Analysis of the subunit assembly of the type IC restriction–modification enzyme EcoR124I

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Received June 2, 1998; Revised and Accepted August 14, 1998

ABSTRACT

Type I restriction–modification (R–M) enzymes are composed of three different subunits, of which HsdS determines DNA specificity, HsdM is responsible for DNA methylation and HsdR is required for restriction. The HsdM and HsdS subunits can also form an independent DNA methyltransferase with a subunit stoichiometry of M2S1. We found that the purified EcoR124I R–M enzyme was a mixture of two species as detected by the presence of two differently migrating specific DNA–protein complexes in a gel retardation assay. An analysis of protein subunits isolated from the complexes indicated that the larger species had a stoichiometry of R2M2S1 and the smaller species had a stoichiometry of R1M2S1. In vitro analysis of subunit assembly revealed that while binding of the first HsdR subunit to the M2S1 complex was very tight, the second HsdR subunit was bound weakly and it dissociated from the R1M2S1 complex with an apparent Kd of \( \approx 2.4 \times 10^{-7} \) M. Functional assays have shown that only the R2M2S1 complex is capable of DNA cleavage, however, the R1M2S1 complex retains ATPase activity. The relevance of this situation is discussed in terms of the regulation of restriction activity in vivo upon conjugative transfer of a plasmid-borne R–M system into an unmodified host cell.

INTRODUCTION

Type I restriction and modification (R–M) systems protect the bacterial cell against invasion of foreign DNA (such as viruses) by cleaving DNA which lacks a target specific N6-adenine methyl-ation. The second physiological role of these systems is to restore full methylation of the target sites on the host DNA after DNA replication. Type I R–M enzymes are composed of three different subunits (HsdR, HsdM and HsdS) encoded by the three hsd genes (for recent reviews see 1,2). All three subunits are absolutely required for restriction activity, while the HsdM and HsdS subunits are sufficient for modification activity and can also form an independent DNA methyltransferase (MTase). Type I R–M enzymes specifically recognise a non-palindromic DNA sequence (e.g. GAAnnnnnnnRTCG for EcoR124I, where n is any base and R is a purine) but cleave DNA at a non-specific site distant from the recognition sequence using Mg2+, S-adenosylmethionine (AdoMet) and ATP as cofactors (3,4). Binding of the endonuclease to a non-modified recognition site activates a powerful ATPase activity (5), which fuels DNA translocation past the DNA–enzyme complex, while the enzyme remains bound to the recognition site (6,7). DNA is cleaved at positions where the DNA translocation stops either due to a collision of two translocating enzyme molecules on two-site, linear DNA substrates (8,9), or due to the build-up of topological strain on circular molecules (10). The endonuclease does not turnover in the cleavage reaction, however, the ATPase activity continues for a long period of time after the cleavage is completed (3,5). DNA methylation activity of the type I R–M systems results in a transfer of a methyl group from AdoMet to the N-6 position of a specific adenine in each strand of the recognition sequence (11,12).

Type I R–M systems are grouped into four families based on allelic complementation, protein homologies and biochemical properties of the enzymes. Types IA, IB and ID R–M systems are chromosomally encoded (13,14) while most type IC R–M systems are carried on large conjugal plasmids (15–17). The type IA family is typified by the EcoKI and EcoBI enzymes, type IB by EcoAI and type IC by EcoR124I. EcoKI forms a stable R2M2S1 complex; the independent EcoKI MTase (M2S1) is, however, a relatively weak complex, dissociating into an inactive M1S1 species and free HsdM subunit (18–20). The purified EcoBI restriction endonuclease has been described as existing in a number of different stoichiometric forms including R3M2S1, R2M2S1 and R1M2S1 (21). The type IB restriction endonuclease EcoAI was found to be a weak complex that dissociated into MTase and HsdR subunit when purified, and the active endonuclease could only be studied following assembly in vitro (22). We have recently purified the EcoR124I restriction endonuclease from a cell culture harbouring a recombinant, two-plasmid system, which over-produces all three subunits. The stoichiometry of this endonuclease preparation appeared to be R3M2S1 (23). The EcoR124I MTase has also been over-produced and purified to homogeneity (12,24). This enzyme was found to exist only in the M2S1 stoichiometry and no M1S1 complex has been detected (12).

The hsd genes of all R–M systems can be transferred into an unmodified host by conjugation, transformation or transfection despite the fact that the presence of a restriction enzyme in the recipient cell would be a lethal event (17,23,25–31). It has been reported that the appearance of restriction activity was delayed

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after conjugative transfer of EcoKI genes into an unmodified host, while the modification activity was expressed immediately after conjugative transfer (29). It was proposed that restriction and modification activities of the EcoKI–R system are regulated by a gene distant from the hsd locus since the conjugative transfer of EcoKI hsd genes into a mutant form of Escherichia coli C (designated as hsdC) had a lethal effect on the cell (29). The same regulation mechanism was found to be employed in the type IB R–M systems (32). However, the conjugal transfer of EcoR124I (IC) hsd genes into the hsdC mutant was not lethal and full restriction activity was detected six generations after the start of conjugation (32). Recently, it has been shown that ClpX and ClpP, components of the ClpXP protease, are necessary for efficient transmission of genes encoding the EcoKI and EcoAl R–M systems, suggesting protease-mediated post-translational control of restriction and modification (33). The involvement of protease control of restriction and modification would support the proposal that post-translational control of restriction activity of EcoKI could be related to subunit assembly (20).

In this paper we show that the purified EcoR124I restriction endonuclease is a mixture of two species, which have a subunit stoichiometry of R2M2S1 and R1M2S1, respectively. Only the former species was found to have endonuclease activity. However, the R2M2S1 complex is relatively weak, dissociating into free HsdR subunit and the restriction-deficient R1M2S1 assembly intermediate, which appears to be a very tight complex. We discuss the relevance of this situation with respect to the propagation of a conjugal plasmid-borne R–M system into an unmodified host cell. We suggest that control of EcoR124I endonuclease activity in vivo is at the level of subunit assembly in a manner similar to that observed for EcoKI (20).

**MATERIALS AND METHODS**

**DNA substrates**

Complementary HPLC-purified oligonucleotides used in the gel retardation assays were purchased from Cnachem Ltd. The EcoR124I recognition site is shown in bold (top strand: 5’-CTACCGTAACCGAAACCGCTGTCCGGCCCGCAAGC-3’), DNA concentration was determined from the absorption at 260 nm. The extinction coefficients of the oligonucleotides and the annealed oligoduplex were calculated as a sum of the contributions from individual nucleotides, taking into account hyperchromeicity observed after digestion of the DNA to completion with snake venom phosphodiesterase (34). The oligoduplex (usually 2.5 µg) was 5’-end-labelled using [γ-32P]ATP (25 µCi) and T4 polynucleotide kinase. Unincorporated label was removed using ‘Nuctrap’ columns (Stratagene).

A 2891 bp plasmid containing a single EcoR124I recognition site, pDRM-1R, (23) used in DNA cleavage assays was produced from *E.coli* HB101 (35) and its covalently closed form was purified by an equilibrium centrifugation in a CsCl–ethidium bromide gradient (36).

**Protein preparations**

The EcoR124I restriction endonuclease, MTase (M2S1) and HsdR subunit were over-produced and purified as described previously (12,23). Either the plasmid pACR124 or pBGSRI24 containing the hsdR gene under the control of the PPr promoter (23), were used together with the MTase plasmid pJS4M (24) to over-produce the endonuclease. The latter plasmid combination produced a higher concentration of HsdR than the former, presumably due to the higher copy number of pBGSRI24. Both two-plasmid systems produced the same concentrations of the HsdS and HsdM subunits (data not shown). To obtain a stable clone containing the pJS4M-pBGSRI24 plasmid-expression system, the cells were first transformed with the pJS4M plasmid (MTase) and then with pBGSRI24 plasmid (HsdR). Transformation of pJS4M and pBGSRI24 at the same time was found to be lethal. The molar concentration of purified proteins was determined from the absorbance at 280 nm using molar extinction coefficients calculated as a sum of contributions from tyrosines and tryptophans in the predicted amino acid sequences [HsdR subunit; 91 900/M/cm, MTase (M2S1); 160 400/M/cm, endonuclease, 252 300/M/cm for R1M2S1, 344 200/M/cm for R2M2S1].

**Reconstitution of the endonuclease**

Fully active reconstituted EcoR124I endonuclease, used in the DNA cleavage and gel retardation assays, was simply produced by mixing of the purified MTase (M2S1) and HsdR subunit in assay reaction buffers (see below). The concentration of active endonuclease was taken as the input concentration of the MTase.

**Determination of subunit ratio in the endonuclease–DNA complexes**

The two DNA–protein complexes observed with purified and reconstituted EcoR124I endonucleases (Fig. 1) were separated in preparative amounts (2 µM 39mer DNA in 10 µl binding reaction) on a non-denaturing polyacrylamide gel. After staining the gel in ethidium bromide solution, fluorescent bands were excised from the gel, mashed and protein extracted with SDS–PAGE loading buffer. Protein extracts were subjected to SDS–PAGE together with a series of R/MTase mixtures of known molar ratios. After staining with Coomassie Blue R250, the gel was quantified by densitometry using Image Grabber 2.05b1 and NIH image 1.52 software. The ratio of densitometer traces of individual subunits (S:M:R) was calculated for each sample. To determine the stoichiometry of the species isolated of the DNA–protein complexes, the best fit between their S:M:R ratios and one of the values obtained with the R/MTase mixtures was determined.

**HPLC gel filtration**

HPLC gel filtration used a Rainin Dynamax 4.6 × 250 mm Hydropropore-5-SEC column and a guard column. Samples of 20 or 50 µl, in 20 mM Tris, 20 mM 2-[N-morpholino] ethane sulphonic acid (MES), 0.2 M NaCl, 10 mM MgCl2, 7 mM β-mercaptoethanol, 0.1 mM EDTA, pH 6.5, were injected onto the column. The samples were equilibrated at room temperature for varying time periods up to 24 h prior to injection. The flow rate was 0.5 ml/min and detection was most sensitive and stable at 254 nm. The column was calibrated with a series of globular proteins of known molecular weight giving a linear calibration curve of log (molecular weight) as a function of elution time. Most of our samples after buffer exchange into the column buffer using PD10 Sephadex G50 columns (Pharmacia), contained trace amounts of a small molecule which we believe to be glycerol. This gave rise to a ‘solvent peak’ after ~6.5 min, the elution time of which served as an internal standard to correct for slight run to run variation in protein elution times. The apparent molecular weight of the proteins eluting from the column was calculated. In the
titration of MTase with HsdR subunit, the HsdR subunit–MTase complex was present only as a partially resolved shoulder when HsdR subunit was in large excess of the MTase.

**DNA cleavage and ATPase assays**

DNA cleavage and ATPase activities of EcoR124I restriction endonuclease were assayed using plasmid pDRM-IR (single EcoR124I site) as described previously (23). DNA, usually at a concentration of 200 nM, was incubated either with an equimolar concentration of the MTase and increasing concentration of the HsdR, or with a range of concentrations of reconstituted endonuclease. After addition of ATP, the reactions were incubated for 5 min (sufficient time to reach the reaction endpoint). DNA cleavage activity (concentration of full-length linear plasmid DNA) and ATPase activity (concentration of inorganic phosphate) were measured as described previously (23).

**Gel retardation assay**

DNA binding reactions were usually performed in a volume of 10 µl in the presence of a buffer consisting of 50 mM Tris–HCl (pH 8.0), 50 mM NaCl, 10 mM MgCl₂, 1 mM DTT and 10% (v/v) glycerol. The end-labelled 39mer oligoduplex (see above) of known concentration was incubated with purified endonuclease, MTase and MTase/HsdR or endonuclease (ENase)/HsdR mixtures over a range of various concentrations and/or protein:protein ratios, respectively, at room temperature for 10 min. Unbound DNA and DNA–protein complexes were separated on a 5% non-denaturing polyacrylamide gel run, at 4°C, at 100 V in 40 mM Tris-acetate pH 8.0, 1 mM EDTA buffer (TAE). After electrophoresis, gels were dried and subjected to autoradiography or quantified using a PhosphorImager and Image Quant software.

**RESULTS**

**Gel retardation analysis of EcoR124I endonuclease reveals two molecular species**

We have previously purified the EcoR124I endonuclease following over-production of the protein by a recombinant, two-plasmid system consisting of pJS4M (MTase) and pACR124 (HsdR). Using a combination of data from gel filtration and densitometric scanning SDS–polyacrylamide gels stained with Coomassie Blue, we concluded that this enzyme had a stoichiometry of R₁M₂S₁ (23).

To investigate the DNA-binding properties of the over-produced EcoR124I endonuclease, the technique of gel retardation, with a 39mer oligoduplex carrying the EcoR124I recognition site, was utilised. Unexpectedly, two DNA–protein complexes were observed after separation of bound and unbound DNA on a non-denaturing polyacrylamide gel (Fig. 1, lane 2). The complex with faster mobility (complex I) was preferentially formed (88% of bound DNA). When the plasmid pBGSRI24 (23), a higher copy number plasmid, which produces a significantly higher level of HsdR than the plasmid pACR124, was used with the plasmid pJS4M in the two-plasmid over-producing system, the enzyme preparation showed a different ratio between the two complexes; 53% of complex I and 47% of complex II (Fig. 1, lane 4). To exclude non-specific DNA binding, the experiment was repeated in the presence of excess competitor DNA (poly dI–dC), but no change was observed in the two retarded DNA bands (data not shown).

Another possibility was that there were two stoichiometric species present in the endonuclease preparations. This was confirmed by the following results. Addition of excess HsdR to both endonuclease preparations resulted in the disappearance of the complex I and increased the concentration of the complex II (Fig. 1, lanes 3 and 5). A 1:1 mixture of HsdR and the MTase (M₂S₁) produced only complex I, while the 6:1 mixture of HsdR and the MTase produced mostly complex II (Fig. 1, lanes 8 and 9). Finally, addition of the MTase to the mixture of the endonuclease and DNA led to the disappearance of complex II and increased the amount of complex I (Fig. 1, lane 6). These results indicate that the two observed species vary in the number of HsdR subunits, and the lower molecular weight species is preferentially formed at HsdR:MTase ratios <1.

**Analysis of subunit stoichiometry in protein–DNA complexes of EcoR124I**

To investigate the stoichiometry of the two species present in the endonuclease preparations, the proteins bound in complexes I and II were extracted from the retardation gel and subjected to an SDS–PAGE analysis together with a series of R/MTase mixtures, including ratios 0.5:1, 1:1, 2:1, 3:1 and 4:1. The relative ratio of the three endonuclease subunits, in each sample, was determined from a densitometer scan of the polyacrylamide gel after staining the gel with Coomassie Blue (23). Analysis of subunit stoichiometry in protein–DNA complexes of EcoR124I

**Analysis of interaction between the MTase and HsdR in the presence of DNA**

To investigate the endonuclease assembly in vitro, we performed a reconstitution titration of the MTase (M₂S₁) with the HsdR subunit, and followed this by a gel retardation assay using a 39mer oligoduplex carrying the EcoR124I recognition site.
The apparent molecular weight of the complexes is given in Table 1. The expected molecular weight of the complexes is also given. The apparent molecular weight of the complexes is determined by HPLC gel filtration, and the expected molecular weight of the complexes is determined by gel retardation assay.

**Table 1.** Molecular weights of complexes found for the EcoR124I system

<table>
<thead>
<tr>
<th>Sample</th>
<th>Apparent molecular weight by HPLC gel filtration (kDa)</th>
<th>Expected molecular weight (kDa)</th>
</tr>
</thead>
<tbody>
<tr>
<td>M&lt;sub&gt;2&lt;/sub&gt;S&lt;sub&gt;1&lt;/sub&gt; MTase</td>
<td>220–230</td>
<td>162.2</td>
</tr>
<tr>
<td>HsdR subunit</td>
<td>137–148</td>
<td>119.6</td>
</tr>
<tr>
<td>In vivo assembled R&lt;sub&gt;1&lt;/sub&gt;M&lt;sub&gt;2&lt;/sub&gt;S&lt;sub&gt;1&lt;/sub&gt;</td>
<td>280–300</td>
<td>281.8</td>
</tr>
<tr>
<td>In vivo assembled mixture of R&lt;sub&gt;1&lt;/sub&gt;M&lt;sub&gt;2&lt;/sub&gt;S&lt;sub&gt;1&lt;/sub&gt; + R&lt;sub&gt;2&lt;/sub&gt;M&lt;sub&gt;2&lt;/sub&gt;S&lt;sub&gt;1&lt;/sub&gt;</td>
<td>333–350</td>
<td>281.8 + 401.5</td>
</tr>
<tr>
<td>In vitro assembled R&lt;sub&gt;2&lt;/sub&gt;M&lt;sub&gt;2&lt;/sub&gt;S&lt;sub&gt;1&lt;/sub&gt; (freshly prepared)</td>
<td>290–315</td>
<td>401.5</td>
</tr>
<tr>
<td>In vitro assembled R&lt;sub&gt;2&lt;/sub&gt;M&lt;sub&gt;2&lt;/sub&gt;S&lt;sub&gt;1&lt;/sub&gt; (after 24 h incubation)</td>
<td>380–420</td>
<td>401.5</td>
</tr>
</tbody>
</table>

<sup>a</sup>Produced by the pJS4M-pACR124 plasmid system.

<sup>b</sup>Produced by the pJS4M-pBGS124 plasmid system.
Analysis of interactions between the MTase and HsdR in the absence of DNA

To analyse endonuclease assembly in the absence of DNA, we used HPLC gel filtration. The apparent molecular weights of various subunit complexes of EcoR124I from an HPLC gel filtration column were determined from their elution times and are given in Table 1. It can be seen that the MTase and HsdR subunit eluted from the column earlier than expected, giving a slightly higher weight. Similar behaviour was observed for the EcoKI enzyme (20). The endonuclease purified from cells containing the pJS4M-pACR124 plasmid-expression system eluted with an apparent molecular weight similar to that expected for R1M2S1. This is in agreement with previously published data (23). The endonuclease purified from cell culture containing the pJS4M-pBGSR124 plasmid-expression system had an apparent mass intermediate between R1M2S1 and R2M2S1 and formed a slightly asymmetric peak suggesting that it was a mixture of these two species. A freshly prepared mixture of MTase with a 5-fold excess of HsdR subunit, eluted with an apparent weight close to that expected for R1M2S1. However, the same mixture after a 24 h incubation at room temperature showed the formation of a higher molecular weight species with a weight equal to that expected for the R2M2S1 endonuclease. Figure 5a shows the elution profiles of the HsdR subunit, MTase and a mixture of these two components freshly prepared and after 24 h. The titration of a constant concentration of MTase with the HsdR subunit also showed the dramatic change in molecular weight of the complex upon prolonged incubation (Fig. 5b). Therefore, HPLC data indicated that the formation of the R2M2S1 complex is a slow process under the conditions used. One possible explanation of this observation is that the presence of 5% glycerol in the samples slows diffusion; although it seems unlikely that this would be a large effect. Alternatively, it appears likely that the presence of DNA in the sample grossly changes the rate of attainment of thermodynamic equilibrium. This idea is supported by the fact that DNA cleavage appeared in a very short time following mixing of MTase and HsdR in the presence of DNA (see the next section).

DNA cleavage and ATPase activities of MTase–HsdR complexes

To investigate enzyme activities of the R1M2S1 and R2M2S1 species, DNA cleavage and ATP hydrolysis were assayed following a reconstitution titration of the MTase with HsdR in the presence of a plasmid DNA substrate. The DNA used contained a single EcoR124I recognition site. This DNA readily undergoes primary cleavage to produce full-length linear DNA. Secondary processing of the linear product is observed only at very high enzyme:DNA ratios (23). Reaction products (linear DNA and inorganic phosphate, respectively) were quantified by densitometric scanning of agarose gels and malachite green assay as described elsewhere (23). A gradually increasing concentration of HsdR subunit was added into a 1:1 mixture of the MTase and DNA, which were present at a concentration of 200 nM to ensure full DNA binding. The same concentration conditions were also used in the reconstitution titration followed by a gel retardation assay (Fig. 3). At an HsdR:MTase ratio <1, where only the R1M2S1 complex was detected by a gel retardation assay, no DNA cleavage occurred (Fig. 6a). However, ATPase activity gradually increased with HsdR concentration (Fig. 6b). It is important to note that the HsdR subunit by itself shows only a weak DNA/Mg2+-dependent ATPase activity which is 100 times lower than that of the endonuclease (37). DNA cleavage appeared only when the R2M2S1-DNA complex of the total bound DNA was plotted against protein concentration (b).

DISCUSSION

The gel retardation and DNA cleavage experiments presented in this work revealed that the active form of the type IC EcoR124I restriction endonuclease has a subunit stoichiometry of R2M2S1. It is assembled from the trimeric MTase (M2S1) and the HsdR subunits through a stable intermediate with a stoichiometry of R1M2S1, which does not cleave DNA. While binding between the MTase and the first HsdR subunit is apparently very stable,
Figure 5. Elution profiles from the HPLC gel filtration column and titration of MTase with HsdR followed by HPLC gel filtration. (a) From left to right the traces show MTase (420 nM), HsdR subunit (1 µM), MTase (420 nM) freshly mixed with a 5-fold excess of HsdR subunit, and MTase (420 nM) incubated with a 5-fold excess of HsdR subunit for 24 h before application to the column. The numbers are the elution times, in minutes, of the peaks. (b) The molecular weight, determined by HPLC gel filtration, of the complex formed between MTase and HsdR subunit as the HsdR subunit concentration is increased. The MTase concentration was 420 nM throughout. The samples were applied to the column immediately after mixing ($) or after 24 h incubation (C). The lines drawn are only to guide the eye.

interaction between the R1M2S1 intermediate and the second HsdR subunit is much weaker. The R2M2S1 complex dissociated into the R1M2S1 complex and the HsdR subunit with an apparent Kd of $\sim 2.4 \times 10^{-7}$ M (Fig. 4).

In contrast to EcoR124I, the R2M2S1 complex of EcoKI endonuclease, a member of the type IA family, is relatively stable and both HsdR subunits appear to be bound to the MTase with similar affinities (20; L.Powell and N.Murray, personal communication).

Our results indicate that the two HsdR subunits interact differently with the MTase in the assembly of the EcoR124I endonuclease. A simple model for assembly would be that there are two equivalent HsdR-binding sites on the MTase, but that the presence of one bound HsdR subunit affects the binding of the second HsdR subunit perhaps by partially blocking the second site, or by means of a conformational change at the second site. This model would require that the R1M2S1 complex should be a 1:1 mixture of two species with occupation of either of the two HsdR-binding sites. A second model would require that the two sites are not equivalent and that the assembly is inherently asymmetric with only one type of R1M2S1 complex being formed. However, the data presented in this work are not sufficient to definitely determine which model is applicable.

Since binding of the first HsdR subunit to the MTase is much stronger than binding the second HsdR subunit to the R1M2S1 intermediate, the in vivo formation of the R2M2S1 complex will be dependent on the ratio between the HsdR subunit and the MTase in the cell. At an HsdR:MTase ratio of <1, only the R1M2S1 complex will be present. The R2M2S1 complex will be formed only when the HsdR:MTase ratio is >1. Indeed, when the ratio between MTase and HsdR was varied in vivo by using different recombinant two-plasmid systems, the resulting endonuclease preparations differed in ratio between the species (Fig. 1). The purified endonuclease produced from hsd genes under the control of their natural promoters was also found to be a mixture of R1M2S1 and R2M2S1 species (J.Dreier, personal communication). This shows that the heterogeneity of the stoichiometry of EcoR124I endonuclease assembled in vivo was not a result of using an artificial expression system. The presence of two non-equivalent binding sites for HsdR has also been shown using the technique of surface plasmon resonance (38). This
clearly supports the data presented in this paper and has shown that the two sites differ in binding affinity by at least two orders of magnitude. Since two separate techniques have demonstrated the equilibrium between the RfM2S1 and R1M2S1 forms of the endonuclease, we believe the slow assembly of the R2 complex in the absence of DNA, as measured by HPLC gel filtration, either reflects the experimental conditions used, or the absence of DNA greatly decreases the rate for attainment of thermodynamic equilibrium.

Why should the restriction endonuclease of the EcoR124I R–M system be such a weak complex and the stable assembly intermediate R1M2S1 not be able to cleave DNA? We suggest this may reflect a very sensitive mechanism for regulation of restriction activity following conjugal transfer of the hsd genes into a non-modified recipient cell. The EcoR124I R–M system was originally found on the conjugative plasmid R124 (16) and this presents an unusual problem for the recipient cell. If the R124 plasmid entered a new host and immediately produced an active endonuclease, the unmodified host DNA would be restricted and the host chromosome is modified following conjugation. Despite this, when the R124 plasmid entered a new host and immediately produced an active endonuclease, the unmodified host DNA would be restricted and the host chromosome is modified following conjugation. Despite these attempts to show transcriiptional or post-transcriptional control of this phenomenon for the EcoR124I R–M system (32; K.Firman, unpublished observations), there is no evidence of such control. Nor is there any evidence of such control for the other type I R–M systems. Therefore, we propose that this control is exercised at the level of subunit assembly. The M2S1 MTase is a very tight complex and it will be assembled first in the cell. Since the MTase can form a tight complex with one HsdR subunit, which does not cleave DNA, all the HsdR subunit will initially be trapped in this inactive complex. Therefore, the MTase (and possibly the R1M2S1 complex) can modify and protect the host DNA. The weak endonuclease complex RfM2S1 can be formed only when a large amount of HsdR has built up in the cell, which is unlikely during the early stage of establishing the R–M system in a new host. When cellular levels of individual subunits were altered by using different promoters and expression vectors, it was observed that excess HsdR over MTase was lethal, unless the MTase was already expressed in the cell (23). This implies that the function of the EcoR124I endonuclease in the cell is finely balanced and absolutely dependent upon subunit concentration. Control of restriction and modification by subunit assembly has also been recently proposed for the EcoK1 restriction endonuclease (20), suggesting this mechanism may be common to all type I systems. In addition, the EcoR124I system shows a relatively high methylation activity with non-modified DNA (23) when compared to that of EcoK1 (11). This will also ensure that a non-modified chromosome in the recipient cell can be readily modified following conjugal transfer of the hsd genes.

ACKNOWLEDGEMENTS

The authors would like to thank Lynn Powell and Noreen Murray (University of Edinburgh) for communicating results prior to publication. PJ was supported by an International Fellowship from the Wellcome Trust. D.D. is supported by a Royal Society University Research Fellowship.

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