

## Short Communication

# Modulation of Human Serum Glutathione S-Transferase A1/2 Concentration by Cruciferous Vegetables in a Controlled Feeding Study Is Influenced by *GSTM1* and *GSTT1* Genotypes

Sandi L. Navarro,<sup>1</sup> Jyh-Lurn Chang,<sup>1</sup> Sabrina Peterson,<sup>2</sup> Chu Chen,<sup>1</sup> Irena B. King,<sup>1</sup> Yvonne Schwarz,<sup>1</sup> Shuying S. Li,<sup>1</sup> Lin Li,<sup>1</sup> John D. Potter,<sup>1</sup> and Johanna W. Lampe<sup>1</sup>

<sup>1</sup>Cancer Prevention Program, Division of Public Health Sciences, Fred Hutchinson Cancer Research Center, Seattle, Washington and <sup>2</sup>Department of Food Science and Nutrition, University of Minnesota, St. Paul, Minnesota

### Abstract

Glutathione S-transferases (GST) detoxify a wide range of carcinogens. Isothiocyanates (ITC), from cruciferous vegetables, are substrates for and inducers of GST. GST variants may alter ITC clearance such that response to crucifers varies by genotype. In a randomized cross-over trial, we tested the hypothesis that changes in serum GSTA1/2 concentration in response to cruciferous vegetable feeding depends on *GSTM1/GSTT1* genotype. Thirty-three men and 34 women (age 20-40 years) ate four 14-day controlled diets—basal (vegetable-free), basal supplemented with two different doses of crucifers (“single dose” and “double dose”), and single-dose cruciferous-plus-apiaceous vegetables—fed per kilogram of body weight. Fasting bloods from days 0, 7, 11, and 14 of each diet period were analyzed for serum GSTA1/2 by ELISA. GSTA1/2 increased with single- and double-dose cruciferous compared with basal diet

(10% and 13%, respectively;  $P = 0.02$  and  $0.004$ ), but cruciferous-plus-apiaceous did not differ from basal ( $P = 0.59$ ). Overall, GSTA1/2 was higher in *GSTM1*-null/*GSTT1*-null than *GSTM1*+/*GSTT1*+ individuals ( $4,198 \pm 338$  and  $3,372 \pm 183$  pg/mL;  $P = 0.03$ ). The formal interaction of genotype-by-diet was not statistically significant, but the GSTA1/2 increase during the single-dose cruciferous diet was among *GSTM1*-null/*GSTT1*-null individuals (by 28%;  $P = 0.008$ ), largely explained by *GSTM1*-null/*GSTT1*-null men (by 41%;  $P = 0.01$ ). GSTA1/2 increased during the double-dose cruciferous diet in both *GSTM1*-null/*GSTT1*-null men (by 35%;  $P = 0.04$ ) and *GSTM1*+/*GSTT1*+ men (by 26%;  $P = 0.01$ ) but not in women. In summary, cruciferous vegetable supplementation increased GSTA1/2, but the effect was most marked in *GSTM1*-null/*GSTT1*-null men. (Cancer Epidemiol Biomarkers Prev 2009;18(11):2974–8)

### Introduction

Cruciferous vegetables contain high amounts of glucosinolates (1), which, upon hydrolysis, form biologically active compounds such as indoles and isothiocyanates (ITC). These compounds may exert chemoprotective effects through several mechanisms, including induction of detoxification enzymes. Glutathione S-transferases (GST) are enzymes that detoxify a broad range of electrophiles by conjugation with glutathione. ITCs are also substrates for GST, particularly *GSTM1* (2). Null genotypes for *GSTM1* and *GSTT1* result in the absence of their respective enzymes; thus, among *GSTM1*-null and *GSTT1*-null individuals, ITC may be metabolized more slowly and thus increase the likelihood of upregulation of other GST isoenzymes (3, 4). GSTA1 is the major hepatic GST (5). Despite overlap in substrate specificity,

*GSTA1* has a higher affinity than other GSTs for many carcinogens, particularly polycyclic aromatic hydrocarbons, including the activated heterocyclic amine, 2-amino-1-methyl-6-phenylimidazo[4,5-*b*]pyridine, produced in well-cooked meats and implicated in the etiology of colorectal cancer (6).

We previously reported that, compared with a diet devoid of fruit and vegetables, a cruciferous vegetable diet fed for 7 days statistically significantly increased serum GSTA1/2 concentrations, particularly in *GSTM1*-null women (7). We also found that GSTA1/2 concentrations measured at day 7 were significantly higher than on day 6, suggesting that the response to diet had not reached a steady state after 1 week. Our objectives in this follow-up study were to test (a) the combined effect of *GSTM1/GSTT1* genotypes on serum GSTA1/2 concentrations in response to three defined vegetable diets compared with a vegetable-free diet and (b) whether there was a dose-response effect. Secondary aims were to (a) evaluate the difference in serum GST- $\alpha$  concentrations between 1 and 2 weeks of cruciferous vegetable feeding and (b) determine the additional effect of *GSTT1* genotype on serum GSTA1/2 response to diet among *GSTM1*-null individuals.

Received 7/14/09; revised 8/27/09; accepted 9/11/09; published online 11/9/09.

Grant support: NIH grant R01CA070913.

Note: S.L. Navarro and J.-L. Chang contributed equally to this work.

Requests for reprints: Johanna W. Lampe, Cancer Prevention Program, Fred Hutchinson Cancer Research Center, 1100 Fairview Avenue North, M4-B402, Seattle, WA 98109. Phone: 206-667-6580, Fax: 206-667-7850. E-mail: jlampe@fhcc.org

Copyright © 2009 American Association for Cancer Research.

doi:10.1158/1055-9965.EPI-09-0701

## Materials and Methods

We used a randomized, controlled, crossover design with four experimental diets as described previously (8). Participants were recruited based on sex and *GSTM1/GSTT1* and *CYP1A2* genotypes; each participant received the four diets in computer-generated random sequence, blocked on genotype and sex. Each diet was consumed for 14 d with a 3-wk washout period between the diets. Exclusion criteria included factors known to influence biotransformation-enzyme induction, e.g., medications, alcohol, and smoking.

Of the 73 participants randomized, two had *GSTM1+/GSTT1*-null (versus *GSTM1*-null/*GSTT1*+) genotypes because they were recruited for their *CYP1A2*(*C*<sup>734</sup>*A*) genotype and were not included in this analysis. Three additional participants were not included in the analysis due to an insufficient serum sample or extreme *GSTA1/2* values (>20,000 pg/mL). Four participants dropped out after the first feeding period, five after the second, and three after the third. Data for all completed diet periods were included in the analysis, even if a participant did not complete all four diet periods, except for one individual who completed only the basal diet. Sixty-seven participants were included in the final analysis.

Participants consumed four different diets with vegetable doses based on a per kilogram of body weight (BW) calculation to minimize confounding by BW between sexes: a basal, fruit- and vegetable-free diet; basal diet supplemented with ~7 g cruciferous vegetables (a mixture of broccoli, cabbage, cauliflower, and radish sprouts) per kilogram of BW ("single dose"); basal diet supplemented with ~14 g cruciferous vegetables per kilogram of BW ("double dose"); and basal diet supplemented with ~7 g cruciferous vegetables plus ~4 g apiaceous vegetables (a mixture of carrots, celery, dill weed, parsley, and parsnips) per kilogram of BW. Study diet details have been published previously (8).

Biological samples were collected at baseline and during each 2-wk feeding period at days 0, 7, 11, and 14 in the morning after a 12-h overnight fast (8). Buccal cells, collected before randomization, were isolated and DNA was extracted for determination of *GSTM1/GSTT1* genotype and participant eligibility.

*GSTM1* and *GSTT1* genotyping (present versus null) was conducted on buccal cell DNA (8), using primers outlined by Arand et al. (9). *GSTA1* was amplified using primer sequences 5'-TGTTGATTGTTTGCCTGAAATT-CAC-3' and 5'-GTAAACGCTGTCACCGTCCTG-3' under the following PCR conditions: 1 cycle at 95°C for 5 min, 40 cycles at 94°C for 1 min, 63°C for 1 min, 72°C for 2 min, 1 cycle at 72°C for 5 min. The resulting PCR fragment was digested with the restriction enzyme *EarI* for 2 h at 37°C. The reaction was then run on a 2% agarose gel and the genotype was determined by fragments of different sizes (10).

Serum *GSTA1/2* concentrations were measured using a commercially available, enzyme-linked immunoassay kit (High Sensitivity Alpha GST EIA Hepkit, Biotrin International), which measures a mixture of *GSTA1* and *GSTA2* subunits (7). Intra- and inter-assay coefficients of variation on quality control serum (mean 3,510 pg/mL) were 2.7% and 16.2%, respectively. Using high performance

liquid chromatography (11), we measured urinary total ITC in 24-h urines collected on day 13 to assess diet adherence.

**Statistical Analysis.** Before analysis, natural logarithmic transformations were performed on *GSTA1/2* concentrations to normalize distributions. A linear mixed model was used, including sex, *GSTM1/GSTT1* genotypes, feeding periods, diet treatments, feeding order, sampling day, and interaction terms as fixed effects and participants as a random effect. Observations at day 0 and habitual diet were covariates adjusted in the model. Analyses by *GSTA1* genotype were carried out using the same model. Pearson correlation was used to evaluate the correlations between *GSTA1/2* concentrations and 24-h total ITC. All statistical analyses were done using the Statistical Analysis System Program (version 8.2; SAS Institute). Data are presented as back-transformed least squares (LS) means + SEMs, unless otherwise indicated. Because there were no statistically significant differences between analyses with and without adjustment for vegetable amount, the data are presented without adjustment. The two-sided *P* value for statistical significance was set at <0.05.

## Results

Of the 67 participants, two completed only three diet periods, five completed two, and three completed one diet period. There were no differences in demographic and baseline characteristics across genotypes (Table 1). Eighty-seven percent or more of the prescribed dose of study vegetables was consumed on each vegetable-supplemented diet. Based on daily food check-off forms, participants consumed nonstudy food items <3% of study days. Total vegetable intake ranged from 284 to 662 g for the single-dose cruciferous, 568 to 1,324 g for the double-dose cruciferous, and 458 to 1,065 g for the single-dose cruciferous-plus-apiaceous diet.

Overall (days 7, 11, and 14, and all diets combined), *GSTA1/2* concentrations were higher among *GSTM1*-null/*GSTT1*-null individuals than *GSTM1+/GSTT1+* individuals (4,198 ± 338 and 3,372 ± 183 pg/mL, respectively; *P* = 0.03), but did not differ between men and women (*P* = 0.4; Table 2). Among *GSTM1*-null individuals, there was no additional effect of *GSTT1*-null genotype (3,573 ± 190 pg/mL versus 4,198 ± 338 pg/mL for *GSTM1*-null/*GSTT1*-null; *P* = 0.1).

*GSTA1/2* concentrations were higher on the single-dose and double-dose cruciferous diets than on the basal diet (by 10% and 13%, respectively; *P* = 0.02 and 0.004); however, there was no dose-response effect (*P* = 0.5). Consumption of the single-dose cruciferous-plus-apiaceous diet did not increase *GSTA1/2* concentrations compared with the basal diet.

When evaluating response to diet stratified by genotype and sex, increases in *GSTA1/2* concentrations during the single-dose cruciferous diet were exclusively among *GSTM1*-null/*GSTT1*-null individuals (by 28%; *P* = 0.008), largely explained by *GSTM1*-null/*GSTT1*-null men (by 41%; *P* = 0.01). During the double-dose cruciferous diet, *GSTA1/2* concentrations increased in both *GSTM1*-null/*GSTT1*-null men (by 35%; *P* = 0.04) and *GSTM1+/GSTT1+* men (by 26%; *P* = 0.01), but not in women (Table 2). Although there was no overall effect

**Table 1. Characteristics of study participants stratified by sex and GSTM1/GSTT1 genotypes**

	Men			Women		
	GSTM1+/GSTT1+ (n = 14)	GSTM1-/GSTT1+ (n = 14)	GSTM1-/GSTT1- (n = 5)	GSTM1+/GSTT1+ (n = 12)	GSTM1-/GSTT1+ (n = 13)	GSTM1-/GSTT1- (n = 9)
Age (y)	33.9 ± 6.1	30.4 ± 7.0	29.7 ± 6.0	28.2 ± 5.8	30.3 ± 5.3	28.7 ± 3.5
Height (m)	177 ± 0.07	178 ± 0.07	174 ± 0.08	162 ± 0.07	165 ± 0.07	162 ± 0.08
Weight (kg)	83.6 ± 12.1	77.6 ± 11.6	76.3 ± 11.4	59.7 ± 9.1	61.2 ± 9.2	61.9 ± 12.8
BMI (kg/m <sup>2</sup> )	26.7 ± 3.3	24.4 ± 2.3	25.3 ± 3.4	22.8 ± 2.6	22.5 ± 2.5	23.3 ± 4.0
Race						
Caucasian	11 (79%)	9 (64%)	3 (60%)	8 (67%)	9 (69%)	3 (33%)
Asian	2 (14%)	5 (36%)	2 (40%)	4 (33%)	2 (15%)	4 (44%)
Other	1 (7%)	0	0	0	2 (15%)	2 (22%)
Baseline GST-α (pg/mL)	5,070 ± 374	8,379 ± 1,106	5,270 ± 587	4,923 ± 415	2,692 ± 150	4,239 ± 416

NOTE: There were no significant differences in baseline characteristic means ± SD across genotypes.

of cruciferous-plus-apiaceous vegetables compared with the basal diet, increases in GSTA1/2 concentrations were observed in GSTM1+/GSTT1+ men (by 20%;  $P = 0.03$ ; Table 2), but were related to lower GSTA1/2 concentrations during the basal diet. Compared with the single- and double-dose cruciferous diets, the cruciferous-plus-apiaceous diet decreased GSTA1/2 concentrations in GSTM1-null/GSTT1-null men (by 35% and 33%, respectively;  $P = 0.003$  and  $0.009$ ).

Overall, a statistically significant effect of the single-dose cruciferous diet on GSTA1/2 concentrations (compared with basal diet) was observed at day 7 (by 13%;  $P = 0.04$ ) but not at day 11 (by 11%;  $P = 0.07$ ) or day 14 (by 5%;  $P = 0.4$  Table 3). The double-dose cruciferous diet increased GSTA1/2 concentrations at days 7 and 11 (by 14% and 15%, respectively;  $P = 0.04$  and  $0.03$ ) but only marginally by day 14 (by 11%;  $P = 0.08$ ).

When examining the diet effects measured at different sampling days by genotype, the greatest effect of single-dose cruciferous vegetables was observed among

GSTM1-null/GSTT1-null individuals at day 7 (by 50%;  $P = 0.002$ ) and day 11 (by 31%;  $P = 0.04$ ), but not at day 14 (by 6%;  $P = 0.6$ ). Compared with the basal diet, the double-dose cruciferous diet did not differ by genotype at any sampling day, except for an increase in GSTA1/2 concentrations among GSTM1+/GSTT1+ individuals at day 11 (by 23%;  $P = 0.02$ ), a result of lower GSTA1/2 concentrations during the basal diet for this group.

The -69C>T polymorphism in the promoter region of the GSTA1 gene has been associated with 3- to 4-fold lower GSTA1/2 enzyme expression (10). We therefore evaluated whether serum GSTA1/2 concentrations differed by GSTA1 genotype. The overall interaction term for genotype-by-diet was not statistically significant nor were there any statistically significant differences in GSTA1/2 concentrations by GSTA1 within the diet (data not shown).

Mean ± SD total ITC concentrations for the basal, single-dose cruciferous, double-dose cruciferous, and cruciferous-plus-apiaceous diets were  $7.0 \pm 36.2$ ,  $130.7 \pm 57.1$ ,  $270.0 \pm 185.3$ , and  $107.9 \pm 49.8$  μmol/24 hours,

**Table 2. Serum GST-α concentrations by GSTM1/GSTT1 genotype, sex, and diet: the ratio between response to basal and vegetable diets**

Genotype	Diet periods*						
	GST-α, pg/mL	Ratios <sup>†</sup>					
		Basal <sup>‡</sup>	Single/basal	Double/basal	Double/single	Single + apiaceous/basal	Single + apiaceous/single
Overall (n = 67)	3,480 ± 155	1.10 ± 0.05 <sup>§</sup>	1.13 ± 0.05 <sup>§</sup>	1.03 ± 0.04	1.02 ± 0.04	0.93 ± 0.04	0.90 ± 0.04 <sup>§</sup>
GSTM1+/GSTT1+ (n = 26)	3,161 ± 212	1.02 ± 0.06	1.16 ± 0.07 <sup>§</sup>	1.14 ± 0.07 <sup>§</sup>	1.10 ± 0.07	1.08 ± 0.07	0.95 ± 0.06
GSTM1-null/GSTT1+ (n = 31)	3,500 ± 223	1.02 ± 0.06	1.07 ± 0.06	1.05 ± 0.06	1.00 ± 0.06	0.98 ± 0.06	0.93 ± 0.05
GSTM1-null/GSTT1-null (n = 14)	3,811 ± 371	1.28 ± 0.12 <sup>‡</sup>	1.18 ± 0.12	0.92 ± 0.09	0.98 ± 0.10	0.77 ± 0.08 <sup>§</sup>	0.83 ± 0.09
GSTM1+/GSTT1+ Men (n = 14)	2,863 ± 272	1.06 ± 0.09	1.26 ± 0.11 <sup>§</sup>	1.19 ± 0.10 <sup>§</sup>	1.20 ± 0.11 <sup>§</sup>	1.13 ± 0.10	0.96 ± 0.08
Women (n = 12)	3,490 ± 336	0.98 ± 0.08	1.06 ± 0.09	1.09 ± 0.10	1.00 ± 0.09	1.02 ± 0.09	0.94 ± 0.08
GSTM1-null/GSTT1+ Men (n = 16)	3,836 ± 342	0.96 ± 0.08	1.02 ± 0.08	1.06 ± 0.08	1.01 ± 0.08	1.05 ± 0.08	0.99 ± 0.08
Women (n = 15)	3,192 ± 303	1.08 ± 0.09	1.13 ± 0.09	1.05 ± 0.09	0.98 ± 0.08	0.91 ± 0.08	0.87 ± 0.07
GSTM1-null/GSTT1-null Men (n = 5)	3,923 ± 578	1.41 ± 0.19 <sup>§</sup>	1.35 ± 0.19 <sup>§</sup>	0.96 ± 0.14	0.91 ± 0.13	0.65 ± 0.09 <sup>§</sup>	0.67 ± 0.10 <sup>§</sup>
Women (n = 9)	3,701 ± 468	1.16 ± 0.14	1.02 ± 0.14	0.88 ± 0.11	1.05 ± 0.14	0.91 ± 0.12	1.03 ± 0.14

\*Basal, fruit/vegetable-free; single, single-dose cruciferous; double, double-dose cruciferous; all vegetable diets adjusted per kilogram of BW.

<sup>†</sup>The difference of the back-transformed LS means between diets as indicated.

<sup>‡</sup>LS means ± SEM, adjusted for baseline and feeding period day 0 serum GST-α concentrations.

<sup>§</sup>Significantly different at  $P < 0.05$ .

**Table 3. Serum GST- $\alpha$  concentrations by *GSTM1/GSTT1* genotype, sampling day, and diet: the ratio between response to basal and vegetable diets**

Genotype	Diet periods*						
	GST- $\alpha$ , pg/mL	Ratios <sup>†</sup>					
		Basal <sup>‡</sup>	Single/ basal	Double/ basal	Double/ single	Single + apiaceous/ basal	Single + apiaceous/ single
Overall							
Day 7	3,433 $\pm$ 185	1.13 $\pm$ 0.07 <sup>§</sup>	1.14 $\pm$ 0.07 <sup>§</sup>	1.00 $\pm$ 0.06	1.00 $\pm$ 0.06	0.88 $\pm$ 0.05 <sup>§</sup>	0.88 $\pm$ 0.05 <sup>§</sup>
Day 11	3,520 $\pm$ 190	1.11 $\pm$ 0.07	1.15 $\pm$ 0.07 <sup>§</sup>	1.03 $\pm$ 0.06	1.07 $\pm$ 0.07	0.96 $\pm$ 0.06	0.93 $\pm$ 0.06
Day 14	3,488 $\pm$ 188	1.05 $\pm$ 0.06	1.11 $\pm$ 0.07	1.06 $\pm$ 0.06	1.01 $\pm$ 0.06	0.96 $\pm$ 0.06	0.90 $\pm$ 0.06
<i>GSTM1+/GSTT1+</i>							
Day 7	3,263 $\pm$ 265	1.00 $\pm$ 0.09	1.12 $\pm$ 0.10	1.11 $\pm$ 0.10	1.10 $\pm$ 0.10	1.09 $\pm$ 0.10	0.98 $\pm$ 0.09
Day 11	3,081 $\pm$ 250	1.03 $\pm$ 0.09	1.23 $\pm$ 0.11 <sup>§</sup>	1.20 $\pm$ 0.11 <sup>‡</sup>	1.11 $\pm$ 0.10	1.08 $\pm$ 0.09	0.90 $\pm$ 0.08
Day 14	3,143 $\pm$ 255	1.03 $\pm$ 0.09	1.13 $\pm$ 0.10	1.09 $\pm$ 0.10	1.09 $\pm$ 0.10	1.06 $\pm$ 0.09	0.97 $\pm$ 0.09
<i>GSTM1-null/GSTT1+</i>							
Day 7	3,637 $\pm$ 278	0.96 $\pm$ 0.08	1.08 $\pm$ 0.09	1.12 $\pm$ 0.10	0.93 $\pm$ 0.08	0.97 $\pm$ 0.08	0.86 $\pm$ 0.07
Day 11	3,550 $\pm$ 272	1.03 $\pm$ 0.09	0.99 $\pm$ 0.08	0.96 $\pm$ 0.08	1.01 $\pm$ 0.08	0.99 $\pm$ 0.08	1.03 $\pm$ 0.09
Day 14	3,319 $\pm$ 257	1.07 $\pm$ 0.09	1.15 $\pm$ 0.10	1.08 $\pm$ 0.09	1.05 $\pm$ 0.09	0.98 $\pm$ 0.08	0.91 $\pm$ 0.08
<i>GSTM1-null/GSTT1-null</i>							
Day 7	3,409 $\pm$ 402	1.50 $\pm$ 0.19 <sup>§</sup>	1.21 $\pm$ 0.17	0.81 $\pm$ 0.11	0.97 $\pm$ 0.14	0.65 $\pm$ 0.09 <sup>§</sup>	0.80 $\pm$ 0.12
Day 11	3,989 $\pm$ 470	1.31 $\pm$ 0.17 <sup>§</sup>	1.25 $\pm$ 0.17	0.96 $\pm$ 0.13	1.09 $\pm$ 0.15	0.84 $\pm$ 0.12	0.88 $\pm$ 0.13
Day 14	4,068 $\pm$ 480	1.06 $\pm$ 0.14	1.07 $\pm$ 0.15	1.01 $\pm$ 0.14	0.89 $\pm$ 0.13	0.84 $\pm$ 0.12	0.83 $\pm$ 0.12

\*Basal, fruit/vegetable-free; single, single-dose cruciferous; double, double-dose cruciferous; all vegetable diets adjusted per kilogram of BW.

<sup>†</sup>The difference of the back-transformed LS means between diets as indicated.

<sup>‡</sup>LS means  $\pm$  SE, adjusted for baseline and feeding period day 0 serum GST- $\alpha$  concentrations.

<sup>§</sup>Significantly different at  $P < 0.05$ .

respectively, indicating a dose-dependent increase in ITC excretion over the basal-diet period. Correlations between GSTA1/2 concentrations and 24-hour urinary ITC excretion were not statistically significant ( $P = 0.46$ ).

## Discussion

In response to cruciferous vegetable feeding, GSTA1/2 concentrations were increased among individuals with combined *GSTM1-null/GSTT1-null* genotypes compared to their wild-type counterparts. Few human intervention trials have evaluated the ability of the *GST* genotype to modulate response to cruciferous vegetable intake on biomarkers. In one controlled feeding trial, *GSTM1* genotype-related changes were reported in transforming growth factor- $\beta$ 1 and epidermal growth factor signaling pathways in prostate tissue after 11 men consumed 400 g broccoli per week for 6 months (12); *GSTM1+* individuals showed greater diet-induced changes in prostate tissue gene expression. In our prior feeding study, the GSTA1/2 response to cruciferous vegetable feeding was greatest among *GSTM1-null* women (7).

Lack of consistent *GSTM1* modulation of crucifer effects across intervention studies is probably due to multiple factors, including tissue-specific responses, differences in end points measured, and the type and amount of crucifers fed. In our studies, we used a mixture of crucifers, previously  $\sim$ 400 g/d for 1 week (7) and currently  $\sim$ 300 to 1,300 g/d for 2 weeks, whereas Traka et al. (12) used only broccoli (400 g/wk). Glucosinolate composition, both amount and type, varies substantially among different cruciferous vegetables (13, 14). It is unknown whether these differences in glucosinolate profiles, and therefore ITC, lead to different biological effects in humans; however, several laboratories have shown differences in potency and function of ITC *in vitro* (15–17). Longer-term, chronic consumption of cooked

broccoli may also lead to changes in gut microbial enzymes and altered ITC exposure (18).

There were differences in response to crucifers between our prior study and the present one. Previously, we found that GST- $\alpha$  response was greatest among *GSTM1-null* women. Here, testing *GSTM1-null/GSTT1-null* genotypes combined, increases in GSTA1/2 concentrations were most marked in men. This may reflect a difference in dose. In our prior feeding trial, all participants received the same amount of vegetables daily. Consequently, the vegetable dose per BW was different between men and women ( $\sim$ 7 g/kg BW for women and  $\sim$ 6 g/kg BW for men,  $P = 0.001$ ). In the present study, vegetable amounts were dosed by BW to determine whether our previous observation was due to a dose difference or other sex-related physiologic effects. The lower dose in men relative to that in women in the original study may partially explain why women responded to a greater extent previously whereas men had a greater response here. There were also differences in baseline GSTA1/2 concentrations between sexes, between the studies. *GSTM1-null* women had lower basal serum GST- $\alpha$  concentrations than men of both genotypes in the initial study, and *GSTM1-null/GSTT1-null* women had the higher basal serum GSTA1/2 concentrations in the present study.

These differences in concentrations during the control diet influence the comparisons of diets between men and women in both trials. In either case, individuals with one or more null alleles responded to a greater extent than individuals with both intact alleles. These results also suggest that the intact *GSTT1* allele may be compensating for the lack of active *GSTM1* enzyme activity by playing a larger role in ITC metabolism among *GSTM1-null* individuals; when both alleles are absent, this compensation is no longer possible. Overlap in substrate specificity has been observed between different GST enzymes (6). Thus, it is possible that other GST enzymes compensate for polymorphic isoforms that result in lower activity.

Supplementation of apiaceous vegetables also affected GSTA1/2 concentrations, decreasing GSTA1/2 concentrations when consumed alone compared with the basal diet among *GSTM1+/GSTT1+* men in the first study (7) and attenuating the effects of the cruciferous vegetables in the present study. This underscores the challenge in interpreting the relationship between a complex, mixed diet and phenotype in the context of observational studies.

Contrary to our hypothesis, there was no dose response between the single- and double-dose cruciferous diets nor was there a significant difference in response between 1 and 2 weeks of supplementation. Overall, GSTA1/2 concentrations increased significantly by day 7 relative to the basal diet on both the single- and double-dose cruciferous diets; then, by day 11, GSTA1/2 concentrations were lower for the single-dose cruciferous diet but were still increasing for the double-dose cruciferous diet. These data are consistent with evidence of adaptation to crucifers (19). However, it is not clear why GSTA1/2 concentrations started to decrease after day 11. Perhaps there is an adaptation of hepatic enzymes, as well as gut microbial enzymes, in the presence of chronic crucifer consumption.

The strengths of this study include the controlled feeding-study design, recruitment of participants based on *GSTM1* and *GSTT1* genotypes, the 2-week duration of each study diet, blood collection at multiple time points during each feeding period, and dosing based on BW. Further, the stringent exclusion criteria minimized potential confounding due to other factors that may influence GST enzyme activity.

A limitation of the study is our reliance on serum GSTA1/2 concentrations. Because GSTA1 is mainly found in the liver, the actual change in hepatic enzyme activity in response to vegetable feeding may be greater than what can be measured using circulating GSTA1/2 concentrations. Another potential limitation is generalizability. The average intake of cruciferous vegetables in the United States is ~25 to 30 g/d (20). Although the cruciferous vegetables used in our study are commonly consumed in the United States, they are not usually consumed in the amounts fed in this study (e.g., 5-10 servings per day or ~300-1,300 g). Finally, although we had sufficient power to detect overall diet and genotype differences, we were not sufficiently powered to evaluate sex-by-genotype-by-diet interactions. We based the sample size estimate for the current study on results from our previous GST study, which included a similar study population (7), and determined that we would have 80% to 96% power with a sample size of 64. A post hoc calculation based on our present results indicates that our power was lower, ranging from 60% to 81% for overall effects. Therefore, it is possible that significant results may also be explained by chance.

In summary, cruciferous vegetable supplementation increased serum GSTA1/2 concentrations, but the effect was most marked in *GSTM1*-null/*GSTT1*-null men. In addition, the combination of apiaceous vegetables and cruciferous vegetables attenuated the effects of cruciferous vegetables alone.

## Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed.

## Acknowledgments

The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked *advertisement* in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

We thank Karen Noar, Kara Breymer, and their staff in the Human Nutrition Laboratory for their dedicated work, and JoAnn Prunty, Sherianne Fish, and Maureen Downey for their technical support.

## References

- Shapiro TA, Fahey JW, Dinkova-Kostova AT, et al. Safety, tolerance, and metabolism of broccoli sprout glucosinolates and isothiocyanates: a clinical phase I study. *Nutr Cancer* 2006;55:53-62.
- Kolm RH, Danielson H, Zhang Y, Talalay P, Mannervik B. Isothiocyanates as substrates for human glutathione transferases: structure-activity studies. *Biochem J* 1995;311:453-9.
- Ketterer B. Dietary isothiocyanates as confounding factors in the molecular epidemiology of colon cancer. *Cancer Epidemiol Biomarkers Prev* 1998;7:645-6.
- Lin HJ, Probst-Hensch NM, Louie AD, et al. Glutathione transferase null genotype, broccoli, and lower prevalence of colorectal adenomas. *Cancer Epidemiol Biomarkers Prev* 1998;7:647-52.
- Coles BF, Kadlubar FF. Human  $\alpha$  class glutathione S-transferases: genetic polymorphism, expression, and susceptibility to disease. *Methods Enzymol* 2005;401:9-42.
- Hayes JD, Flanagan JU, Jowsey IR. Glutathione transferases. *Annu Rev Pharmacol Toxicol* 2005;45:51-88.
- Lampe JW, Chen C, Li S, et al. Modulation of human glutathione S-transferases by botanically defined vegetable diets. *Cancer Epidemiol Biomarkers Prev* 2000;9:787-93.
- Navarro SL, Peterson S, Chen C, et al. Cruciferous vegetable feeding alters UGT1A1 activity: diet- and genotype-dependent changes in serum bilirubin in a controlled feeding trial. *Cancer Prev Res* 2009;2:345-52.
- Arand M, Muhlbauer R, Hengstler J, et al. A multiple polymerase chain reaction protocol for the simultaneous analysis of the glutathione S-transferase *GSTM1* and *GSTT1* polymorphisms. *Anal Biochem* 1996;236:184-6.
- Coles BF, Morel F, Rauch C, et al. Effect of polymorphism in the human glutathione S-transferase A1 promoter on hepatic GSTA1 and GSTA2 expression. *Pharmacogenetics* 2001;11:663-9.
- Chung FL, Jiao D, Getahun SM, Yu MC. A urinary biomarker for uptake of dietary isothiocyanates in humans. *Cancer Epidemiol Biomarkers Prev* 1998;7:103-8.
- Traka M, Gasper AV, Melchini A, et al. Broccoli consumption interacts with *GSTM1* to perturb oncogenic signalling pathways in the prostate. *PLoS ONE* 2008;3:e2568.
- Kushad MM, Brown AF, Kurilich AC, et al. Variation of glucosinolates in vegetable crops of *Brassica oleracea*. *J Agric Food Chem* 1999;47:1541-8.
- Vermeulen M, Van den Berg R, Freidig AP, Van Bladeren PJ, Vaes WHJ. Association between consumption of cruciferous vegetables and condiments and excretion in urine of isothiocyanate mercapturic acids. *J Agric Food Chem* 2006;54:5350-8.
- Zhang Y, Talalay P. Mechanism of differential potencies of isothiocyanates as inducers of anticarcinogenic Phase 2 enzymes. *Cancer Res* 1998;58:4632-9.
- Ye L, Zhang Y. Total intracellular accumulation levels of dietary isothiocyanates determine their activity in elevation of cellular glutathione and induction of Phase 2 detoxification enzymes. *Carcinogenesis* 2001;22:1987-92.
- Jakubikova J, Bao Y, Sedlak J. Isothiocyanates induce cell cycle arrest, apoptosis and mitochondrial potential depolarization in HL-60 and multidrug-resistant cell lines. *Anticancer Res* 2005;25:3375-86.
- Shapiro TA, Fahey JW, Wade KL, Stephenson KK, Talalay P. Human metabolism and excretion of cancer chemoprotective glucosinolates and isothiocyanates of cruciferous vegetables. *Cancer Epidemiol Biomarkers Prev* 1998;7:1091-100.
- Sreerama L, Hedge MW, Sladek NE. Identification of a class 3 aldehyde dehydrogenase in human saliva and increased levels of this enzyme, glutathione S-transferases, and DT-diaphorase in the saliva of subjects who continually ingest large quantities of coffee or broccoli. *Clin Cancer Res* 1995;1:1153-63.
- International Agency for Research on Cancer. Cruciferous vegetables, isothiocyanates and indoles. IARC handbooks of cancer prevention. Lyon (France): International Agency for Research on Cancer; 2004.