

## A Five-Gene Molecular Grade Index and *HOXB13:IL17BR* Are Complementary Prognostic Factors in Early Stage Breast Cancer

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**Abstract Purpose:** Histologic tumor grade is a well-established prognostic factor for breast cancer, and tumor grade-associated genes are the common denominator of many prognostic gene signatures. The objectives of this study are as follows: (a) to develop a simple gene expression index for tumor grade (molecular grade index or MGI), and (b) to determine whether MGI and our previously described *HOXB13:IL17BR* index together provide improved prognostic information.

**Experimental Design:** From our previously published list of genes whose expression correlates with both tumor grade and tumor stage progression, we selected five cell cycle-related genes to build MGI and evaluated MGI in two publicly available microarray data sets totaling 410 patients. Using two additional cohorts ( $n = 323$ ), we developed a real-time reverse transcription PCR assay for MGI, validated its prognostic utility, and examined its interaction with *HOXB13:IL17BR*.

**Results:** MGI performed consistently as a strong prognostic factor and was comparable with a more complex 97-gene genomic grade index in multiple data sets. In patients treated with endocrine therapy, MGI and *HOXB13:IL17BR* modified each other's prognostic performance. High MGI was associated with significantly worse outcome only in combination with high *HOXB13:IL17BR*, and likewise, high *HOXB13:IL17BR* was significantly associated with poor outcome only in combination with high MGI.

**Conclusions:** We developed and validated a five-gene reverse transcription PCR assay for MGI suitable for analyzing routine formalin-fixed paraffin-embedded clinical samples. The combination of MGI and *HOXB13:IL17BR* outperforms either alone and identifies a subgroup (~30%) of early stage estrogen receptor-positive breast cancer patients with very poor outcome despite endocrine therapy.

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The most recent (2005) St. Gallen consensus guidelines for treatment selection for early stage breast cancer consider both risk of recurrence and endocrine responsiveness to better balance risk and benefit of systemic adjuvant therapies (1). To better define risk stratification, genome-wide expression profiling studies have created multiple prognostic gene signatures for breast cancer (2, 3). An important issue is whether these signatures overlap in their prognostic information and whether combining several of these signatures would provide more accurate prognosis. In one comparative study, four signatures (the intrinsic subtypes, 70-gene signature, wound response signature, and Recurrence Score) were found to be highly concordant in classifying patients into low and high-risk groups (4). Notably, combining these signatures did not yield significant improvement in predictive accuracy, suggesting that the prognostic information provided by these signatures is largely overlapping (4). Sotiriou et al. (5) showed that a 97-gene tumor grade signature was comparable with the 70-gene signature and the Recurrence Score algorithm (6) in independent cohorts, and they hypothesized that most of the prognostic power of these

signatures comes from genes associated with cellular proliferation (7).

Previously, we conducted a microarray analysis of 60 patients with hormone receptor-positive breast cancer treated with standard 5 years of adjuvant tamoxifen therapy (8). To facilitate discovery of novel biomarkers predictive of clinical outcome beyond standard prognostic factors, patients who developed recurrences were matched to those who did not with respect to tumor stage and grade. We identified three genes associated with clinical outcome, *HOXB13*, *IL17BR*, and *CHDH*, none of which had been previously implicated in breast cancer. Because high *HOXB13* and low *IL17BR* expression levels are associated with recurrence, we proposed that a simple *HOXB13:IL17BR* two-gene ratio could serve as a novel biomarker for predicting recurrence in breast cancer patients receiving adjuvant tamoxifen therapy. Subsequent studies by us (9, 10) and others (11, 12) have shown that *HOXB13:IL17BR* is both prognostic (i.e., predicts the risk of breast cancer recurrence) and predictive of tamoxifen benefit (i.e., tamoxifen response/resistance).

Given the well-established prognostic importance of histologic tumor grade (13) and tumor grade-associated genes (7) in breast cancer, a simple robust gene expression assay for tumor grade would be a valuable clinical tool. Herein, we developed a simple 5-gene tumor grade signature (MGI for molecular grade index) that recapitulates tumor grade and show that it predicts clinical outcome with comparable performance to the well-characterized 97-gene tumor grade index. We implemented a robust real-time reverse transcription-PCR (RT-PCR) assay for MGI and show that MGI together with *HOXB13:IL17BR* provides more accurate prognosis than either biomarker alone.

## Patients and Methods

**Patients and tumor samples.** The designation of training and test sets, and the work flow of data analyses are outlined in Fig. 1. Previously published microarray data sets (accessions GSE3494 and GSE1456) are described in Supplementary Data. Two additional patient cohorts were analyzed by real-time RT-PCR. The first cohort [Massachusetts General Hospital (MGH) cohort] used a retrospective case-cohort design (14) and was derived from 683 stage I, II, and III patients with estrogen receptor-positive primary breast cancer treated at the MGH from 1991 to 1999. Clinical follow-up data were obtained from tumor registry and hospital records. Cases were all patients who developed distant metastasis during follow-up; controls were randomly selected from patients who remained disease free at last follow-up to achieve a 2:1 ratio of controls to cases. In addition, controls were matched to cases with respect to adjuvant therapy and time of diagnosis. For ~80% of the cases and controls, both clinical outcome data and formalin-fixed paraffin-embedded tumor blocks were successfully retrieved. The final cohort consisted of 79 cases and 160 controls, and its patient and tumor characteristics were summarized in Supplementary Table S1. The second cohort (Oxford cohort; see Supplementary Table S2) consisted of 84 of the Oxford series described previously (6); all patients had estrogen receptor-positive primary breast cancer and were treated with adjuvant tamoxifen therapy. This study used portions of RNA from frozen tumor samples isolated previously (6).

**Gene expression analysis by real-time RT-PCR.** We used primer/probe sequences for *HOXB13*, *IL17BR*, *ESR1*, *PGR*, *CHDH*, *ACTB*, *HMBS*, *SDHA*, and *UBC* described previously (9), and designed primer/probe sequences for the five molecular grade genes (*BUB1B*, *CENPA*, *NEK2*, *RACGAP1*, and *RRM2*) and *ERBB2* (*HER2*; Supplementary Table S2), using Primer Express (Applied Biosystem, Inc.). For

each formalin-fixed paraffin-embedded sample, two 7- $\mu$ m tissue sections were subject to gross macrodissection to enrich for tumor content. RNA extraction, reverse transcription, and Taq Man RT-PCR using the ABI 7900HT instrument (Applied Biosystem, Inc.) were done as previously described (9). The cycling threshold numbers were normalized to the mean cycling threshold of four reference genes (*ACTB*, *HMBS*, *SDHA*, and *UBC*).

**Calculation of gene expression indices.** Normalized expression levels for the five molecular grade genes from microarrays or RT-PCR were standardized to a mean of 0 and a SD of 1 across samples within each data set and then combined into a single index per sample as the first principal component (15). Standardization of the primary data within each data set was necessary to account for the different platforms (microarrays and RT-PCR) and sample types (frozen and formalin-fixed paraffin-embedded).

*HOXB13:IL17BR* was calculated as described previously (9); the means and SDs for *HOXB13* and *IL17BR* used for standardizing the MGH cohort were derived from an analysis of 190 formalin-fixed paraffin-embedded samples from a separate population-based cohort of estrogen receptor-positive lymph node-negative breast cancer patients (data not shown).

Genomic grade index (GGI) was calculated from microarray data using the 128 Affymetrix probe sets representing 97 genes and scaled within each data set to have a mean of -1 for grade 1 tumors and +1 for grade 3 tumors as described before (5).

**Statistical analysis.** The cutpoint for MGI was determined as follows. Initial analysis of MGI in the Uppsala cohort indicated good discrimination of grade 1 and grade 3 tumors using the mean (0) as cutpoint. This cutpoint was further supported by receiver operating characteristic analysis (Supplementary Fig. S1; ref. 16). GGI was dichotomized at the cutpoint of 0 as described previously (5). The cutpoint of 0.06 for *HOXB13:IL17BR*, previously defined to stratify patients treated with adjuvant tamoxifen into low and high risk of recurrence (9), was applied directly in this study.

Kaplan-Meier analysis with log-rank test and Cox proportional hazards regression were done to assess the association of gene expression indices with clinical outcome. Multivariate Cox regression models were done to assess the prognostic capacity of gene

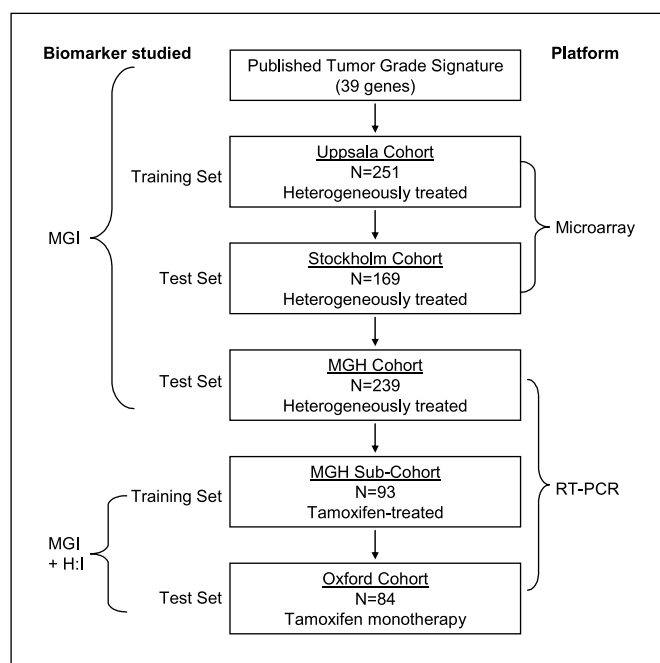
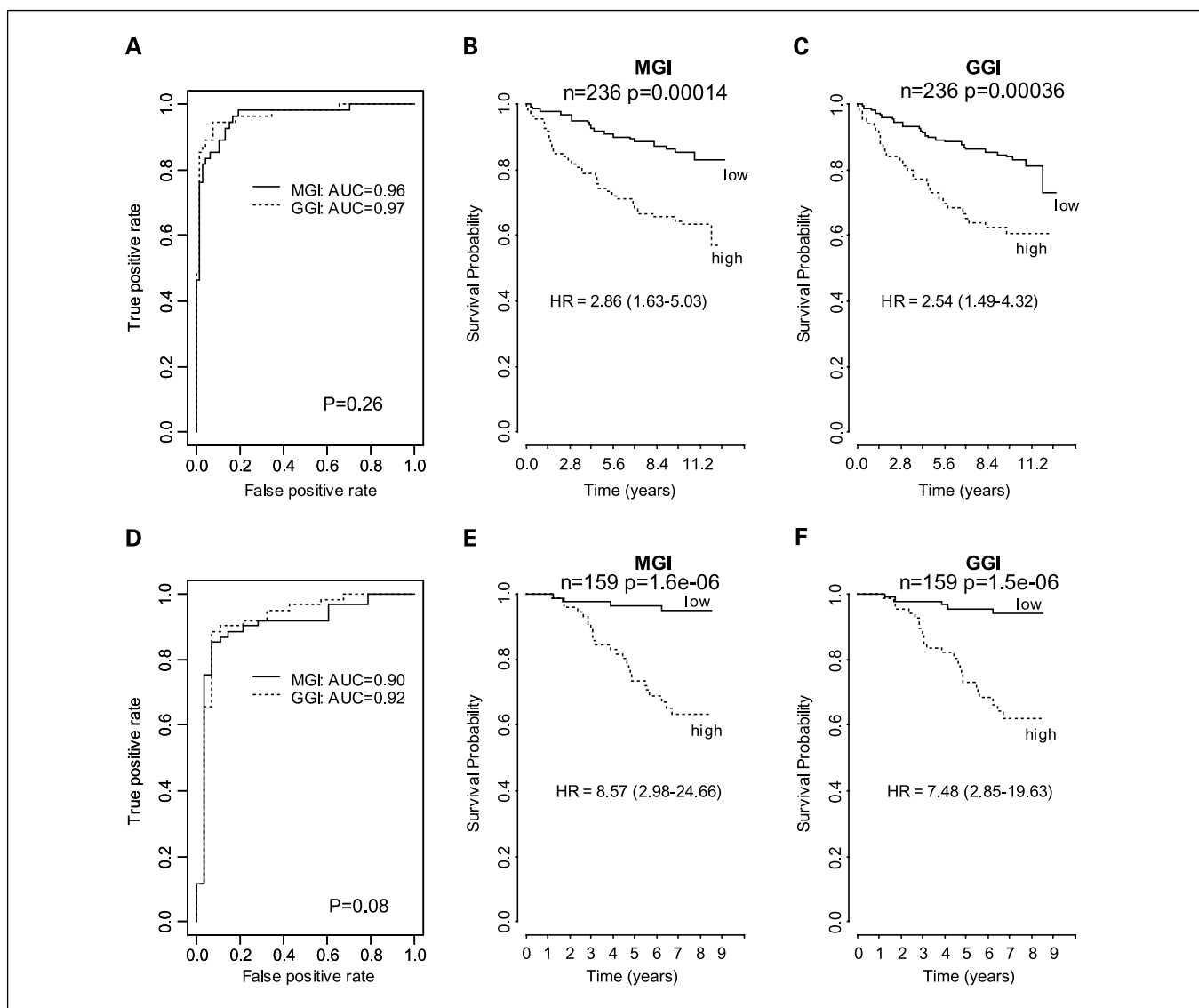


Fig 1. Flow chart of study design with data sets and analysis.



**Fig. 2.** Comparison of MGI and GGI for correlation with tumor grade and clinical outcome. *A-C*, Uppsala cohort; *D-F*, Stockholm cohort. Note that in Kaplan-Meier analysis of the Uppsala cohort, 15 patients with no clinical follow-up were excluded, resulting in 236 patients. *A* and *D*, receiver operating characteristic curve analysis of MGI and GGI for discriminating grade 1 and grade 3 tumors. *B-F*, Kaplan-Meier survival curves showing probability of breast cancer-specific death according to MGI or GGI status (high versus low). HR, hazard ratio.

expression indices after adjusting for known prognostic factors. Proportional hazard assumption was checked by scaled Schoenfeld residuals; variables violating proportional hazard assumption were adjusted for in the model through stratification. To account for the case-cohort design of the MGH cohort, we used weighed Kaplan-Meier analysis and Cox regression models with modifications to handle case-cohort designs (17, 18) as implemented in the survey package in R.<sup>6</sup> To test for interaction between dichotomized MGI and *HOXB13:IL17BR* in Cox regression models, the Wald statistic was used in the MGH cohort and the likelihood ratio test was used in the Oxford cohort.

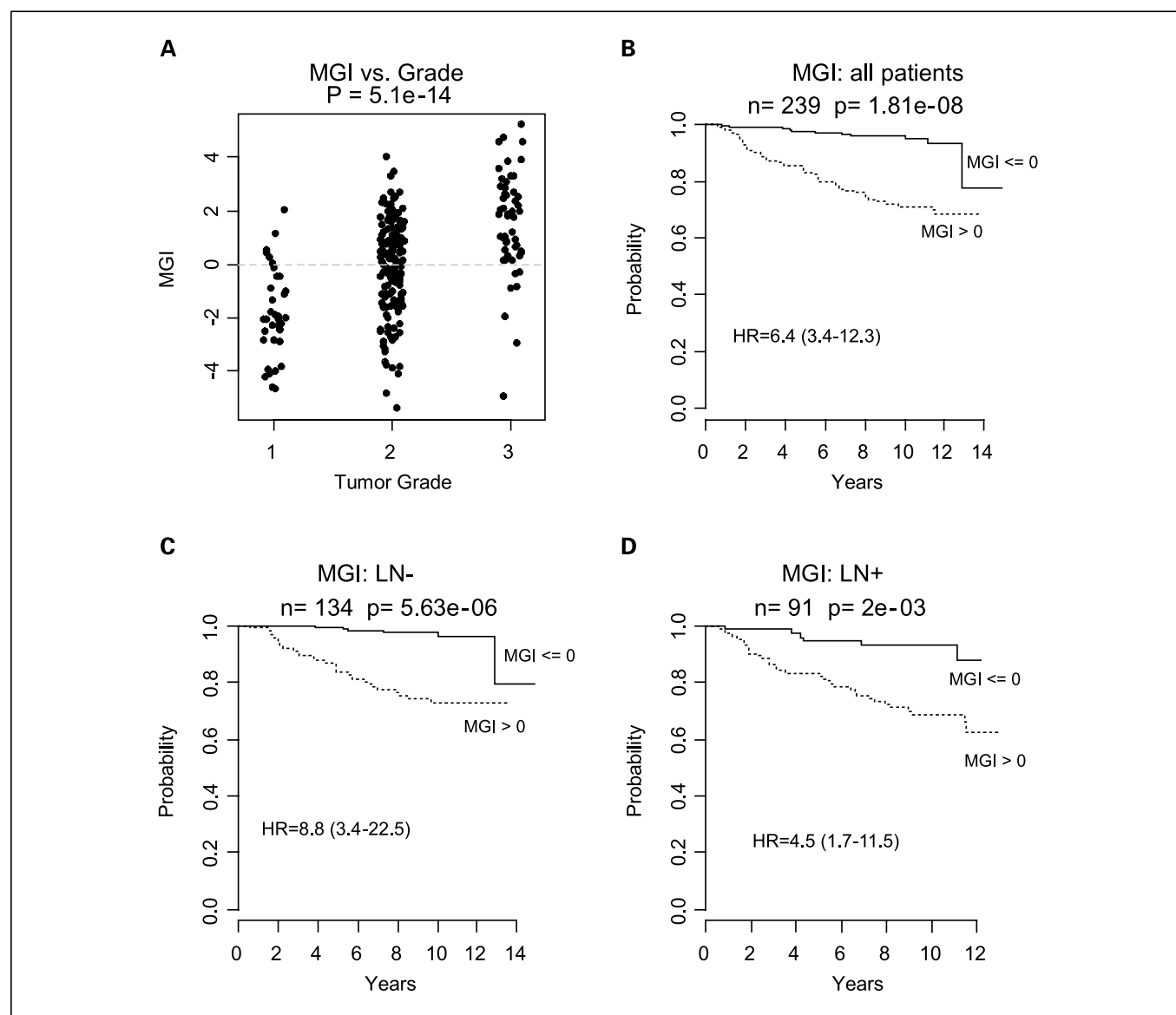
Correlations of continuous variables with categorical factors were examined using nonparametric two-sample Wilcoxon test or Kruskal-Wallis test for factors with more than two levels. All statistical analyses

were done in R. All significance tests were two sided, and a *P* value of <0.05 was considered significant.

## Results

**Gene selection for MGI.** In our previous study of gene expression associated with breast cancer progression, we showed hundreds of genes differentially expressed between tumors of low and high histologic grade (19). Moreover, we showed that a subset of 39 genes with increased expression in high-grade tumors were also more highly expressed in the invasive compared with the preinvasive stage of breast cancer, suggesting a role for these genes in invasive growth (19). Thus, we focused on this subset of 39 genes to develop a prognostic tumor grade signature. First, we confirmed their high correlation with tumor grade in a large publicly available microarray

<sup>6</sup> <http://www.r-project.org>



**Fig. 3.** MGI determined by the RT-PCR Taq Man assay in the MGH cohort. *A*, correlation of MGI with tumor grade. *B-D*, Kaplan-Meier analysis of distant metastasis-free survival according to MGI using all patients (*B*), lymph node – negative (*C*), or lymph node – positive patients (*D*).

data set (Uppsala cohort,  $n = 251$ ; ref. 20). Next, we narrowed this list to five genes based on functional annotation of the genes [genes involved in different cell cycle phases (21) and processes], association with clinical outcome (Uppsala cohort,  $n = 236$ ), and correlation with tumor grade in another independent 60-patient cohort (see Supplementary Tables S3-5; ref. 8).

Through the use of unsupervised (i.e., without using any clinical outcome data) principal component analysis, we combined the five-gene expression pattern into a single MGI score and showed that MGI strongly correlated with tumor grade (Supplementary Fig. S1). Using the mean value (0) of MGI as a cutpoint, most of the grade 1 and grade 3 tumors were classified correctly (89% overall accuracy), and grade 2 tumors were stratified into two groups (59% and 41% in the low and high MGI group, respectively).

**Prognostic performance of MGI in breast cancer patients.** Using two independent publicly available microarray data sets (Uppsala and Stockholm data sets; Fig. 1), we examined the capacity of MGI to predict clinical outcome in breast cancer patients and compared MGI with the previously described 97-gene GGI (5). In the Uppsala data set, receiver operating characteristic analysis indicated that MGI and GGI were comparable in discriminating grade 1 and grade 3 tumors, and Kaplan-Meier analysis (MGI dichotomized at a cutpoint of 0) separated patients into two subgroups with significantly different risks of breast cancer death (Fig. 2A-B). The survival curves and hazard ratios were comparable with those generated by GGI (Fig. 2C). Similar results were obtained in the independent Stockholm test data set (Fig. 2D-F). Therefore, our results showed that a simple five-gene index could reproduce the prognostic performance of the more complex 97-gene

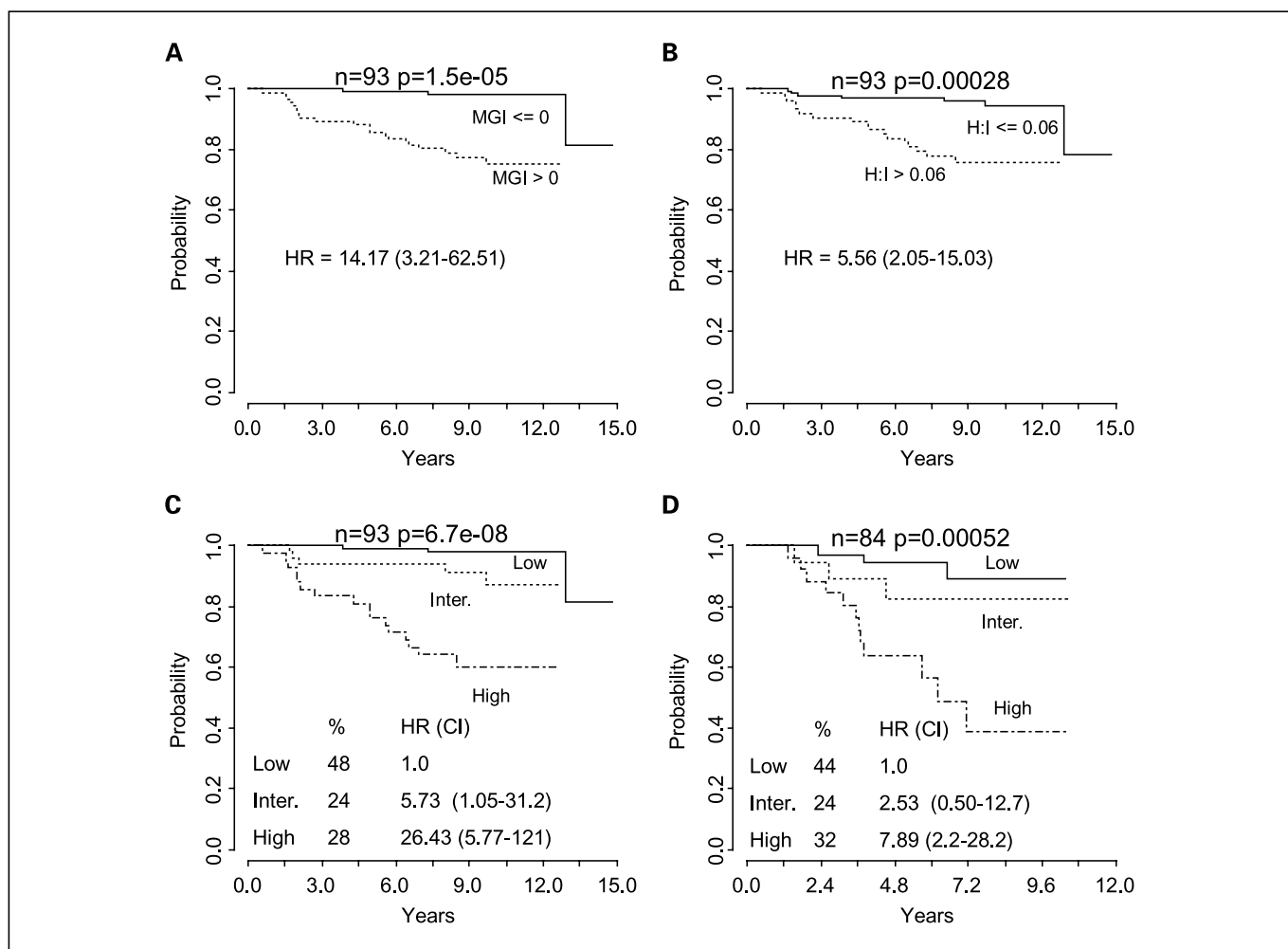
**Table 1.** Multivariate Cox proportional hazards analysis of MGI in entire cohort

| Variable    |                      | Hazard ratio (95% confidence interval) | P      |
|-------------|----------------------|--|--------|
| MGI         | High vs low          | 4.7 (2.1-10.8)                         | 0.0002 |
| Tumor size  | >2 cm vs <2 cm       | 0.8 (0.4-1.5)                          | 0.4580 |
| Tumor grade |                      |  | 0.0011 |
|             | II vs I              | 1.6 (0.5-5.2)                          | 0.4331 |
|             | III vs I             | 5.6 (1.5-20.6)                         | 0.0105 |
| Age         | ≥35 y vs <35 y       | 0.7 (0.2-1.9)                          | 0.4687 |
| Node status | Pos. vs neg.         | 1.2 (0.6-2.3)                          | 0.5581 |
| Treatment   |                      |  | 0.5733 |
|             | Chemo vs none        | 0.9 (0.4-2.4)                          | 0.8837 |
|             | Endo vs none         | 1.5 (0.5-4.5)                          | 0.4406 |
|             | Chemo + endo vs none | 1.0 (0.3-3.5)                          | 0.9939 |

Abbreviations: Pos, positive; Neg, negative; Chemo, chemotherapy; Endo, endocrine therapy.

signature; we note that although MGI was developed entirely independently of GGI, four (*BUB1B*, *CENPA*, *RACGAP1*, and *RRM2*) of the five genes were among the 97-gene signature, and the fifth gene, *NEK2*, was only 2 positions down from the 112 grade 3-associated probe sets included in GGI (5).

**Development and validation of an RT-PCR assay for MGI.** We designed primers and probes for the five MGI genes in the TaqMan real-time PCR (RT-PCR) assay format (Supplementary Table S3). To validate the RT-PCR-based MGI assay, we carried out a retrospective case-cohort study. The cases were patients



**Fig. 4.** Kaplan-Meier analysis of distant metastasis-free survival according to MGI (A), *HOXB13:IL17BR* (B), or the three groups (low, intermediate, and high risk) generated by combing MGI and *HOXB13:IL17BR* (C) in the MGH cohort, or the same three risk groups in the Oxford cohort (D).

**Table 2.** Multivariate Cox proportional hazards model of combining MGI and *HOXB13:IL17BR* in node-negative patients treated with endocrine therapy or endocrine therapy + chemotherapy

| Variables                  |                      | Hazard ratio (95% confidence interval) | P      |
|----------------------------|----------------------|--|--------|
| MGI + <i>HOXB13:IL17BR</i> |                      |  | 0.0007 |
|                            | Intermediate vs low  | 5.5 (0.9-34.6)                         | 0.0720 |
|                            | High vs low          | 24.2 (4.3-135.2)                       | 0.0003 |
| Tumor size                 | >2 cm vs ≤2 cm       | 1.0 (0.3-2.9)                          | 0.9804 |
| Age                        | ≥ 35 y vs <35 y      | 0.1 (0.0-0.4)                          | 0.0036 |
| Treatment                  | Endo vs chemo + endo | 11.5 (2.2-59.4)                        | 0.0034 |

NOTE: Tumor grade was adjusted for by stratification.

who were treated at the MGH between 1991 and 1999 and developed distant metastasis during follow-up. Controls were randomly selected from patients treated during the same period and were disease free at their most recent follow-up (Supplementary Table S1). Patients were treated variably with the therapy of best choice by their medical oncologist including the following: no systemic therapy, hormonal therapy, and chemotherapy. Because we were most interested in determining the therapy-independent prognostic utility of MGI, we matched controls with cases with respect to systemic therapy.

Similar to the microarray data sets analyzed above, the RT-PCR-based MGI also accurately discriminated grade 1 and grade 3 tumors (86% accuracy) using the same cutpoint of 0 as described above (Fig. 3A). Kaplan-Meier analysis indicated that high MGI was significantly associated with high risk of distant metastasis irrespective of nodal status (Fig. 3B-D). To remove potential confounding effect from mixed treatments in this cohort, we further performed Kaplan-Meier analysis of MGI within each treatment group. Indeed, MGI was significantly associated with distant metastasis-free survival in each of the untreated, endocrine-, and endocrine + chemotherapy-treated subgroups (Supplementary Fig. S2); the lack of significance in the chemotherapy-alone group might be due to its small sample size ( $n = 20$ ). In addition, MGI was significantly prognostic in both postmenopausal and premenopausal patients and in patients with small tumors (<2 cm) or with intermediate tumor grade (grade 2; Supplementary Fig. S3). Finally, in a multivariate Cox regression model adjusting for age, tumor size, tumor grade, lymph node status, and systemic therapy, MGI remained highly significant with a hazard ratio of 4.7 (2.1-10.8; Table 1).

**Complementary prognostic value of MGI and *HOXB13:IL17BR*.** To explore whether *HOXB13:IL17BR* provides additional prognostic information to MGI and vice versa, we analyzed both indices in patients with lymph node-negative tumors who received adjuvant endocrine therapy ( $n = 93$ ). In this patient group, MGI and *HOXB13:IL17BR* each was strongly associated with risk of distant metastasis (Fig. 4A-B). When both were considered together, MGI was highly significant in stratifying patients into low and high-risk groups only when the tumors had high *HOXB13:IL17BR*, and likewise, *HOXB13:IL17BR* was only significant in stratifying patients with tumors having high MGI ( $P_{\text{interaction}} = 0.09$ ; Supplementary Fig. S4). We therefore combined MGI and *HOXB13:IL17BR* to stratify patients into three risk groups (low risk, low for MGI and low or high *HOXB13:IL17BR*; intermediate risk, high MGI and low *HOXB13:IL17BR*; and high risk, high for

both MGI and *HOXB13:IL17BR*, accounting for 48%, 24%, and 28% of the patients, respectively). Kaplan-Meier analysis of these three groups indicated that high MGI and high *HOXB13:IL17BR* together predicted very poor outcome for the high-risk group (hazard ratio versus low-risk group = 26.4; 95% CI, 5.8-121; Fig. 4C). The Kaplan-Meier estimate of the 10-year distant metastasis-free survival probability was 98% (96-100%), 87% (77-99%), and 60% (47-78%) for the low, intermediate, and high-risk groups, respectively. Furthermore, after adjusting for systemic therapy and standard prognostic factors (age, tumor size, and grade) in a multivariate Cox regression model, the combined index remained highly statistically significant (Table 2), demonstrating the strong independent prognostic value of combining MGI and *HOXB13:IL17BR*.

To further substantiate the prognostic power of combining MGI and *HOXB13:IL17BR*, we examined these two indices in an independent test cohort of 84 patients with estrogen receptor-positive breast cancer uniformly treated with adjuvant tamoxifen therapy (Oxford cohort; ref. 6). After applying the same cutpoints to these two indices and the same combination algorithm as described above, the resulting low-, intermediate-, and high-risk groups consisted of 44%, 24%, and 32% of the patients, respectively, in keeping with their proportions seen in the MGH cohort. Again, Kaplan-Meier analysis indicated that the high-risk group with tumors high for both indices had the worst clinical outcome [hazard ratios versus low-risk group = 7.9 (2.2-28.2); Fig. 4D], and the likelihood ratio test indicated a statistically significant interaction between these two indices ( $P = 0.036$ ; see interaction plots in Supplementary Fig. S5).

## Discussion

Given the importance of tumor grade in prognosis and the existence of hundreds of genes whose expression levels highly correlate with tumor grade and proliferation (5, 19), it is not surprising that a multitude of seemingly distinct prognostic signatures could be developed. Furthermore, the prognostic robustness and redundancy of these genes suggest that a much simpler assay involving a few genes may be sufficient. For example, Ivshina et al. (22) showed that a 264-gene tumor grade signature can be reduced to 6 genes *in silico*, whereas Sotiriou et al. (6) noted that only a fraction of the 97 genes for GGI are needed for prognosis. In this study, through a combined data- and knowledge-driven approach, we developed

a five-gene tumor grade signature and implemented it in a robust RT-PCR assay. One important characteristic of MGI is that its calculation does not involve complex weighting trained on clinical outcome. Instead, it is a molecular correlate of tumor grade and derives its prognostic capacity from the latter (so-called “bottom-up” approach; ref. 3). The advantage of MGI over histologic tumor grade is 2-fold. First, such as GGI (5), it classifies grade 2 tumors to be either grade 1-like or grade 3-like, removing most of the ambiguity of pathologic tumor grading. Second, because an RT-PCR-based assay can be standardized in the clinical laboratory, it also removes the subjectivity and inter/intraobserver variability associated with pathologic grading (13).

Most notably, our results show that the prognostic accuracy of MGI can be augmented by also considering *HOXB13:IL17BR* and vice versa, suggesting a simple algorithm that stratifies patients into three risk groups. MGI and *HOXB13:IL17BR* seem to represent two distinct prognostic modules in breast cancer, as suggested by the observation that *HOXB13:IL17BR* but not MGI is strongly associated with the estrogen receptor signaling pathway (Supplementary Fig. S6). A recent small interfering RNA study of *HOXB13* in breast cancer cell lines suggests that expression of *HOXB13* blocks apoptotic cell death.<sup>7</sup> Therefore, a simple model to explain the interaction between these two indices is that a high proliferation rate (high MGI) and a decreased cell death (high *HOXB13:IL17BR*) promote aggressive tumor growth in a synergistic manner. We also note that MGI values are not statistically different between the intermediate (high MGI but low *HOXB13:IL17BR*) and high (high MGI AND high *HOXB13:IL17BR*) risk groups in our classification scheme. This is in contrast to the Recurrence Score algorithm, which defines the intermediate-risk group as having intermediate recurrence scores compared with the low and high-risk groups (23). It would be interesting to determine whether tumors with intermediate recurrence scores can be further stratified using our combination algorithm.

The St. Gallen guidelines classify estrogen receptor-positive node-negative breast cancer patients into low and interme-

diated-risk groups, with the majority falling into the latter. For example, applying these guidelines to the MGH cohort resulted in the classification of 86% of the patients into the intermediate-risk group. An important treatment decision is whether to withhold chemotherapy for some of the patients in the intermediate-risk group, a question targeted by the TAILORx (24) and MINDACT (25) prospective clinical trials (24, 25). Significantly, this large intermediate-risk group could be reclassified by MGI and *HOXB13:IL17BR* into low (43%), intermediate (26%), or high (31%) risk (Supplementary Fig. S7). Therefore, the use of MGI and *HOXB13:IL17BR* could identify a large subgroup of low-risk women that may be spared from toxic chemotherapy and a subgroup at high risk for whom more intense chemotherapy regimens or new therapeutic agents should be considered.

High tumor grade or mitotic index predicts benefit from chemotherapy in node-negative breast cancer patients (26). Indeed, high MGI predicted complete pathologic response to preoperative paclitaxel followed by 5-fluorouracil, doxorubicin, and cyclophosphamide in estrogen receptor-positive patients.<sup>8</sup> *HOXB13:IL17BR* has been shown to predict tamoxifen response in the metastatic setting (12) and benefit from prolonged tamoxifen monotherapy in the adjuvant setting (11). Thus, beyond risk stratification, MGI and *HOXB13:IL17BR* may prove to be important predictors of benefit from endocrine and chemotherapy agents.

In summary, we developed and validated MGI as a powerful prognostic factor in estrogen receptor-positive breast cancer. Furthermore, MGI and *HOXB13:IL17BR* can be combined to provide more accurate prognostic information than either alone. The identification of a subset of patients with very poor outcome using these two biomarkers should facilitate clinical trial designs and drug development to target those cancers with both high MGI and high *HOXB13:IL17BR*.

### Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed.

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<sup>8</sup> X. Ma, M. Erlander, D. Sgroi, unpublished data.

## References

- Goldhirsch A, Glick JH, Gelber RD, Coates AS, Thurlimann B, Senn HJ. Meeting highlights: international expert consensus on the primary therapy of early breast cancer 2005. *Ann Oncol* 2005;16:1569–83.
- Massague J. Sorting out breast-cancer gene signatures. *N Engl J Med* 2007;356:294–7.
- Sotiriou C, Piccart MJ. Taking gene-expression profiling to the clinic: when will molecular signatures become relevant to patient care? *Nat Rev Cancer* 2007;7:545–53.
- Fan C, Oh DS, Wessels L, et al. Concordance among gene-expression-based predictors for breast cancer. *N Engl J Med* 2006;355:560–9.
- Sotiriou C, Wirapati P, Loi S, et al. Gene expression profiling in breast cancer: understanding the molecular basis of histologic grade to improve prognosis. *J Natl Cancer Inst* 2006;98:262–72.
- Loi S, Haibe-Kains B, Desmedt C, et al. Definition of clinically distinct molecular subtypes in estrogen receptor-positive breast carcinomas through genomic grade. *J Clin Oncol* 2007;25:1239–46.
- Desmedt C, Sotiriou C. Proliferation: the most prominent predictor of clinical outcome in breast cancer. *Cell Cycle* 2006;5:2198–202.
- Ma XJ, Wang Z, Ryan PD, et al. A two-gene expression ratio predicts clinical outcome in breast cancer patients treated with tamoxifen. *Cancer Cell* 2004;5:607–16.
- Ma XJ, Hilsenbeck SG, Wang W, et al. The *HOXB13:IL17BR* expression index is a prognostic factor in early-stage breast cancer. *J Clin Oncol* 2006;24:4611–9.
- Goetz MP, Suman VJ, Ingle JN, et al. A two-gene expression ratio of homeobox 13 and interleukin-17B receptor for prediction of recurrence and survival in women receiving adjuvant tamoxifen. *Clin Cancer Res* 2006;12:2080–7.
- Jerevall PL, Brommesson S, Strand C, et al. Exploring the two-gene ratio in breast cancer-independent roles for *HOXB13* and *IL17BR* in prediction of clinical outcome. *Breast Cancer Res Treat* 2008;107(2):225–34. Epub 2007 Apr 24 2007.
- Jansen MP, Sieuwerts AM, Look MP, et al. *HOXB13*-to-*IL17BR* expression ratio is related with tumor aggressiveness and response to tamoxifen of recurrent breast cancer: a retrospective study. *J Clin Oncol* 2007;25:662–8.
- Cianfrocca M, Goldstein LJ. Prognostic and predictive factors in early-stage breast cancer. *Oncologist* 2004;9:606–16.
- Rundle AG, Vineis P, Ahsan H. Design options for molecular epidemiology research within cohort studies. *Cancer Epidemiol Biomarkers Prev* 2005;14:1899–907.
- Jolliffe IT. *Principal Component Analysis*. 2nd ed. New York: Springer-Verlag; 2002.
- Sing T, Sander O, Beerwinkel N, Lengauer T. ROCR: visualizing classifier performance in R. *Bioinformatics* 2005;21:3940–1.

17. Borgan O, Langholz B, Samuelsen SO, Goldstein L, Pogoda J. Exposure stratified case-cohort designs. *Lifetime Data Anal* 2000;6:39–58.
18. Barlow WE. Robust variance estimation for the case-cohort design. *Biometrics* 1994;50:1064–72.
19. Ma XJ, Salunga R, Tuggle JT, et al. Gene expression profiles of human breast cancer progression. *Proc Natl Acad Sci U S A* 2003;100:5974–9.
20. Miller LD, Smeds J, George J, et al. An expression signature for p53 status in human breast cancer predicts mutation status, transcriptional effects, and patient survival. *Proc Natl Acad Sci U S A* 2005;102:13550–5.
21. Whitfield ML, Sherlock G, Saldanha AJ, et al. Identification of genes periodically expressed in the human cell cycle and their expression in tumors. *Mol Biol Cell* 2002;13:1977–2000.
22. Ivshina AV, George J, Senko O, et al. Genetic reclassification of histologic grade delineates new clinical subtypes of breast cancer. *Cancer Res* 2006;66:10292–301.
23. Paik S, Shuk S, Tang G, et al. A multigene assay to predict recurrence of tamoxifen-treated, node negative breast cancer. *N Engl J Med* 2004;351:2817–26.
24. Sparano JA. TAILORx: trial assigning individualized options for treatment (Rx). *Clin Breast Cancer* 2006;7:347–50.
25. Bogaerts J, Cardoso F, Buyse M, et al. Gene signature evaluation as a prognostic tool: challenges in the design of the MINDACT trial. *Nat Clin Pract Oncol* 2006;3:540–51.
26. Page DL, Gray R, Allred DC, et al. Prediction of node-negative breast cancer outcome by histologic grading and S-phase analysis by flow cytometry: an Eastern Cooperative Oncology Group Study (2192). *Am J Clin Oncol* 2001;24:10–8.