



MALT Lymphomas

Franco Cavalli, Peter G. Isaacson, Randy D. Gascoyne, and Emanuele Zucca

This review addresses the biology and the treatment of lymphomas arising from mucosa-associated lymphoid tissue (MALT). This entity, first described in 1983, represents about 8% of all non-Hodgkin's lymphomas and was recently re-classified as "extranodal marginal zone lymphomas of MALT-type." The term marginal zone lymphoma (MZL) encompasses the three closely related lymphoma subtypes of nodal, primary splenic and extranodal lymphomas of MALT type: the latter represent the vast majority of MZL. These lymphomas arise at different anatomic sites, are composed of mature B-cells lacking expression of CD5 and CD10, often present with overlapping morphologic features, but typically quite distinct clinical behaviors. Only very recently cytogenetic/molecular genetic observations have underlined the distinctiveness of these three lymphoid neoplasms, which in both the R.E.A.L. and WHO-classifications are included in the general term of MZL. MALT lymphomas arise in numerous extranodal sites, but gastric MALT lymphoma is the most common and best studied and is, therefore, the paradigm for the group as a whole.

Dr. Isaacson describes the principal histological features of these lymphomas, including criteria to distinguish this entity from other small B-cell

lymphomas. Several lines of evidence suggest that gastric lymphoma arises from MALT acquired as the result of a *H. pylori* infection. However, at least 1/3 of cases do not respond to eradication of *H. pylori*. Very recent data suggest that both t(11;18) (q21;q21) and bcl10 nuclear expression are associated with failure to respond to this treatment.

Dr. Gascoyne discusses the biologic function of proteins deregulated through the different translocations, which play a role in pathogenesis of MALT lymphomas, emphasizing particularly their influence in disrupting the apoptotic pathway.

Dr. Zucca reviews findings suggesting that MALT lymphoma is an antigen driven neoplasm. He also presents specific guidelines for treatment of gastric lymphomas trying to shed some light on the amazingly inconsistent and confusing data in the literature.

Taking advantage on the more than 300 non-gastric MALT lymphomas collected by the International Extranodal Lymphoma Study Group (ILES), Dr. Cavalli compares gastric lymphomas with those arising in many other sites.

Overall, the data presented in this session will underline the fact, that MALT lymphomas are characterized by some unique biological properties.

I. THE PATHOLOGY OF GASTRIC MALT LYMPHOMA AND ITS RESPONSE TO ERADICATION OF *HELICOBACTER PYLORI*

*Peter G. Isaacson, MBChB**

The observation that the histology of certain extranodal lymphomas was related to mucosa associated lymphoid tissue (MALT) rather than that of peripheral lymph nodes was first made by Isaacson and Wright in 1983.¹ These authors noted similarities between the histology of the condition known as immunoproliferative small intesti-

nal disease (IPSID), a subtype of primary intestinal B-cell lymphoma, and primary low-grade gastric B-cell lymphoma. However, their histology differed from that of comparable low-grade nodal B-cell lymphomas in that their overall structure and cytology resembled those of MALT rather than lymph nodes. These observations were later extended to include other extranodal low-grade B-cell lymphomas, especially those of salivary gland, lung and thyroid.² The collective histological features of these lymphomas recapitulated the features of the principal B-cell component of MALT as exemplified by the Peyer's patch.^{3,4} MALT lymphomas, recently re-classified as "extranodal marginal zone lymphomas of MALT-type,"⁵ arise in numerous extranodal sites, but gastric MALT lymphoma is the commonest and best studied and is, therefore, the paradigm for the group as a whole.

*University College London Medical School, Department of Histopathology, Rockefeller Building, University Street, London WC1E 6JJ, United Kingdom

Pathology

Gastric lymphoma, like carcinoma, most often involves the antrum but may occur in any part of the stomach. The histological features of gastric MALT lymphoma, which by definition is a low grade lymphoma, closely simulate those of the Peyer's patch.^{6,7} The lymphoma infiltrates around reactive follicles in the region corresponding to the Peyer's patch marginal zone, spreading diffusely into the surrounding mucosa. The tumor cells may resemble germinal center centrocytes (centrocyte-like cells) or small lymphocytes, or assume a monocytoid appearance. Scattered transformed blasts are common and may cause problems in grading the lymphoma (see below). An important feature of MALT lymphomas is the presence of lymphoepithelial lesions formed by invasion of individual glands by aggregates of lymphoma cells. Certain additional histological features suggest that the cells of low-grade gastric MALT lymphoma may be participating in an immune response. These include the presence of scattered transformed blasts, plasma cell differentiation, which is maximal beneath the surface epithelium, and follicular colonization⁸. The latter phenomenon occurs when the lymphoma cells in the marginal zone extend centrally to replace the follicles with dispersion of the mantle zone cells resulting in a nodular appearance, or when lymphoma cells selectively colonize germinal centers, often with preservation of the mantle zone. Within the germinal centers the cells may transform, which can lead to difficulty in distinguishing MALT lymphoma from follicular lymphoma, or undergo plasma cell differentiation.

Immunophenotype and Normal Cell Counterpart

MALT lymphoma cells typically surround reactive B-cell follicles in the distribution of the marginal zone and show a tendency to involve this zone when they disseminate to lymph nodes and spleen (see below). The B-cells of MALT lymphoma share the cytological features and immunophenotype (CD20+, CD21+, CD35+, IgM+, IgD-) of marginal zone B-cells.⁴

Dissemination

Gastric MALT lymphoma disseminates both locally and systemically more frequently than was originally believed. Indirect evidence for this comes from the observation of recurrent MALT lymphomas in the gastric stump after partial gastrectomy in patients in whom clear resection margins were documented by histological examination.^{9,10} Wotherspoon et al¹¹ used the "Swiss roll" technique to examine gastrectomy specimens of gastric MALT lymphomas and showed that numerous small tumor foci with identical immunoglobulin (Ig) light chain restriction to the main tumor mass were distributed throughout the gastric mucosa including macroscopically

normal regions. Subsequently, the clonal identity of these multiple tumor foci was confirmed by sequence analysis of the rearranged Ig heavy chain genes¹². Microdissection studies using clone specific PCR demonstrated that tumor cells were frequently present in reactive lymphoid tissue that showed no histological evidence of lymphoma.¹³

MALT lymphoma cells tend to localize in the marginal zone of regional lymph nodes without disturbing the lymph node architecture, and the incidental discovery of secondary small intestinal MALT lymphoma during gastrectomy for MALT lymphoma has been reported. Gastric MALT lymphoma has been shown preferentially to disseminate to the splenic marginal zone where it is usually undetectable by conventional histopathology.¹⁴

The tendency of gastric MALT lymphoma to disseminate to other parts of the gastrointestinal mucosa and the splenic marginal zone reflects the homing properties of MALT lymphoma cells. Dogan et al¹⁵ found that the mucosal homing receptor $\alpha_4\beta_7$ integrin was strongly expressed by a secondary intestinal MALT lymphoma but not by the primary gastric lesion. However, a high level of $\alpha_4\beta_7$ expression could be induced in gastric lymphoma cells following activation by a *H. pylori* generated T-cell response. This suggests that $\alpha_4\beta_7$ + tumor cells could be similarly generated in gastric lymphoma in vivo and thus become "programmed" to home to an appropriate microenvironment where the mucosal addressin cell adhesion molecule (MAdCAM-1), the ligand for $\alpha_4\beta_7$, is expressed. This includes sites such as the intestinal mucosa^{15,16} and splenic marginal zone.¹⁷

Differential Diagnosis

Because of differences in clinical behavior and management, it is important to differentiate MALT lymphoma from the other small B-cell lymphomas that may present in the stomach. These include mantle cell lymphoma, lymphocytic lymphoma (chronic lymphocytic leukemia) and follicular lymphoma. The cytological features of mantle cell lymphoma can closely simulate those of MALT lymphoma even to the extent that occasional lymphoepithelial lesions may be present. However, absence of transformed blasts together with expression of CD5, IgD and, importantly, intranuclear expression of cyclin D1, a consequence of t(11;14), serve to distinguish mantle cell lymphoma. Lymphocytic lymphoma is characterized by small round lymphocytes together with peripheral blood lymphocytosis. Expression of CD5, CD23 and IgD without nuclear cyclin D1 provide further distinction from MALT lymphoma. Finally, follicular lymphoma, which frequently involves the stomach, can be difficult to distinguish from MALT lymphoma with follicular colonization. The transformed MALT lymphoma cells within follicles may closely resemble

centroblasts but typically are CD10 and BCL6 (nuclear) negative in contrast to the cells of follicular lymphoma, which usually express both antigens both within and between follicles.

Transformation of Gastric MALT Lymphoma

Transformation of MALT lymphoma to diffuse large B-cell lymphoma (DLBCL) is heralded by the emergence of increased numbers of transformed blasts that form sheets or clusters^{18,19} and, finally, grow to confluence effacing the preceding MALT lymphoma. De Jong et al¹⁹ have suggested dividing gastric MALT lymphoma into four categories. Category A refers to classical low-grade MALT lymphoma in which transformed blasts comprise no more than 5% of cells and do not occur in clusters of more than ten cells. In category B, transformed cells may account for 10-20% of cells and occur in clusters of up to 20 cells. Category C is characterized by unequivocal high-grade transformation with sheets of transformed cells that may leave only small foci of low-grade disease. In category D, no MALT component is detectable and it is probably better classified as DLBCL without reference to MALT. In the hands of De Jong et al the separation into the four categories had clinical significance.

Helicobacter Pylori and Gastric Lymphoma

Several lines of evidence suggest that gastric lymphoma arises from MALT acquired as the result of this *H. pylori* infection. *H. pylori* can be demonstrated in the gastric mucosa of the majority of cases of gastric MALT lymphoma.²⁰ In the first study in which this association was examined²¹ the organism was present in over 90% of cases. Subsequent studies have shown a lower incidence²² but also that the density and detectability of *H. pylori* decreases as lymphoma evolves from chronic gastritis.²³ A case control study showed an association between previous *H. pylori* infection and the development of primary gastric lymphoma.²⁴ More direct evidence confirming the importance of *H. pylori* in the pathogenesis of gastric lymphoma has been obtained from two studies that detected the lymphoma B-cell clone in the chronic gastritis that preceded the lymphoma and from a series of in vitro studies²⁵ showing that lymphoma growth could be stimulated in culture by *H. pylori* strain specific T-cells when crude lymphoma cultures were exposed to the organism. Finally, following the initial study by Wotherspoon et al, several groups have shown that eradication of *H. pylori* with antibiotics together results in regression of gastric MALT lymphoma in 75% of cases.²⁶

PCR Detection of Ig Gene Rearrangement in Gastric MALT Lymphoma

Because of the difficulty in assessing gastric biopsies both before and after eradication of *H. pylori* there has been a tendency to rely on molecular evidence of monoclonality detected by the polymerase chain reaction (PCR) for the diagnosis of lymphoma both before and after antibiotic therapy for *H. pylori*. This technique may fail to detect monoclonality in up to 30% of cases of overt lymphoma and thus produce false negative results.²⁷ There are also reports of apparently spurious monoclonality in biopsies showing only chronic gastritis²⁸⁻³⁰ and in biopsies following antibiotic induced regression of lymphoma where there is no histological evidence of malignancy.³¹ The frequency of this spurious monoclonality varies between laboratories,³² which suggests that technique may be a factor. These findings serve to emphasize that gastric MALT lymphoma should not be diagnosed in the absence of clear histological evidence.

PCR evidence of B-cell monoclonality may persist in 50% to 75% of cases that have regressed histologically following eradication of *H. pylori*. The persistence of monoclonality in post-treatment biopsies without histological evidence of lymphoma is consistent with the notion that eradication of *H. pylori* suppresses but does not necessarily eradicate the neoplastic clone in all cases. In keeping with this, in rare cases re-infection with *H. pylori* has been associated with relapse of the lymphoma. It should be stressed, however, that persistence of the neoplastic clone is not, on its own, an indication for further treatment. There is some evidence that with time the neoplastic clone becomes undetectable.

Prediction of Response of Gastric MALT Lymphoma to Eradication of *H. pylori*

The follow-up of MALT lymphoma patients following eradication of *H. pylori* is rather complex, requiring repeated gastroscopy, and it would be extremely useful to be able to identify the approximately 25% of cases of gastric MALT lymphoma that do not respond to eradication of *H. pylori*. Studies using endoscopic ultrasound have suggested that if the tumor has invaded beyond the submucosa it will not respond.^{33,34} More recently, two MALT lymphoma-associated translocations that have a bearing on the response to *H. pylori* eradication have been cloned.^{35,36} The first of these, t(1;14)(p22;q14), present in rare cases, has been shown to involve a novel gene, bcl10, that is strongly expressed in the nucleus of the neoplastic lymphocytes.³⁷ The second, t(11;18)(q21;q21), is present in up to 40% of cases and is strongly associated with failure to respond to eradication of *H. pylori*.³⁸ Interestingly, t(11;18) is also associated with nuclear expression of bcl10, albeit more weakly than t(1;14).³⁹ More-

over, the frequency of both t(11;18)(q21;q21) and nuclear BCL10 expression is significantly higher in tumors that have disseminated beyond the stomach (78% and 93%, respectively) than those confined to the stomach (10% and 38%).³⁹ These findings in part explain the results based on the use of endoscopic ultrasound^{33,34} and suggest that both t(11;18)(q21;q21) and BCL10 nuclear expression are associated with failure to respond to *H. pylori* eradication and with more advanced MALT lymphoma. This suggests that their oncogenic activities may be related.

II. THE MOLECULAR BIOLOGY OF MALT LYMPHOMA

Randy D. Gascoyne, MD, FRCPC*

In the recent past, the term marginal zone lymphoma (MZL) was considered to define a specific disease entity that included the three closely related lymphoma subtypes of nodal, primary splenic and extranodal lymphomas of MALT type.^{1,2} These lymphomas arise at different anatomic sites, are similarly composed of mature B cells lacking expression of CD5 and CD10, often reveal overlapping morphologic features, but typically display distinct clinical behaviors.³⁻⁶ As recently as 1996, there were developing concepts suggesting that all three lymphomas shared similar cytogenetic alterations, including whole or partial trisomy 3, trisomy 18 and structural rearrangements of chromosome 1 involving breakpoints at 1q21 and 1p34.^{2,7} Thus, a concept was emerging suggesting that MZL at different sites shared a common pathogenesis. Importantly, no evidence of a recurrent balanced translocation had yet been substantiated, a finding thought to play an important pathogenic role in the development of many B cell lymphomas including Burkitt's lymphoma (*cmyc*), follicular lymphoma (*bcl2*) and mantle cell lymphoma (*cyclin D1*).¹

Since that time, several important cytogenetic/molecular genetic observations have shed light on the distinctiveness of these lymphoid neoplasms, and each is now considered a unique lymphoma subtype in both the REAL and World Health Organization (WHO) classifications.^{1,8} Nodal MZL and primary splenic MZL will be discussed only briefly here, principally to distinguish these disorders from extranodal marginal zone lymphoma of MALT-type. Although this latter disorder can occur in many different anatomic sites, this discussion will focus primarily on gastric MALT lymphomas as the most common extranodal site of involvement.⁹

Cytogenetics

Only a small number of publications have described the karyotypic abnormalities associated with MALT lymphoma, owing in part to the extranodal nature of the biopsy specimens, their small size in many cases and the difficulty in producing analyzable metaphases. Originally described in 1989, the most common structural abnormality associated with MALT lymphomas is the t(11;18)(q21;q21).¹⁰ The possibility that it might play an important role in MALT lymphoma was first suggested in 1992.¹¹ This was confirmed by two studies published simultaneously in 1997.^{12,13} It is present in approximately one third of cases, has been found in most anatomic sites associated with MALT lymphomas and is often the sole cytogenetic alteration.¹² The latter finding suggests a major role for this translocation as a disease initiation event, although cryptic, sub-cytogenetic changes cannot be excluded as contributing to the development of lymphoma. A second nonrandom cytogenetic alteration associated with MALT lymphoma is t(1;14)(p22;q32), which is rarely encountered.¹⁴⁻¹⁶ This alteration has only been described in association with gastric and pulmonary MALT lymphoma.^{14,15} Thus, MALT lymphoma is unusual in comparison to most other small B cell lymphomas, having now been associated with two seemingly unrelated translocations. However, very recent data have provided insights into the mechanisms that unify these two observations (discussed in detail below).

Important negative findings in MALT lymphoma include the lack of evidence for involvement of either t(11;14)(q13;q32) or t(14;18)(q32;q21), implicating the *cyclin D1* and *bcl2* oncogenes, respectively. *Bcl6* rearrangements involving chromosome 3q27 have been described infrequently in MALT lymphoma and are more often found in association with extranodal diffuse large B cell lymphoma (DLBCL).^{17,18} Moreover, there are no reported cases of nodal MZL or primary splenic MZL harboring either a t(11;18) or t(1;14), a testament to the distinct biology of these related neoplasms.^{19,20} A characteristic balanced translocation has not been described in nodal MZL, while splenic MZL appears to be associated with an interstitial deletion of the long arm of chromosome 7 (del 7q31-32) and trisomy 3.²¹⁻²³

The most common numerical cytogenetic abnormality in MALT lymphoma is trisomy 3.^{12,24} It is present in approximately 60% of cases by classical cytogenetic studies, but is not specific for this lymphoma subtype. It has been described in other lymphomas, including both nodal MZL and primary splenic MZL.^{23,25} The same can be said for trisomy 18 and chromosomal breakpoints involving bands 1q21 and 1p34-36, as all three alterations are common clonal evolution events in many subtypes of lymphoma.^{26,27} More recent studies employing fluo-

* Department of Pathology, British Columbia Cancer Agency, 600 West 10th Avenue, Vancouver BC V5Z 4E6, Canada

rescence in situ hybridization (FISH) techniques have reported a much lower incidence of trisomy 3 in MALT lymphoma, ranging between 20 and 46%.^{28,29} The reason for this difference may reflect a combination of variable patient populations and disease definition.

Cmyc oncogene alterations may be involved as early events in the genesis of MALT lymphoma but do not involve translocation of the gene as is typical of Burkitt's lymphoma. Peng and colleagues described the presence of point mutations involving either the first exon or intron in 17% of 54 cases and suggested the role of this molecular alteration as an early event in the development of MALT lymphoma.³⁰ The significance of genetic instability in MALT lymphomas remains controversial. Microsatellite instability (MSI; replication error phenotype) results from defects in the DNA mismatch repair pathway and is generally considered uncommon in non-Hodgkin's lymphomas (NHL).³¹ Peng et al reported this to be a frequent finding in MALT lymphoma, but more recent data are contradictory, finding a very low frequency of MSI in both low-grade and high-grade MALT lymphomas.³²⁻³⁴

The concept of histological transformation of MALT lymphoma to DLBCL was described in the Section I. Genetic events that may herald this change are similar to those recognized in nodal lymphoma and include mutations of *p53* and altered methylation status or homozygous deletions of *p16*.^{35,36} Importantly, the relationship between gastric MALT lymphoma and DLBCL at the same site remains controversial, particularly as the t(11;18) is not found in gastric DLBCL.^{19,20,37} These data suggest two possibilities. Firstly, MALT lymphomas with a t(11;18) have a reduced frequency of transformation; secondly, some cases of de novo DLBCL involving the stomach may have a distinct histogenesis and are best considered de novo extranodal DLBCL rather than "high-grade" MALT lymphomas. Finally, the translocation breakpoint involved in t(6;14)(p21;q32) has been described in association with MALT lymphoma undergoing histological transformation and implicates deregulated expression of cyclin D3 in the pathogenesis of this event (manuscript submitted). Interestingly, this same balanced translocation involving the immunoglobulin heavy chain locus (IgH) has been described in myeloma.³⁸

Molecular Genetics

Immunoglobulin Heavy Chain

MALT lymphoma B cells are related to normal marginal zone cells. As such, their IgH variable region gene sequence reveals a molecular signature characteristic of a post germinal center B cell, demonstrating evidence of somatic hypermutation.^{39,40} Intracлонаl variation is also

found in many cases, suggesting that expansion of the clone is occurring in the presence of antigen. Studies of both gastric and salivary gland MALT lymphomas have revealed preferential use of germline V_H segments associated with autoantibody formation, in keeping with the observation that MALT lymphomas are commonly derived from a background of chronic inflammatory disorders, including autoimmune diseases.³⁹ The limited repertoire of V_H genes and conserved complementary determining region sequences in salivary gland MALT lymphomas suggests that a unique antigen may be driving the B cell proliferation.⁴¹

API2-MALT1

The t(11;18)(q21;q21) represents the most common structural abnormality in MALT lymphoma.^{12,13,42} In 1999, Dierlamm and colleagues showed that the t(11;18) produced a fusion of *API2* on chromosome 11q21 with *MLT* (for MALT lymphoma-associated translocation) on chromosome 18q21.⁴³ Two other groups simultaneously characterized the same gene on 18q21, now known as *MALT1*.^{44,45} In a subset of cases the translocation was associated with a cryptic deletion involving the 3' portion of *API2*, resulting in loss of the reciprocal transcript. Thus, the 5' *API2-MALT1* 3' fusion localized to the derivative chromosome 11 was shown to be the pathogenetically important event.⁴⁶

API2, also known as *cIAP2*, belongs to a family of inhibitors of apoptosis (IAPs) first identified in baculoviruses. They contain a BIR (*baculovirus inhibitor of apoptosis repeat*) motif in one to three copies, a caspase recruitment domain (CARD) and a C-terminal zinc-binding RING finger domain (absent in the IAP survivin). Several of the IAP family proteins have been shown to be potent inhibitors of activated caspases, mediated through their interaction with TNF-associated factor (TRAF) proteins, thus fulfilling a role as inhibitors of apoptosis. The BIR domain, left intact in the *API2-MALT1* fusion, is thought to be important for this function.⁴⁷

Until very recently, the function of the MALT1 protein was unknown. It was hypothesized that the fusion protein resulting from the t(11;18), owing to the presence of the *API2* component, inhibited apoptosis and thereby conferred a survival advantage to MALT lymphomas and allowed for antigen-independent proliferation.^{42,43} Uren et al have now identified MALT1 as a "paracaspase," a caspase-like protease found in several species.⁴⁸ This class of proteins has altered substrate specificity in comparison to caspases. Death domains (DDs), death effector domains (DEDs) and caspase recruitment domains (CARDs) are protein modules found exclusively in proteins that mediate apoptotic signaling.⁴⁹ MALT1 possesses a DD, two adjacent immunoglobulin

lin-like domains and a caspase-like domain (but lacks a CARD domain). Similar to CARDs and DEDs, the DD of MALT1 acts as a homotypic interaction module.⁴⁸ In an attempt to illuminate signaling pathways involving MALT1, yeast two-hybrid screens were performed in order to identify molecules that bind the prodomains of MALT1.⁴⁸ Surprisingly, Bcl10, the protein involved in the other MALT lymphoma translocation, binds to MALT1 and appears to do so through an interaction with the two Ig-like domains (**Figure 1**). Under physiological conditions, Bcl10 and MALT1 form a tight bond and synergize to increase activation of NF- κ B.⁵⁰ Over-expression of full-length or mutant forms of Bcl10 has been shown to weakly activate NF- κ B. Full-length MALT1 protein does not significantly activate NF- κ B, nor does full-length API2. However, the fusion protein produced by the *API2-MALT1* translocation significantly increases NF- κ B activation.⁴⁸ Truncated versions of the fusion fail to activate NF- κ B, suggesting that both the BIR domains contributed by API2 and the paracaspase domain of the chimeric protein are required for complete activation of NF- κ B.⁴⁸

NF- κ B, a member of the *rel* family, plays a central role in the activation of genes involved in immunity, in-

flammation and apoptosis.^{51,52} In unstimulated cells, NF- κ B is sequestered in the cytoplasm through interactions with inhibitory I κ B proteins. In response to a variety of stimuli, I κ B α is phosphorylated and targeted for degradation by the ubiquitin pathway. The degradation of I κ B α results in the translocation of NF- κ B to the nucleus, where it binds to specific promoters and activates transcription. Thus, the mechanism by which Bcl10 activates NF- κ B has now been elucidated. These new data demonstrate that under normal conditions, Bcl10 and MALT1 form a tight complex that serves to oligomerize and activate the caspase-like domain of MALT1, leading to induction of NF- κ B.⁵⁰ Unlike wild type MALT1, which appears to be dependent upon an interaction with Bcl10 as a mechanism for oligomerization and auto-activation, the API2-MALT1 fusion protein may possess a mechanism for self-oligomerization, possibly due to the three BIR domains contributed by the chimera.⁵⁰ Thus, in normal cells the basal level of expression maintains the signaling pathway but can be perturbed by either a t(1;14) or a t(11;18) in MALT lymphoma that results in marked over-expression of Bcl10 or the API2-MALT1 fusion proteins, respectively. The resultant dramatic increase in NF- κ B activity is likely critical to lymphoma progression.

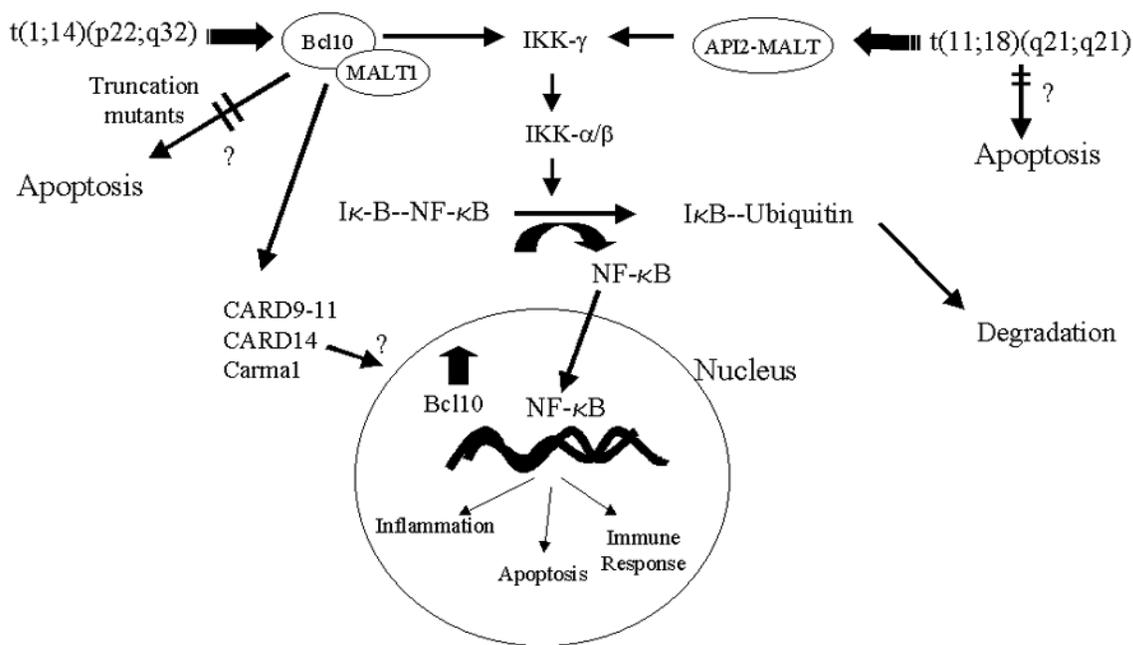


Figure 1. An improved understanding of the paracaspase MALT1 has brought clarity to the mechanisms underlying the molecular biology of MALT lymphomas.

When *bcl10* expression is increased by translocation, it binds to its normal partner, MALT1 and together they increase the activity of IK kinase, resulting in the increased translocation of NF- κ B into the nucleus. Alternatively, this same pathway can be activated by the novel fusion, *API2-MALT1*, which does not require *bcl10*. Therefore, both the t(1;14) resulting in deregulation of *bcl10* and the t(11;18) resulting in a novel fusion *API2-MALT1* appear capable of activating IK kinase, which leads to the phosphorylation and subsequent degradation of I κ B and translocation of NF- κ B into the nucleus. NF- κ B induces the transcription of a number of different genes, including those involved in inflammation, cell viability and the immune response. Although both translocations were thought to promote lymphomagenesis by directly inhibiting apoptosis, this now seems less likely. However, this does not preclude that both translocations may affect the apoptotic signaling pathway indirectly by altering downstream apoptotic events resulting from increased nuclear NF- κ B.

Bcl10

The other recurrent translocation in MALT lymphomas is t(1;14)(p22;q32), which is significantly less frequent than t(11;18).¹⁴⁻¹⁶ This rearrangement results in the juxtaposition of the entire coding region of *bcl10* to chromosome 14 under control of the Ig enhancer element. All *bcl10* breakpoints thus far characterized cluster within the 5' promoter region of the gene.¹⁵ The human *bcl10* gene encodes a protein of 233 amino acids containing an N-terminal caspase recruitment domain (CARD), characteristic of both antiapoptotic and proapoptotic proteins. In normal tissues, Bcl10 is ubiquitously expressed, but at low levels. The highest Bcl10 expression levels are found in spleen, lymph node, testis and developing central nervous system.^{15,53} Forced expression of wild type Bcl10 in cell lines induces both apoptosis and activates NF- κ B.^{54,55} The translocation leads to over-expression of the Bcl10 protein but is frequently associated with frameshift mutations causing C-terminal truncations distal to the CARD.^{15,16} CARD-truncation mutants lose apoptosis activity and fail to induce NF- κ B, whereas mutants with C-terminal truncations retain NF- κ B activation but do not induce apoptosis.¹⁶ It was thus hypothesized that Bcl10 might induce tumors via two mechanisms: mutant Bcl10 would lose its proapoptotic function conferring a growth advantage on MALT B cells, and constitutive NF- κ B activation could provide both anti-apoptotic and proliferative signals by up-regulating transcription of specific targets.¹⁶

The initial report of the cloning of the *bcl10* gene indicated a high frequency of truncating *bcl10* mutations independent of the presence of the t(1;14) (~45% of B and T cell lymphomas).¹⁵ These cases were analyzed using cDNA preparations from a variety of tumors. However, a comparable rate of *bcl10* mutation was not detected when genomic DNA from a wide range of tumors was analyzed.⁵⁶⁻⁶¹ Some of these differences may be due to technical artifacts arising from the study of cDNA versus genomic DNA, resulting from "molecular misreading."^{62,63} Nonetheless, subsequent reports have confirmed the presence of *bcl10* mutations in a small percentage of MALT lymphomas (6.7%), follicular lymphomas (9.5%) and DLBCLs (4.3%).⁶³ Moreover, mutations of *bcl10* were slightly more common in "high-grade" MALT lymphomas, suggesting that mutations may underlie histological progression. An intriguing finding from this work was the over-expression of wild type Bcl10 in some MALT lymphoma cases with a t(1;14).⁶³

Murine models used to study the functional role of Bcl10 have recently been described.^{64,65} Transgenic mice in which *bcl10* is linked to an immunoglobulin enhancer construct that directs expression to lymphoid cells only develop splenomegaly due to a marked increase in splenic marginal zone B cells, similar to splenic MZL.

Interestingly, these studies also revealed that mice over-expressing Bcl10 mutants in their lymphocytes have no apparent abnormalities of lymphocyte development or function, suggesting that deregulated expression of the normal Bcl10 protein by the t(1;14) rather than *bcl10* mutation contributes to MALT tumorigenesis.⁶⁵

Significant improvement in our understanding of the role of Bcl10 in normal intracellular signaling and tumorigenesis has resulted directly from the study of knockout mice with targeted disruption of *bcl10* (*bcl10*^{-/-}).⁵³ One third of *bcl10*^{-/-} embryos developed exencephaly, leading to embryonic lethality, but the mice retained their susceptibility to apoptotic stimuli. Importantly, the mice were markedly immunodeficient due to defective NF- κ B activation via the antigen receptor complex of both B and T cells. These data allowed the authors to conclude that Bcl10 is unlikely to be an essential component of the mammalian cell death machinery. Truncating mutations of the *bcl10* gene were postulated to be important in the pathogenesis of MALT lymphomas by conferring a relative resistance to apoptosis. The *bcl10*^{-/-} data reveal that complete disruption of both *bcl10* alleles fails to promote cell survival and does not result in tumor formation in the mice.⁵³ It is well established that most MALT lymphomas originate in the setting of inflammation triggered by infection or autoimmune disorders. Moreover, gastric MALT lymphomas are in part dependent on *H. pylori*-specific T cells to provide the immunological stimulus for their early development and proliferation.⁶⁶ Thus, translocation and up-regulated expression of either wild type or truncated Bcl10 could mimic this antigen-receptor signaling, producing constitutive activation of NF- κ B, antibiotic-resistant growth and progression of the lymphoma.

Since the discovery of Bcl10, a number of proteins have been described, including a new family of proteins called the membrane-associated guanylate kinase family (MAGUK).⁶⁷ Thus far, several members of this family have been described, including CARD9, CARD10, CARD11 and CARD14 and Carma 1.⁶⁸⁻⁷⁰ Briefly, this class of proteins appears to function as upstream regulators of Bcl10. MAGUK proteins function to organize signaling complexes at plasma membranes. Each appears to bind Bcl10 at its CARD motif, resulting in NF- κ B activation, but variable consequences result from this interaction including phosphorylation of Bcl10, translocation of Bcl10 from the cytoplasm to perinuclear structures and a molecular scaffolding function for the assembly of Bcl10 signaling complexes.⁶⁷⁻⁷⁰

Bcl10 protein expression, molecular and clinical correlates

Monoclonal antibodies raised against the Bcl10 protein demonstrate expression of Bcl10 in lymphoid tissue and

breast epithelium.⁷¹ In normal B cell follicles, Bcl10 is expressed in the cytoplasm of germinal center B cells and marginal zone cells, but only weakly in mantle zone cells. MALT lymphomas characterized by the t(1;14) express Bcl10 in both the nucleus and cytoplasm. Cases that lack the translocation have also been shown to express nuclear Bcl10. A preliminary study of MALT lymphomas has revealed a strong correlation between the presence of the t(11;18) and nuclear Bcl10 expression.⁷² These data suggest that nuclear localization of Bcl10 can occur as the result of two apparently independent cytogenetic events. Thus, these two seemingly disparate translocations that target *bcl10* and *MALT1* appear to affect the same signaling pathway, the result of which is the activation of NF- κ B and the nuclear localization of Bcl10 protein. This latter finding may not only be important for the pathogenesis of MALT lymphomas but may serve as a surrogate marker for identifying more aggressive clinical disease and resistance to antibiotic therapy.^{72,73}

Conclusion

In summary, our understanding of the molecular biology of MALT lymphoma has significantly improved following the recent cloning of two balanced translocations involved in this disease. Although both genes code for novel proteins with distinct protein interaction modules, suggesting involvement in the cell death pathway, the inhibition of apoptotic signaling may not be central to the pathogenesis of either *bcl10* or *API2-MALT1*. Instead, constitutive activation of the NF- κ B pathway may mimic engagement of the antigen receptor complex, driving antigen-independent growth and lymphoma progression. Curiously, two unrelated translocations appear to impact the same signal transduction pathway, giving rise to the development of a single clinicopathologic entity we recognize as MALT lymphoma. Much excitement will await the experimental data that determine the precise role of nuclear Bcl10 and how this contributes to lymphomagenesis.

III. LOW GRADE GASTRIC MALT LYMPHOMA

Emanuele Zucca, MD, and Franco Cavalli, MD**

The term low-grade MALT lymphoma identifies a group of extranodal B-cell lymphomas, composed mostly of small cells, that share similar clinical, pathological, and molecular features; these lymphomas are defined as extranodal marginal zone B-cell lymphomas of MALT type in the Revised European-American Classification

of Lymphoid Neoplasms (REAL)/World Health Organization (WHO) Classification of Lymphoid Neoplasms. The stomach is by far the most common localization.¹⁻²

The onset of MALT lymphoma in the stomach, where lymphocytes are not normally present, is preceded by the acquisition of a mucosa-associated lymphoid tissue (MALT) as a result of *H. pylori* infection.¹⁻² The microorganism can be found in the gastric mucosa in nearly all instances of gastric MALT lymphoma, with several lines of evidence suggesting a link between *H. pylori*-chronic gastritis and the lymphoma. A close association has been reported in epidemiologic studies between *H. pylori* infection and gastric lymphomas of either low-grade or high-grade histology.³ In vitro experiments have demonstrated that the neoplastic cells of low-grade gastric MALT lymphoma proliferate in a strain-specific response to *H. pylori* and that this response is dependent on specific T-cell activation.⁴ The presence of the B-cell clone that would become predominant in the transformation to MALT lymphoma has been demonstrated in *H. pylori* gastritis specimens taken several years before development of the lymphoma.⁵ A regression of gastric MALT lymphoma after antibiotic eradication of *H. pylori* has been reported in more than half of the treated patients.⁶⁻¹² This close association of *H. pylori* with gastric MALT lymphoma has led to the hypothesis that the microorganism may provide the antigenic stimulus for sustaining the growth of the lymphoma in the stomach.¹³ However, the exact mechanism of the transition from *H. pylori* infection to low-grade MALT lymphoma is still unclear. Most patients with *H. pylori* gastritis do not develop lymphoma; therefore, it is widely accepted that additional environmental and microbial or host genetic factors may play a role in gastric lymphomagenesis.¹⁻²

Diagnosis and Staging

The most common presenting symptoms of low-grade gastric MALT lymphomas are nonspecific dyspepsia and epigastric pain. Constitutional B symptoms are exceedingly uncommon. Endoscopy usually reveals nonspecific gastritis or peptic ulcer, with mass lesions being unusual.^{6,14} Few patients present with elevated lactate dehydrogenase (LDH) or β 2-microglobulin levels.⁶

The best staging system is still controversial.¹⁵⁻¹⁶ We currently use the revised version of the Blackledge staging system that was recommended for general use by an international workshop held in Lugano, Switzerland, in 1993.¹⁶ When ultrasound endoscopy is available, the TNM system (using the criteria initially proposed for gastric carcinoma by the American Joint Committee on Cancer and Union International Contre le Cancer) can also be employed, based on the echoendoscopic extent of the gastric wall involvement.¹⁰

The initial staging should comprise a gastroduode-

* Oncology Institute of Southern Switzerland, Department of Medical Oncology, 6500 Bellinzona, TI, Switzerland

nal endoscopy with multiple biopsies from each area of the gastric map and from all the abnormal sites. Upper airway examination is required as well as all of the usual procedures performed for nodal lymphomas, including bone marrow biopsy. The presence of active *H. pylori* infection must be always ruled out by histology; serology studies are useful when results of histology are negative.

Unlike most low-grade B-cell lymphomas of peripheral lymph nodes, low-grade MALT lymphoma is usually a very indolent disease, often remaining localized for a prolonged period; in some cases, no progression is seen during several years without treatment. A few patients present with systemic disease, often due to the simultaneous involvement of multiple mucosal sites and it has been postulated that this pattern of dissemination may be related to specific homing properties similar to those of the normal B-cells of MALT.¹ Bone marrow involvement is reported in 0-15% of cases.^{6,17,18} Prognosis seems particularly poor in the few cases presenting with advanced stages¹⁷⁻²⁰ or with an unfavorable IPI score.²¹ Patients with primary gastrointestinal presentation might have a better survival than those with non-gastrointestinal MALT lymphoma.²² A deep infiltration of the gastric wall by the lymphoma has been reported to be strongly associated with spread to the regional lymph nodes, analogous to findings in gastric carcinoma. It has therefore been recommended that the depth of infiltration be included in pathology reports concerning primary gastric lymphoma of the MALT.²³ Endoscopic ultrasound allows evaluation of the depth of infiltration. Moreover, it might be useful to distinguish benign lymphoid aggregate from malignant lymphoma and should be included in both the initial and the follow-up procedures whenever possible.²⁴⁻²⁷

Treatment

Despite abundant literature on histologic, clinical, and biological features of MALT lymphoma, final results of controlled trials to define the optimal therapy have not yet been published. Published data are confusing: insufficient staging and outdated histologic classifications are a major problem of the older reports, and more recent studies often refer to retrospective series of patients not uniformly staged and treated.

Few published studies specifically report treatment outcome for localized gastric MALT lymphoma. The patients have been treated with a variety of combinations of surgery, radiotherapy, and chemotherapy, and the overall survival rates range from 80% to 95% at 5 years.^{6,14,17,20,29} Therefore, while prognosis of patients with MALT lymphoma seems excellent regardless of treatment, optimal therapy remains to be determined.

Increasing evidence indicates that eradication of *H. pylori* with antibiotics can be effectively employed as

the sole initial treatment.^{6-12,26-28} In a series of 93 patients from northern Italy and southern Switzerland with low-grade gastric MALT lymphoma, no statistically significant difference was apparent in overall survival or event-free survival between patients who received different initial treatments (chemotherapy alone, surgery alone, surgery with additional chemotherapy or radiation therapy, or antibiotics against *H. pylori*).⁶ The actuarial 5-year overall survival was 82% (95% confidence interval [CI], 67%-91%) in the series as a whole. At a median follow-up of 3 years, 10 of 93 patients had died, all but 1 from a second (solid) tumor. The unexpectedly high incidence of additional neoplasms was not treatment related and has been described in other series, but its significance is controversial.³⁰⁻³² In this series, 49 patients with stage I disease were given antibiotics alone as initial treatment; eradication of *H. pylori* was achieved in 97% of patients (95% CI, 88.2%-99.9%), and histologic regression of the MALT lymphoma was documented in 67% of the patients (95% CI, 51%-80%) after the eradication.⁶ The median time required to achieve histologic regression was 5 months (range, 3 to 18 months). A German multicenter study confirms, with a median follow-up of 2 years, the efficacy of antibiotics in inducing apparently durable lymphoma remission. Moreover, this study demonstrated that patients in whom the lymphoma did not respond to *H. pylori* eradication may have harbored high-grade lesions that had not initially been recognized.¹¹

An American trial of 34 patients with stage I-II disease showed that the antibiotic efficacy is higher in early lesions: 70% (95% CI, 35%-93%) of the cases with disease confined to the mucosa and submucosa achieved a complete remission (CR), whereas those with locally advanced disease infiltrating the muscularis mucosae, the serosa, or the perigastric lymph nodes had a significantly lower CR rate (38%; 95% CI, 17%-64%).¹⁰ A recent French series of 46 patients reported a CR rate of 43% with no response in the 10 *H. pylori*-negative cases. In this study the absence of nodal involvement was the strongest predictor of regression with a CR rate of 79% for the *H. pylori*-positive cases without any lymph node involvement.¹² A preliminary response evaluation has been performed in the first 170 patients with localized low-grade lymphoma of the stomach enrolled in the ongoing international controlled clinical trial LY03 of chlorambucil versus observation after antibiotic therapy; it confirmed that at least half of the treated cases can achieve a histologic CR.³³ A subset of patients has undergone a molecular follow-up by the PCR assay for the detection of a monoclonal rearrangement of the immunoglobulin gene: approximately half of the patients with a histologic CR also had a molecular complete response that sometimes required a long time (up to 2 years) to be

demonstrated.³⁴ Similar molecular data have been found by other authors.^{10-11,28,35} These data demonstrate that PCR-detectable B-cell monoclonality may persist after the disappearance of histologic evidence of MALT lymphoma, suggesting that *H. pylori* eradication suppresses but does not eradicate the lymphoma clones. Whether the persistence of PCR-detected B-cell monoclonality is associated with a higher risk of lymphoma relapse remains to be determined. Thus, careful histologic examination of multiple gastric biopsies remains the cornerstone for the follow-up of gastric MALT lymphoma patients.²⁸

In our opinion, the indolent nature of the disease in most cases of localized MALT lymphoma makes a conservative approach advisable, with antibiotic therapy as the sole initial treatment provided that strict oncohematologic and endoscopic follow-up is conducted. The use of antibiotics as first-line therapy may avert the indication for surgical resection in most patients, and we recommend eradication of *H. pylori* before consideration of further therapeutic options. Any of the highly effective antibiotic regimens proposed can be used.¹ A strict endoscopic follow-up is recommended, with multiple biopsies taken 2 months after treatment to document *H. pylori* eradication and, subsequently, at least twice per year for 2 years to monitor the histologic regression of the lymphoma. In case of unsuccessful *H. pylori* eradication, a second-line anti-*Helicobacter* therapy should be attempted with alternative triple- or quadruple-therapy regimens of proton-pump inhibitor plus antibiotics. However, it is still unknown whether *H. pylori* eradication will definitely cure the lymphoma; therefore, long-term follow-up of antibiotic-treated patients is mandatory. Some cases of documented tumor recurrence following *H. pylori* reinfection have been reported, suggesting that residual dormant tumor cells can be present despite clinical and histologic remission. Relapses have also been documented in the absence of *H. pylori* reinfection, indicating the presence of B-cell lymphoma clones that have escaped the antigenic drive.¹¹ The efficacy of antibiotic therapy is reduced in locally advanced disease, with bulky masses or deep infiltration of the gastric wall, and in disease associated with increased numbers of large cells. In our experience, however, eradication of *H. pylori* is worthwhile even in these cases but usually cannot be the unique therapeutic approach. In addition to antibiotic therapy, chemotherapy (or radiotherapy) should be given to these patients as well as to those with regional nodal involvement.¹

No treatment guidelines exist for the management of patients after antibiotics failure and for the subset of cases in which no evidence of *H. pylori* can be found. It has been shown that the chance of a response to antibiotics is dramatically reduced in the latter group.^{10,12} A

choice can be made between conventional oncologic modalities, including chemotherapy, radiotherapy, and surgery, alone or in combination. Unfortunately, there are no published randomized studies to help the decision.

Chemotherapy has never been adequately evaluated in gastric MALT lymphomas because it was usually not administered or given after surgery or radiotherapy. Some scanty data suggesting the efficacy of chlorambucil in low-grade gastric lymphoma can be found in the older literature,³⁶ but only one nonrandomized trial has thus far tested the activity of chemotherapy with single alkylating agents in MALT lymphomas.³⁷ In this study, 24 patients, 17 with stage I_E and 7 with stage IV were given continuous oral administration of cyclophosphamide, 100 mg/day, or chlorambucil, 6 mg/day (median treatment duration, 18 months; range, 8-24 months). A 75% CR rate was reported. Five patients relapsed (2 with stage I and 3 with stage IV) at 12 to 96 months, all in initial sites, and 1 with large-cell transformation. The projected 5-year event-free and overall survivals were 50% and 75%, respectively.³⁷

A preliminary report from a phase II trial of the International Extranodal Lymphoma Study Group suggest that the anti-CD20 antibody rituximab may also have a significant clinical activity in relapsing or *H. pylori*-negative gastric MALT lymphoma.³⁸

Analogous to chemotherapy, the efficacy of local radiotherapy has also not been extensively studied in trials that take account of the MALT concept.^{1,29,39} In a small American study, 17 patients with stage I-II MALT lymphoma of the stomach without evidence of *H. pylori* infection or with persistent lymphoma after antibiotics were treated with radiation alone (1.5-Gy fractions in 4 weeks to the stomach and the adjacent lymph nodes, with a median total dose of 30 Gy). The results are encouraging, with 100% biopsy-confirmed CR and 100% event-free survival (at a median follow-up of 27 months).²⁹

Surgery has been widely used in the past. Cogliatti et al²⁰ reported a series of histologically reviewed cases of low-grade MALT lymphoma (48 patients with stage I_E and 21 with stage II_E disease): 45 had surgery alone; 12 surgery and adjuvant chemotherapy; 11 surgery and irradiation; 1 surgery, chemotherapy, and radiotherapy. The 5-year overall survival was 91% (95% for stage I_E and 82% for stage II_E) with no significantly different survival rates between gastrectomy alone versus additional treatment.

While the use of local treatment is evidently associated with an excellent disease control, the precise role for surgical resection must nowadays be redefined. Follow-up endoscopy may reveal the reappearance of lymphoepithelial lesions in the remaining gastric mucosa that can be responsible for local recurrence. Indeed, the fact that MALT lymphoma is often a multifocal disease

suggests that clear excision margins are not necessarily a guarantee of radical resection. If surgery is chosen, a total gastrectomy may offer greater chances of cure, but this operation carries a risk of mortality and may severely impair the patient's quality of life.⁴⁰⁻⁴¹

IV. NON-GASTRIC MARGINAL ZONE B-CELL LYMPHOMA OF MALT TYPE

Franco Cavalli, MD, and Emanuele Zucca, MD

Extranodal marginal zone B-cell lymphoma is a discrete clinico-pathological entity arising in MALT. Two types of MALT can be identified in disparate organs that do not correspond to peripheral sites of the immune system. The native type consists of lymphoid tissue physiologically present in the gut (e.g., Peyer's patches), whereas acquired MALT develops in sites of inflammation in response to either infectious conditions, such as *H. pylori* gastritis, or autoimmune processes, such as Hashimoto's thyroiditis or myoepithelial sialadenitis (MESA) associated with Sjögren's syndrome.¹⁻³ In the context of these prolonged lymphoid reactive proliferations, the growth of a pathological clone can progressively replace the normal lymphoid population, giving rise to a MALT lymphoma.⁴⁻⁷

Outside the stomach the role of infectious agents is less clearly defined. There is some evidence indicating that *Borrelia burgdorferi* may be implicated in the pathogenesis of some cutaneous lymphoma.⁸ The hepatitis C virus (HCV) has been linked to B-cell lymphoproliferation and autoimmunity, and has been localized in several tissues, including the gastric mucosa. Several recent studies have reported a high rate of previous HCV infection in patients with non-Hodgkin's lymphoma (NHL), suggesting a possible pathogenetic link between HCV and certain histologic lymphoma subtypes, including MALT lymphomas and splenic MZL. However, it appears that there are marked geographical differences in the prevalence of HCV among NHL patients, and its role in the development of MZLs needs to be further investigated.⁹⁻¹⁰

The group of lymphomas classified as low-grade MALT lymphomas include a number of extranodal B-cell lymphomas defined as extranodal marginal zone B-cell lymphomas of MALT type in the Revised European-American Classification of Lymphoid neoplasms (REAL classification)¹¹ and in the new WHO Classification of Neoplastic Diseases of the Hematopoietic and Lymphoid Tissue.¹² The histologic features of low-grade B-cell lymphomas of MALT type are similar regardless of site of origin.

Far from being rare, extranodal marginal zone B-cell lymphomas account for approximately 8% of all

NHL, being the third most frequent histologic subtype (after diffuse large B-cell lymphoma and follicular lymphoma).¹³ The stomach is by far the most common and best-studied site; however, MALT lymphomas have also been described in various non-gastrointestinal sites, such as salivary gland, thyroid, skin, conjunctiva, orbit, larynx, lung, breast, kidney, liver, prostate, and even in the intracranial dura.^{6-8,14-30} Non-gastric MALT lymphomas have been difficult to characterize because these tumors, numerous when considered together, are distributed so widely throughout the body that it is difficult to assemble adequate series of any given site. **Table 1** shows the main localizations of MALT lymphomas arising outside the stomach.

Gastric MALT lymphoma usually remains localized for long periods within the tissue of origin, where it can present with multifocal lesions.¹⁻² Sometimes involvement of multiple mucosal sites is present; some cases with simultaneous gastric and intestinal involvement have been reported, and thyroid and salivary gland MALT lymphomas may also disseminate to the gastrointestinal tract.³¹⁻³² Bone marrow involvement at presentation is uncommon.¹⁻²

Disseminated disease appear to be more common in non-gastrointestinal MALT lymphomas, where one-quarter of cases has been reported to present with involvement of multiple mucosal sites and/or an involvement of non-mucosal sites such as bone marrow.¹⁶⁻¹⁷ It has been postulated that this dissemination may be due to the specific expression of special homing receptors or adhesion on the surface of the B cells of MALT. Indeed, the integrin $\alpha_4\beta_7$ has been identified as a main mucosal homing receptor that regulates the traffic of circulating lymphocytes to the mucosal tissues by binding to the addressin MAdCAM-1, a vascular recognition molecule selectively expressed on the mucosal endothelium in Peyer's patches, mesenteric lymph nodes, and sinus lining cells of the splenic marginal sinus. The $\alpha_4\beta_7$ integrin

Table 1. Most frequent primary localizations of non-gastric mucosa-associated lymphoid tissue (MALT) lymphoma (data pooled from the International Extranodal Lymphoma Study Group [IELSG] study and other published series).

| Primary site | Percentage |
|---------------------|------------|
| Head and neck | 30% |
| Ocular adnexa | 24% |
| Lung | 12% |
| Skin | 12% |
| Intestinal tract | 8% |
| Thyroid | 7% |
| Breast | 2% |
| Genitourinary tract | 1% |

is expressed by most MALT lymphoma cells and normal B-cells of MALT.³³⁻³⁴

Usually MALT lymphomas behave as a clinically indolent disease. In the few series thus far published of non-gastrointestinal MALT lymphomas^{16-17, 35-36} patients appear to have a good outcome, with a 5-year overall survival ranging from 86% to 100%.

The International Extranodal

Lymphoma Study Group Survey

The International Extranodal Lymphoma Study Group (IELSG) conducted an analysis of a large series of patients who were diagnosed as non-gastric MALT lymphoma with the aim of better characterizing this disease entity. Preliminary data have been presented at the 2000 meeting of the European Society for Medical Oncology (ESMO).³⁷

The study population initially included 365 patients from 20 centers. A histological review was performed by a panel of expert pathologists; 119 of 365 patients were excluded from the study, 108 because of non-confirmed histology (20 diffuse large cell lymphomas with a low grade MALT component, 9 reactive lymphoid proliferations, 9 mantle cell lymphomas, 8 follicular lymphomas, 3 plasmacytomas, 2 peripheral T cell lymphomas, 1 lymphoplasmacytoid lymphoma, and 56 where pathological material was not available or inadequate for histologic confirmation), 2 cases because of their primary gastric localization, and 9 cases because of incomplete data or inadequate follow-up. Therefore, only 246 patients were included in the analysis.

Table 2 shows the main clinical features at presentation. The median age was 60 years (range 21-92 years). One hundred and sixty-five (67%) patients presented with Ann Arbor stage I disease; 17 (7%) patients had disease involving loco-regional nodes to the primary extranodal site of disease (stage II), whereas 64 (26%) patients presented stage IV disease. B symptoms were documented in 17 patients (7%), whereas 13 patients (5%) had an impaired performance status (ECOG PS score = 2). The serum lactate dehydrogenase (LDH) level was elevated in 23% of the 213 patients who had it measured at presentation. HCV positive serology was documented in 14% of 135 tested cases. According to the International Prognostic Index (IPI) (applicable to 243 patients), 82% of patients ranked in the low or low-intermediate risk groups.

The primary site was operationally defined as the clinically dominant extranodal component, which requires diagnostic investigation and to which primary treatment must often be directed. Most of the cases (approximately 25% each) had the lymphoma primarily in the salivary glands or in the ocular adnexa. The lung was the primary site in 14% of the cases, and 12% had

skin lymphoma. Upper airways and Waldeyer's ring accounted for 8% of cases, thyroid and intestinal tract for 7% each. The disease presented with concomitant involvement of multiple MALT sites in 13% of cases; 16% of patients had nodal involvement with either loco-regional or disseminated adenopathy.

Two hundred and forty (98%) patients have been treated; in the remaining a wait-and see policy was adopted. Primary treatment included chemotherapy in 80 patients, radiotherapy in 54 patients or a combined modality approach (chemo-radiotherapy) in 17 patients. Eighty-three patients had a surgical resection, followed by chemotherapy (n = 20), radiotherapy (n = 24) or both (n = 6). Five patients received interferon- α therapy and one, with a skin localization, had a tumor regression after antibiotic therapy against *Borrelia burgdorferi*.⁸ Of the 123 patients who received chemotherapy, 62 had a single alkylating agent or CVP (cyclophosphamide, vincristine, prednisone), and 63 were treated with an anthracycline-containing regimen. At a median follow-up of 3.6 years there was no evidence of a survival advantage for any type of therapy. However, this finding must be taken cautiously: patients were treated according to the current policy of each institution at the time of diagnosis, and the presence of organ-specific problems presumably had also a role in the choice of treatment.

At 5 years the overall survival for the entire population (n = 246) was 93%, with a cause-specific survival of 96% and a progression-free survival of 73%. At univariate analysis, elevated LDH, advanced stage, poor IPI score were found to be significantly predictive of a poorer outcome in terms of either overall, cause-specific or progression-free survival. As shown in **Figure 2**, the 5-year overall survival was 99% for the IPI low

Table 2. Patient characteristics in the International Extranodal Lymphoma Study Group (IELSG) series of non-gastric mucosa-associated lymphoid tissue (MALT 0 lymphoma (n = 246).

| Clinical feature | Percentage |
|--|------------|
| Median Age | 60 yrs |
| Median age range | 20-92 yrs |
| Male | 39% |
| PS (ECOG)>0 | 35% |
| Stage IV | 26% |
| Multiple MALT Sites | 13% |
| Lymph Node involvement | 16% |
| Bone marrow involvement | 13% |
| High serum lactate dehydrogenase (LDH) (n=213) | 23% |
| High β -2 microglobulin (n=125) | 36% |
| Hepatitis C infection (n=135) | 16% |
| B-symptoms | 7% |

risk group, 85%-88% for the intermediate risk groups and 72% for the high risk patients ($p = 0.0008$). The patients without lymph node involvement at presentation had a 5-year survival of 97%, while it was 75% in those with nodal involvement ($p = 0.0002$). However, in agreement with the finding of Thieblemont et al,¹⁶ the involvement of multiple mucosal sites at diagnosis did not appear to change the outcome.

In conclusion, non-gastric MALT lymphomas, despite presenting with stage IV disease in approximately one quarter of cases, usually have indolent course (5-year survival of 93%). Patients at high risk according to the IPI and those with lymph node involvement at presentation, but not those with involvement of multiple MALT sites, have a worse prognosis. Localization can be an important factor because of organ-specific problems, which result in particular management strategies. However, whether different sites have a different natural history remains an open question. In the IELSG series the patients with the disease initially presenting in the upper airways appeared to have a slightly poorer outcome (5-year overall survival, 82%), but their small number prevented any definitive conclusion.

The optimal management of extranodal marginal zone B-cell lymphomas of MALT type has not yet been clearly defined. Surgery, chemotherapy, and radiotherapy alone or in combination have been employed. Based on preliminary data anti-CD20 antibodies may also have significant clinical activity.³⁸ The treatment choice should be "patient-tailored," taking into account the site, the stage and the clinical characteristics of the individual patient.

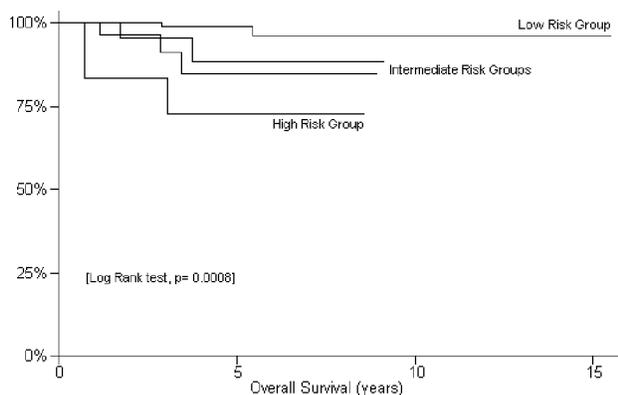


Figure 2. The International Extranodal Lymphoma Study Group (IELSG) study of non-gastric MALT lymphomas: Kaplan-Meier estimate of overall survival according to the International Prognostic Index risk groups (defined considering age, Ann Arbor stage, LDH levels, number of extranodal sites and performance status)

REFERENCES

I. The Pathology of Gastric MALT Lymphoma and Its Response to Eradication of *H. Pylori*

1. Isaacson P, Wright DH. Malignant lymphoma of mucosa-associated lymphoid tissue. A distinctive type of B-cell lymphoma. *Cancer*. 1983;52:1410-1416.
2. Isaacson P, Wright DH. Extranodal malignant lymphoma arising from mucosa associated lymphoid tissue. *Cancer*. 1984;53:2515-2524.
3. Spencer J, Finn T, Isaacson PG. Human Peyer's patches: an immunohistochemical study. *Gut*. 1986;27:405-410.
4. Spencer J, Finn T, Pulford KAF et al. The human gut contains a novel population of B-lymphocytes which resemble marginal zone cells. *Clin Exp Immunol*. 1985;62:607-610.
5. Harris NL, Jaffe ES, Diebold J et al. The World Health Organization classification of neoplastic diseases of haematopoietic and lymphoid tissues: report of the Clinical Advisory Committee Meeting, Airlie House, Virginia, November 1997
6. Isaacson PG, Norton AJ. Extranodal Lymphomas. New York: Churchill Livingstone; 1994.
7. Isaacson PG, Spencer J. Malignant lymphoma of mucosa-associated lymphoid tissue. *Histopathology*. 1987;11:445-462.
8. Isaacson PG, Wotherspoon AC, Diss T et al. Follicular colonization in B-cell lymphoma of mucosa-associated lymphoid tissue. *Am J Surg Pathol*. 1991;15:819-828.
9. Green JA, Dawson AA, Jones PF et al. The presentation of gastrointestinal lymphoma: study of a population. *Br J Surg*. 1979;66:798-801.
10. Cogliatti SB, Schmid U, Schumacher U, et al. Primary B-cell gastric lymphoma: a clinicopathological study of 145 patients. *Gastroenterology*. 1991;101:1159-1170.
11. Wotherspoon AC, Dogliani C, Isaacson PG. Low-grade gastric B-cell lymphoma of mucosa-associated lymphoid tissue (MALT): a multifocal disease. *Histopathology*. 1992;20:29-34.
12. Du MQ, Diss TC, Xu CF et al. Clonal origin of micro-lymphomas in low grade B-cell gastric MALT lymphoma. *J Pathol*. 1997; 181:57(Abstract)
13. Du MQ, Diss TC, Dogan A, et al. Clone specific PCR reveals wide dissemination of gastric malt lymphoma to the gastrointestinal mucosa. *J Pathol*. 1 2000;192:488-493.
14. Du MQ, Peng HZ, Dogan A, et al. Preferential dissemination of B-cell gastric mucosa-associated lymphoid tissue (MALT) lymphoma to the splenic marginal zone. *Blood*. 1997;90:4071-4077.
15. Dogan A, Du M, Koulis A et al. Expression of lymphocyte homing receptors and vascular addressins in low-grade gastric B-cell lymphomas of mucosa-associated lymphoid tissue. *Am J Pathol*. 1997;151:1361-1369.
16. Briskin MJ, Winsor-Hines D, Shyjan AM, et al. Human mucosal addressin cell adhesion molecule-1 is preferentially expressed in intestinal tract and associated lymphoid tissues. *Am J Pathol*. 1997;151:97-110.
17. Kraal G, Schornagel K, Streeter PR et al. Expression of the mucosal vascular addressin, MAdCAM-1 on sinus lining cells in the spleen. *Am J Pathol*. 1995;147:763-771.
18. Chan JKC, Ng CS, Isaacson PG. Relationship between high-grade lymphoma and low-grade B cell mucosa associated lymphoid tissue lymphoma (MALToma) of the stomach. *Am J Pathol*. 1990;136:1153-1164.
19. De Jong D, Boot H, Van Heerde P et al. Histological grading and gastric lymphoma: pre-treatment criteria and clinical relevance. *Gastroenterology*. 1997;112:1466-1474.
20. Wotherspoon AC, Ortiz-Hidalgo C, Falzon MR et al. Helicobacter pylori-associated gastritis and primary B-cell

- gastric lymphoma. *Lancet*. 1991;338:1175-1176.
21. Nakamura S, Yao T, Aoyagi K et al. *Helicobacter pylori* and primary gastric lymphoma. A histopathologic and immunohistochemical analysis of 237 patients. *Cancer*. 1997;79:3-11.
 22. Nakamura S, Aoyagi K, Fruruse M et al. B-cell monoclonality precedes the development of gastric MALT lymphoma in *Helicobacter pylori*-associated chronic gastritis. *Am J Pathol*. 1998;152:1271-1279.
 23. Doglioni C, Wotherspoon AC, Moschini A et al. High incidence of primary gastric lymphoma in Northeastern Italy. *Lancet* 1992;339:834-835.
 24. Parsonnet J, Hansen S, Rodriguez L et al. *Helicobacter pylori* infection and gastric lymphoma. *N Engl J Med*. 1994;330:1267-71.
 25. Hussell T, Isaacson PG, Crabtree JE et al. The response of cells from low-grade B-cell gastric lymphomas of mucosa-associated lymphoid tissue to *Helicobacter pylori*. *Lancet*. 1993;342:571-574.
 26. Wotherspoon AC, Doglioni C, Diss TC, et al. Regression of primary low-grade B-cell gastric lymphoma of mucosa-associated lymphoid tissue type after eradication of *Helicobacter pylori*. *Lancet*. 1993;342:575-577.
 27. Diss TC, Pan L. Polymerase chain reaction in the assessment of lymphomas. *Cancer Surv*. 1997;30:21-44.
 28. Hsi E, Siddiqui J, Alkan S. Analysis of immunoglobulin heavy chain gene rearrangement in myoepithelial sialadenitis by polymerase chain reaction. *Mod Pathol*. 1994;7:111A.
 29. Sorrentino D, Ferraccili G, DeVita S et al. B-cell clonality and infection with *Helicobacter pylori*: Implications for development of gastric lymphoma. *Gut*. 1996;38:837-840.
 30. Soni M, Shabbab I, Fitzgerald M et al. Detection of clonality in B-cell proliferations in *Helicobacter pylori* induced chronic gastritis in pediatric patients. *Mod Pathol*. 1997;10:65A.
 31. Savio A, Franzin G, Wotherspoon AC et al. Diagnosis and posttreatment follow-up of *Helicobacter Pylori*-positive gastric lymphoma of mucosa-associated lymphoid tissue: Histology, polymerase chain reaction or both? *Blood*. 1996;87:1255-1260.
 32. De Mascarel A, Dubus P, Belleanne G et al. Low prevalence of monoclonal B-cells in *Helicobacter pylori* gastritis patients with duodenal ulcer. *Hum Pathol*. 1998;29:784-790.
 33. Sackmann M, Morgner A, Rudolph B et al. Regression of gastric MALT lymphoma after eradication of *Helicobacter pylori* is predicted by endosonographic staging. MALT Lymphoma Study Group. *Gastroenterology*. 1997;113:1087-1090.
 34. Nakamura S, Matsumoto T, Suekane H et al. Predictive value of endoscopic ultrasonography for regression of gastric low grade and high grade MALT lymphomas after eradication of *Helicobacter pylori*. *Gut*. 2001;48:454-460
 35. Willis TG, Jaydayel DM, Du MQ et al. *Bcl10* is involved in t(1;14)(p22;q32) of MALT B cell lymphoma and mutated in multiple tumor types. *Cell*. 1999;96:35-45.
 36. Dierlamm J, Baens M, Wlodarska I et al. The apoptosis inhibitor gene API2 and a novel 18q gene, MLT, are recurrently rearranged in the t(11;18)(q21;q21)p6s associated with mucosa-associated lymphoid tissue lymphomas. *Blood*. 1999;93:3601-3609.
 37. Ye H, Dogan A, Karran L et al. BCL10 expression in normal and neoplastic lymphoid tissue. Nuclear localization in MALT lymphoma. *Am J Pathol*. 2000;157:1147-54.
 38. Liu H, Ruskone-Fourmestraux A, Lavergne-Slove A et al. Gastric MALT Lymphoma with t(11;18)(q21;q21) fails to respond to *Helicobacter pylori* eradication therapy. *Lancet*. 2001;357:39-40
 39. Liu H, Ye H, Dogan A et al. T(11;18)(q21;q21) is associated with more advanced MALT lymphoma that expresses nuclear BCL10. *Blood*. In press.

II. The Molecular Biology of MALT Lymphoma

1. Harris NL, Jaffe ES, Stein H, et al. A revised European-American classification of lymphoid neoplasms: a proposal from the International Lymphoma Study Group. *Blood*. 1994;84:1361-1392
2. de Wolf-Peters C, Pittaluga S, Dierlamm J, Wlodarska I, Van Den Berghe H. Marginal zone B-cell lymphomas including mucosa-associated lymphoid tissue type lymphoma (MALT), monocytoid B-cell lymphoma and splenic marginal zone cell lymphoma and their relation to the reactive marginal zone. *Leuk Lymphoma*. 1997;26:467-478.
3. Campo E, Miquel R, Krenacs L, Sorbara L, Raffeld M, Jaffe ES. Primary nodal marginal zone lymphomas of splenic and MALT type. *Am J Surg Pathol*. 1999;23:59-68
4. Mollejo M, Menarguez J, Lloret E, et al. Splenic marginal zone lymphoma: a distinctive type of low-grade B-cell lymphoma. A clinicopathological study of 13 cases. *Am J Surg Pathol*. 1995;19:1146-1157
5. Isaacson PG, Matutes E, Burke M, Catovsky D. The histopathology of splenic lymphoma with villous lymphocytes. *Blood*. 1994;84:3828-3834
6. Nathwani BN, Anderson JR, Armitage JO, et al. Marginal zone B-cell lymphoma: A clinical comparison of nodal and mucosa-associated lymphoid tissue types. Non-Hodgkin's Lymphoma Classification Project. *J Clin Oncol*. 1999;17:2486-2492.
7. Dierlamm J, Pittaluga S, Wlodarska I, et al. Marginal zone B-cell lymphomas of different sites share similar cytogenetic and morphologic features [see comments]. *Blood*. 1996;87:299-307
8. Harris NL, Jaffe ES, Diebold J, et al. World Health Organization classification of neoplastic diseases of the hematopoietic and lymphoid tissues: report of the Clinical Advisory Committee meeting-Airlie House, Virginia, November 1997. *J Clin Oncol*. 1999;17:3835-3849
9. Isaacson PG. Gastric MALT lymphoma: from concept to cure [In Process Citation]. *Ann Oncol*. 1999;10:637-645
10. Levine EG, Arthur DC, Machnicki J, et al. Four new recurring translocations in non-Hodgkin lymphoma. *Blood*. 1989;74:1796-1800.
11. Horsman D, Gascoyne R, Klasa R, Coupland R. t(11;18)(q21;q21.1): a recurring translocation in lymphomas of mucosa-associated lymphoid tissue (Malt)? *Genes Chromosomes Cancer*. 1992;4:183-187
12. Auer IA, Gascoyne RD, Connors JM, et al. t(11;18)(q21;q21) is the most common translocation in Malt lymphomas. *Ann Oncol*. 1997;8:979-985
13. Ott G, Katzenberger T, Greiner A, et al. The t(11;18)(q21;q21) chromosome translocation is a frequent and specific aberration in low-grade but not high-grade malignant non-Hodgkin's lymphomas of the mucosa-associated lymphoid tissue (MALT-) type. *Cancer Res*. 1997;57:3944-3948
14. Wotherspoon AC, Pan LX, Diss TC, Isaacson PG. Cytogenetic study of B-cell lymphoma of mucosa-associated lymphoid tissue. *Cancer Genet Cytogenet*. 1992;58:35-38.
15. Willis TG, Jaydayel DM, Du MQ, et al. Bcl10 is involved in t(1;14)(p22;q32) of MALT B cell lymphoma and mutated in multiple tumor types. *Cell*. 1999;96:35-45
16. Zhang Q, Siebert R, Yan M, et al. Inactivating mutations and overexpression of BCL10, a caspase recruitment domain-containing gene, in MALT lymphoma with t(1;14)(p22;q32). *Nat Genet*. 1999;22:63-68.
17. Dierlamm J, Pittaluga S, Stul M, et al. BCL6 gene rearrangements also occur in marginal zone B-cell lymphoma. *Br J Haematol*. 1997;98:719-725
18. Offit K, Lo Coco F, Louie DC, et al. Rearrangement of the bcl-6 gene as a prognostic marker in diffuse large-cell lymphoma. *N*

- Engl J Med. 1994;331:74-80
19. Remstein ED, James CD, Kurtin PJ. Incidence and subtype specificity of API2-MALT1 fusion translocations in extranodal, nodal, and splenic marginal zone lymphomas. *Am J Pathol.* 2000;156:1183-1188.
 20. Rosenwald A, Ott G, Stilgenbauer S, et al. Exclusive detection of the t(11;18)(q21;q21) in extranodal marginal zone B cell lymphomas (MZBL) of MALT type in contrast to other MZBL and extranodal large B cell lymphomas. *Am J Pathol.* 1999;155:1817-1821
 21. Sole F, Woessner S, Florensa L, et al. Frequent involvement of chromosomes 1, 3, 7 and 8 in splenic marginal zone B-cell lymphoma. *Br J Haematol.* 1997;98:446-449
 22. Mateo M, Mollejo M, Villuendas R, et al. 7q31-32 allelic loss is a frequent finding in splenic marginal zone lymphoma. *Am J Pathol.* 1999;154:1583-1589.
 23. Sole F, Salido M, Espinet B, et al. Splenic marginal zone B-cell lymphomas: two cytogenetic subtypes, one with gain of 3q and the other with loss of 7q. *Haematologica.* 2001;86:71-77.
 24. Wotherspoon AC, Finn TM, Isaacson PG. Trisomy 3 in low-grade B-cell lymphomas of mucosa-associated lymphoid tissue. *Blood.* 1995;85:2000-2004.
 25. Brynes RK, Almaguer PD, Leathery KE, et al. Numerical cytogenetic abnormalities of chromosomes 3, 7, and 12 in marginal zone B-cell lymphomas. *Modern Pathology.* 1996;9:995-1000
 26. Offit K, Jhanwar SC, Ladanyi M, Filippa DA, Chaganti RS. Cytogenetic analysis of 434 consecutively ascertained specimens of non-Hodgkin's lymphoma: correlations between recurrent aberrations, histology, and exposure to cytotoxic treatment. *Genes Chromosomes Cancer.* 1991;3:189-201
 27. Horsman DE, Connors JM, Pantzar T, Gascoyne RD. Analysis of secondary chromosomal alterations in 165 cases of follicular lymphoma with t(14;18). *Genes Chromosomes Cancer.* 2001;30:375-382.
 28. Ott G, Kalla J, Steinhoff A, et al. Trisomy 3 is not a common feature in malignant lymphomas of mucosa-associated lymphoid tissue type. *American Journal of Pathology.* 1998;153:689-694
 29. Hoeve MA, Gisbertz IA, Schouten HC, et al. Gastric low-grade MALT lymphoma, high-grade MALT lymphoma and diffuse large B cell lymphoma show different frequencies of trisomy. *Leukemia.* 1999;13:799-807.
 30. Peng H, Diss T, Isaacson PG, Pan L. c-myc gene abnormalities in mucosa-associated lymphoid tissue (MALT) lymphomas. *J Pathol.* 1997;181:381-386.
 31. Gamberi B, Gaidano G, Parsa N, et al. Microsatellite instability is rare in B-cell non-Hodgkin's lymphomas. *Blood.* 1997;89:975-979
 32. Peng H, Chen G, Du M, Singh N, Isaacson PG, Pan L. Replication error phenotype and p53 gene mutation in lymphomas of mucosa-associated lymphoid tissue. *Am J Pathol.* 1996;148:643-648.
 33. Furlan D, Bertoni F, Cerutti R, et al. Microsatellite instability in gastric MALT lymphomas and other associated neoplasms. *Ann Oncol.* 1999;10:783-788.
 34. Starostik P, Greiner A, Schwarz S, Patzner J, Schultz A, Muller-Hermelink HK. The role of microsatellite instability in gastric low- and high-grade lymphoma development. *Am J Pathol.* 2000;157:1129-1136.
 35. Du M, Peng H, Singh N, Isaacson PG, Pan L. The accumulation of p53 abnormalities is associated with progression of mucosa-associated lymphoid tissue lymphoma. *Blood.* 1995;86:4587-4593.
 36. Martinez-Delgado B, Fernandez-Piqueras J, Garcia MJ, et al. Hypermethylation of a 5' CpG island of p16 is a frequent event in non-Hodgkin's lymphoma. *Leukemia.* 1997;11:425-428.
 37. Baens M, Maes B, Steyls A, Geboes K, Marynen P, De Wolf-Peeters C. The product of the t(11;18), an API2-MLT fusion, marks nearly half of gastric MALT type lymphomas without large cell proliferation. *Am J Pathol.* 2000;156:1433-1439.
 38. Qi Y, Gabrea A, Sawyer J, et al. The t(6;14)(p21;q32) translocation causes dysregulation of cyclin D3 in multiple myeloma. *Blood.* 2000;96:86a
 39. Du M, Diss TC, Xu C, Peng H, Isaacson PG, Pan L. Ongoing mutation in MALT lymphoma immunoglobulin gene suggests that antigen stimulation plays a role in the clonal expansion. *Leukemia.* 1996;10:1190-1197
 40. Bertoni F, Cazzaniga G, Bosshard G, et al. Immunoglobulin heavy chain diversity genes rearrangement pattern indicates that MALT-type gastric lymphoma B cells have undergone an antigen selection process. *Br J Haematol.* 1997;97:830-836.
 41. Miklos JA, Swerdlow SH, Bahler DW. Salivary gland mucosa-associated lymphoid tissue lymphoma immunoglobulin V(H) genes show frequent use of V1-69 with distinctive CDR3 features. *Blood.* 2000;95:3878-3884.
 42. Dierlamm J, Wlodarska I, Michaux L, et al. Genetic abnormalities in marginal zone B-cell lymphoma. *Hematol Oncol.* 2000;18:1-13.
 43. Dierlamm J, Baens M, Wlodarska I, et al. The apoptosis inhibitor gene API2 and a novel 18q gene, MLT, are recurrently rearranged in the t(11;18)(q21;q21) associated with mucosa-associated lymphoid tissue lymphomas [In Process Citation]. *Blood.* 1999;93:3601-3609
 44. Morgan JA, Yin Y, Borowsky AD, et al. Breakpoints of the t(11;18)(q21;q21) in mucosa-associated lymphoid tissue (MALT) lymphoma lie within or near the previously undescribed gene MALT1 in chromosome 18. *Cancer Res.* 1999;59:6205-6213
 45. Akagi T, Motegi M, Tamura A, et al. A novel gene, MALT1 at 18q21, is involved in t(11;18)(q21;q21) found in low-grade B-cell lymphoma of mucosa-associated lymphoid tissue. *Oncogene.* 1999;18:5785-5794.
 46. Baens M, Steyls A, Dierlamm J, De Wolf-Peeters C, Marynen P. Structure of the MLT gene and molecular characterization of the genomic breakpoint junctions in the t(11;18)(q21;q21) of marginal zone B-cell lymphomas of MALT type. *Genes Chromosomes Cancer.* 2000;29:281-291.
 47. Motegi M, Yonezumi M, Suzuki H, et al. API2-MALT1 chimeric transcripts involved in mucosa-associated lymphoid tissue type lymphoma predict heterogeneous products. *Am J Pathol.* 2000;156:807-812.
 48. Uren GA, O'Rourke K, Aravind L, et al. Identification of paracaspases and metacaspases: two ancient families of caspase-like proteins, one of which plays a key role in MALT lymphoma. *Mol Cell.* 2000;6:961-967.
 49. Reed JC. Mechanisms of apoptosis. *Am J Pathol.* 2000;157:1415-1430.
 50. Lucas PC, Yonezumi M, Inohara N, et al. Bcl10 and MALT1, independent targets of chromosomal translocation in MALT Lymphoma, cooperate in a novel NF- κ B signaling pathway. *J Biol Chem.* 2001;276:19012-19019.
 51. Ghosh S, May MJ, Kopp EB. NF- κ B and Rel proteins: evolutionarily conserved mediators of immune responses. *Annu Rev Immunol.* 1998;16:225-260
 52. Karin M, Ben-Neriah Y. Phosphorylation meets ubiquitination: the control of NF- κ B activity. *Annu Rev Immunol.* 2000;18:621-663
 53. Ruland J, Duncan GS, Elia A, et al. Bcl10 is a positive regulator of antigen receptor-induced activation of NF- κ B and neural tube closure. *Cell.* 2001;104:33-42.
 54. Thome M, Martinon F, Hofmann K, et al. Equine herpesvirus-2 E10 gene product, but not its cellular homologue, activates NF- κ B transcription factor and c-Jun N-terminal kinase. *J Biol*

Chem. 1999;274:9962-9968.

55. Yan M, Lee J, Schilbach S, Goddard A, Dixit V. mE10, a novel caspase recruitment domain-containing proapoptotic molecule. *J Biol Chem.* 1999;274:10287-10292.
56. Fakruddin JM, Chaganti RS, Murty VV. Lack of BCL10 mutations in germ cell tumors and B cell lymphomas. *Cell.* 1999;97:683-684; discussion 686-688.
57. Gill S, Broni J, Jefferies S, et al. BCL10 is rarely mutated in human prostate carcinoma, small-cell lung cancer, head and neck tumours, renal carcinoma and sarcomas. MPT Collaborators, St George's Hospital Collaborators. *Br J Cancer.* 1999;80:1565-1568.
58. Lambers AR, Gumbs C, Ali S, et al. Bcl10 is not a target for frequent mutation in human carcinomas. *Br J Cancer.* 1999;80:1575-1576.
59. van Schothorst EM, Mohkamsing S, van Gurp RJ, Oosterhuis JW, van der Saag PT, Looijenga LH. Lack of Bcl10 mutations in testicular germ cell tumours and derived cell lines. *Br J Cancer.* 1999;80:1571-1574.
60. Yuille MR, Stone JG, Bradshaw PS, Houlston RS. Bcl10 in chronic lymphocytic leukaemia and T-cell prolymphocytic leukaemia. *Br J Haematol.* 1999;107:384-385.
61. Shih LY, Fu JF, Shurtleff SA, Morris SW, Downing JR. Lack of BCL10 mutations in multiple myeloma and plasma cell leukemia. *Genes Chromosomes Cancer.* 2001;30:402-406.
62. Dyer MJ. Bcl10 mutations in malignancy. *Br J Cancer.* 1999;80:1491.
63. Du MQ, Peng H, Liu H, et al. BCL10 gene mutation in lymphoma. *Blood.* 2000;95:3885-3890.
64. Yoneda T, Imaizumi K, Maeda M, et al. Regulatory mechanisms of TRAF2-mediated signal transduction by Bcl10, a MALT lymphoma-associated protein. *J Biol Chem.* 2000;275:11114-11120.
65. Zhang Q, Cui X, Sangster MY, et al. Selective hyperexpansion of marginal zone B cells in E_μ-BCL10 mice. *Blood.* 2000;96:822a
66. Wotherspoon AC, Diss TC, Pan L, Singh N, Whelan J, Isaacson PG. Low grade gastric B-cell lymphoma of mucosa associated lymphoid tissue in immunocompromised patients. *Histopathology.* 1996;28:129-134
67. Bertin J, Wang L, Guo Y, et al. CARD11 and CARD14 are novel caspase recruitment domain (CARD)/membrane-associated guanylate kinase (MAGUK) family members that interact with BCL10 and activate NF-κB. *J Biol Chem.* 2001;276:11877-11882.
68. Bertin J, Guo Y, Wang L, et al. CARD9 is a novel caspase recruitment domain-containing protein that interacts with BCL10/CLAP and activates NF-κB. *J Biol Chem.* 2000;275:41082-41086.
69. Wang L, Guo Y, Ke X, et al. CARD10 Is a Novel CARD/MAGUK Family Member That Interacts with BCL10 and Activates NF-κB. *J Biol Chem.* 2001;20:20
70. Gaide O, Martinon F, Micheau O, Bonnet D, Thome M, Tschopp J. Carma1, a CARD-containing binding partner of Bcl10, induces Bcl10 phosphorylation and NF-κB activation(1). *FEBS Lett.* 2001;496:121-127.
71. Ye H, Dogan A, Karran L, et al. BCL10 expression in normal and neoplastic lymphoid tissue. Nuclear localization in MALT lymphoma. *Am J Pathol.* 2000;157:1147-1154.
72. Ye H, Liu H, Dogan A, et al. MALT lymphoma with t(11;18)(q21;q21) expresses nuclear BCL10. *Blood.* 2000;96:468a
73. Liu H, Ruskone-Fourmesttraux A, Lavergne-Slove A, et al. Gastric MALT lymphoma with t(11;18)(q21;q21) fails to respond to *Helicobacter pylori* eradication therapy. *Blood.* 2000;96:468a

III. Low Grade Gastric MALT Lymphoma

1. Zucca E, Bertoni F, Roggero E, Cavalli F. The gastric marginal zone B-cell lymphoma of MALT type. *Blood.* 2000;96:410-419.
2. Zucca E, Roggero E, Pileri S. B-cell lymphoma of MALT type: a review with special emphasis on diagnostic and management problems of low-grade gastric tumours. *Br J Haematol.* 1998;100:3-14.
3. Parsonnet J, Hansen S, Rodriguez L, et al. *Helicobacter pylori* infection and gastric lymphoma. *N Engl J Med.* 1994;330:1267-1271.
4. Hussell T, Isaacson PG, Crabtree JE, Spencer J. *Helicobacter pylori*-specific tumour-infiltrating T cells provide contact dependent help for the growth of malignant B cells in low-grade gastric lymphoma of mucosa-associated lymphoid tissue. *J Pathol.* 1996;178:122-127
5. Zucca E, Bertoni F, Roggero E, et al. Molecular analysis of the progression from *Helicobacter pylori*-associated chronic gastritis to mucosa-associated lymphoid-tissue lymphoma of the stomach. *N Engl J Med.* 1998;338:804-810.
6. Pinotti G, Zucca E, Roggero E, et al. Clinical features, treatment and outcome in a series of 93 patients with low-grade gastric MALT lymphoma. *Leuk Lymphoma.* 1997;26:527-537.
7. Roggero E, Zucca E, Pinotti G, et al. Eradication of *Helicobacter pylori* infection in primary low-grade gastric lymphoma of mucosa-associated lymphoid tissue. *Ann Intern Med.* 1995;122:767-769.
8. Bayerdorffer E, Neubauer A, Rudolph B, et al. Regression of primary gastric lymphoma of mucosa-associated lymphoid tissue type after cure of *Helicobacter pylori* infection: MALT Lymphoma Study Group. *Lancet.* 1995;345:1591-1594.
9. Montalban C, Manzanal A, Boixeda D, et al. *Helicobacter pylori* eradication for the treatment of low-grade gastric MALT lymphoma: follow-up together with sequential molecular studies. *Ann Oncol.* 1997;8(suppl 2):37-40.
10. Steinbach G, Ford R, Globler G, et al. Antibiotic treatment of gastric lymphoma of mucosa-associated lymphoid tissue: an uncontrolled trial. *Ann Intern Med.* 1999;131:88-95.
11. Neubauer A, Thiede C, Morgner A, et al. Cure of *Helicobacter pylori* infection and duration of remission of low-grade gastric mucosa-associated lymphoid tissue lymphoma. *J Natl Cancer Inst.* 1997;89:1350-1355.
12. Ruskone Formesttraux A, Lavergne A, Aegerter PH et al. Predictive factors for regression of gastric MALT lymphoma after anti-*Helicobacter pylori* treatment. *Gut.* 2001;48:297-303.
13. Bertoni F, Cazzaniga G, Bosshard G, et al. Immunoglobulin heavy chain diversity genes rearrangement pattern indicates that MALT-type gastric lymphoma B cells have undergone an antigen selection process. *Br J Haematol.* 1997;97:830-836.
14. Taal BG, Boot H, van Heerde P, de Jong D, Hart AA, Burgers JM. Primary non-Hodgkin lymphoma of the stomach: endoscopic pattern and prognosis in low versus high grade malignancy in relation to the MALT concept. *Gut.* 1996;39:556-561.
15. de Jong D, Aleman BM, Taal BG, Boot H. Controversies and consensus in the diagnosis, work-up and treatment of gastric lymphoma: an international survey. *Ann Oncol.* 1999;10:275-280.
16. Rohatiner A, d'Amore F, Coiffier B, et al. Report on a workshop convened to discuss the pathological and staging classifications of gastrointestinal tract lymphoma. *Ann Oncol.* 1994;5:397-400.
17. Montalban C, Castrillo JM, Abaira V, et al. Gastric B-cell mucosa-associated lymphoid tissue (MALT) lymphoma: clinicopathological study and evaluation of the prognostic factors in 143 patients. *Ann Oncol.* 1995;6:355-362.

18. Raderer M, Vorbeck F, Formanek M, et al. Importance of extensive staging in patients with mucosa-associated lymphoid tissue (MALT)-type lymphoma. *Br J Cancer*. 2000;83:454-7.
19. Fisher RI, Dahlborg S, Nathwani BN, Banks PM, Miller TP, Grogan TM. A clinical analysis of two indolent lymphoma entities: mantle cell lymphoma and marginal zone lymphoma (including the mucosa-associated lymphoid tissue and monocytoid B-cell subcategories): a Southwest Oncology Group study. *Blood*. 1995;85:1075-1082.
20. Cogliatti SB, Schmid U, Schumacher U, et al. Primary B-cell gastric lymphoma: a clinicopathological study of 145 patients. *Gastroenterology*. 1991;101:1159-1170.
21. The Non-Hodgkin's Lymphoma Classification Project. A clinical evaluation of the International Lymphoma Study Group classification of non-Hodgkin's lymphoma. *Blood*. 1997;89:3909-3918
22. Thieblemont C, Bastion Y, Berger F, et al. Mucosa-associated lymphoid tissue gastrointestinal and nongastrointestinal lymphoma behavior: analysis of 108 patients. *J Clin Oncol*. 1997;15:1624-1630
23. Eidt S, Stolte M, Fischer R. Factors influencing lymph node infiltration in primary gastric malignant lymphoma of the mucosa-associated lymphoid tissue. *Pathol Res Pract*. 1994;190:1077-1081.
24. Pavlick AC, Gerdes H, Portlock CS. Endoscopic ultrasound in the evaluation of gastric small lymphocytic mucosa-associated lymphoid tumors. *J Clin Oncol*. 1997;15:1761-1766.
25. Sackmann M, Morgner A, Rudolph B, et al. Regression of gastric MALT lymphoma after eradication of *Helicobacter pylori* is predicted by endosonographic staging: MALT Lymphoma Study Group. *Gastroenterology*. 1997;113:1087-1090
26. Nobre-Leitao C, Lage P, Cravo M, et al. Treatment of gastric MALT lymphoma by *Helicobacter pylori* eradication: a study controlled by endoscopic ultrasonography. *Am J Gastroenterol*. 1998;93:732-736.
27. Weston AP, Banerjee SK, Horvat RT, Zoubine MN, Campbell DR, Cherian R. Prospective long-term endoscopic and histologic follow-up of gastric lymphoproliferative disease of early stage I_e low-grade B-cell mucosa-associated lymphoid tissue type following *Helicobacter pylori* eradication treatment. *Int J Oncol*. 1999;15:899-907.
28. Savio A, Franzin G, Wotherspoon AC, et al. Diagnosis and posttreatment follow-up of *Helicobacter pylori*-positive gastric lymphoma of mucosa-associated lymphoid tissue: histology, polymerase chain reaction, or both? *Blood*. 1996;87:1255-1260.
29. Schechter NR, Portlock CS, Yahalom J. Treatment of mucosa-associated lymphoid tissue lymphoma of the stomach with radiation alone. *J Clin Oncol*. 1998;16:1916-1921.
30. Zucca E, Pinotti G, Roggero E, et al. High incidence of other neoplasms in patients with low-grade gastric MALT lymphoma. *Ann Oncol*. 1995;6:726-728.
31. Montalban C, Castrillo JM, Lopez-Abente G, et al. Other cancers in patients with gastric MALT lymphoma. *Leuk Lymphoma*. 1999;33:161-168.
32. Au WY, Gascoyne RD, Le N, et al. Incidence of second neoplasms in patients with MALT lymphoma: no increase in risk above the background population. *Ann Oncol*. 1999;10:317-321.
33. Zucca E, Roggero E, Traulle C, et al. Early interim report of the LY03 randomised cooperative trial of observation vs chlorambucil after anti *Helicobacter* therapy in low-grade gastric lymphoma. *Ann Oncol*. 1999;10(suppl 3):25.
34. Zucca E, Bertoni F, Roggero E, Gisi M, Cavalli F. Patient-specific molecular monitoring of MALT-lymphoma after antibiotic treatment. *Proc Am Soc Clin Oncol*. 1999;18:10a.
35. Thiede C, Wundisch T, Alpen B, et al. Long-term persistence of monoclonal B cells after cure of *Helicobacter pylori* infection and complete histologic remission in gastric mucosa-associated lymphoid tissue B-cell lymphoma. *J Clin Oncol*. 2001;19:1600-9.
36. Richards MA, Gregory WM, Hall P, et al. Management of localized non-Hodgkin's lymphoma: the experience at St. Bartholomew's Hospital 1972-1985. *Hematol Oncol*. 1989;7:1-18.
37. Hammel P, Haioun C, Chaumette MT, et al. Efficacy of single-agent chemotherapy in low-grade B-cell mucosa-associated lymphoid tissue lymphoma with prominent gastric expression. *J Clin Oncol*. 1995;13:2524-2529.
38. Conconi A, Thieblemont C, Martinelli G, et al. An International Extranodal Lymphoma Study group Phase II study of Rituximab in extranodal marginal zone B-cell lymphomas (MZL). *Proc ASCO* 2001; 20(part1):296a (abs#1183).
39. Fung CY, Grossbard ML, Linggood RM, et al. Mucosa-associated lymphoid tissue lymphoma of the stomach: long term outcome after local treatment. *Cancer*. 1999;85:9-17.
40. Zucca E, Roggero E, Bertoni F, Cavalli F. Primary extranodal non-Hodgkin's lymphomas Part 1: gastrointestinal, cutaneous and genitourinary lymphomas. *Ann Oncol*. 1997;8:727-737.
41. Coiffier B, Salles G. Does surgery belong to medical history for gastric lymphomas? *Ann Oncol*. 1997;8:419-421.

IV. Non-Gastric Marginal Zone B-Cell Lymphoma of MALT Type

1. Zucca E, Bertoni F, Roggero E, Cavalli F: The gastric marginal zone B-cell lymphoma of MALT type. *Blood*. 2000;96:410-419.
2. Isaacson P. Gastric MALT lymphoma: from concept to cure. *Ann Oncol*. 1999;10:637-645.
3. Isaacson P. Mucosa-associated lymphoid tissue lymphoma. *Semin Hematol*. 1999;36:139-147
4. Zucca E, Bertoni F, Roggero E, et al. Molecular analysis of the progression from *Helicobacter pylori*-associated chronic gastritis to mucosa-associated lymphoid-tissue lymphoma of the stomach. *N Engl J Med*. 1998;338:804-810.
5. Jonsson V, Wiik A, Hou-Jensen K, et al. Autoimmunity and extranodal lymphocytic infiltrates in lymphoproliferative disorders. *J Intern Med*. 1999;245:277-286.
6. Hyjek E, Isaacson PG. Primary B cell lymphoma of the thyroid and its relationship to Hashimoto's thyroiditis. *Hum Pathol*. 1988;19:1315-1326.
7. Hyjek E, Smith WJ, Isaacson PG. Primary B-cell lymphoma of salivary glands and its relationship to myoepithelial sialadenitis. *Hum Pathol*. 1988;19:766-776.
8. Roggero E, Zucca E, Mainetti C, et al. Eradication of *Borrelia burgdorferi* infection in primary marginal zone B-cell lymphoma of the skin. *Hum Pathol*. 2000;31:263-8.
9. De Re V, De Vita S, Marzotto A, et al. Sequence analysis of the immunoglobulin antigen receptor of hepatitis C virus-associated non-Hodgkin lymphomas suggests that the malignant cells are derived from the rheumatoid factor-producing cells that occur mainly in type II cryoglobulinemia. *Blood*. 2000;96:3578-3584.
10. Zucca E, Roggero E, Maggi-Solca N, et al. Prevalence of *Helicobacter pylori* and hepatitis C virus infections among non-Hodgkin's lymphoma patients in Southern Switzerland. *Haematologica*. 2000;85:147-53.
11. Harris NL, Jaffe ES, Stein H, et al. A revised European-American classification of lymphoid neoplasms: a proposal from the International Lymphoma Study Group. *Blood*. 1994;84:1361-1392.
12. Harris NL, Jaffe ES, Diebold J, et al. The World Health Organization classification of neoplastic diseases of the hematopoietic and lymphoid tissues: report of the clinical

- advisory committee meeting, Airlie House, Virginia, November, 1997. *Ann Oncol.* 1999;10:1419-1432.
13. The Non-Hodgkin's Lymphoma Classification Project. A clinical evaluation of the International Lymphoma Study Group classification of non-Hodgkin's lymphoma. *Blood.* 1997;89:3909-3918.
 14. Thieblemont C, Bastion Y, Berger F, et al. Mucosa-associated lymphoid tissue gastrointestinal and nongastrointestinal lymphoma behavior: Analysis of 108 patients. *J Clin Oncol.* 1997;15:1624-1630.
 15. Zinzani PL, Magagnoli M, Ascani S, et al. Nongastrointestinal mucosa-associated lymphoid tissue (MALT) lymphomas: Clinical and therapeutic features of 24 localized patients. *Ann Oncol.* 1997;8:883-886.
 16. Thieblemont C, Berger F, Dumontet C, et al. Mucosa-associated lymphoid tissue lymphoma is a disseminated disease in one third of 158 patients analyzed. *Blood.* 2000;95:802-806.
 17. Zinzani P, Magagnoli M, Galieni P, et al. Nongastrointestinal low-grade mucosa-associated lymphoid tissue lymphoma: analysis of 75 patients. *J Clin Oncol.* 1999;17:1254-1258.
 18. Zucca E, Roggero E, Bertoni F, Conconi A, Cavalli F. Primary extranodal non-Hodgkin's lymphomas. Part 2: Head and neck, central nervous system and other less common sites. *Ann Oncol.* 1999;10:1023-1033.
 19. Zucca E, Roggero E, Bertoni F, Cavalli F. Primary extranodal non-Hodgkin's lymphomas. Part 1: Gastrointestinal, cutaneous and genitourinary lymphomas. *Ann Oncol.* 1997;8:727-37.
 20. Kaplan MA, Pettit CL, Zukerberg LR, et al. Primary lymphoma of the trachea with morphologic and immunophenotypic characteristics of low-grade B-cell lymphoma mucosa-associated lymphoid tissue. *Am J Surg Pathol.* 1992;16:71-75.
 21. Nicholson AG, Wotherspoon AC, Diss TC, et al. Pulmonary B-cell non-Hodgkin's lymphomas: The value of immunohistochemistry and gene analysis in diagnosis. *Histopathology.* 1995;26:395-403.
 22. Isaacson PG, Chan JKC, Tang C, et al. Low-grade B-cell lymphoma of mucosa-associated lymphoid tissue arising in the thymus: A thymic lymphoma mimicking myoepithelial sialadenitis. *Am J Surg Pathol.* 1990;14:342-351.
 23. Mattia AR, Ferry JA, Harris NL. Breast lymphoma: A B-cell spectrum including the low grade B-cell lymphoma of mucosa associated lymphoid tissue. *Am J Surg Pathol.* 1993;17:574-587.
 24. Bailey EM, Ferry JA, Harris NL, et al. Marginal zone lymphoma (low-grade B-cell lymphoma of mucosa-associated lymphoid tissue type) of skin and subcutaneous tissue. *Am J Surg Pathol.* 1996;20:1011-1023.
 25. Isaacson PG, Banks PM, Best PV, et al. Primary low-grade hepatic B-cell lymphoma of mucosa-associated lymphoid tissue (MALT)-type. *Am J Surg Pathol.* 1995;19:571-575.
 26. Wotherspoon AC, Hardman-Lea S, Isaacson PG. Mucosa-associated lymphoid tissue (MALT) in the human conjunctiva. *J Pathol.* 1994;174:33-37.
 27. Calvo R, Ribera JM, Vaquero M, et al. Low-grade, MALT-type, primary B-cell lymphoma of the conjunctiva. *Leuk Lymphoma.* 1997;28:203-207.
 28. Nicholson AG, Wotherspoon AC, Jones AL, et al. Pulmonary B-cell non-Hodgkin's lymphoma associated with autoimmune disorders: A clinicopathological review of six cases. *Eur Respir J.* 1996;9:2022-2025.
 29. Parveen T, Navarro-Roman L, Medeiros LJ, Raffeld M, Jaffe ES. Low-grade B-cell lymphoma of mucosa-associated lymphoid tissue arising in the kidney. *Arch Pathol Lab Med.* 1993;117:780-3.
 30. Kumar S, Kumar D, Kaldjian EP, et al. Primary low-grade B-cell lymphoma of the dura: a mucosa associated lymphoid tissue-type lymphoma. *Am J Surg Pathol.* 1997;21:81-7.
 31. Du MQ, Xu CF, Diss TC, et al. Intestinal dissemination of gastric mucosa-associated lymphoid tissue lymphoma. *Blood.* 1996;88:4445-445.
 32. Du M, Peng H, Dogan A, et al. Preferential dissemination of B-cell gastric mucosa-associated lymphoid tissue (MALT) lymphoma to the splenic marginal zone. *Blood.* 1997;90:4071-4077.
 33. Drillenburger P, van der Voort R, Koopman G, et al. Preferential expression of the mucosal homing receptor integrin alpha 4 beta 7 in gastrointestinal non-Hodgkin's lymphomas. *Am J Pathol.* 1997;150:919-927.
 34. Dogan A, Du M, Koulis A, Briskin M, Isaacson P. Expression of lymphocyte homing receptors and vascular addressing in low-grade gastric B-cell lymphomas of mucosa-associated lymphoid tissue. *Am J Pathol.* 1997;151:1361-1369.
 35. Sancho JM, Ribera JM, Vaquero M, et al. Non-gastrointestinal malt lymphomas: a study of 10 cases and comparison with 27 patients with gastrointestinal MALT lymphoma. *Haematologica.* 2000;85:557-9.
 36. Ferrer A, Lopez-Guillermo A, Bosch F, et al. [Non-gastric mucosa-associated lymphoid tissue (MALT) lymphomas: analysis of 14 patients]. *Med Clin (Barc).* 1999;112(15):577-80.
 37. Zucca E, Conconi A, Roggero E, et al. Non-gastric MALT lymphomas: a survey of 369 European patients. The International Extranodal Lymphoma Study Group. *Ann Oncol.* 2000;11:99.
 38. Conconi A, Thieblemont C, Martinelli G, et al. An International Extranodal Lymphoma Study Group (IELSG) phase II study of rituximab in extranodal marginal zone B-cell lymphomas (MZL). *Proc Amer Soc Clin Oncol.* 2001;20:269a.