



Hemophilia gene therapy: ushering in a new treatment paradigm?

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After 3 decades of clinical trials, repeated proof-of-concept success has now been demonstrated in hemophilia A and B gene therapy. Current clinical hemophilia gene therapy efforts are largely focused on the use of systemically administered recombinant adeno-associated viral (rAAV) vectors for *F8* or *F9* gene addition. With multiple ongoing trials, including licensing studies in hemophilia A and B, many are cautiously optimistic that the first AAV vectors will obtain regulatory approval within approximately 1 year. While supported optimism suggests that the goal of gene therapy to alter the paradigm of hemophilia care may soon be realized, a number of outstanding questions have emerged from clinical trial that are in need of answers to harness the full potential of gene therapy for hemophilia patients. This article reviews the use of AAV vector gene addition approaches for hemophilia A and B, focusing specifically on information to review in the process of obtaining informed consent for hemophilia patients prior to clinical trial enrollment or administering a licensed AAV vector.

LEARNING OBJECTIVES

- Understand information important to review with hemophilia patients prior to consenting for a clinical trial or approved AAV vector
- Differentiate information that is well understood from information that remains incompletely understood

CLINICAL CASE

A 62-year-old man with severe hemophilia A (HA) with a history of an inhibitor, eradicated by immune tolerance induction, on emicizumab prophylaxis is interested in enrolling in a HA gene therapy trial or receiving a licensed adeno-associated viral (AAV) vector that will be available soon. His comorbidities include 3 target joints, a prior 2-decade history of the hepatitis C virus (HCV) infection with stage 2 liver fibrosis, and HIV well controlled on antiretroviral medications. He wants to know what is known and unknown about hemophilia gene therapy.

Introduction: What are the current approaches for hemophilia gene therapy?

Clinical approaches in hemophilia gene therapy nearly exclusively use systemically delivered recombinant AAV (rAAV) vectors to target hepatocyte expression of an episomally maintained transgene expressed under a hepatocyte promoter (Table 1). The vector is engineered from naturally

occurring AAV, which is nonpathogenic in humans and replication defective. rAAV vectors are produced in cell culture using either HEK293 mammalian cells or baculovirus/Sf9 insect cell culture. The design and manufacturing of rAAV vectors was recently reviewed by Li and Samulski.¹

While hepatocytes are the natural site of factor IX (FIX) synthesis, liver sinusoidal endothelial cells are the predominate site of factor VIII (FVIII) synthesis.² Most HA rAAV vectors contain a B-domain deleted FVIII, FVIII-SQ transgene that retains full procoagulant function and accommodates AAV vector packaging constraints (4.7 kB).³ This exact amino acid sequence has been used for recombinant protein HA therapy for 2 decades (eg, Xyntha/ReFacto®, Pfizer) without evidence of increased risk of inhibitor formation. A single trial is using an alternative B-domain deleted FVIII, FVIII-V3 that consists of the SQ linker with 6 additional N-linked glycosylation sites to improve expression.^{4,5} All enrolling hemophilia B (HB) trials have adapted the use of the *FIX-Padua* (*FIX-R338L*) mutation, which is a naturally occurring missense mutation with approximately 8-fold greater specific activity relative to wild-type FIX.⁶⁻¹⁰

Table 1. Ongoing hemophilia A and B gene therapy trials

Sponsor	Clinicaltrials.gov	Manufacturing platform	Transgene	Capsid serotype	Dose (vg/kg)	Phase	Ref
HB							
UCL/St. Jude	NCT00979238	Mammalian	scFIX	AAV8	2×10 ¹¹ to 2×10 ¹²	1/2	16,45
Pfizer/Spark	NCT03681273 NCT03307980	Mammalian	ssFIX-R338L	SPK100	5×10 ¹¹	1/2, 3	9
uniQure	NCT02396342	Sl9 (insect)	ssFIX-R338L	AAV5	2×10 ¹³	3	8
Freeline	NCT03369444	Mammalian	scFIX-R338L	AAVS3	7.5×10 ¹¹ 9.5×10 ¹¹		10
HA							
BioMarin	NCT02576795 NCT03392974 NCT03370913	Sl9 (insect)	ssFVIII-SQ	AAV5	6×10 ¹³	1/2, 3	11,15
Sangamo/Pfizer	NCT03061201	Sl9 (insect)	ssFVIII-SQ	AAV6	3×10 ¹³	1/2, 3	18
Spark	NCT03003533 NCT03432520	Mammalian	ssFVIII-SQ	LK03	5×10 ¹¹ to 2×10 ¹²	1/2	14
UCL/St. Jude	NCT03001830	Mammalian	ssFVIII-V3	AAV8	6×10 ¹¹ to 6×10 ¹²	1/2	4,5
Bayer/Ultragenyx	NCT03588299	Mammalian	ssFVIII-SQ	AAVhu37	5×10 ¹² to 2×10 ¹³	1/2	58
Takeda	NCT03370172	Mammalian	ssFVIII-SQ	AAV8		1/2	

sc, self-complementary; ss, single stranded; UCL, University College London.

What are the short- and long-term rAAV safety considerations?

In the 300-fold range of rAAV vector doses (2×10¹¹ to 6×10¹³ vg/kg) thus far evaluated in hemophilia clinical trials, there have been no major safety concerns.

In the short term, a small number of subjects across 3 trials at rAAV vector doses ranging from 2×10¹² to 6×10¹³ vg/kg have experienced acute vector infusion reactions, including a single incidence of anaphylaxis and a couple of incidences of either myalgias, fever, and/or hypotension of unclear etiology that may be consistent with an innate immune response to rAAV¹¹⁻¹⁴; these events have not clearly correlated with clinical outcomes post vector.

Because AAV efficiently targets the liver, the bulk of safety considerations of systemic rAAV delivery have focused on hepatotoxicity. Within hemophilia trials, a single study outlined multimonth transaminase elevations post vector of unclear etiology that were not associated with liver dysfunction and resolved.^{11,15} Beyond this, most transaminase elevations observed in hemophilia trials occurred in the setting of an immune response to the rAAV capsid.^{9,14,16-18} This AAV capsid immune response is hypothesized to occur when CD8⁺ T cells recognize capsid peptides on the surface of transduced hepatocytes, triggering a cellular immune response that results in clearance of the transduced cells.¹⁹ While the AAV capsid immune response has not posed safety concerns in hemophilia, it has diminished or prevented efficacy in some trials.^{9,16-18} Close monitoring for a capsid immune response within the first 3 months or so post vector classically includes factor assays, liver function studies, and peripheral blood mononuclear cell interferon-γ anticapsid enzyme-linked immunosorbent spot assays that, when interpreted as consistent with a capsid immune response, trigger intervention with immune-modulating therapies to maintain transgene expression; glucocorticoids have been used predominantly,^{9,16-18} but other immune-modulating

agents have been used concurrently with glucocorticoids or in isolation to maintain transgene expression.^{10,13,14} Additional study is needed to understand which regimen best maintains transgene expression. Further still, because some studies have employed glucocorticoids while simultaneously reporting that participants had no cellular immune response, additional study is needed to distinguish a cellular immune response vs other potential etiologies for elevated transaminases.^{12,13,15}

While safety in hemophilia has been excellent, rAAV vectors developed for other monogenic disorders suggest there may be dose-limiting hepatic toxicities with systemic rAAV vector doses >1×10¹⁴ vg/kg. Specifically, Zolgensma (Novartis), an rAAV9 vector and the first licensed systemic rAAV vector, delivered at a dose of 1.1×10¹⁴ vg/kg demonstrated not only remarkable efficacy for spinal muscular atrophy but also evidence of hepatic toxicity in clinical trial such that it was approved with a boxed safety warning for acute liver injury. Subsequently, a case report outlined 2 children presenting 6 to 8 weeks post Zolgensma infusion with acute liver failure that resolved with glucocorticoid intervention.²⁰ While the exact etiology is unclear, the timing post vector, liver biopsy specimens demonstrating CD8⁺ T-cell infiltration, and responsiveness to steroids suggest an immune-mediated process that is possibly consistent with an rAAV capsid immune response. These observations support the possibility that the rAAV capsid immune response has safety implications and support using the lowest possible rAAV vector dose to minimize safety concerns and maximize efficacy. Additionally, a study using an rAAV8 vector for X-linked myotubular myopathy reported that 3 of 17 boys in its 3.5×10¹⁴ vg/kg dose cohort developed liver failure approximately 6 weeks post vector and ultimately succumbed to complications.²¹ The gene therapy community eagerly awaits a composite analysis of these observations. Nonetheless, these tragic events support possible dose-limiting toxicities of systemic rAAV vectors at doses >1×10¹⁴

vg/kg, which are >1.6- to 500-fold higher than the rAAV doses used in hemophilia.

Finally, dorsal root ganglia (DRG) toxicity has recently emerged as a potential short- or long-term safety concern. Overwhelmingly, these observations are histological findings in large-animal models. Specifically, a meta-analysis of NHP data (n=33 vectors, n=256 NHPs) by a single laboratory following heterogeneous routes of administration (intrathecal, intracisternal magna, and systemic), vector doses, AAV capsids, and transgenes reported no or mild histological evidence of DRG toxicity in most animals that appeared to be dose dependent and more common with direct central nervous system administration.²² Importantly, clinical symptomatology in only 2 animals was attributed to DRG toxicity. Subsequent NHP studies by the same group suggested DRG toxicity may be related to an unfolded protein response abrogated when inhibiting transgene expression in the DRG with, as yet, unclear clinical translation.²³ Thus far, in the full cadre of clinical AAV trials spanning 3 decades, various routes of administration for a variety of disorders, and an approximate 1000-fold range of rAAV vector doses,²⁴ no DRG toxicity clinical symptoms have been observed outside of a single recently reported case. A trial participant who received an rAAVrh10 vector (a capsid not currently used for hemophilia) by intrathecal injection to deliver microRNA for amyotrophic lateral sclerosis presented with symptoms and imaging consistent with DRG toxicity; symptoms diminished with glucocorticoid and supportive care intervention but had not been completely ameliorated at the time of publication.²⁵ Whether DRG toxicity will be route of administration, capsid, or rAAV vector dose dependent and/or a potential rAAV vector platform toxicity is unclear.

The major identified long-term safety concerns of systemic rAAV vectors are the risks of hepatotoxicity and genotoxicity. Direct sequencing data in animals and humans demonstrate that AAV integration can occur and has a propensity for sites of active transcription.²⁶⁻³⁰ Data in neonatal mice demonstrated AAV integration that disrupted the murine *Rian* locus (that does not have a human ortholog) and resulted in near 100% penetrance of hepatocellular carcinoma (HCC).²⁹ A subsequent murine study demonstrated that the risk of HCC correlated directly with the rAAV vector dose and degree of cellular division and that hepatocyte-specific enhancer and promoter elements protected against oncogenesis, while non-hepatocyte-specific elements increased HCC risk.²⁷ This would thus support using the lowest possible effective vector dose with hepatocyte-specific promoter and enhancer elements to minimize genotoxicity risk. Recently, studies in HA dogs followed for 10 years post rAAV vector demonstrated evidence of clonal proliferation at sites of AAV integration in liver tissue without tumorigenesis but were the first large-animal data to support the theoretical risk of genotoxicity.³⁰

Nearly all severe hemophilia patients >40 years of age contracted iatrogenic hepatitis B virus and/or HCV, which are known risk factors for HCC such that the hemophilia population has an increased risk of HCC relative to the general population.³¹ Published 15-year follow-up data of the first participants to receive a systemic rAAV vector (HB subjects) demonstrated no evidence of long-term hepatotoxicity, but the report is limited in interpretation due to small cohort size and the absence of detectable transgene expression or direct sequencing data to confirm the persistence of transduced hepatocytes.³² A single clinical trial participant in an HB trial with prior long-term HCV

Similar inactivation

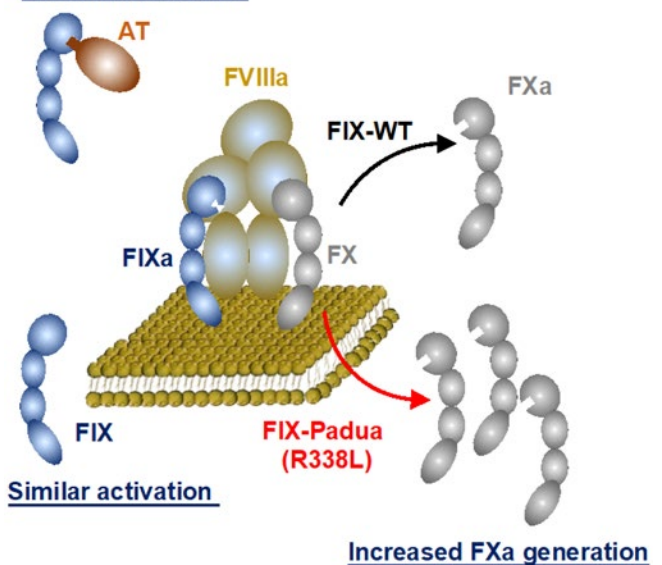


Figure 1. Biochemical characterization of *FIX-Padua* (*FIX-R338L*). Data support that *FIX-R338L* requires factor VIIIa (FVIIIa) cofactor function for enzymatic activity to generate factor Xa (FXa) from FX and is similarly activated for factor XIa (FIXa) and inactivated by antithrombin (AT). Modified with permission from Samelson-Jones et al.⁷

infection and other risk factors for HCC developed HCC approximately 1 year following vector infusion. Recently presented tumor sequencing data demonstrated the expected random, low-frequency AAV integrations without evidence of clonality as well as well-described HCC-associated genetic changes, suggesting that rAAV vector administration did not contribute to HCC development.³³ There is thus far no clinical evidence to suggest that systemic rAAV vector delivery is a risk factor for the development of HCC. Ultimately, cohorts outside the adult hemophilia population may more closely mimic the risk factors identified in mice for the development of HCC post systemic rAAV vector infusion. Specifically, infants and toddlers (who have inherent greater degrees of cellular division than adults) who receive higher systemic rAAV vector doses that contain ubiquitous promoters will be important populations to follow to evaluate long-term AAV genotoxicity.

What are the transgene safety considerations?

The generally identified safety concerns related to transgene expression include an immunological response to the expressed protein and direct transgene toxicity. Taking the latter first, epidemiological studies have outlined that supraphysiologic FIX:C has a moderate, independent risk of venous thrombosis (odds ratio, 1.8-4.0) relative to supraphysiologic FVIII:C (OR, 8.8-21.3).³⁴ Biochemical data support that *FIX-R338L* function, like wild-type FIX, requires FVIIIa cofactor function for enzymatic activity and is similarly activated by FIXa and inactivated by antithrombin (Figure 1), supporting that *FIX-R338L* is regulated the same as wild-type *FIX*⁷ and is not inherently not prothrombotic. Consistent with this, observed thrombotic events in mice expressing *wt-FIX* or *FIX-R338L* correlated with the degree of supratherapeutic FIX:C,

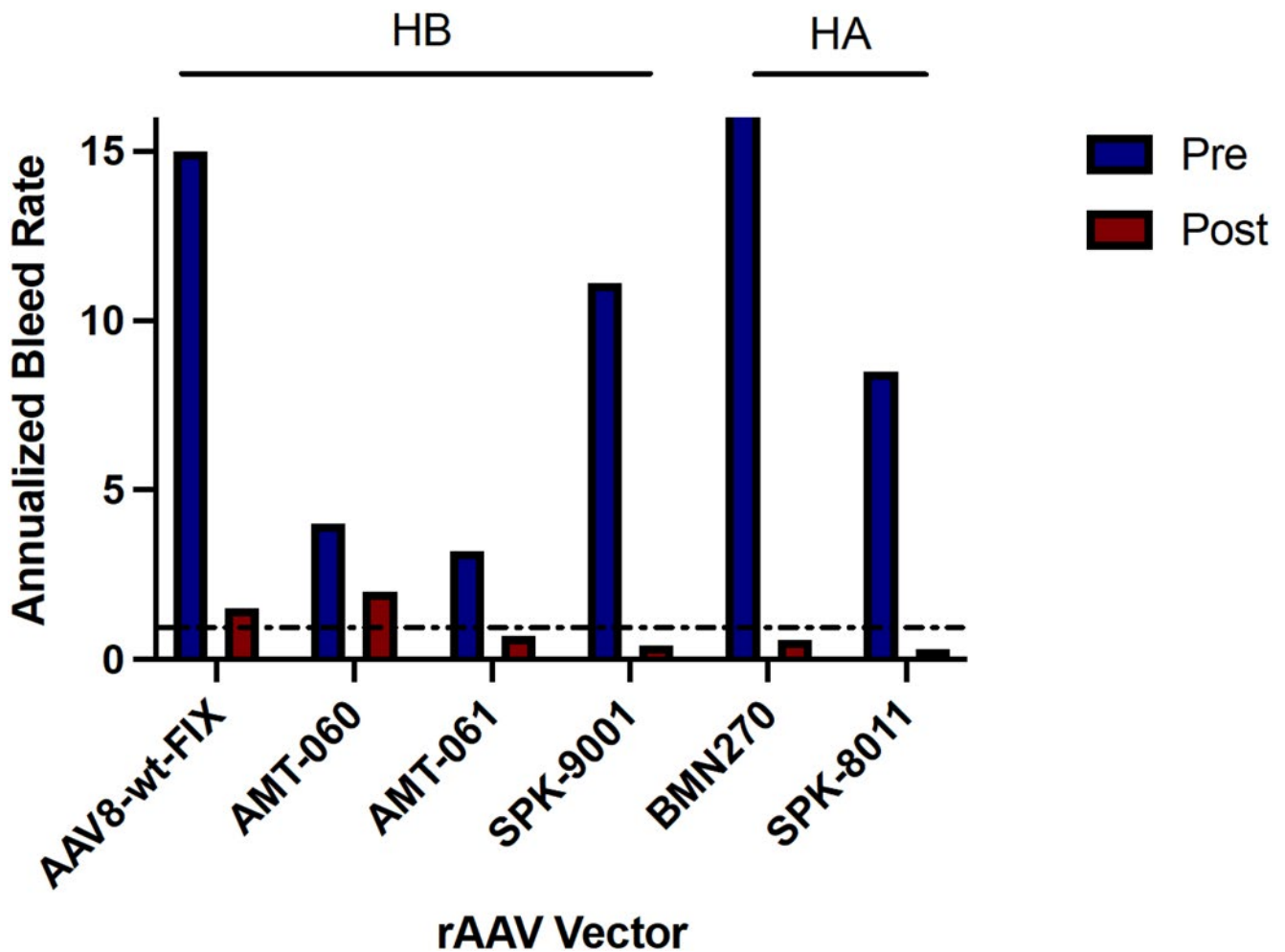


Figure 2. Aggregated reported annualized bleeding rate data for hemophilia A and B clinical trials before (blue) and after (maroon) receiving the described recombinant AAV vectors. The dotted line denotes an annualized bleeding rate of 1. AAV8-wt-FIX, also known as scAAV2/8-LP1-hFIXco. Data from Nathwani et al.⁴⁵ AMT-060 data from Miesbach et al.¹² AMT-061, now etranocogene dezaparovec, data from Pipe et al.⁸ SPK-9001, now fidancogene elaparovec, data from George et al.⁹ BMN270, now valoctocogene roxaparovec, data from Pasi et al.¹¹ SPK-8011 data from George et al.¹⁴

independent of the expressed transgene.³⁵ Concurrently, HB gene therapy trials using the *FIX-R338L* transgene achieved FIX:C in the range of mild or normal HB without safety concerns. A single *FIX-R338L* trial reported thrombosis in a subject who achieved supraphysiologic FIX:C with multiple prothrombotic comorbidities (obesity, kidney failure with an arteriovenous fistula).¹⁰ This observation when paired with epidemiological data of supraphysiologic factor activity and venous thrombosis risk underscores the importance of maintaining expression within a therapeutic window. In addition to thrombosis risk, the expression of FVIII or *FIX-R388L* could impart direct cellular toxicity, which is one possible cause of the unexpected declining FVIII expression observed in the first HA gene therapy trial detailed below.

While multiple immunological responses to the expressed transgene are possible, the most concerning is the development of inhibitory antibodies, which have not been observed in clinical trial. However, current trial enrollment includes patients who are the least likely to develop an inhibitor (eg, participants with prior or current inhibitors are excluded and must have >50-150

factor exposures). Nonetheless, inhibitor risk post gene therapy is generally thought to be unlikely because of small- and large-animal data that demonstrate the ability of hepatocyte-directed gene therapy to induce tolerance to FVIII, FIX, or *FIX-R338L*.³⁵⁻³⁷ These animal studies show that future gene transfer may ultimately be an effective means for FVIII tolerance induction in HA patients with inhibitors; initiating clinical trials for tolerance induction was recently supported by consensus recommendations following a National Heart, Lung, and Blood Institute State of the Science of FVIII inhibitors.³⁸

What are the preliminary efficacy data?

The approximate homogeneous and remarkable phenotypic amelioration that has been observed with heterogeneous transgene-derived factor activity supporting factor activity itself should not be the sole distinguishing feature of clinical programs (Figure 2). Specifically, currently available data are consistent with HA natural history studies that suggest factor activity >10% ameliorates spontaneous bleeding; however, higher factor

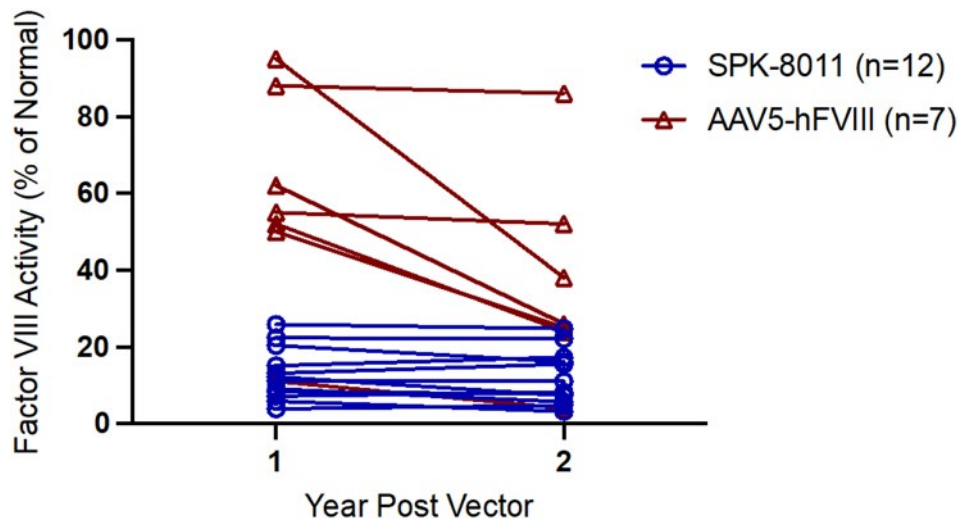


Figure 3. FVIII activity 1 and 2 years post vector infusion in trial participants who received a therapeutic vector dose (6×10^{13} vg/kg of AAV5-hFVIII vs 5×10^{11} to 2×10^{12} vg/kg of SPK 8011) that maintained expression and were followed >2 years post vector. FVIII activity is reported by 1-stage assay for SPK-8011 participants and CSA in participants who received AAV5-hFVIII. Data for AAV5-hFVIII are from Pasi et al¹¹ and data for SPK-8011 are from George et al.¹⁴

activity is likely required among patients with marked preexisting joint disease. Most trials report an unpredictable range of expression of 5- to 10-fold variability (some trials up to 50- to 100-fold) such that it is not currently possible to target expression based on individual patient needs.^{9,10,11,13,14,39} Correspondingly, an up to 7-fold variability in transduction was demonstrated in a chimeric murine model of human hepatocytes.⁴⁰ This degree of variable expression in a clonal mouse population is consistent with the multiple variables that exist from vector infusion to transgene expression¹ that may necessitate tolerance for a range of transgene expression, albeit hopefully to a lesser degree than has thus far been reported. This underscores the need for an accurate measurement of in vivo activity of transgene hemostatic function to define the range of tolerable safe and efficacious expression.

In HA and HB efforts, there is a discrepancy in transgene-derived factor activity by one-stage assay (OSA) and chromogenic assay (CSA). Transgene-derived FVIII:C by OSA measures approximately 1.6-fold higher than CSA in humans^{15,39-42} and mice,⁴³ which differs from the experience with recombinant protein products of the same amino acid sequence. Which assay correlates with in vivo hemostatic benefit is unclear. Current trials target exogenous FVIII expression, and whether this mildly alters biochemical properties of the protein is hypothesized (eg, changes in von Willebrand factor affinity or FVIII cleavage by thrombin or factor Xa) but not determined. Similarly, FIX-R338L determined that FIX:C measures higher by OSA vs CSA FIX:C; however, unlike HA, this observation is also maintained with recombinant FIX-R338L and consistent with the available biochemical understanding of FIX-R338L.⁴⁴ Additionally, unlike HA trials, OSA determined that FIX:C from FIX-R338L expression differs by initiating reagents.⁴⁴

How long will the therapy last?

Thus far, clinical trial efforts in HB gene transfer mirror hemophilia canine models of rAAV-mediated gene transfer that demonstrated no decline in FVIII or FIX expression for up to

8 years post gene transfer.^{16,30,42,45,46} In contrast, the first phase 1/2 HA gene therapy trial of valoctocogene roxaparvovec at a dose of 6×10^{13} vg/kg demonstrated an average loss of approximately 40% of transgene expression from year 1 to year 2 post vector ($n=7$)^{11,15}; similar observations were observed among the 17 phase 3 participants administered the same vector dose who were followed for >2 years.¹³ Available phase 1/2 data of participants 5 years post valoctocogene roxaparvovec demonstrated a continued, but lesser, decline in FVIII expression in years 2 to 5.⁴⁷ In contrast, a phase 1/2 study of SPK-8011 for HA at vector doses 5×10^{11} to 2×10^{12} vg/kg demonstrated different pharmacokinetics of expression such that the 12 participants with sustained expression outside the cellular immune response and followed for >2 years demonstrated no apparent decline in expression from year 1 to year 2 (Figure 3).¹⁴ Some of these subjects have been followed for up to 4 years and demonstrate generally stable FVIII expression after 1 year post vector; these preliminary SPK-8011 data generally support the current strategy of targeting hepatocytes for multiyear stable FVIII expression. Given that the transgene is maintained episomally, some degree of declining expression over multiyear follow-up may be anticipated, although it is unclear how rapidly, but the mechanism of loss or the more marked decline in FVIII expression from year 1 to year 2 observed with valoctocogene roxaparvovec is unknown. Possible hypotheses include (1) an unfolded protein response, which has been described in mammalian expression systems of recombinant FVIII production and in mouse models of supratherapeutic but not low levels of FVIII expression post rAAV⁴⁸⁻⁵⁰; (2) promoter silencing; (3) an ongoing undetected/unmitigated immune response to AAV or the transgene; or (4) failure to form stable concatemeric episomal DNA due to properties of the vector.

What do AAV neutralizing antibodies mean?

AAV neutralizing antibodies (NAb) are incorporated into most eligibility criteria in hemophilia and other disease models

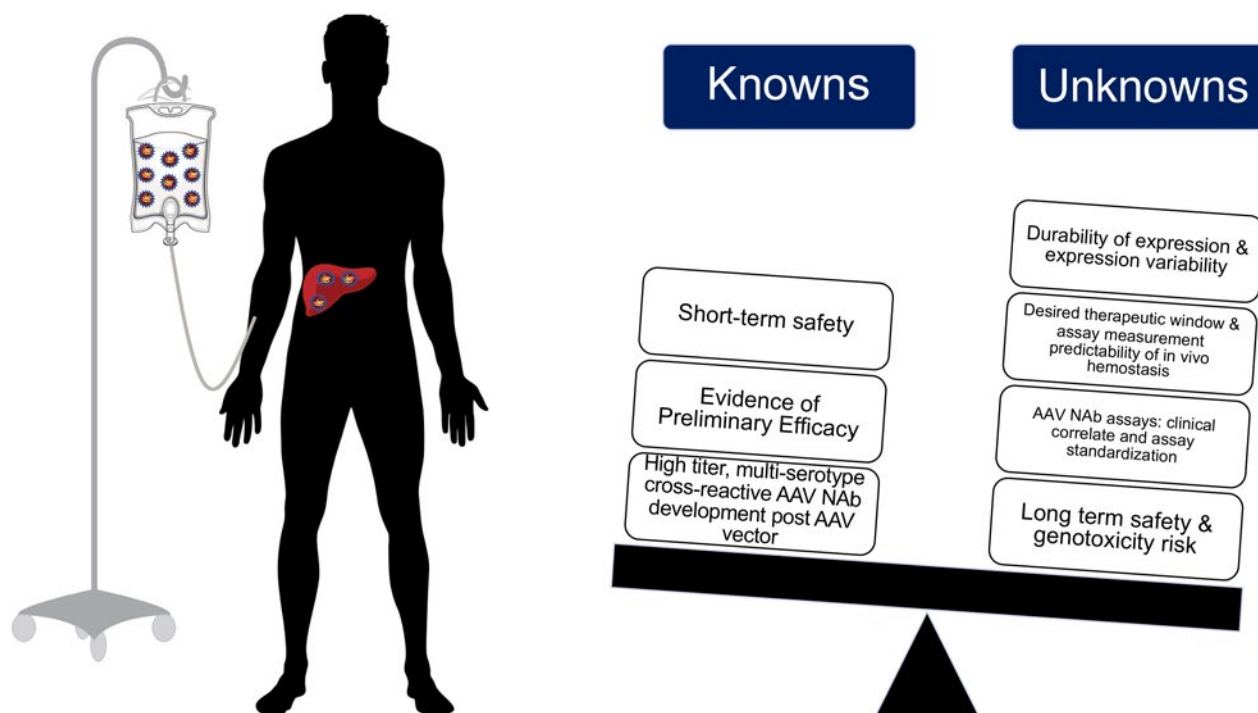


Figure 4. Summary of known and unknown variables for recombinant AAV gene editing clinical efforts for in hemophilia A and B.

using systemic AAV vectors. While results of the assay have an impact on eligibility, the assays are not standardized, can yield highly variable results depending on the assay methods (eg, transduction vs enzyme-linked immunosorbent assay based, employed multiplicity of infection, etc), and do not always have a clear correlation with clinical outcome. Nonetheless, most available data support the fact that high-titer AAV NAb preclude target tissue transduction of a systemically administered rAAV vector and thus limit efficacy. A single AAV5 trial using a dose of 2×10^9 vg/kg is enrolling participants independent of AAV NAb status and has demonstrated transgene expression in all participants except the participant with the highest measured AAV NAb ($>1:3000$).^{8,13} These data challenge existing notions that would otherwise predict preexisting AAV NAb to preclude efficacy. Details of the AAV NAb, thus far unreported, are necessary to interpret the data and whether these data are specific to rAAV5 and/or the vector dose used or may be partially explained by methods used to perform the AAV NAb.

Following rAAV vector administration, patients universally develop persistent, multiserotype cross-reactive AAV NAb that available data suggest would preclude repeat rAAV vector efficacy.^{32,51} This underscores that patients interested in gene therapy must recognize that the current state of clinical development likely only permits 1 systemic rAAV vector infusion. Thus, outlining the current known and unknown information regarding rAAV gene therapy for hemophilia is essential to provide true informed consent for both clinical trials and anticipated soon-to-be licensed vectors (Figure 4); this will be necessary to avoid "buyer's remorse" should one receive an rAAV vector that is ultimately found to be inferior to another rAAV vector.

What other approaches are in development?

Safely achieving sustained, stable, and predictable FVIII or FIX expression, even in the presence of an immune response, is an unrealized goal of hemophilia gene therapy. Beyond gene addition approaches, gene editing using lipid nanoparticles to deliver mRNA encoding Cas9 and gRNA and a donor FIX cDNA template via an rAAV vector to knockin *F9* into the albumin locus have demonstrated normal levels of FIX expression in an NHP model.⁵² In addition, lentiviral vectors for either systemic infusion,⁵³ ex vivo transduction of hematopoietic stem cells,⁵⁴⁻⁵⁶ or induced pluripotent stem cells are being pursued in preclinical investigation and early-phase clinical trial (clinicaltrials.gov; NCT03818763).⁵⁷

Will hemophilia gene therapy ever be mainstream?

Building on the remarkable successes the field has seen over the past approximately 5 years will be predicated on the thoughtful investigation of questions that have emerged from clinical trials. Optimism in the field will continue to progress, buoyed by 3 general strengths: (1) a strong understanding of the molecular and biochemical basis of HA and HB, (2) an organized patient advocacy and physician provider network that fosters basic research and clinical trials, and (3) a rapidly expanding knowledge base in gene therapy with significant financial investment. Collectively, these strengths ensure that gene therapy will fulfill its promise to alter the paradigm of hemophilia care.

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Conflict-of-interest disclosure

Lindsey A. George: scientific advisory board member: STRM.Bio; data safety monitoring committee member: AvroBio; consultancy: Intellia, BiomarIn, Pfizer, Bayer.

Off-label drug use

Lindsey A. George: glucocorticoids were discussed.

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References

- Li C, Samulski RJ. Engineering adeno-associated virus vectors for gene therapy. *Nat Rev Genet*. 2020;21(4):255-272.
- Shahani T, Covens K, Lavend'homme R, et al. Human liver sinusoidal endothelial cells but not hepatocytes contain factor VIII. *J Thromb Haemost*. 2014;12(1):36-42.
- Pittman DD, Alderman EM, Tomkinson KN, Wang JH, Giles AR, Kaufman RJ. Biochemical, immunological, and in vivo functional characterization of B-domain-deleted factor VIII. *Blood*. 1993;81(11):2925-2935.
- McIntosh J, Lenting PJ, Rosales C, et al. Therapeutic levels of FVIII following a single peripheral vein administration of rAAV vector encoding a novel human factor VIII variant. *Blood*. 2013;121(17):3335-3344.
- Nathwani AC, Tuddenham E, Chowdhury P, McIntosh J, et al. GO-8: preliminary results of a phase I/II dose escalation trial of gene therapy for hemophilia A using a novel human factor VIII variant. *Blood*. 2018;132(suppl 1):489.
- Simioni P, Tormene D, Tognin G, et al. X-linked thrombophilia with a mutant factor IX (factor IX Padua). *N Engl J Med*. 2009;361(17):1671-1675.
- Samelson-Jones BJ, Finn JD, George LA, Camire RM, Arruda VR. Hyperactivity of factor IX Padua (R338L) depends on factor VIIIa cofactor activity. *JCI Insight*. 2019;4(14):e128683.
- Pipe SW, Recht M, Key NS, et al. HOPE-B investigators 52 week efficacy and safety of etranacogene dezaparovec in adults with severe or moderate-severe hemophilia B: data from the phase 3 HOPE-B gene therapy trial [abstract]. *Res Pract Thromb Haemost*. 2021;5(suppl 2).
- George LA, Sullivan SK, Giermasz A, et al. Hemophilia B gene therapy with a high-specific-activity factor IX variant. *N Engl J Med*. 2017;377(23):2215-2227.
- Chowdhury P, Makris M, Evans G, et al. A novel adeno associated virus (AAV) gene therapy (FLT180a) achieves normal FIX activity levels in severe hemophilia B (HB) patients (B-AMAZE study). *Res Pract Thromb Haemost*. 2020;4(suppl 2):17.
- Pasi KJ, Rangarajan S, Mitchell N, et al. Multiyear follow-up of AAV5-hFVIII-SQ gene therapy for hemophilia A. *N Engl J Med*. 2020;382(1):29-40.
- Miesbach W, Meijer K, Coppens M, et al. Gene therapy with adeno-associated virus vector 5-human factor IX in adults with hemophilia B. *Blood*. 2018;131(9):1022-1031.
- Ozelo M, Mahlangu J, Pasi K, Giermasz A, et al; GENER8-1 Trial Group. Efficacy and safety of valoctocogene roxaparovec adeno-associated virus gene transfer for severe hemophilia A: results from the phase 3 GENER8-1 trial [abstract]. *Res Pract Thromb Haemost*. 2021;5(suppl 1). Abstract OC 26.1.
- George LA, Monahan PE, Eyster ME, Sullivan SK, et al. Phase I/II trial of SPK-8011: stable and durable FVIII expression after AAV gene transfer for hemophilia A [abstract]. *Res Pract Thromb Haemost*. 2021;5(suppl 1). Abstract OC 67.2.
- Rangarajan S, Walsh L, Lester W, et al. AAV5-factor VIII gene transfer in severe hemophilia A. *N Engl J Med*. 2017;377(26):2519-2530.
- Nathwani AC, Tuddenham EG, Rangarajan S, et al. Adenovirus-associated virus vector-mediated gene transfer in hemophilia B. *N Engl J Med*. 2011;365(25):2357-2365.
- Manno CS, Pierce GF, Arruda VR, et al. Successful transduction of liver in hemophilia by AAV-factor IX and limitations imposed by the host immune response. *Nat Med*. 2006;12(3):342-347.
- Konkle BA, Walsh CE, Escobar MA, et al. BAX 335 hemophilia B gene therapy clinical trial results: potential impact of CpG sequences on gene expression. *Blood*. 2021;137(6):763-774.
- Mingozzi F, Maus MV, Hui DJ, et al. CD8(+) T-cell responses to adeno-associated virus capsid in humans. *Nat Med*. 2007;13(4):419-422.
- Feldman AG, Parsons JA, Dutmer CM, et al. Subacute liver failure following gene replacement therapy for spinal muscular atrophy type 1. *J Pediatr*. 2020;225(October):252-258.e1258e1.
- Shieh PB, Bönemann CG, Müller-Felber W, et al. Re: "Moving Forward After Two Deaths in a Gene Therapy Trial of Myotubular Myopathy" by Wilson and Flotte. *Hum Gene Ther*. 2020;31(15-16):787.
- Hordeaux J, Buza EL, Dyer C, et al. Adeno-associated virus-induced dorsal root ganglion pathology. *Hum Gene Ther*. 2020;31(15-16):808-818.
- Hordeaux J, Buza EL, Jeffrey B, et al. MicroRNA-mediated inhibition of transgene expression reduces dorsal root ganglion toxicity by AAV vectors in primates. *Sci Transl Med*. 2020;12(569):eaba9188.
- Wang D, Tai PWL, Gao G. Adeno-associated virus vector as a platform for gene therapy delivery. *Nat Rev Drug Discov*. 2019;18(5):358-378.
- Mueller C, Bery JD, McKenna-Yasek DM, et al. SOD1 suppression with adeno-associated virus and microRNA in familial ALS. *N Engl J Med*. 2020;383(2):151-158.
- Nault JC, Datta S, Imbeaud S, et al. Recurrent AAV2-related insertional mutagenesis in human hepatocellular carcinomas. *Nat Genet*. 2015;47(10):1187-1193.
- Chandler RJ, LaFave MC, Varshney GK, et al. Vector design influences hepatic genotoxicity after adeno-associated virus gene therapy. *J Clin Invest*. 2015;125(2):870-880.
- Nakai H, Montini E, Fuess S, Storm TA, Grompe M, Kay MA. AAV serotype 2 vectors preferentially integrate into active genes in mice. *Nat Genet*. 2003;34(3):297-302.
- Donsante A, Miller DG, Li Y, et al. AAV vector integration sites in mouse hepatocellular carcinoma. *Science*. 2007;317(5837):477.
- Nguyen GN, Everett JK, Kafle S, et al. A long-term study of AAV gene therapy in dogs with hemophilia A identifies clonal expansions of transduced liver cells. *Nat Biotechnol*. 2021;39(1):47-55.
- Mazepa MA, Monahan PE, Baker JR, Riske BK, Soucie JM; US Hemophilia Treatment Center Network. Men with severe hemophilia in the United States: birth cohort analysis of a large national database. *Blood*. 2016;127(24):3073-3081.
- George LA, Ragni MV, Rasko JEJ, et al. Long-term follow-up of the first in human intravascular delivery of AAV for gene transfer: AAV2-hFIX16 for severe hemophilia B. *Mol Ther*. 2020;28(9):2073-2082.
- Schmidt M, Foster GM, Coppens M, Thomsen H, et al. Liver safety case report from the phase 3 HOPE-B gene therapy trial in adults with hemophilia B [abstract]. *Res Pract Thromb Haemost*. 2021;5(suppl 1). Abstract OC 67.4.
- Rietveld IM, Lijfering WM, le Cessie S, et al. High levels of coagulation factors and venous thrombosis risk: strongest association for factor VIII and von Willebrand factor. *J Thromb Haemost*. 2019;17(1):99-109.
- Crudele JM, Finn JD, Siner JI, et al. AAV liver expression of FIX-Padua prevents and eradicates FIX inhibitor without increasing thrombogenicity in hemophilia B dogs and mice. *Blood*. 2015;125(10):1553-1561.
- Finn JD, Ozelo MC, Sabatino DE, et al. Eradication of neutralizing antibodies to factor VIII in canine hemophilia A after liver gene therapy. *Blood*. 2010;116(26):5842-5848.
- Mingozzi F, Hasbrouck NC, Basner-Tschakarjan E, et al. Modulation of tolerance to the transgene product in a nonhuman primate model of AAV-mediated gene transfer to liver. *Blood*. 2007;110(7):2334-2341.
- Ragni MV, George LA; for Members of Working Group 1, the National Heart, Lung, and Blood Institute State of the Science Workshop on Factor VIII Inhibitors: Generating a National Blueprint for Future Research. The national blueprint for future factor VIII inhibitor clinical trials: NHLBI State of the Science (SOS) Workshop on factor VIII inhibitors. *Haemophilia*. 2019;25(4):581-589.
- Konkle BA, Stine K, Visweshwar N, Harrington TJ, et al. Updated follow-up of the Alta study, a phase 1/2, open-label, adaptive, dose-ranging study to assess the safety and tolerability of SB-525 gene therapy. *Blood*. 2019;134(suppl 1):2060.
- Zou C, Vercauteren KOA, Michailidis E, et al. Experimental variables that affect human hepatocyte AAV transduction in liver chimeric mice. *Mol Ther Methods Clin Dev*. 2020;18(September):189-198.
- Rosen S, Tiefenbacher S, Robinson M, et al. Activity of transgene-produced B-domain-deleted factor VIII in human plasma following AAV5 gene therapy. *Blood*. 2020;136(22):2524-2534.
- Nathwani AC, Tuddenham E, Chowdhury P, et al. Adeno-associated mediated gene transfer for hemophilia B: 8 year follow up and impact of removing "empty viral particles" on safety and efficacy of gene transfer. *Blood*. 2019;132(suppl 1):491.

43. Sternberg AR, Davidson RJ, Samelson-Jones BJ, George L. In vitro and in vivo models to understand one-stage and chromogenic factor VIII activity assay discrepancy of hepatocyte-derived factor VIII [abstract]. *Res Pract Thromb Haemost*. 2021;5(suppl 1). Abstract OC 75.3.
44. Robinson MM, George LA, Carr ME, et al. Factor IX assay discrepancies in the setting of liver gene therapy using a hyperfunctional variant factor IX-Padua. *J Thromb Haemost*. 2021;19(5):1212-1218.
45. Nathwani AC, Reiss UM, Tuddenham EG, et al. Long-term safety and efficacy of factor IX gene therapy in hemophilia B. *N Engl J Med*. 2014;371(21):1994-2004.
46. Niemeyer GP, Herzog RW, Mount J, et al. Long-term correction of inhibitor-prone hemophilia B dogs treated with liver-directed AAV2-mediated factor IX gene therapy. *Blood*. 2009;113(4):797-806.
47. Pasi KR, Rangarajan S, TM Robinson, Lester W, et al. Hemostatic response is maintained for up to 5 years following treatment with valoctocogene roxaparvovec, an AAV5-hFVIII-SQ gene therapy for severe hemophilia A [abstract]. *Res Pract Thromb Haemost*. 2021;5(suppl 1). Abstract OC 671.
48. Lange AM, Altynova ES, Nguyen GN, Sabatino DE. Overexpression of factor VIII after AAV delivery is transiently associated with cellular stress in hemophilia A mice. *Mol Ther Methods Clin Dev*. 2016;3(28 September):16064.
49. Zolotukhin I, Markusic DM, Palaschak B, Hoffman BE, Srikanthan MA, Herzog RW. Potential for cellular stress response to hepatic factor VIII expression from AAV vector. *Mol Ther Methods Clin Dev*. 2016;3(28 September):16063.
50. Poothong J, Pottekat A, Siirin M, et al. Factor VIII exhibits chaperone-dependent and glucose-regulated reversible amyloid formation in the endoplasmic reticulum. *Blood*. 2020;135(21):1899-1911.
51. Long BR, Veron P, Kuranda K, et al. Early phase clinical immunogenicity of valoctocogene roxaparvovec, an AAV5-mediated gene therapy for hemophilia A. *Mol Ther*. 2021;29(2):597-610.
52. Huang H MEC, Bialek P, Wang C, et al. CRISPR/Cas9-mediated targeted insertion of human F9 achieves therapeutic circulating protein levels in mice and non-human primates [abstract]. *Mol Ther*. 2019;27(April):7. Abstract 11.
53. Cantore A, Ranzani M, Bartholomae CC, et al. Liver-directed lentiviral gene therapy in a dog model of hemophilia B. *Sci Transl Med*. 2015;7(277):277ra28.
54. Du LM, Nurden P, Nurden AT, et al. Platelet-targeted gene therapy with human factor VIII establishes haemostasis in dogs with haemophilia A. *Nat Commun*. 2013;4(November):2773.
55. Shi Q, Wilcox DA, Fahs SA, et al. Factor VIII ectopically targeted to platelets is therapeutic in hemophilia A with high-titer inhibitory antibodies. *J Clin Invest*. 2006;116(7):1974-1982.
56. Greene TK, Wang C, Hirsch JD, et al. In vivo efficacy of platelet-delivered, high specific activity factor VIII variants. *Blood*. 2010;116(26):6114-6122.
57. Follenzi A, Benten D, Novikoff P, Faulkner L, Raut S, Gupta S. Transplanted endothelial cells repopulate the liver endothelium and correct the phenotype of hemophilia A mice. *J Clin Invest*. 2008;118(3):935-945.
58. Pipe S, Becka M, Detering E, Vanevski K, Lissitchkov T. First-in-human gene therapy study of AAVhu37 capsid vector technology in severe hemophilia A. *Blood*. 2019;134(suppl 1):4630.

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