

Favored Use of Immunoglobulin V_H4 Genes in AIDS-Associated B-Cell Lymphoma

By Alberto Bessudo, Vladimir Cherepakhin, Todd A. Johnson, Laura Z. Rassenti, Ellen Feigal, and Thomas J. Kipps

We examined the Ig heavy chain variable region genes (Ig V_H genes) expressed in biopsy specimens of 10 patients with acquired immunodeficiency syndrome (AIDS)-associated lymphoma. Eight expressed Ig V_H genes of the V_H4 group, indicating a bias toward expression of Ig V_H genes of this subgroup. Sequence analyses of Ig V_H genes isolated from any one lymphoma did not reveal evidence for intraclonal diversity. However, some lymphomas express Ig V_H genes that apparently have undergone somatic diversification and selection. In addition, we found that the sequence encoding

each examined third complementarity determining region most likely resulted from D-D fusion, a process that ordinarily contributes to the generation of a relatively small proportion of the Ig heavy chain genes expressed by normal adult B cells. The noted restriction in the use of Ig V_H genes by AIDS-associated B-cell lymphomas suggests that antigenic stimulation contributes to lymphomagenesis in patients with AIDS.

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PATIENTS WITH acquired immunodeficiency syndrome (AIDS) have a high incidence of B-cell lymphoma.¹⁻⁴ Multiple factors are hypothesized to contribute to this high incidence, including opportunistic viral infections, chronic antigenic stimulation, an acquired tendency for chromosomal translocations, and the human immunodeficiency virus (HIV) itself.⁵⁻⁸ Persons infected with HIV typically develop a persistent antibody response to external viral glycoproteins and internal core antigens.⁹⁻¹¹ Moreover, patients with AIDS often have B-lymphoid hyperplasia and/or hypergammaglobulinemia.¹² Finally, HIV-infected individuals can develop B cells with an altered surface-antigen phenotype¹³ or an impaired *in vitro* response to polysaccharides or pokeweed mitogens,^{14,15} suggesting that these patients may acquire intrinsic B-cell abnormalities that also may play a pathogenic role.

Structural analysis of the Ig genes expressed in AIDS-associated lymphoma can provide insight into the mechanisms involved in B-cell neoplasia. During B-cell development, Ig genes are formed by recombination and assembly of several genes selected from a larger pool of related gene segments. To generate an Ig heavy chain, rearrangements involving a heavy chain variable region gene (Ig V_H gene), a diversity (D) segment(s), and a heavy chain joining (J_H) segment must occur. There are ~50 functional Ig V_H genes in the human haploid genome that are grouped into seven Ig V_H gene subgroups based on similarities in primary structure.^{16,17} Ig V_H genes of the Ig V_H3 subgroup encode the largest portion of Ig in the normal human adult repertoire,¹⁸

in part because of the relatively large number of functional Ig V_H3 genes in the human haploid genome.¹⁹ There also may be nonstochastic use of certain Ig V_H genes^{20,21} because of advantageous accessibility of particular Ig V_H genes to recombinase, enhanced transcriptional activation of particular Ig V_H genes, and/or homology directed joining, as reviewed.²² Selection for or against particular Ig encoded by certain Ig V_H genes also may alter the distribution of Ig V_H gene subgroups expressed in the Ig repertoire. For this reason, finding an abnormal distribution of Ig V_H genes expressed in AIDS-associated lymphomas could reveal selective forces acting on the expressed Ig repertoire that may contribute to the development of lymphoid neoplasia.

In addition, some Ig V_H genes innately encode antibodies that may have binding activity for HIV glycoproteins. HIV gp120, for example, has been found to bind B cells that express Ig encoded by a subset of Ig V_H3 genes.²³ However, patients infected with HIV can develop anti-gp120 antibodies encoded by Ig V_H genes of other Ig V_H gene subgroups,²⁴ as well as make antibodies to other HIV glycoproteins, such as gp41 and p24.^{25,26} Currently, it is not known whether the repertoire of Ig expressed in AIDS-associated lymphomas is biased, let alone biased toward or against Ig V_H3 genes that encode antibodies that innately have binding activity for HIV glycoproteins. This study was performed to evaluate the Ig V_H genes expressed by random cases of AIDS-associated B-cell lymphoma.

MATERIALS AND METHODS

Patient material. We obtained residual tissue from fresh frozen biopsy specimens of lymph nodes or extra-lymphatic lymphoid tissue from AIDS patients with clinical and pathologic diagnosis of high grade lymphoma. The tissue samples were embedded in Tissue-Tek Optimal Cutting Temperature (OCT) compound 4583 (Miles Inc, Elkhart, IN) and preserved at -80°C until RNA and/or DNA was extracted, as described.²⁷

DNA hybridization. Restriction enzyme digested genomic DNA from each lymphoma specimen or from normal human placenta were applied to separate wells of a 0.8% agarose gel for electrophoresis. Electrophoretically size-separated DNA fragments were transferred onto nylon (Amersham, Arlington Heights, IL) for Southern blot analyses, as described.²⁸ Heavy chain Ig gene rearrangements were detected by hybridizing the filters with a ³²P-radiolabeled human J_H gene segment probe, as described.²⁹ Samples were considered to have detectable clonal Ig heavy chain gene rearrangements when they had one or more bands hybridizing with the J_H gene probe that were distinct from the expected germline band(s) present in enzyme-restricted normal human placental DNA.

From the Division of Hematology/Oncology, the Department of Medicine, University of California at San Diego, La Jolla, CA; and the Division of Cancer Treatment, National Cancer Institute, Bethesda, MD.

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Address reprint requests to Thomas J. Kipps, MD, PhD, Division of Hematology/Oncology, Department of Medicine, University of California at San Diego, La Jolla, CA 92093-0663.

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Polymerase chain reaction (PCR). Genomic DNA from lymphoma specimens having detectable Ig heavy chain gene rearrangements were used as templates for PCR, as described.²⁷ Consensus J_H antisense oligonucleotide primer 5'-ggcgaattcACCTGAGGAGACRGTGACC-3' was used in combination with one of the following Ig V_H subgroup-specific leader sense primers: 5'-gtggaattcaagcttGTTCHTACCATGGACTG-3' (Ig V_{H1}); 5'-gtgaagcttATGGACATACTTTGTTCCACG-3' (Ig V_{H2}); 5'-gtgctgagaagcttATGGAGTTKGGGCTGAGCTGG-3' (Ig V_{H3}); 5'-gtggaattcaagcttATGAAACAYCTGTGGTTCTTC-3' (Ig V_{H4}); 5'-gcaagcttatcgatCCATCATGGGGTCAACCG-3' (Ig V_{H5}); 5'-gtggaattcaagcttCATGTCTGTCTCCTCC-3' (Ig V_{H6}) (NC-IUB nomenclature,³⁰ linkers are shown in lower case).

RNA isolation, cDNA synthesis, and reverse transcription (RT)-PCR. Total cellular RNA was isolated from cryopreserved AIDS-associated B-cell lymphoma specimens using RNAzol B (TELTEST, Inc, Friendswood, TX). First strand cDNA was synthesized from 5 µg total RNA using an oligo-dT primer and Superscript II reverse transcriptase (Life Technologies, Grand Island, NY). RT was performed for 1 hour at 42°C. Afterwards, the mixture was heated to 70°C for 15 minutes, cooled on ice 10 minutes, and then incubated with RNase H (Life Technologies) at 37°C for 20 minutes. Five microliters of this reaction mixture was used for a 100 µL total volume PCR reaction, as previously described. Primers consisted of an Ig heavy chain constant region oligonucleotide and Ig V_H subgroup oligonucleotides. Where indicated the RT-PCR product was size separated via agarose gel electrophoresis, blotted onto nylon, and hybridized with an Ig heavy chain variable region third framework (FR3) oligonucleotide specific for each of the different V_H subgroups.

Anchored RT-PCR enzyme linked immunosorbent assay (ELISA). An anchored RT-PCR assay was performed to assess the relative levels of different Ig V_H genes expression in blood or lymphoid tissue, as described.³¹ dG tailed first strand cDNA is subjected to primary aPCR amplification followed by a second step "nested PCR," using 5' biotinylated primers. The primers consists of an anchored primer and C_μ antisense oligonucleotides. The antisense biotin labeled V_H gene is added to an avidin coated ELISA plate. The presence of specific V_H genes is probed by adding digoxigenin labeled oligonucleotides specific for the different V_H gene leaders and developed by adding antidigoxigenin bound to peroxidase to produce a colored reaction.

Sequence analysis of Ig V_H genes. Amplified Ig V_H DNA fragments were ligated into Bluescript (Stratagene, San Diego, CA) that subsequently was used to transform XL1-Blue strain of *Escherichia coli* (Stratagene). Screening of transformed bacterial colonies and sequencing of the Ig variable region inserts were performed as described,³² using dideoxy chain termination (Version 2 DNA sequencing kit; USB, Cleveland, OH). We determined the Ig variable region nucleotide sequence of at least two colonies of transformed bacteria. Where indicated, direct dsDNA sequencing of the amplified PCR product was performed using the *fmol* DNA Sequencing System (Promega, Madison, WI), as per manufacturer's instructions.

All sequences were analyzed using the GenBank database and the Sequence Analysis Software Package, 1991 (Genetics Computer Group, Inc, Madison, WI). Ig V_H gene sequences were aligned with the published germline Ig V_H gene sequences having the highest homology using the local homology algorithm of Smith and Waterman.³³ Differences between the V_H gene sequence and its putative germline counterpart were defined as being a replacement (R) mutation if the observed difference results in a change in the deduced amino acid sequence. Nucleotide differences that do not result in an amino acid substitution were defined as being silent (S). Calculations of the expected ratio of R mutations to S mutations (R:S ratio) for any given codon were derived from analyses of the actual codons

used in the putative germline V_H gene sequence. This takes into consideration the observation that codons encoding the first or second complimentary determining region (CDR) generally are more susceptible to having replacement mutations from any given single nucleotide change than codons within the framework regions.³⁴

CDR3 analysis. Each sequence was searched for homology against all published D genes available in the GenBank database. To best-fit D genes we accepted reverse homology, and gave priority to the homology of the J_H segment. We avoided introducing gaps and base additions to the alignment and selected D segments that had the highest homology and the longest stretch of shared nucleic acid sequence.

RESULTS

Southern blot analyses. We examined the biopsy tissue of 10 patients with AIDS-associated lymphoma (samples AL1 through AL10) for Ig heavy chain gene rearrangements by Southern blot analyses. Three had evidence for Ig gene rearrangements involving both alleles (patients AL4, AL5 and AL7), whereas the remaining 7 samples each had only one Ig gene rearrangement and a nonrearranged Ig gene fragment similar to that seen in enzyme restricted human placental DNA (data not shown).

Analysis of Ig V_H gene subgroups expressed by AIDS-associated lymphomas. RNA was extracted from tissue samples AL1, AL2, AL4, AL5, AL7, AL9, and AL10, for use in an anchored RT-PCR/ELISA, as described.³¹ AL1, AL2, AL4, AL5, AL9, and AL10, generated a PCR product identified only by an oligonucleotide specific for rearranged Ig V_H genes of the Ig V_{H4} gene subgroup, but not with probes for rearranged Ig V_H genes of any one of the other Ig V_H gene subgroups. Similarly, RT-PCR of AL7 generated a product that exclusively hybridized with an oligonucleotide probe specific for Ig V_H genes of the Ig V_{H3} subgroup.

The Ig V_H genes expressed by cases AL3, AL6, and AL8, were examined by PCR on isolated genomic DNA. PCR with DNA of AL3 or AL6 generated a 660 bp PCR product using oligonucleotides specific for J_H and the Ig V_{H4} subgroup, but not with primers for J_H and any one of the other 6 Ig V_H gene subgroups. Similarly, PCR with the DNA of AL8 generated a 660 bp product only when the J_H primer was used in conjunction with sense-strand primers specific for Ig V_{H5} gene consensus leader sequence. The Ig V_H gene subgroup was verified by Southern blot analyses of the PCR products using probes specific for each of the Ig V_H gene subgroups (data not shown). Collectively, these studies indicated that only 1 of the 10 AIDS-associated lymphoma samples used Ig V_{H3} genes. On the other hand, 8 of the remaining samples each used Ig V_H genes of the V_{H4} subgroup.

Structural analysis of the rearranged Ig variable region genes. The PCR products of samples AL1, AL3, AL4, AL5, AL6, AL7, and AL8, were each cloned for double stranded DNA sequence analysis. This revealed each to have a rearranged Ig V_H gene of the Ig V_{H4} (AL1, AL3, AL4, AL5, and AL6), Ig V_{H3} (AL7), or Ig V_{H5} (AL8) subgroup (Figs 1 and 2). Analyses of at least 2 separate clones of each PCR product did not reveal significant nucleotide sequence variation (less than 0.33% variation in each case). Nucleic acid sequence comparison of each cloned Ig V_H gene with known germline Ig V_H genes revealed that the clones from

A. VH3

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+-----LEADER-----+-----INTERVENING SEQUENCE-----+
DP58 ATGGAGTGTGGGGCTGTCTGGGTTTCCTTGTCTATTATTAGAGGTGATTCATGGAAACCTAGAGAGATTAGTGTGTGGATATGATGAGAGAAACAGTGGATATGTGTGGCAAGTTCTGACCTTGGTGTCTTTTGTTCAGGT
AL7.1 ..... (----- Intron deleted -----) .....

AL7.0 .....A.....A.....T.G.....A.....G.A.....G.....A.....C.....*****.....T.....G.....
VH3-9 .....A.....A.....A.....T.G.....A.....G.A.....G.....A.....C.....*****.....T.....G.....

              1          10          20          30 +----CDR1----+36          40
DP58 GluValGlnLeuValGlnSerGlyGlyLeuValGlnProGlyGlySerLeuArgLeuSerCysAlaAlaSerGlyPheThrPheSerSerTyrGlnMetAsnTrpValArgGlnAlaProGlyIysGlyLeuGluTrpV
GTCCAGTGTGAGGTGACAGCTGGAGAGCTGGGGAGGCTGGTACAGCCTGGAGGTCCTGACACTCTCCCTGTCAGCCCTCGGATCACCTTCAGTGTATGAATGAACTGGTCCGCGAGCCTCCAGCGAAGGGCTGGAGTGGG
AL7.1 .....T.....
VH3-9 .....A.....C.....CA.....TGA.GA.....CC.C.....G.A.....C.....
VH3-9 .....A.....CA.....TGA.GA.....CC.C.....G.A.....C.....

49 +-----52A-----+-----CDR2-----+66          70          80 82 A B C          90          94
DP58 alSerTyrIleSerSerSerGlySerThrIleTyrTyrAlaAspSerValIysGlyArgPheThrIleSerArgAspAsnAlaIysAsnSerLeuTyrLeuGlnMetAsnSerLeuArgAlaGluAspThrAlaValTyrTyrCysAlaArg
TTCATACAGATTAGTAGTAGTACCATATACTACGACAGACTGTGTGAAGGGGATTCACCATCTCCAGAGACACCGCAAGAATCACTGTATCTGCAATGACAGAGCTGAGAGCGGACACGGCTGTTTATTACTGTGCAGAA
AL7.1 .....T.T.TG.....
VH3-9 .....T.T.TG.....
VH3-9 .....C.....G.T.....G.A.A.G.G.....G.....G.....T.G.....C.....T.....T.....T.....CT.G.....A.A.
VH3-9 .....C.....G.T.....T.G.A.A.G.G.....G.....G.....T.G.....C.....T.....T.....T.....CT.G.....A.A.

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B. VH4

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+-----LEADER-----+-----INTERVENING SEQUENCE-----+
VH4-59 ATGAAACATCTGTGTCTCTCTCTCTGTGTCAGCTCCAGATGTGATTCAGGGATCCAGACATGGGGATATGGGA*GGTGCCTCTGATCCAGGCTCAGTGTGGTCTCTCTCTCACAGGGTCTCTCCAGTGG
AL1 .....A.....G.....*.....
AL2 .....G.....A.....
AL4 .....C.....A.....
AL6 .....C.....
F105 ..... (----- Intron deleted -----) .....
268-D .....

AL5 .....C.....G.....G.....*.....
VH4-61 .....C.....G.....G.....*.....

AL3 .....G.....A.G.G.TG.AA.C.TA.T.....T.T.....C.T.T.AA.A
VH4-34 .....C.....C.....G.....A.G.G.T.AA.C.T.....T.T.....T.T.....

              10          20          30 +----CDR1----+36          40          49
VH4-59 GlnLeuGlnGlnSerGlyProGlyLeuValIysProSerGluThrLeuSerLeuTyrCysThrValSerGlyGlySerValSerSerTyrTyrTrpSer*****TrpIleArgGlnProProGlyIysGlyLeuGluTrpIleGly
CAGCTCCAGAGTGTGGGCTCCAGGACTGGTGAAGCCTCGGAGACCTTGTCCCTCACTGCACACTGTCTCTGTGGCTCCGTCAGTAGTTACTACTGGAG*****TGGATCCCGCTCAGCCCGAGGAGGGACTGGATGGATGGG
AL1 .....A.....C.A.C.....C.....G.....CA
AL2 .....AC.....T.....A.....C.....T.....*****G.....
AL4 .....A.....A.....T.....A.....CC.A.....T.....*****A.G.....C.....
AL6 .....G.....A.....C.....*****.....
F105 .....A.....A.....C.....*****.....
268-D .....T.....T.....C.A.A.A.GC.....CA*****.....A.....G.A.....ACC.A.A

AL5 .....C.....C.A.ACTAGT.ACTA.TGGAGC.....
VH4-61 .....C.....G.TAGT.ACTA.TGGAGC.....

AL3 .....T.AA.AC.A.G.....G.....C.....A.C.....AA.TG.....T.A.....GC.C.....G.....T.....T*****.....A.....T.TA.AGG.....C.....C.A
VH4-34 .....A.AC.....G.....G.....T.....G.....A.....G.....T.....G.....*****.....C.....TGGGGAGGCTGGATGGATGGG

+-----CDR2-----+66          70          80 82 A B C          90          94
VH4-59 TyrIleTyrTyrSerGlySerThrAsnTyrAsnProSerLeuLysSerArgValThrIleSerValAspThrSerLysAsnGlnPheSerLeuLysLeuSerSerValThrAlaAlaAspThrAlaValTyrTyrCysAlaArg
TMTATCTMTATCAGTGGAGCCACACTACACACCCCTCCCTCAGAGTCGATCCACTATGACATGACATGACAGCATCCAGACACCGCTTCCTCCCTGAGCTGAGCCTGTGTGACCGCTCCGGACCGGCTGTATCTGTCCAGAA
AL1 .....C.G.....AG.GC.....TT.T.....T.TT.....C.G.....G.....G.....CG.....G.....G.A.....T.....T.AG
AL4 .....GA.....CA.....GT.....C.....A.....T.....T.C.A.....C.A.....G.A.....
AL6 .....C.....G.....C.....*****.....
F105 .....C.....G.....G.....C.....A.....C.....
268-D .....G.....C.T.C.....AGT.....T.....C.GC.....A.A.....G.....G.....A.....C.GC.....AG.T.....T.....A.....T.....

AL5 .....T.....A.....
VH4-61 .....

AL3 .....G.A.A.G.TG.C.AG.....T.GT.G.T.....G.....GA.C.....T.....T.....CT.A.....T.....G.A.....C.....TACA.....T.....
VH4-34 .....G.A.A.C.T.....G.....G.....T.....G.....C.....A.....GATG.....A.....

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C. VH5

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+-----LEADER-----+-----INTERVENING SEQUENCE-----+
VH5-51 ATGGGGTCAACCCATCTCTCGCCCTCTCTCTCTCTCTCTCAAGGTCAAGTCTGTCCGAGGGCTTGGAGTCAACAGGAGAAAGGGTGGAAAGGAGCCCTGATTCAAAATTTTGTGTCTCCCCACAGAGGTCTGTGCGGAGG
AL8 .....A.....

              10          20          30 +----CDR1----+36          40          49 +--
VH5-51 GlnLeuValGlnSerGlyAlaGluValIysLysProGlyGluSerLeuLysIleSerCysLysGlySerGlyTyrSerPheThrSerTyrTrpThrGlyTrpValArgGlnMetProGlyIysGlyLeuGluTrpMetGlyIle
CAGCTGGTCCAGTCTGGAGCAGAGGTGAAAGAGCCCGGAGTCTGTGAAGATCTCCCTGTAGGGTCTCTGGATACAGCTTACAGCTACTGGACCGGCTGGTGTGACCGCTCCGGACCGGCTGTATCTGTCCAGAA
AL8 .....G.....G.....C.T.T.....C.C.T.C.CG.....

+-----52A-----+-----CDR2-----+66          70          80 82 A B C          90          94
VH5-51 IleTyrProGlyAspSerAspThrArgTyrSerProSerPheGlnGlyGlnValThrIleSerAlaAspLysSerIleSerThrAlaTyrLeuGlnTrpSerSerLeuLysAlaSerAspThrAlaMetTyrTyrCysAlaArg
ATCTATCCGTGACTCTGATACAGATACACAGCCCTCTCCAGGCCAGGTCAACATCTCAGCCGACAAAGTCCATACAGCCAGCTACTCCAGCTACTGGACCGGCTGGATGAGCTGGAGCGGCTGTATCTGTCCAGAA
AL8 .....T.....T.....T.....G.....G.....T.....C.....A.....GATG.....A.....

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A. V_H3

	1	10	20	29+CDR1→36	40	48+---CDR2-----+66	82ABC	90	←-----CDR3-----++--FR4--→
DP58	EVQLVDSGGGLVQPGGSLRLS	CAASGFTFS	SYEMN**	WVROAPGKGL	EWVSYISSG	STIYADSVKGRFTIS	SRNAKISLYLQNS	SLRAEDTAV	YYCAR
AL7.1
AL7.0	R.....	R.....	DD·A·H**	G·WNSGS·G	L·K	DTVGVATAMASYIII
V _H 3-9	R.....	R.....	DD·A·H**	G·WNSGS·G	L·K

B. V_H4

	1	10	20	29+CDR1→36	40	48+---CDR2-----+66	82ABC	90	←-----CDR3-----++--FR4--→
V _H 4-59	QVQLQESGPGGLVQPKSEITLS	LTCTVSGGSVSS	YIWS**	WIRQPPGKGL	EWIGYIY*	YSGSTINYNPSL	KSRVITISVDT	SKNQFSLK	LSSVTAADTAVYYCAR
AL1	S.....	ITNS·T**	A·S*D·K·KH·S	SL·L·S	R·T·S·G·VKMGD·KRAG·RSET
AL4	V·ITN·F·**	T·D·*	R·V·V·L·T·H	R·G
AL6	R.....	R.....	I·H·**	S·Q·*	S·E
F105	I·H·**	S·Q·*	S·E	T·M
268-D	V.....	PIINNA·T**	YL·V·HT·V	L·TI
AL5	NT·WS	F*	Q
V _H 4-61	WS	*	Q
V _H 4-34	QVQLQCSGAGLLKPKSEITLS	LTCAVYGGSP	SGYIWS*	WIRQPPGKGL	EWIGYIY*	HSGSTINYNPSL	KSRVITISVDT	SKNQFSLK	LSSVTAADTAVYYCAR
AL2	I·K	K·PL	ST·G	DG·G

C. V_H5

	1	10	20	29+CDR1→36	40	48+---A---CDR2-----+66	82ABC	90	←-----CDR3-----++--FR4--→
V _H 5-51	EVQLVDSGAEVQPKPGESLK	LSCKRGGSYPT	SYWIG*	WVROAPGKGL	EWIGYIY*	PGSDIRYSP	SPQGVITISADK	SISTAYLQNS	SLKASDTAMYYCAR
AL7	R·QVF·T·TR	A·I	E·R

Fig 2. Amino acid sequences deduced from the Ig V_H genes of AIDS-associated lymphomas. (·) Indicate sequence homology and asterisks are used to introduce gaps that maximize sequence homology. Related germline sequences and patient samples are indicated as in Fig 1.

AL1, AL4, and AL6, each had the highest homology with V_H4-59 (V71-4, DP-71), clones from AL3 had the highest homology with V_H4-34, clones from AL5 had the highest homology with V_H4-61 (V71-2), clones from AL7 were most homologous to V_H3-9, and AL8 clones had the highest homology with V_H5-51 (Fig 1 and Table 1).³⁵

We determined the DNA sequence of the Ig V_H gene expressed by sample AL2 (Fig 1), AL9, and AL10, by direct dsDNA sequence analysis of the RT-PCR product in the region contiguous to the deduced CDR1. This confirmed that AL2, AL9, and AL10, each expressed an Ig V_H gene of the Ig V_H4 subgroup (data not shown). Moreover, the deduced sequence of AL2 or AL10 have highest homology with V_H4-59 (V71-4) whereas AL9 has highest homology with germline V_H4-61 (V71-2) (Fig 1, and data not shown).

Comparison of the sequences with those reported in GenBank revealed that the Ig V_H genes used by some AIDS-associated lymphomas have relatively high homology to certain Ig V_H genes noted to encode anti-HIV antibodies of HIV-seropositive donors (Figs 1 and 2). For example, the

Ig V_H gene encoding the anti-gp120 antibody F105,²⁴ displayed 87, 96, 88, or 96, percent nucleic acid sequence homology with AL1, AL2, AL4, or AL6, respectively. However, the degrees of homology noted between the lymphoma-derived Ig V_H genes invariably were higher with their putative germline counterpart than with these rearranged and functional Ig V_H genes. As some sites, there are base differences from the putative germline gene that are shared between certain members of these expressed Ig V_H genes. For example, compared to V_H4-59 there is a nonconservative shared substitution in the third position of codon 29 of AL1, AL3, AL4, F105, or 268-D (Fig 1), resulting in a valine → isoleucine substitution (Fig 2). Such nonconservative shared substitutions most likely are due to genetic polymorphism in the Ig V_H genes, rather than somatic selection.³⁶

All cloned genes had functional Ig gene rearrangements, except for the Ig V_H isolated from the genomic DNA of AL7. Although this Ig V_H gene had 99.6% nucleic sequence homology with a functional germline Ig V_H3 gene, V_H3-9, sequence analyses revealed a termination codon in the region

Fig 1. Ig V_H gene nucleotide sequences of the rearranged Ig V_H genes cloned from AIDS-associated lymphoma tissue samples. The deduced CDR3 and FR4 regions are presented in Fig 3. The clones are designated with the prefix AL and are assigned the number corresponding to the patient number provided in the text. Sequence number AL7.0 is the sequence of the genomic PCR product of patient 7 that contains a stop codon in the deduced CDR3 (Fig 3). Sequence AL7.1 is the sequence of the RT-PCR product of the expressed Ig V_H gene from the same patient sample. Each sequence is compared to that of the most closely homologous germline Ig V_H sequence. (A) Ig V_H3: DP58¹⁷ and V3-9,³⁵ (B) Ig V_H4: 4-59^{35,67} and 4-61^{35,67}; and (C) Ig V_H5: 5-51.^{35,68} Also listed for comparison are the sequences of anti-HIV antibody heavy chains F105²⁴ and V_H268-D.⁹⁰ (·) indicate nucleotide sequence homology and (*) are used to introduce gaps that maximize sequence homology. The numbering of amino acids is according to Kabat et al.³⁷

Table 1. Structural Analysis of the Ig V_H Genes in AIDS-Associated Lymphomas

Sample	Germline V _H Gene	% Homology	FR				CDR			
			S	R	R:S	Innate R:S	S	R	R:S	Innate R:S
AL1	V4-59	89.7	8	14	1.8	2.6	2	12	6.0	4.6
AL3	V4-34	85.6	19	23	1.2	2.7	4	7	1.8	4.5
AL4	V4-59	93.0	6	15	2.5	2.6	0	5	5.0	4.6
AL5	V4-61	98.6	1	1	1.0	2.6	0	4	4.0	4.5
AL6	V4-59	99.3	0	2	2.0	2.6	0	0	0.0	4.6
AL7.1	DP58	92.8	3	2	0.6	ND	2	6	3.0	ND
AL8	V5-51	91.7	4	13	3.2	3.0	1	7	7.0	3.5
F105	V4-59	95.9	2	6	3.0	3.0	1	2	2.0	4.6
268-D	V4-59	86.1	9	18	2.0	3.0	3	11	3.6	4.6

Listed for comparison is the analysis of anti-HIV antibody heavy chains F105²⁴ and V_H268-D.⁶⁰ The germline Ig V_H genes having the highest homology to lymphoma isolated sequences are presented using the revised nomenclature for germline Ig V_H genes.⁶⁹ The column marked "% Homology" provides the percent nucleotide sequence homology between the expressed Ig V_H gene and its putative germline counterpart. FR labels the columns providing the analysis of segments FR1, FR2 and FR3, and CDR labels the columns providing the analysis of CDR1 and CDR2. Columns marked "S," "R," or "R:S", respectively, indicate the number of deduced silent mutations, replacement mutations, or ratio of replacement to silent mutations in each region. The columns labeled "innate R:S" provide the total possible replacement mutations to total possible silent mutations for the codons in the specified regions, as calculated by Chang and Casali.⁵⁸

Abbreviation: ND, not determined.

deduced to encode the CDR3 (Fig 3). As mentioned earlier, this patient had Ig gene rearrangements involving both alleles, making it possible that this clone contained the Ig V_H gene of the nonexpressed allele. Because of this, we determined the nucleic acid sequence of the RT-PCR product of tissue sample 7, designated AL7.1 (Fig 1). We found that AL7.1 also contained an Ig V_H3 gene. However, this Ig V_H3 gene differed from that obtained through genomic DNA PCR, and had the highest sequence homology with another germline Ig V_H3 gene, DP58 (Fig 1). Consistent with this clone representing the expressed allele, AL7.1 had a functional Ig V_H gene rearrangement (Fig 2).

The CDR3 and FR4 were deduced for each clone, as per Kabat et al³⁷ (Fig 3). Each variable region gene encoded a long CDR3 of 10 to 22 amino acid residues. Comparison of the CDR3 with known germline D segments revealed that each probably arose through D-D fusion (Fig 3). By comparing the CDR3 and FR4 with known J_H gene segments, we deduce that AL5 and AL8 probably use J_H2, AL1 and AL6 use J_H3b, AL3 and AL7.1 use J_H4b, whereas sample AL4 most likely uses J_H5b (Fig 3).³⁸⁻⁴³

DISCUSSION

We examined the Ig V_H genes expressed in 10 cases of AIDS-associated lymphoma. We found that such B-cell lymphomas expressed a highly skewed repertoire of Ig V_H genes. First, despite the finding that a subset of Ig V_H3 genes can encode antibodies that innately have binding activity for HIV gp120,²³ only one of the ten samples examined (sample AL7.1) expressed Ig V_H genes of this subgroup. Ig V_H3 genes ordinarily encode most of the Ig expressed in the normal adult B-cell repertoire.^{18,44-47} Some groups have noted HIV-infected patients may have a selective depletion of B cells expressing certain Ig V_H3 genes⁴⁸ and a concomitant increase in the relative use of Ig V_H1 genes.⁴⁹ However, using a quantitative RT-PCR/ELISA to examine the distribution

of expressed Ig V_H genes,³¹ we have not noted significant reduction in the relative expression of Ig V_H3 genes by blood B cells of patients infected with HIV compared to that of normal adults.⁵⁰ That only one of the lymphoma samples examined expressed Ig V_H genes of this subgroup is significantly less than what would be expected if the repertoire of Ig V_H genes in AIDS-associated lymphoma reflected the repertoire of Ig V_H genes expressed by normal B cells, χ -square of 4.17, and a *P* value of .04 (*df* = 1).

Furthermore, the examined B-cell lymphomas predominantly expressed Ig V_H genes of the Ig V_H4 subgroup. Ordinarily, Ig V_H4 genes encode between 12% to 22% (average 18%) of the Igs expressed by normal adult B cells.^{18,44-47} However, we found that eight of ten samples expressed Ig V_H genes of this subgroup. This proportion is significantly higher than what would be expected if the repertoire of Ig V_H genes in AIDS-associated lymphoma reflected the repertoire of Ig V_H genes expressed by normal B cells, with a χ -square 6.8, *P* = .009, *df* = 1. Furthermore, the overall distribution of Ig V_H genes in AIDS-associated lymphoma compared to normal adults is significantly different with a χ -square of 27.4 and a *P* value of .0001, *df* = 6.

The noted frequent expression of Ig V_H4 genes in AIDS-associated lymphomas appears similar to that recently reported for large cell lymphomas of patients not infected with HIV.⁵¹ Hsu and Levy⁵¹ noted that large cell lymphomas frequently express Igs encoded by Ig V_H4-34, an Ig V_H4 gene frequently found to encode IgM cold agglutinins.⁵² Indeed, two AIDS-associated lymphomas found to produce cold agglutinins were noted to express Ig encoded by this particular Ig V_H gene.⁵³ However, in our series of 10 AIDS-associated lymphomas not selected for antigen binding activity, only one expressed an Ig V_H4 gene that had the highest homology to Ig V_H4-34 germline gene (AL3) (Fig 1).

Rather, most AIDS-associated lymphomas we examined used a sub-subgroup of Ig V_H4 genes distinct from Ig V_H4-

N - D - N Segments		JH Segment	CDR3 length	N additions
AL1	MetGlyAspProLysArgAlaGlyArgArgSerGluThr ATGGGAGATTCCGAAAAGGGCCGGCTCGAAGTGA	GlyPheAspIleTrpGlyGlnGlyThrMetValThrValSerSer GGTATTGATATCTGGGGCCAAGGGACAAATGGTCACTGCTCCTCA	17	7
DIR	· · · · T · · · · ·	· C · · · · · · · · · · · · · · · ·	J _{H3b}	
DIR	· C · · · · · · CC · · · · · · · G · T ·			
AL3	GlyArgAspTyrTyrAsnHisThrThrLeuThrGlnGluPheThr GGCCGTGATTACTACAATCACACGACTCTGACTCAGGAATCACT	PheAspThrTrpGlyGlnGlyAsnLeuValThrValSerSer TTTGACACATGGGGCCAGGGAACTTGGTCACTGCTCCTCA	18	15
DLR4-C	· · T · GT · C · · · · · · · · · · · · · · · ·	· · · · · · TAC · · · · · · · · · · · · C · C · ·	J _{H4b}	
DIR	· · · G · · · · · · · G · · ·			
AL4	HisValArgGlyGlyArgLeuGlyAspLeuSerSerAla CATGTGAGGGGGGGCCCTTGGGGACTTATCTCCGGCG	AspSerTrpGlyProGlyThrLeuValThrValSerSer GACTCTGGGGCCCGGGAACTTGGTCACTGCTCCTCA	15	10
D6	· · · · TC · · · · ·	· · · · · · · · · · · A · · · · · · · · · ·	J _{H5b}	
D21/10	· · T · · · · · · · · G · · · · G · · ·			
AL5	AspThrValLeuValIleGlyAlaAlaGly GACACTGTACTAGTGAATGGCGCGCAGCC	TrpTyrPheAspLeuTrpGlyArgGlyThrLeuValThrValSerSer TGGTATTTCGATCTCTGGGGCCGTGGCACCCCTGGTCACTGCTCCTCA	15	12
DLR1B	· · T · T · · · · · · G · · · G ·	· ·	J _{H2}	
DN1	· · A · · · ·			
AL6	AspAsnLysPheLeuArg GATAATAAATTTTGGCT	AlaPheAspIleTrpGlyGlnGlyThrMetValThrValSerSer GCTTTTGATATCTGGGGCCAAGGGACAAATGGTCACTGCTCCTCA	10	4
Dxp 1-C	· · · · · · · · · · · · · · · ·	· ·	J _{H3b}	
Dxp5	· · · · · · · · · · · · · · · ·			
AL7.0	AspThrValGlyAlaTyrThrAlaMetAlaSerTyrIleIleIleEnd GATACTGTCTGGGGCTTATACAGCTATGGCTTCGTATATCATCATAGTACCGGGTCCCGAGGTGTACA	ACTGGTTCGACCCCTGGGGCCAGGGAACTTGGTCA		
DIR-C	· · · · · · · · · · · · · · · ·	· ·	J _{H5b}	
DK4	· T · A ·			
DN1	· · · · · G · · G · · G · · G · · ·			
AL7.1	ValLeuArgProProAsnAspGlnArgAspCysGlyGlyGlyCysTyrSerPro GTCTTAGGCCCCCGAATGACCAAGAGATTGTGGTGGTGGTGTCTATTCCGCC	TyrPheAspCysTrpGlyGlnGlyThrLeuValThrValSerSer TACTTTGATCTCTGGGGCCAGGGAACTTGGTCACTGCTCCTCA	22	18
DIR	· · · · T · · · · G · ·	· · · · · · · · A · · · · · · · · · · · · · · ·	J _{H4b}	
DLR3	· · T · · · · · · · · · · A · · · · · · · · · ·			
AL8	HisTrpGlyAspGlyTyrAsnTyrLysThr CATGGGAGATGGCTACAATTATAAACT	TrpSerPheAspLeuTrpGlyArgGlyThrLeuValThrValSerSer TGGCTTCGATCTCTGGGGCCCGGGCACTTGGTCACTGCTCCTCA	15	10
D6	· C · · · · ·	· · · · A · · · · · · · · · · · T · · · · · · · · · ·	J _{H2b}	
DK1	· · · · · · · · G · · · ·			

Fig 3. CDR3 and FR4 regions. Lines one and two display the amino acid and nucleotide sequence of the CDR3 of each Ig of AIDS-associated lymphomas. Lines 3 and 4 are the germline J_H and diversity (D) segments that have the highest homology to that of the lymphoma-derived Ig gene. (·) Indicate sequence homology and asterisks are used to introduce gaps that maximize sequence homology. The CDR3 and FR4 are defined as per Kabat et al.³⁷ Indicated for sample AL7.0 is the termination codon in the genomic DNA PCR product. AL7.1 is the RT-PCR sequence of the RNA isolated from the same patient. The underlined portion of the J_H segment identifies the sequence of the oligonucleotide primer.

34. Five samples (AL1, AL2, AL4, AL6, and AL9) expressed Ig V_{H4} genes that each had the highest nucleic acid sequence homology with V_{H4}-59 (V71-4), and two additional cases (AL5 and AL10) used an Ig V_H with highest homology with V_{H4}-61 (V71-2) (Fig 1). Ig encoded by these Ig V_{H4} genes generally have a high degree of structural similarity, as shown by the fact that they frequently share cross-reactive idiotypic determinants, such as those detected by the anti-idiotypic monoclonal antibody Lc1.²⁷ Future studies should examine for the frequent expression of this supratypic cross-reactive idio type in AIDS-associated lymphoma specimens.

The observed restriction in the Ig V_H genes expressed by AIDS-associated lymphomas may be because of selection for B cells that express Ig with certain binding specificities. Evidence for this may be derived from the structural analyses of the Ig V_H genes expressed by the AIDS-associated lymphomas. First, we deduce that most of the AIDS lymphomas express Ig V_H genes that have incurred somatic mutations. Second, several of these Ig V_H genes have relatively high ratios of deduced replacement mutations (R) to silent mutations (S) in regions that encode part of the antigen

combining site(s) (namely CDR1 and CDR2) (Table 1). Moreover, the deduced R:S ratios of the segments encoding the FR portion of the antibody variable region are consistently lower than those of CDR1 and CDR2, and lower than the average R:S ratio of 2.9 calculated for random codons.⁵⁴ Higher R:S ratios in the CDR1 and CDR2 compared to those in the FR are observed commonly in Ig V genes expressed by B cells selected in a secondary antigen-driven immune response.^{16,22,55-57}

However, Chang and Casali observed that the codons for the CDR of many germline Ig V genes are more prone to replacement mutations than are those used in most other proteins.⁵⁸ Conversely, the codons used in the FR of many germline V genes are slightly less prone to replacement mutations than are random codons.⁵⁸ As such, finding higher R:S ratios for the CDR than for the FR segments could result from random nonselected nucleotide base substitutions. Nevertheless, the Ig V_H 4-59 genes used by AL1, AL4, and AL8, still have deduced R:S ratios in the CDR1/CDR2 that exceed that expected from random base substitutions. Furthermore, most of the Ig V_H genes characterized in this study have lower R:S ratios than that expected for random base substitutions in the FR (Table 1).

It should be noted that anti-HIV antibodies from patients with AIDS may be encoded by Ig V_H genes that have deduced somatic mutations resulting in R:S ratios that are less than that expected for random base substitutions in the CDR (eg, Table 1, and data not shown). This may reflect a reduced capacity of B cells to undergo antibody affinity maturation in the HIV-infected host. If found to be the case, then the immune status of patients with AIDS may favor use of Ig V_H genes in immune responses that selectively can incur advantageous replacement mutations in the CDR with minimal numbers of somatic mutations. Conceivably, this could enhance the expression of such Ig V_H genes in AIDS-associated lymphomas. In this regard it is noteworthy that the V_H4 genes that we found frequently expressed in AIDS-associated lymphomas, namely V_H 4-59 and V_H 4-61, have among the highest average innate R:S ratios for codons within CDR1/CDR2, namely 4.486 and 4.613, respectively. (The mean innate R:S ratio for CDR1/CDR2 of all Ig V_H genes is 4.152 (± 0.449 SD)).⁵⁸ Moreover, these two Ig V_H genes have the lowest average innate R:S ratio for segments encoding the antibody FR of all characterized germline Ig V_H genes, namely 2.593 (The mean innate R:S ratio for FR of all Ig V_H genes is 2.845 (± 0.169 SD)).⁵⁸

Analyses of the CDR3 of the Ig heavy chains expressed in AIDS-associated lymphomas provides additional evidence for selection. The heavy chain CDR3 of Ig genes expressed by all lymphomas except AL6 have nonconservative base differences from the deduced D and J_H gene segments that encode the CDR3 (Fig 3). Furthermore, the length of each CDR3 ranged from 10 to 22 amino acids ($m = 15 \pm 4$). This is longer than the noted range of 8 to 18 amino acids ($m = 13 \pm 4$) for the CDR3 of the Ig heavy chains used by normal adult B cells.⁴³ The probable number of N base additions in the CDR3 (range 4-18, $m = 10$, Fig 3) appears similar to that reported for Igs expressed by normal adult blood B cells.⁴³ However, we deduce that each CDR3 except AL1 probably is encoded by 2 or 3 different D genes segments (Fig 3). The CDR3 of AL1 probably represents D-D fusion of DIR genes, genes of approximately 170 bp in length with irregular recombination signal sequences that are present in multiple copy in the human haploid genome.⁵⁹ In contrast, only approximately 10% of Ig heavy chains expressed by normal adult B cells have CDR3 with evidence for D-D fusion.⁴³ In this regard, it is noteworthy that the characterized anti-HIV antibodies of patients infected with HIV each has a heavy chain CDR3 that also probably results from D-D fusion.^{24,60,61} Moreover, the heavy chain CDR3 of recombinant antibodies selected for binding to HIV from combinatorial libraries also display features similar to the CDR3 of the lymphoma-derived immunoglobulin genes described in this study.^{62,63} Conceivably, such D-D fusions may enhance the expression probability for antibodies capable of binding HIV glycoproteins. However, we note that the nonproductive allele of AL7 (Fig 3), also has evidence for having been formed through D-D fusion. As the nonsense mutation is the only identified defect in this Ig gene, it is possible that it was acquired during a process of Ig somatic mutation. Alternatively, the D-D fusion deduced for this nonfunctional gene may reflect an enhanced propensity of

B cells from patients infected with HIV to undergo D-D fusion. Further studies on the functional and nonfunctional Ig gene rearrangements of normal B cells from patients with AIDS are necessary to resolve this issue.

Persons infected with HIV develop a persistent humoral immune response to HIV.^{9-11,64} It is noteworthy that several previously characterized anti-HIV-1 antibodies made by HIV-seropositive donors are similar in primary structure to those described in this study (Figs 1 and 2, and data not shown).^{9-11,64} Furthermore, characterized antibodies from cDNA of HIV-seropositive donors also have identified Ig V_H genes similar to those described in this study.^{9-11,63-66} Finally, Ng et al²⁰ recently reported that cell lines established by cloning single cells from biopsies obtained from two AIDS patients with Burkitt's lymphoma produced IgM antibodies reactive with HIV gp160. Collectively, these studies and the findings reported here support a model proposing that chronic antigenic stimulation in HIV-infected individuals can lead to selective proliferation of B cells that ultimately may undergo neoplastic transformation.

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