involving vessel destabilization, endothelial cell migration and proliferation, sprouting, and resolution with vessel stabilization (46). Tumor angiogenesis differs in that the resolution phase generally fails and the vessel network is highly disordered. However, blood vessel development is critical for tumor growth because it provides essential nutrients and growth factors while also providing a conduit for waste, and sustained angiogenesis has long been considered a hallmark of cancer (47).

The accumulation of Tregs at tumor sites has been correlated with biomarkers of accelerated angiogenesis such as VEGF overexpression and increased microvessel density in endometrial (48) and breast cancers (49), providing clinical cues for an association between Tregs and angiogenesis. Tregs can contribute to tumor angiogenesis through both indirect and direct mechanisms. Tregs promote angiogenesis indirectly

by suppressing the activities of Th1 effector T cells that release angiostatic cytokines like TNF- $\alpha$  and IFN- $\gamma$ , as well as interferon-induced chemokines such as CXCL9, 10, and 11. Indeed, Tregs have been shown to promote tumor angiogenesis by specifically inhibiting tumor-reactive T cells (52). However, we have also shown that Tregs can make significant contributions to the direct promotion of tumor angiogenesis (ref. 3, Fig. 1). We showed that tumor hypoxia in ovarian cancer leads to the recruitment of Tregs via CCL28 upregulation (3). The forced expression of CCL28 in mouse ovarian carcinoma resulted in striking increase of *in vivo* tumor growth. CCL28 expression also resulted in robust Treg accumulation, increased VEGF levels, and significantly increased blood vessel development. Of importance, depletion of CD25<sup>+</sup> or CCR10<sup>+</sup> cells eliminated Treg cells from the tumor microenvironment and significantly suppressed VEGF expression and angiogenesis at these sites (3). We showed that CD4<sup>+</sup>CD25<sup>+</sup> Treg cells secreted higher amounts of VEGF in the steady state as well as under hypoxic conditions when compared with CD4<sup>+</sup>CD25<sup>-</sup> T cells, and media conditioned by Tregs in hypoxia promoted capillary tube formation *in vitro*, an effect that was dependent on VEGF signaling. Further, using an entirely cell-free Matrigel implant, we showed that supernatants of hypoxic Tregs were able to significantly promote angiogenesis *in vivo* (3). Our results are supported by early observations that T cells exposed to hypoxia express VEGF, and T cells within tumors express VEGF (53). Thus, we established a new mechanism whereby tumor hypoxia recruits Tregs to tumor sites, leading to substantial, direct contributions to the proangiogenic tumor microenvironment.

### Tregs as Targets for Cancer Immunotherapy

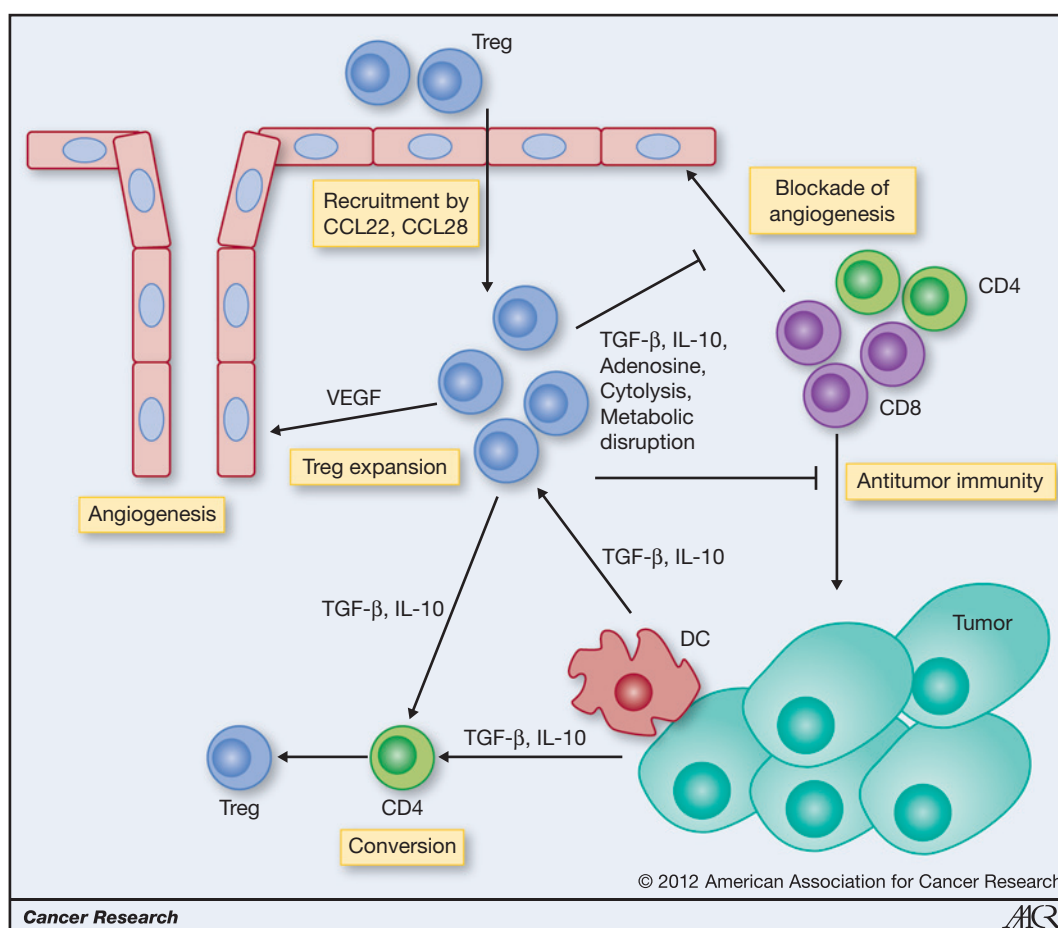
On the basis of the information provided above, it is apparent that Tregs make important contributions to tumor immune escape and have newly described functions in angiogenesis. Therefore, the elimination of Tregs in cancer patients, and particularly within the tumor microenvironment, should be considered to be an essential component of any successful cancer therapy. Several available therapeutics can either reduce the number of Tregs or disrupt their functions. Of interest, several chemotherapeutic drugs that interfere with Tregs, such as methotrexate and cyclophosphamide, have well-described immunostimulatory and antiangiogenic effects in cancer patients.

#### Nonspecific targeting of Tregs

A number of commonly used chemotherapeutics have been shown to reduce either the number or the immunosuppressive capacity of Tregs. These drugs include antimetabolites such as cyclophosphamide, gemcitabine, mitoxantrone, and fludarabine, as well as thalidomide analogues and cyclooxygenase 2 (COX-2) inhibitors. Thus, it is intriguing to speculate that these drugs have some off-target antitumor effects mediated through the modulation of Tregs.

Cyclophosphamide has been shown to preferentially deplete CD4<sup>+</sup>CD25<sup>+</sup> Tregs in rats, and in one study (54), only a single injection prior to tumor challenge with a rat colon cancer line was sufficient to delay tumor growth. Cyclophosphamide





**Figure 1.** Role of Tregs in tumor progression. Tregs are recruited to tumors from the periphery by tumor-derived, hypoxia-induced CCL28, but also by DC and tumor-derived CCL22. Within the tumor microenvironment, Tregs can be expanded by TGF- $\beta$  and possibly IL-10, which can also convert CD4<sup>+</sup>-naïve precursors into induced Tregs. Tregs promote tumor progression by direct inhibition of antitumor effector CD4<sup>+</sup> and CD8<sup>+</sup> T cells through inhibitory cytokines, cytolysis, and metabolic disruption. Further, Tregs that have been recruited to hypoxic areas directly stimulate angiogenesis through production of VEGF. Tregs also indirectly encourage angiogenesis by blocking effector-cell-derived angiostatic cytokines such as IFN- $\gamma$  and CXCL-10.

alkylates DNA, resulting in crosslinks between (interstrand) and within (intrastrand) DNA strands, which leads to cell death, and it has been suggested that Tregs are more sensitive to cyclophosphamide-induced apoptosis (55). For example, Ercolini and colleagues (56) showed that low-dose cyclophosphamide given to HER-2/neu transgenic mice with HER-2/neu-expressing mammary tumors selectively depleted Tregs that were progressing through the cell cycle. Of interest, in untreated tumor-bearing mice, Tregs were shown to be the predominant cycling T-cell population, although fewer than half of the Foxp3<sup>+</sup>CD4<sup>+</sup>CD25<sup>+</sup> Treg cells were cycling and thus susceptible to depletion by cyclophosphamide. Redmond and colleagues (57) reported that effector cells that are in the process of being tolerized or deleted are also cycling and proliferating, indicating that depletion of cycling cells with cyclophosphamide may be beneficial for clearing out tolerizing cells. Further, low-dose cyclophosphamide also disrupts the homeostatic proliferation of Tregs and decreases their immunosuppressive functionality by decreasing FoxP3 and glucocorticoid-induced TNF receptor (GITR; ref. 55). Recent evidence also suggests that

inhibition of Treg function by low-dose cyclophosphamide may be the result of selective depletion of intracellular stores of ATP caused by increased surface expression of CD39 (an ATP-to-adenosine conversion enzyme; ref. 58). However, in one study (59), cyclophosphamide was shown to deplete effector T cells in addition to Tregs. Therefore, cyclophosphamide disrupts Tregs via a multitude of mechanisms, but it may also have unintended effects on tumor-reactive effector T cells.

Two drugs that inhibit DNA synthesis, fludarabine and gemcitabine, have also been shown to disrupt Tregs. Fludarabine administration as a standard 5-day course induces lymphopenia, which has been shown to be favorable for patients with chronic lymphocytic leukemia and other hematologic malignancies. In a clinical setting, fludarabine treatment of chronic lymphocytic leukemia resulted in surprising increases of Treg apoptosis and decreases of Treg inhibitory functions (60). Further, fludarabine blocked the expansion of IL-10-producing CD4<sup>+</sup> Tregs *in vitro*, which was associated with higher numbers of antigen-specific CTLs (61). In a phase I study in non-small cell lung cancer, gemcitabine

administration induced lymphopenia with a decrease in effector T-cell populations (62). However, another phase I study of patients with colon cancer showed that gemcitabine caused an increase in CTLs with a concomitant decrease in CD25<sup>+</sup>CD4<sup>+</sup> T cells in clinical responders (63). Although both studies showed positive results, the direct effects of this drug on lymphocyte populations is unknown. Gemcitabine affects a variety of immunosuppressive cells, including myeloid-derived suppressor cells (MDSC). When given at a clinically equivalent dose, gemcitabine resulted in a dramatically reduced number of MDSCs in animal spleens, accompanied by an increase in the antitumor activity of CD8<sup>+</sup> T cells and activated NK cells (64). In light of the observation that MDSCs are capable of converting naïve CD4 cells to Tregs, it is entirely plausible that gemcitabine limits Tregs through its effects on MDSCs (65). Although both fludarabine and gemcitabine show little to no direct specificity for Tregs, it is possible that a particular dosing regimen could provide optimal disruption of Tregs while concomitantly inhibiting tumor growth.

Thalidomide and thalidomide derivatives have been used in the treatment of various nonmalignant diseases, including cutaneous and systemic inflammatory disorders (66–68). Lenalidomide is a U.S. Food and Drug Administration (FDA)-approved oral thalidomide analogue that is used to treat multiple myeloma and low- to intermediate-risk myelodysplastic syndrome caused by deletion of chromosome 5q (5q syndrome). Lenalidomide induces myeloma cell apoptosis directly and indirectly by inhibition of bone marrow stromal cell support, by antiangiogenic and antiosteoclastogenic effects, and through immunomodulatory activity. Lenalidomide has a broad range of immunomodulatory properties that can be exploited to treat many hematologic and solid cancers. Lenalidomide inhibits human Treg cell proliferation in response to IL-2 and downregulates FoxP3 expression (69). It was also shown to significantly reduce Treg cells in mouse lymph nodes (69). In a recent clinical trial in chronic leukocyte leukemia, the administration of lenalidomide resulted in a decrease of Tregs and an increase of Th17 cells in peripheral blood (70), supporting a potential role for thalidomide analogues in the elimination of Tregs in patients (69). Of importance, lenalidomide has also been shown to exert costimulatory effects on T cells and enhance T-cell proliferation, effector function (71–73), and Th1 reprogramming (74), and in combination therapy it has augmented tumor lysate vaccines (75).

COX enzymes, and particularly COX-2, are known to contribute to many facets of tumor progression. Patients who take a nonsteroidal anti-inflammatory drug (NSAID), such as aspirin, are significantly less likely to develop colorectal cancer (76, 77), and several investigators have shown an adjuvant property of COX-2 inhibitors in combination with cancer vaccines (78–80). Experimentally, COX-2 inhibition was shown to reduce Treg cell frequency and suppressive activity, attenuate FoxP3 expression in tumor-infiltrating lymphocytes, and decrease tumor burden *in vivo* (81). In patients with colon cancer, treatment with an oral NSAID significantly increased CD8<sup>+</sup> tumor-infiltrating T cells and decreased expression of FoxP3 and IL-10 (82). Patients with colorectal cancer have increased concentrations of prostaglandin E<sub>2</sub> (PGE<sub>2</sub>) in the

peripheral blood, and although tumors express large amounts of PGE<sub>2</sub>, it has been shown that Treg cells also express COX-2 and produce PGE<sub>2</sub> in a manner that suppresses effector T cells. A role for Treg-derived PGE<sub>2</sub> in immune suppression is supported by the observation that indomethacin (a COX-2 inhibitor) reverses Treg-mediated antitumor suppression *in vitro* (83).

### Specific targeting of Tregs

In light of the important role played by Tregs in different kinds of tumors and other pathologies, investigators have developed several compounds (often depletion antibodies) that directly target Tregs through recognition of Treg markers such as CD25, CTLA-4, and GITR.

A large number of Treg-targeting strategies rely on specific recognition of CD25. In several mouse tumor models, CD4<sup>+</sup>CD25<sup>+</sup> Treg depletion via antibodies targeting CD25 produced significant antitumor activity, although it was often associated with an increased incidence of autoimmunity. Combinatorial approaches using monoclonal antibodies and vaccines have been investigated in murine models, and the positive results of these preclinical studies clearly highlight the potential of the Treg-depletion approach in cancer immunotherapy (84). In an early phase I clinical trial in patients with metastatic breast cancer, the anti-CD25 antibody daclizumab significantly depleted Treg cells and enhanced the immunogenicity of a cancer vaccine. Five of the 10 patients who received the vaccine had stable disease for several months (85, 86). However, in another study using daclizumab in combination with a DC vaccine, investigators noted a detrimental role of daclizumab treatment, which may have been due to the timing of administration (87).

Denileukin diftitox (Ontak; Esai, Inc.) is a fusion protein of human IL-2 and diphtheria toxin. The IL-2 portion of the fusion protein binds preferentially to cells expressing intermediate- to high-affinity IL-2 receptors (IL-2R) comprised of IL-2R $\alpha$ (CD25)/ $\beta$ (CD122)/ $\gamma$ (CD132) subunits or IL-2R $\beta$ / $\gamma$  subunits, and results in cell death by interfering with protein synthesis following endocytosis. Denileukin diftitox was shown to be efficacious in advanced chronic T-cell lymphoma with high CD25 expression, where high CD25 expression is associated with clinical response to denileukin diftitox (88). In patients with melanoma, the application of recombinant Ontak significantly but transiently reduced the frequency of Tregs in peripheral blood. However, another study that evaluated the treatment of melanoma patients with denileukin diftitox failed to show any Treg depletion or clinical benefit (89).

LMB-2 is a fusion protein that is obtained by fusing a single-chain variable fragment antibody (scFv) against CD25 to *Pseudomonas* exotoxin A. *In vitro*, treatment of human peripheral blood mononuclear cells (PBMC) with LMB-2 resulted in specific CD4<sup>+</sup>CD25<sup>+</sup> Treg depletion (90). In a phase I clinical trial, treatment of CD25<sup>+</sup> T-cell malignancies with a dose of >60  $\mu$ g/kg of LMB-2 showed encouraging results, with 8 objective responses in a cohort of 20 patients, indicating that LMB-2 is efficacious in patients (91). In patients with melanoma, LMB-2 administration in combination with peptide vaccination

showed a significant decrease of FoxP3<sup>+</sup>CD4<sup>+</sup>CD25<sup>+</sup> Tregs in peripheral blood; however, the effect was transient and the quantity of Tregs returned to pretreatment levels within days. As might be expected, there was no objective clinical response (92). Thus, the utility of LMB-2 is not yet clear.

When CTLA-4 was first cloned in 1987, it was not clear whether CTLA-4 was involved in stimulatory or inhibitory pathways in T cells. However, the generation of CTLA-4 knockout mice allowed investigators to solve this riddle, because the knockout mice developed a progressive and uncontrolled accumulation of activated T cells and died of lymphoproliferative disease (93). The seminal study by Leach and colleagues (94) showed that CTLA-4 blockade could attenuate the growth of several implanted murine tumors, and the mechanism of inhibition was immune mediated. CTLA-4 is expressed on the surface of Tregs, but blockade could actually expand functionally suppressive Tregs (95). Although various CTLA-4 blockade therapies reduce tumor-infiltrating Tregs (84), this effect may be due entirely to the ability of CTLA-4 blockade to promote the generation of memory and promote effector T-cell functions (96).

So far, 2 humanized anti-human CTLA-4 neutralizing antibodies, MDX-010 (ipilimumab) and CP-675206 (tremelimumab), have been tested in phase I-III trials. The first phase I clinical trial with anti-CTLA-4 blocking antibody was carried out in 2002 at the University of California, Los Angeles and The University of Texas MD Anderson Cancer Center. The majority of enrolled patients had measurable metastatic melanoma. This trial tested doses ranging from 0.01 to 15 mg/kg in 7 cohorts. Objective tumor responses were noted in a subset of patients starting at a dose of 3 mg/kg and becoming more frequent at 15 mg/kg (97). Of interest, supporting the immune-modulatory effects of CTLA-4, treatment of metastatic melanoma patients with ipilimumab resulted in tumor regression in 36% of the patients and was associated with autoimmune toxicity, but patients without autoimmune toxicity were less likely to experience tumor regression (98). A further trial combining high-dose IL-2 and varied doses of ipilimumab showed synergy compared with earlier studies that evaluated IL-2 alone in metastatic melanoma (99). Further analysis of PBMCs in patients undergoing anti-CTLA-4 treatment for stage IV metastatic melanoma and renal cell carcinoma revealed, by *in vitro* coculture, no inhibition of the suppressive activity of CD4<sup>+</sup>CD25<sup>+</sup> T cells per se, but a probable enhancement of effector T-cell function. The results of a phase III clinical trial that included 502 untreated patients with metastatic melanoma were recently reported (100). Ipilimumab (10 mg/kg) in combination with dacarbazine, as compared with dacarbazine plus placebo, improved overall survival in the patients, leading to its FDA approval for the treatment of metastatic melanoma (100). Tremelimumab has been shown not only to suppress Treg activity but also to induce expansion of effector and memory CD4<sup>+</sup> and CD8<sup>+</sup> T cells, with antitumor efficacy (101). Thus, it has been suggested that depletion of Tregs may be of secondary importance for modulating the ratio of CD8<sup>+</sup> effector cells to Tregs, which may be mediated through direct interactions of anti-CTLA-4 antibody with effector cells.

GITR is a TNF receptor family member that is expressed at low levels on resting CD8<sup>+</sup> and CD4<sup>+</sup>Foxp3<sup>-</sup> T cells but is constitutively expressed at high levels on CD4<sup>+</sup>CD25<sup>+</sup>Foxp3<sup>+</sup> Tregs (102). Treg cells express even higher levels of GITR in tumors than elsewhere (103). Although it may not affect systemic Treg, GITR ligation specifically depletes Tregs in tumors, increasing tumor Teff/Treg ratios (104). DTA-1, a GITR agonistic Ab, may disable Treg, depletes intratumoral Treg, and enhances T-cell immunity against tumors (105–108). Of importance, it also costimulates CD4<sup>+</sup> and CD8<sup>+</sup> T-cell proliferation and effector functions, renders Teff cells resistant to Tregs, and enhances a variety of T-cell responses (109, 110). However, some studies have shown that DTA-1 does not affect the number or functions of Tregs. In one study (104), administration of an agonistic antibody prevented the infiltration of Tregs into the tumor microenvironment, promoting a high CD8:Treg ratio and resulting in the control of tumor growth in mice. Thus, although targeting Tregs through GITR is an interesting approach, it may require additional therapeutics to promote systemic antitumor immune responses.

We believe our recent studies have added a new possible target to this list: CCR10. Tumor hypoxia induced CCL28 expression, leading to the recruitment of CCR10<sup>+</sup> Tregs, whereas the depletion of CCR10<sup>+</sup> cells by an anti-CCR10 immunotoxin resulted in complete Treg depletion and loss of the tumor growth advantage conferred by CCL28 overexpression (3). Further, it appears that CCR10 expression on Tregs is associated with a peripheral homing phenotype, and is a marker of highly suppressive Treg cells (111). Although CCR10 expression is not entirely restricted to Tregs, CCR10<sup>+</sup> cell depletion has been shown to be beneficial in a mouse model of ovarian cancer. Thus, CCR10 is an attractive new target for disrupting Tregs in cancer.

## Conclusions

On the basis of the information presented above, it should be apparent that Tregs are instrumental in establishing tumor immune tolerance and are important cellular mediators of tumor progression in patients. In our recent work we expanded this view and showed that, in addition to immune suppression, Tregs can make significant contributions to the direct promotion of tumor angiogenesis. Thus, we believe that Tregs are key orchestrators of tumor development, linking immune suppression and angiogenesis in one biologic program. This highlights the need to specifically target these cells to promote antitumor immunity and tumor regression. Indeed, reducing the functions and/or numbers of Tregs in patients with cancer should allow more effective immune-based therapies, alone or in combination with traditional chemotherapeutics. Here, we have presented numerous preclinical and clinical data that support the notion that the elimination of Tregs should be considered crucial for many cancer therapies. However, a major therapeutic challenge remains because of the paucity of tools available to target Tregs effectively in the clinic. The effort to unravel the complexity of Tregs is only just beginning, and a further understanding of their biology and

characterization of targets will undoubtedly enhance future therapeutic opportunities.

### Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed.

### References

- Motz GT, Coukos G. The parallel lives of angiogenesis and immunosuppression: cancer and other tales. *Nat Rev Immunol* 2011;11:702–11.
- Curotto de Lafaille MA, Lafaille JJ. Natural and adaptive foxp3+ regulatory T cells: more of the same or a division of labor? *Immunity* 2009;30:626–35.
- Facciabene A, Peng X, Hagemann IS, Balint K, Barchetti A, Wang LP, et al. Tumour hypoxia promotes tolerance and angiogenesis via CCL28 and T(reg) cells. *Nature* 2011;475:226–30.
- Woo EY, Chu CS, Goletz TJ, Schlienger K, Yeh H, Coukos G, et al. Regulatory CD4(+)CD25(+) T cells in tumors from patients with early-stage non-small cell lung cancer and late-stage ovarian cancer. *Cancer Res* 2001;61:4766–72.
- Zou W. Regulatory T cells, tumour immunity and immunotherapy. *Nat Rev Immunol* 2006;6:295–307.
- Curiel TJ, Coukos G, Zou L, Alvarez X, Cheng P, Mottram P, et al. Specific recruitment of regulatory T cells in ovarian carcinoma fosters immune privilege and predicts reduced survival. *Nat Med* 2004;10:942–9.
- Bates GJ, Fox SB, Han C, Leek RD, Garcia JF, Harris AL, et al. Quantification of regulatory T cells enables the identification of high-risk breast cancer patients and those at risk of late relapse. *J Clin Oncol* 2006;24:5373–80.
- Sasada T, Kimura M, Yoshida Y, Kanai M, Takabayashi A. CD4+CD25+ regulatory T cells in patients with gastrointestinal malignancies: possible involvement of regulatory T cells in disease progression. *Cancer* 2003;98:1089–99.
- Sato E, Olson SH, Ahn J, Bundy B, Nishikawa H, Qian F, et al. Intraepithelial CD8+ tumor-infiltrating lymphocytes and a high CD8+/regulatory T cell ratio are associated with favorable prognosis in ovarian cancer. *Proc Natl Acad Sci U S A* 2005;102:18538–43.
- Correale P, Rotundo MS, Del Vecchio MT, Remondo C, Migali C, Ginanneschi C, et al. Regulatory (FoxP3+) T-cell tumor infiltration is a favorable prognostic factor in advanced colon cancer patients undergoing chemo or chemoimmunotherapy. *J Immunother* 2010;33:435–41.
- Leffers N, Gooden MJ, de Jong RA, Hoogbeem BN, ten Hoor KA, Hollema H, et al. Prognostic significance of tumor-infiltrating T-lymphocytes in primary and metastatic lesions of advanced stage ovarian cancer. *Cancer Immunol Immunother* 2009;58:449–59.
- Galon J, Costes A, Sanchez-Cabo F, Kirilovsky A, Mlecnik B, Lagorce-Pagès C, et al. Type, density, and location of immune cells within human colorectal tumors predict clinical outcome. *Science* 2006;313:1960–4.
- Pagès F, Berger A, Camus M, Sanchez-Cabo F, Costes A, Molidor R, et al. Effector memory T cells, early metastasis, and survival in colorectal cancer. *N Engl J Med* 2005;353:2654–66.
- Zhang L, Conejo-Garcia JR, Katsaros D, Gimotty PA, Massobrio M, Regnani G, et al. Intratumoral T cells, recurrence, and survival in epithelial ovarian cancer. *N Engl J Med* 2003;348:203–13.
- Onizuka S, Tawara I, Shimizu J, Sakaguchi S, Fujita T, Nakayama E. Tumor rejection by in vivo administration of anti-CD25 (interleukin-2 receptor alpha) monoclonal antibody. *Cancer Res* 1999;59:3128–33.
- Shimizu J, Yamazaki S, Sakaguchi S. Induction of tumor immunity by removing CD25+CD4+ T cells: a common basis between tumor immunity and autoimmunity. *J Immunol* 1999;163:5211–8.
- Yu P, Lee Y, Liu W, Krausz T, Chong A, Schreiber H, et al. Intratumor depletion of CD4+ cells unmasks tumor immunogenicity leading to the rejection of late-stage tumors. *J Exp Med* 2005;201:779–91.
- Antony PA, Piccirillo CA, Akpınarli A, Finkelstein SE, Speiss PJ, Surman DR, et al. CD8+ T cell immunity against a tumor/self-antigen is augmented by CD4+ T helper cells and hindered by naturally occurring T regulatory cells. *J Immunol* 2005;174:2591–601.
- Nishikawa H, Jäger E, Ritter G, Old LJ, Gnjatich S. CD4+ CD25+ regulatory T cells control the induction of antigen-specific CD4+ helper T cell responses in cancer patients. *Blood* 2005;106:1008–11.
- Vignali DA, Collison LW, Workman CJ. How regulatory T cells work. *Nat Rev Immunol* 2008;8:523–32.
- Loser K, Apelt J, Voskort M, Mohaupt M, Balkow S, Schwarz T, et al. IL-10 controls ultraviolet-induced carcinogenesis in mice. *J Immunol* 2007;179:365–71.
- Strauss L, Bergmann C, Szczepanski M, Gooding W, Johnson JT, Whiteside TL. A unique subset of CD4+CD25highFoxp3+ T cells secreting interleukin-10 and transforming growth factor-beta1 mediates suppression in the tumor microenvironment. *Clin Cancer Res* 2007;13:4345–54.
- Grossman WJ, Verbsky JW, Tollefsen BL, Kemper C, Atkinson JP, Ley TJ. Differential expression of granzymes A and B in human cytotoxic lymphocyte subsets and T regulatory cells. *Blood* 2004;104:2840–8.
- Gondek DC, Lu LF, Quezada SA, Sakaguchi S, Noelle RJ. Cutting edge: contact-mediated suppression by CD4+CD25+ regulatory cells involves a granzyme B-dependent, perforin-independent mechanism. *J Immunol* 2005;174:1783–6.
- Ren X, Ye F, Jiang Z, Chu Y, Xiong S, Wang Y. Involvement of cellular death in TRAIL/DR5-dependent suppression induced by CD4(+)CD25(+) regulatory T cells. *Cell Death Differ* 2007;14:2076–84.
- Garín MI, Chu CC, Golshayan D, Cernuda-Morollón E, Wait R, Lechler RI. Galectin-1: a key effector of regulation mediated by CD4+CD25+ T cells. *Blood* 2007;109:2058–65.
- Cao X, Cai SF, Fehniger TA, Song J, Collins LI, Piwnica-Worms DR, et al. Granzyme B and perforin are important for regulatory T cell-mediated suppression of tumor clearance. *Immunity* 2007;27:635–46.
- Pandiyani P, Zheng L, Ishihara S, Reed J, Lenardo MJ. CD4+CD25+Foxp3+ regulatory T cells induce cytokine deprivation-mediated apoptosis of effector CD4+ T cells. *Nat Immunol* 2007;8:1353–62.
- Deaglio S, Dwyer KM, Gao W, Friedman D, Usheva A, Erat A, et al. Adenosine generation catalyzed by CD39 and CD73 expressed on regulatory T cells mediates immune suppression. *J Exp Med* 2007;204:1257–65.
- Bopp T, Becker C, Klein M, Klein-Hessling S, Palmethofer A, Serfling E, et al. Cyclic adenosine monophosphate is a key component of regulatory T cell-mediated suppression. *J Exp Med* 2007;204:1303–10.
- Fallarino F, Grohmann U, Hwang KW, Orabona C, Vacca C, Bianchi R, et al. Modulation of tryptophan catabolism by regulatory T cells. *Nat Immunol* 2003;4:1206–12.
- Cederbom L, Hall H, Ivars F. CD4+CD25+ regulatory T cells down-regulate co-stimulatory molecules on antigen-presenting cells. *Eur J Immunol* 2000;30:1538–43.
- Liang B, Workman C, Lee J, Chew C, Dale BM, Colonna L, et al. Regulatory T cells inhibit dendritic cells by lymphocyte activation gene-3 engagement of MHC class II. *J Immunol* 2008;180:5916–26.
- Ishida T, Ishii T, Inagaki A, Yano H, Komatsu H, Iida S, et al. Specific recruitment of CC chemokine receptor 4-positive regulatory T cells in Hodgkin lymphoma fosters immune privilege. *Cancer Res* 2006;66:5716–22.



35. Yan M, Jene N, Byrne D, Millar EK, O'Toole SA, McNeil CM, et al. Recruitment of regulatory T cells is correlated with hypoxia-induced CXCR4 expression, and is associated with poor prognosis in basal-like breast cancers. *Breast Cancer Res* 2011;13:R47.
36. Campbell DJ, Koch MA. Phenotypical and functional specialization of FOXP3 +regulatory T cells. *Nat Rev Immunol* 2011;11:119–30.
37. Ben-Shoshan J, Maysel-Auslender S, Mor A, Keren G, George J. Hypoxia controls CD4+CD25+ regulatory T-cell homeostasis via hypoxia-inducible factor-1alpha. *Eur J Immunol* 2008;38:2412–8.
38. Valzasina B, Piconese S, Guiducci C, Colombo MP. Tumor-induced expansion of regulatory T cells by conversion of CD4+CD25- lymphocytes is thymus and proliferation independent. *Cancer Res* 2006;66:4488–95.
39. Liu VC, Wong LY, Jang T, Shah AH, Park I, Yang X, et al. Tumor evasion of the immune system by converting CD4+CD25- T cells into CD4+CD25+ T regulatory cells: role of tumor-derived TGF-beta. *J Immunol* 2007;178:2883–92.
40. Seo N, Hayakawa S, Takigawa M, Tokura Y. Interleukin-10 expressed at early tumour sites induces subsequent generation of CD4(+) T-regulatory cells and systemic collapse of antitumour immunity. *Immunology* 2001;103:449–57.
41. Chen W, Jin W, Hardegen N, Lei KJ, Li L, Marinos N, et al. Conversion of peripheral CD4+CD25- naive T cells to CD4+CD25+ regulatory T cells by TGF-beta induction of transcription factor Foxp3. *J Exp Med* 2003;198:1875–86.
42. Zarek PE, Huang CT, Lutz ER, Kowalski J, Horton MR, Linden J, et al. A2A receptor signaling promotes peripheral tolerance by inducing T-cell anergy and the generation of adaptive regulatory T cells. *Blood* 2008;111:251–9.
43. Ghiringhelli F, Puig PE, Roux S, Parcellier A, Schmitt E, Solary E, et al. Tumor cells convert immature myeloid dendritic cells into TGF-beta-secreting cells inducing CD4+CD25+ regulatory T cell proliferation. *J Exp Med* 2005;202:919–29.
44. Coffelt SB, Chen YY, Muthana M, Welford AF, Tai AO, Scholz A, et al. Angiopoietin 2 stimulates TIE2-expressing monocytes to suppress T cell activation and to promote regulatory T cell expansion. *J Immunol* 2011;186:4183–90.
45. Coombes JL, Siddiqui KR, Arancibia-Cárcamo CV, Hall J, Sun CM, Belkaid Y, et al. A functionally specialized population of mucosal CD103+ DCs induces Foxp3 +regulatory T cells via a TGF-beta and retinoic acid-dependent mechanism. *J Exp Med* 2007;204:1757–64.
46. Szekanecz Z, Koch AE. Mechanisms of disease: angiogenesis in inflammatory diseases. *Nat Clin Pract Rheumatol* 2007;3:635–43.
47. Hanahan D, Weinberg RA. The hallmarks of cancer. *Cell* 2000;100:57–70.
48. Giatromanolaki A, Bates GJ, Koukourakis MI, Sivridis E, Gatter KC, Harris AL, et al. The presence of tumor-infiltrating FOXP3+ lymphocytes correlates with intratumoral angiogenesis in endometrial cancer. *Gynecol Oncol* 2008;110:216–21.
49. Gupta S, Joshi K, Wig JD, Arora SK. Intratumoral FOXP3 expression in infiltrating breast carcinoma: Its association with clinicopathologic parameters and angiogenesis. *Acta Oncol* 2007;46:792–7.
50. Müller-Hermelink N, Braumüller H, Pichler B, Wieder T, Mailhammer R, Schaak K, et al. TNFR1 signaling and IFN-gamma signaling determine whether T cells induce tumor dormancy or promote multistage carcinogenesis. *Cancer Cell* 2008;13:507–18.
51. Qin Z, Blankenstein T. CD4+ T cell-mediated tumor rejection involves inhibition of angiogenesis that is dependent on IFN gamma receptor expression by nonhematopoietic cells. *Immunity* 2000;12:677–86.
52. Casares N, Arribillaga L, Sarobe P, Dotor J, Lopez-Diaz de Cerio A, Melero I, et al. CD4+CD25 +regulatory cells inhibit activation of tumor-primed CD4+ T cells with IFN-gamma-dependent antiangiogenic activity, as well as long-lasting tumor immunity elicited by peptide vaccination. *J Immunol* 2003;171:5931–9.
53. Freeman MR, Schneck FX, Gagnon ML, Corless C, Soker S, Niknejad K, et al. Peripheral blood T lymphocytes and lymphocytes infiltrating human cancers express vascular endothelial growth factor: a potential role for T cells in angiogenesis. *Cancer Res* 1995;55:4140–5.
54. Ghiringhelli F, Larmonier N, Schmitt E, Parcellier A, Cathelin D, Garrido C, et al. CD4+CD25+ regulatory T cells suppress tumor immunity but are sensitive to cyclophosphamide which allows immunotherapy of established tumors to be curative. *Eur J Immunol* 2004;34:336–44.
55. Lutsiak ME, Semnani RT, De Pascalis R, Kashmiri SV, Schlom J, Sabzevari H. Inhibition of CD4(+)25+ T regulatory cell function implicated in enhanced immune response by low-dose cyclophosphamide. *Blood* 2005;105:2862–8.
56. Ercolini AM, Ladle BH, Manning EA, Pfannenstiel LW, Armstrong TD, Machiels JP, et al. Recruitment of latent pools of high-avidity CD8(+) T cells to the antitumor immune response. *J Exp Med* 2005;201:1591–602.
57. Redmond WL, Hernandez J, Sherman LA. Deletion of naive CD8 T cells requires persistent antigen and is not programmed by an initial signal from the tolerogenic APC. *J Immunol* 2003;171:6349–54.
58. Zhao J, Cao Y, Lei Z, Yang Z, Zhang B, Huang B. Selective depletion of CD4+CD25+Foxp3 +regulatory T cells by low-dose cyclophosphamide is explained by reduced intracellular ATP levels. *Cancer Res* 2010;70:4850–8.
59. Matsushita N, Pilon-Thomas SA, Martin LM, Riker AI. Comparative methodologies of regulatory T cell depletion in a murine melanoma model. *J Immunol Methods* 2008;333:167–79.
60. Beyer M, Kochanek M, Darabi K, Popov A, Jensen M, Endl E, et al. Reduced frequencies and suppressive function of CD4+CD25hi regulatory T cells in patients with chronic lymphocytic leukemia after therapy with fludarabine. *Blood* 2005;106:2018–25.
61. Hegde U, Chhabra A, Chattopadhyay S, Das R, Ray S, Chakraborty NG. Presence of low dose of fludarabine in cultures blocks regulatory T cell expansion and maintains tumor-specific cytotoxic T lymphocyte activity generated with peripheral blood lymphocytes. *Pathobiology* 2008;75:200–8.
62. Levitt ML, Kassem B, Gooding WE, Miketic LM, Landreneau RJ, Ferson PF, et al. Phase I study of gemcitabine given weekly as a short infusion for non-small cell lung cancer: results and possible immune system-related mechanisms. *Lung Cancer* 2004;43:335–44.
63. Correale P, Cusi MG, Tsang KY, Del Vecchio MT, Marsili S, Placa ML, et al. Chemo-immunotherapy of metastatic colorectal carcinoma with gemcitabine plus FOLFOX 4 followed by subcutaneous granulocyte macrophage colony-stimulating factor and interleukin-2 induces strong immunologic and antitumor activity in metastatic colon cancer patients. *J Clin Oncol* 2005;23:8950–8.
64. Suzuki E, Kapoor V, Jassar AS, Kaiser LR, Albelda SM. Gemcitabine selectively eliminates splenic Gr-1+/CD11b+ myeloid suppressor cells in tumor-bearing animals and enhances antitumor immune activity. *Clin Cancer Res* 2005;11:6713–21.
65. Serafini P, Mgebroff S, Noonan K, Borrello I. Myeloid-derived suppressor cells promote cross-tolerance in B-cell lymphoma by expanding regulatory T cells. *Cancer Res* 2008;68:5439–49.
66. Okafor MC. Thalidomide for erythema nodosum leprosum and other applications. *Pharmacotherapy* 2003;23:481–93.
67. Lazzarini M, Martellosi S, Marchetti F, Scabar A, Bradaschia F, Ronfani L, et al. Efficacy and safety of thalidomide in children and young adults with intractable inflammatory bowel disease: long-term results. *Aliment Pharmacol Ther* 2007;25:419–27.
68. Ossandon A, Cassarà EA, Priori R, Valesini G. Thalidomide: focus on its employment in rheumatologic diseases. *Clin Exp Rheumatol* 2002;20:709–18.
69. Galustian C, Meyer B, Labarthe MC, Dredge K, Klaschka D, Henry J, et al. The anti-cancer agents lenalidomide and pomalidomide inhibit the proliferation and function of T regulatory cells. *Cancer Immunol Immunother* 2009;58:1033–45.
70. Idler I, Giannopoulos K, Zenz T, Bhattacharya N, Nothing M, Döhner H, et al. Lenalidomide treatment of chronic lymphocytic leukaemia patients reduces regulatory T cells and induces Th17 T helper cells. *Br J Haematol* 2010;148:948–50.
71. Bartlett JB, Dredge K, Dalglish AG. The evolution of thalidomide and its IMiD derivatives as anticancer agents. *Nat Rev Cancer* 2004;4:314–22.



72. LeBlanc R, Hideshima T, Catley LP, Shringarpure R, Burger R, Mitsiades N, et al. Immunomodulatory drug costimulates T cells via the B7-CD28 pathway. *Blood* 2004;103:1787-90.
73. Haslett PA, Hanekom WA, Muller G, Kaplan G. Thalidomide and a thalidomide analogue drug costimulate virus-specific CD8+ T cells in vitro. *J Infect Dis* 2003;187:946-55.
74. Xu W, Celeridad M, Sankar S, Webb DR, Bennett BL. CC-4047 promotes Th1 cell differentiation and reprograms polarized human Th2 cells by enhancing transcription factor T-bet. *Clin Immunol* 2008;128:392-9.
75. Dredge K, Marriott JB, Todryk SM, Muller GW, Chen R, Stirling DI, et al. Protective antitumor immunity induced by a costimulatory thalidomide analog in conjunction with whole tumor cell vaccination is mediated by increased Th1-type immunity. *J Immunol* 2002;168:4914-9.
76. Benamouzig R, Uzzan B, Deyra J, Martin A, Girard B, Little J, et al. Prevention by daily soluble aspirin of colorectal adenoma recurrence: 4-year results of the APACC randomised trial. *Gut* 2011;2012;61:255-61.
77. Chan AT, Ogino S, Fuchs CS. Aspirin and the risk of colorectal cancer in relation to the expression of COX-2. *N Engl J Med* 2007;356:2131-42.
78. Toomey D, Conroy H, Jarnicki AG, Higgins SC, Sutton C, Mills KH. Therapeutic vaccination with dendritic cells pulsed with tumor-derived Hsp70 and a COX-2 inhibitor induces protective immunity against B16 melanoma. *Vaccine* 2008;26:3540-9.
79. Mukherjee P, Basu GD, Tindler TL, Subramani DB, Bradley JM, Arefayene M, et al. Progression of pancreatic adenocarcinoma is significantly impeded with a combination of vaccine and COX-2 inhibition. *J Immunol* 2009;182:216-24.
80. Haas AR, Sun J, Vachani A, Wallace AF, Silverberg M, Kapoor V, et al. Cyclooxygenase-2 inhibition augments the efficacy of a cancer vaccine. *Clin Cancer Res* 2006;12:214-22.
81. Sharma S, Yang SC, Zhu L, Reckamp K, Gardner B, Baratelli F, et al. Tumor cyclooxygenase-2/prostaglandin E2-dependent promotion of FOXP3 expression and CD4+ CD25+ T regulatory cell activities in lung cancer. *Cancer Res* 2005;65:5211-20.
82. Lönnroth C, Andersson M, Arvidsson A, Nordgren S, Brevinge H, Lagerstedt K, et al. Preoperative treatment with a non-steroidal anti-inflammatory drug (NSAID) increases tumor tissue infiltration of seemingly activated immune cells in colorectal cancer. *Cancer Immunol* 2008;8:5.
83. Yaqub S, Henjum K, Mahic M, Jahnsen FL, Aandahl EM, Bjørneth BA, et al. Regulatory T cells in colorectal cancer patients suppress anti-tumor immune activity in a COX-2 dependent manner. *Cancer Immunol Immunother* 2008;57:813-21.
84. Quezada SA, Peggs KS, Simpson TR, Shen Y, Littman DR, Allison JP. Limited tumor infiltration by activated T effector cells restricts the therapeutic activity of regulatory T cell depletion against established melanoma. *J Exp Med* 2008;205:2125-38.
85. Rech AJ, Vonderheide RH. Clinical use of anti-CD25 antibody daclizumab to enhance immune responses to tumor antigen vaccination by targeting regulatory T cells. *Ann N Y Acad Sci* 2009;1174:99-106.
86. Rech AJ, Mick R, Recio A, DeMichele A, Tweed CK, Fox KR, et al. Phase I study of anti-CD25 mab daclizumab to deplete regulatory T cells prior to telomerase/survivin peptide vaccination in patients (pts) with metastatic breast cancer (MBC). *J Clin Oncol* 28:15s, 2010 (suppl; abstr 2508).
87. Jacobs JF, Punt CJ, Lesterhuis WJ, Suttmuller RP, Brouwer HM, Scharenborg NM, et al. Dendritic cell vaccination in combination with anti-CD25 monoclonal antibody treatment: a phase I/II study in metastatic melanoma patients. *Clin Cancer Res* 2010;16:5067-78.
88. Talpur R, Jones DM, Alencar AJ, Apisarnthanarax N, Herne KL, Yang Y, et al. CD25 expression is correlated with histological grade and response to denileukin diftitox in cutaneous T-cell lymphoma. *J Invest Dermatol* 2006;126:575-83.
89. Attia P, Maker AV, Haworth LR, Rogers-Freezer L, Rosenberg SA. Inability of a fusion protein of IL-2 and diphtheria toxin (Denileukin Diftitox, DAB389IL-2, ONTAK) to eliminate regulatory T lymphocytes in patients with melanoma. *J Immunother* 2005;28:582-92.
90. Attia P, Powell DJ Jr, Maker AV, Kreitman RJ, Pastan I, Rosenberg SA. Selective elimination of human regulatory T lymphocytes in vitro with the recombinant immunotoxin LMB-2. *J Immunother* 2006;29:208-14.
91. Kreitman RJ, Wilson WH, White JD, Stetler-Stevenson M, Jaffe ES, Giardina S, et al. Phase I trial of recombinant immunotoxin anti-Tac (Fv)-PE38 (LMB-2) in patients with hematologic malignancies. *J Clin Oncol* 2000;18:1622-36.
92. Powell DJ Jr, Felipe-Silva A, Merino MJ, Ahmadzadeh M, Allen T, Levy C, et al. Administration of a CD25-directed immunotoxin, LMB-2, to patients with metastatic melanoma induces a selective partial reduction in regulatory T cells in vivo. *J Immunol* 2007;179:4919-28.
93. Waterhouse P, Penninger JM, Timms E, Wakeham A, Shahinian A, Lee KP, et al. Lymphoproliferative disorders with early lethality in mice deficient in CtlA-4. *Science* 1995;270:985-8.
94. Leach DR, Krummel MF, Allison JP. Enhancement of antitumor immunity by CTLA-4 blockade. *Science* 1996;271:1734-6.
95. Kavanagh B, O'Brien S, Lee D, Hou Y, Weinberg V, Rini B, et al. CTLA4 blockade expands FoxP3+ regulatory and activated effector CD4+ T cells in a dose-dependent fashion. *Blood* 2008;112:1175-83.
96. Pedicord VA, Montalvo W, Leiner IM, Allison JP. Single dose of anti-CTLA-4 enhances CD8+ T-cell memory formation, function, and maintenance. *Proc Natl Acad Sci U S A* 2011;108:266-71.
97. Ribas A, Camacho LH, Lopez-Berestein G, Pavlov D, Bulanahagui CA, Millham R, et al. Antitumor activity in melanoma and anti-self responses in a phase I trial with the anti-cytotoxic T lymphocyte-associated antigen 4 monoclonal antibody CP-675,206. *J Clin Oncol* 2005;23:8968-77.
98. Attia P, Phan GQ, Maker AV, Robinson MR, Quezada MM, Yang JC, et al. Autoimmunity correlates with tumor regression in patients with metastatic melanoma treated with anti-cytotoxic T-lymphocyte antigen-4. *J Clin Oncol* 2005;23:6043-53.
99. Maker AV, Phan GQ, Attia P, Yang JC, Sherry RM, Topalian SL, et al. Tumor regression and autoimmunity in patients treated with cytotoxic T lymphocyte-associated antigen 4 blockade and interleukin 2: a phase I/II study. *Ann Surg Oncol* 2005;12:1005-16.
100. Robert C, Thomas L, Bondarenko I, O'Day S, Weber J, Garbe C, et al. Ipilimumab plus dacarbazine for previously untreated metastatic melanoma. *N Engl J Med* 2011;364:2517-26.
101. Ménard C, Ghiringhelli F, Roux S, Chaput N, Mateus C, Grohmann U, et al. CtlA-4 blockade confers lymphocyte resistance to regulatory T-cells in advanced melanoma: surrogate marker of efficacy of tremelimumab? *Clin Cancer Res* 2008;14:5242-9.
102. Nocentini G, Ronchetti S, Cuzzocrea S, Riccardi C. GITR/GITRL: more than an effector T cell co-stimulatory system. *Eur J Immunol* 2007;37:1165-9.
103. Coe D, Begom S, Addey C, White M, Dyson J, Chai JG. Depletion of regulatory T cells by anti-GITR mAb as a novel mechanism for cancer immunotherapy. *Cancer Immunol Immunother* 2010;59:1367-77.
104. Cohen AD, Schaefer DA, Liu C, Li Y, Hirschhorn-Cymerman D, Kim SC, et al. Agonist anti-GITR monoclonal antibody induces melanoma tumor immunity in mice by altering regulatory T cell stability and intratumor accumulation. *PLoS ONE* 2010;5:e10436.
105. Cohen AD, Diab A, Perales MA, Wolchok JD, Rizzuto G, Merghoub T, et al. Agonist anti-GITR antibody enhances vaccine-induced CD8(+) T-cell responses and tumor immunity. *Cancer Res* 2006;66:4904-12.
106. Ko K, Yamazaki S, Nakamura K, Nishioka T, Hirota K, Yamaguchi T, et al. Treatment of advanced tumors with agonistic anti-GITR mAb and its effects on tumor-infiltrating Foxp3+CD25+CD4+ regulatory T cells. *J Exp Med* 2005;202:885-91.
107. Ko HJ, Kim YJ, Kim YS, Chang WS, Ko SY, Chang SY, et al. A combination of chemioimmunotherapies can efficiently break self-tolerance and induce antitumor immunity in a tolerogenic murine tumor model. *Cancer Res* 2007;67:7477-86.
108. Sharma S, Dominguez AL, Manrique SZ, Cavallo F, Sakaguchi S, Lustgarten J. Systemic targeting of CpG-ODN to the tumor microenvironment with anti-neu-CpG hybrid molecule and T regulatory cell depletion induces memory responses in BALB-neuT tolerant mice. *Cancer Res* 2008;68:7530-40.

109. Shimizu J, Yamazaki S, Takahashi T, Ishida Y, Sakaguchi S. Stimulation of CD25(+)CD4(+) regulatory T cells through GITR breaks immunological self-tolerance. *Nat Immunol* 2002;3:135–42.
110. Moriglan SJ, Ramirez-Montagut T, Alpdogan O, Van Huystee TW, Eng JM, Hubbard VM, et al. GITR activation induces an opposite effect on alloreactive CD4(+) and CD8(+) T cells in graft-versus-host disease. *J Exp Med* 2004;200:149–57.
111. Eksteen B, Miles A, Curbishley SM, Tselepis C, Grant AJ, Walker LS, et al. Epithelial inflammation is associated with CCL28 production and the recruitment of regulatory T cells expressing CCR10. *J Immunol* 2006;177:593–603.