

LYMPHOID NEOPLASIA

Exposure to UV radiation and risk of Hodgkin lymphoma: a pooled analysis

Alain Monnereau,¹⁻³ Sally L. Glaser,^{3,4} Clayton W. Schupp,³ Karin Ekström Smedby,⁵ Silvia de Sanjosé,^{6,7} Eleanor Kane,⁸ Mads Melbye,⁹ Lenka Forétova,¹⁰ Marc Maynadié,¹¹ Anthony Staines,¹² Nikolaus Becker,¹³ Alexandra Nieters,¹⁴ Paul Brennan,¹⁵ Paolo Boffetta,¹⁶ Pierluigi Cocco,¹⁷ Ingrid Glimelius,⁵ Jacqueline Clavel,¹ Henrik Hjalgrim,⁹ and Ellen T. Chang^{3,4,18}

¹Environmental Epidemiology of Cancer Group, Centre for Research in Epidemiology and Population Health, Institut National de la Santé et de la Recherche Médicale U1018, Villejuif, France; ²Registry of Hematological Malignancies in Gironde, Bergonié Institute, Bordeaux, France; ³Cancer Prevention Institute of California, Fremont, CA; ⁴Department of Health Research and Policy, Stanford University School of Medicine, Stanford, CA; ⁵Unit of Clinical Epidemiology, Department of Medicine, Karolinska Institutet, Stockholm, Sweden; ⁶Unit of Infections and Cancer, Cancer Epidemiology Research Program, Catalan Institute of Oncology, Barcelona, Spain; ⁷Centro de Investigación Biomédica en Red de Epidemiología y Salud Pública, Barcelona, Spain; ⁸Epidemiology and Cancer Statistics Group, Department of Health Sciences, University of York, York, United Kingdom; ⁹Department of Epidemiology Research, Statens Serum Institut, Copenhagen, Denmark; ¹⁰Department of Cancer Epidemiology and Genetics, Masaryk Memorial Cancer Institute, Brno, Czech Republic; ¹¹Registre des Hémopathies Malignes de Côte d'Or, Université de Bourgogne and University Hospital, Dijon, France; ¹²School of Nursing and Human Sciences, Dublin City University, Dublin, Ireland; ¹³Department of Cancer Epidemiology, German Cancer Research Center, Heidelberg, Germany; ¹⁴Centre of Chronic Immunodeficiency, Molecular Epidemiology, Universitätsklinikum Freiburg, Freiburg, Germany; ¹⁵Genetics Section, International Agency for Research on Cancer, Lyon, France; ¹⁶The Tisch Cancer Institute and Institute for Translational Epidemiology, Mount Sinai School of Medicine, New York, NY; and International Prevention Research Institute, Lyon, France; ¹⁷Department of Public Health, Clinical and Molecular Medicine, University of Cagliari, Asse Didattico E, Monserrato, Italy; and ¹⁸Health Sciences Practice, Exponent, Inc., Menlo Park, CA

Key Points

- Our pooled analysis found an inverse association between several measures of UVR exposure and Hodgkin lymphoma.
- Significant UVR-related inverse associations of EBV-positive HL with a dose-response relationship support etiologic heterogeneity in HL.

Ultraviolet radiation (UVR) exposure has been inversely associated with Hodgkin lymphoma (HL) risk, but only inconsistently, only in a few studies, and without attention to HL heterogeneity. We conducted a pooled analysis of HL risk focusing on type and timing of UVR exposure and on disease subtypes by age, histology, and tumor-cell Epstein-Barr virus (EBV) status. Four case-control studies contributed 1320 HL cases and 6381 controls. We estimated lifetime, adulthood, and childhood UVR exposure and history of sunburn and sunlamp use. We used 2-stage estimation with mixed-effects models and weighted pooled effect estimates by inverse marginal variances. We observed statistically significant inverse associations with HL risk for UVR exposures during childhood and adulthood, sunburn history, and sunlamp use, but we found no significant dose-response relationships. Risks were significant only for EBV-positive HL (pooled odds ratio, 0.56; 95% confidence interval, 0.35 to 0.91 for the highest overall UVR exposure category), with a significant linear trend for overall exposure ($P = .03$). Pooled relative risk estimates were not heterogeneous across studies. Increased UVR exposure may

protect against HL, particularly EBV-positive HL. Plausible mechanisms involving UVR induction of regulatory T cells or the cellular DNA damage response suggest opportunities for new prevention targets. (*Blood*. 2013;122(20):3492-3499)

Introduction

Classical Hodgkin lymphoma (HL) is a B-cell lymphoma with unusual epidemiologic characteristics.¹ Although rare in most populations, HL is a relatively common cancer of adolescents and young adults, and treatment-related sequelae remain a serious life-long problem.²⁻⁵ Thus, understanding HL etiology is a worthwhile research endeavor with the goal of primary prevention. Numerous epidemiologic studies have identified a variety of putative risk factors as well as substantial etiologic heterogeneity based on age group at diagnosis, tumor histologic subtype, and the presence or absence of Epstein-Barr virus (EBV) in the malignant Hodgkin/Reed-Sternberg cells.⁶⁻¹⁰ However, the causes of HL remain poorly understood, in part

because its low incidence limits the study sample sizes needed for adequate statistical power, particularly for the stratified analyses necessary to accommodate HL heterogeneity. Further, while the presence of EBV in HL tumors in selected patient subsets has been an important etiologic observation, the etiologic role of this virus in EBV-negative HL, which comprises the majority of cases, remains unclear.^{7,9}

Ultraviolet radiation (UVR) exposure has been associated with HL risk, albeit inconsistently and based on only a few studies.¹¹⁻¹⁶ A large population-based case-control study in Sweden and Denmark detected an inverse association between risk of HL and sunbathing or

Submitted April 24, 2013; accepted August 29, 2013. Prepublished online as *Blood* First Edition paper, September 9, 2013; DOI 10.1182/blood-2013-04-497586.

There is an Inside *Blood* commentary on this article in this issue.

The publication costs of this article were defrayed in part by page charge payment. Therefore, and solely to indicate this fact, this article is hereby marked "advertisement" in accordance with 18 USC section 1734.

© 2013 by The American Society of Hematology

sunburn history within the last 5 to 10 years.¹¹ A recent French case-control study reported that having light-colored hair and skin or a high propensity to sunburn—indicators for sun sensitivity and potential avoidance¹⁷—were significantly associated with increased risk of HL.¹³ Similarly, a European case-control study observed a positive association of HL risk with increasing skin sensitivity to sun exposure and a nonsignificant inverse association with more recreational days during childhood and adulthood.¹⁴

No studies have been able to address the role of this common and modifiable behavior in light of the marked heterogeneity of HL. To address this question, we took advantage of an international collaboration under the auspices of the International Lymphoma Epidemiology Consortium (InterLymph) to achieve sufficient numbers of study subjects for meaningful subgroup analyses. We conducted a collaborative pooled analysis of the role of UVR exposure in HL risk focusing on timing and type of exposure and considering epidemiologically distinct disease subtypes defined by age group, histologic subtype, and EBV status of the tumor.

Patients and methods

Study population

Four studies collected data on at least 1 measure of personal UVR exposure and contributed data for this analysis (Table 1).^{11,13,14,18} All 4 studies were conducted during the early 2000s in Europe and included HL patients age 16 to 80 years at diagnosis, with no history of hematologic or other neoplasms, immunosuppression for organ transplantation, or HIV infection. The 4 studies were the Scandinavian Lymphoma Etiology (SCALE) study in Sweden and Denmark, l'Etude des Facteurs Environnementaux et Génétique des Lymphomes de l'Adulte (Engela) in France, the Epidemiology and Cancer Statistics Group Case-Control Study (ELCCS) in the United Kingdom, and the multi-European-center Epilymph study. Three studies (SCALE, ELCCS, and Epilymph [Italy and Germany]) included population-based controls only, whereas the Engela study and some Epilymph centers included hospital-based controls only (Table 1). Controls were frequency-matched to cases in SCALE and at some Epilymph centers and individually matched in the other studies. All tumors had been classified histologically according to either the 2001 World Health Organization Classification of Hematopoietic and Lymphoid Tumors¹⁹ or the International Classification of Diseases for Oncology, Third Edition (ICD-O-3).²⁰ For this analysis, we considered classical HL overall, which includes all histologic subtypes except nodular lymphocyte-predominant HL and comprises the 2 most common subtypes: nodular sclerosis (ICD-O-3 9663/3 to 9667/3) and mixed cellularity (ICD-O-3 9652/3). Three studies characterized tumor EBV status by using standard immunohistochemistry to detect latent membrane protein 1 (LMP-1) and/or Epstein-Barr nuclear antigen or by in situ hybridization for EBV-encoded RNA (EBER) (Table 1). The sources of controls and matching characteristics are described in Table 1.

All studies obtained informed consent from participants in accordance with the Declaration of Helsinki and ethics approval from their local human research ethics committees; the pooled resource project also obtained ethics approval.

Data collection and exposure definition

Each eligible study provided an electronic dataset containing core subject variables, self-reported measures of personal UVR exposure (sunbathing frequency, sunburn history, vacations/holidays to sunny locations, tanning bed/sunlamp exposure, working time outdoors/outdoor occupation, recreational time outdoors, and use of sun protection), and potential confounding or effect-modifying variables (eg, study site, current socioeconomic status, subject's birth order, day care attendance, history of infectious mononucleosis, pigmentation variables, and cigarette smoking status).

Table 1. Characteristics of participating studies

Study name (reference)	Location	Period of interview/diagnosis	Cases (n = 1320)			Controls (n = 6381)				
			Age range (y)	Tumor histology	Classification	Tumor EBV status	No.	% Part	Source	Matching
Epilymph (14)	Spain, France, Italy, Germany, Ireland, Finland, Czech Republic	1998-2004	17-80	195 NS, 71 MC, 1 other	WHO	18 positive, 27 negative (Spain only)	2540	81	Population registers (Italy and Germany) or hospitals (Spain, France, Ireland, Czech Republic)	Individual (Germany, Czech Republic) or frequency (France, Ireland, Italy, Spain) by sex, age (5-y groups), and area of residence
ELCCS	North, East, and West Yorkshire, Lancashire, South Lakeland, Cornwall, South Devon, Dorset, South Hampshire, England	1998-2004	16-67	203 NS, 47 MC, 12 other	ICD-O-3	48 positive, 116 negative	237	69	Randomly selected from population registers	Individual by sex, date of birth, and area of residence
SCALE (11)	Denmark, Sweden	1999-2003	17-75	422 NS, 106 MC, 10 other	WHO	142 positive, 383 negative	3187	71	Randomly selected from population registers	Frequency by country, age (10-y groups), and sex
Engela (13)	Bordeaux, Brest, Caen, Lille, Nantes, Toulouse, France	2000-2004	18-71	106 NS, 18 MC, 3 other	ICD-O-3	No data	417	93	Hospitals, with no history of hematologic neoplasm	Individual by center, sex, age, and area of residence

% part, participation rate; MC, mixed cellularity; NS, nodular sclerosis; WHO, World Health Organization.

Table 2. Measures of UVR exposure categories by study

Study	Time spent outdoors not in shade		Routine outdoor leisure activities hours per day, week, month	Sunbathing in summertime frequency per week or month	Sunlamp use		Age or time intervals in analyses	
	Working day	Non-working day			Ever/never	Frequency	Childhood (age ≤20 y)	Adulthood (age >20 y)
Epilymph	Yes (all but Germany)	Yes (all but Germany)	Yes (only for Germany)		Yes		Ages 10 and 20 y	Ages 30 and 40 y
ELCCS	Yes	Yes			Yes	Yes	Calculated based on age at interview	Calculated based on age at interview
SCALE				Yes	Yes	Yes	Age 20 y	Calculated based on age at interview
Engela			Yes		Yes			Exposure since leaving school

To estimate relative UVR exposure (Table 2), we constructed 5 variables for pooling across studies: overall (ie, lifetime) UVR exposure, childhood UVR exposure, adult UVR exposure, history of sunburn, and history of sunlamp use. Overall UVR exposure was computed as the sum of childhood and adult UVR exposure, except when available data allowed us to include additional lifetime exposure data (eg, the number of times abroad on a sun or beach holiday, as evaluated in SCALE). Variables describing the history of sunburn and the use of a sunlamp/solarium were dichotomized as ever vs never. Childhood UVR exposure (at or before age 20 years) was computed in SCALE, Epilymph, and ELCCS data by summing the non-missing values and creating quartiles for each variable based on available data (eg, hours spent outdoors, hours spent in sun for leisure, or days spent at the water/beach). Because sunlamp users are more likely than nonusers to seek UVR (including sun) exposure,²¹ but because sunlamp use was assessed in the contributing studies only as a binary variable, we elevated a subject's assigned quartile by 1 level if that subject also reported using a sunlamp, sunbed, or solarium at or before the age of 20 years (SCALE and ELCCS studies). Adult UVR exposure (after age 20 years) was measured from data provided by all 4 studies in a similar way, averaging measures that were assessed at multiple ages in adulthood.

Statistical analysis

We used a 2-stage estimation method with mixed-effects models, described by Stukel et al,²² to obtain study-specific odds ratios (ORs) and pooled ORs and 95% confidence intervals (CIs). Since 2 studies (SCALE and Epilymph) were large intercountry efforts that covered study areas of different latitudes, we attempted to better evaluate the impact of between-study variation by producing 2 types of models depending on the definition of study locations. "A" models treated Epilymph and SCALE as single overall studies and adjusted for study location, pooling across 4 study estimates. "B" models treated the 7 study locations of Epilymph and the 2 study locations of SCALE as separate locations, thus pooling across 11 study estimates.

The Stukel approach allows for analysis of each study as originally designed, with study-specific confounder coding. Confounder identification and adjustment procedures were undertaken for each study separately. We used the following criteria, set forth by Rothman et al,²³ to assess a confounding variable: (1) it must be a risk factor for the disease, (2) it must be associated with the exposure, and (3) it must not be on the causal pathway. The first step was to identify variables significantly associated with both outcome and exposure by significance testing with $P \leq .10$. The second step was to narrow these variables to confounders that influence the main effect estimates. We used a 10% change in the OR estimate as a criterion to identify confounders that had a large effect on the relationship between the exposure and outcome and that would result in the most parsimonious model. We used a stepwise selection approach, retaining all identified confounders regardless of P value and additional identified covariates that remained significantly associated with the outcome (but not necessarily the exposure) at $P \leq .20$ in stepwise modeling. As confounders/effect modifiers, we considered pigmentation variables; education; current or childhood socioeconomic status; number of siblings and birth order; daycare or preschool attendance; body size variables; history of infectious mononucleosis, cancer, or

autoimmune disease; and cigarette smoking status. Education for Epilymph and SCALE, a variable combining current socioeconomic status (based on last job held) and education for Engela, and skin pigmentation for Epilymph were included as confounders in final models. The Engela study and 2 centers of the Epilymph study (Germany and the Czech Republic) used individual matching of cases and controls but subsequently analyzed their data ignoring this matching. Because our preliminary analyses for matched and non-matched studies yielded similar results, we opted for non-matched analysis because it provided a larger number of subjects. Data from the ELCCS study allowed us to retain the original individual matching. Effect estimates for each study were obtained by using unconditional logistic models adjusted for age as a continuous variable, sex, and study center for the SCALE, Epilymph, and Engela studies, and conditional logistic models for the ELCCS study.

We used random effects models to quantify between-study variation in effect estimates. The pooled effect estimates were weighted by the inverse marginal variances (ie, the sum of the individual study-specific variances and the variance of the random study effect). Heterogeneity among studies was tested by using the Cochran Q test and the percentage of variation in ORs attributable to heterogeneity between studies (I^2).

We used a weighted regression approach to assess trends in the log-odds estimates with continuous values to represent quartiles. We fit a trend to the final pooled estimates and alternatively fit a trend to each study and pooled the trend estimates for 1 analysis (overall lifetime exposure). Because the results were similar, we present the former. The weights were based on the proportionate sample size of the pooled results at each ordinal level.

We produced effect estimates for the 5 measures of UVR stratified by age group at diagnosis and/or interview (age <40 years, age 40 years and older), sex, tumor histology (nodular sclerosis, mixed cellularity), and tumor-cell EBV status (positive, negative). We also conducted analyses separately by control type (hospital- or population-based). We tested for effect modification of HL risk by UVR exposure variation by latitude by stratifying analyses by study location; as an indicator of the potential for skin damage, we used the average yearly erythemal UVR index, a simple measure of the UVR level at the earth's surface.²⁴

Because the pooled ORs and 95% CIs from both A and B models were consistent for all analyses, we present results only from A models in accordance with the parsimony principle.

To further test hypotheses about etiologic heterogeneity, we performed case analyses to compare exposure-related HL risk across selected histologic subtypes and tumor-cell EBV status. In these comparisons, we adjusted for the same variables as in the case-control analysis. Statistical tests were 2-sided with an α level of .05. All analyses were performed with the open-access software R.

Results

The pooled dataset comprised study data, including UVR information, from 1320 cases and 6381 controls (Table 1). Among all

Table 3. Exposure to UVR and risk of HL by sex and age group

UVR exposure	Sex											
	Overall				Male				Female			
	Cases (n = 1320)	Controls (n = 6381)	OR	95% CI	Cases (n = 722)	Controls (n = 3518)	OR	95% CI	Cases (n = 598)	Controls (n = 2863)	OR	95% CI
									Age group at diagnosis (years)			
									<40		40+	
	Cases	Controls	OR	95% CI	Cases	Controls	OR	95% CI	Cases	Controls	OR	95% CI
Overall*	281	1323	1.00		155	682	1.00		126	641	1.00	
Low	316	1432	0.94	0.76-1.16	167	737	0.95	0.71-1.26	149	695	0.91	0.66-1.25
Low-mid	309	1397	0.87	0.70-1.08	159	795	0.81	0.61-1.09	150	602	0.94	0.68-1.30
Mid-high	312	1688	0.84	0.67-1.04	192	982	0.90	0.68-1.20	120	701	0.77	0.55-1.07
High												
<i>P</i> _{trend} †			.01				.33				.14	
<i>P</i> ‡			.58				.28				.07	
Childhood												
Low	271	1373	1.00		133	610	1.00		138	763	1.00	
Low-mid	306	1543	0.79§	0.59-0.91	157	861	0.61§	0.45-0.83	149	682	0.87	0.64-1.18
Mid-high	215	1090	0.72	0.48-1.07	109	614	0.62§	0.41-0.95	106	476	0.93	0.66-1.29
High	265	1608	0.80§	0.63-1.00	163	984	0.75	0.55-1.01	102	624	0.85	0.60-1.20
<i>P</i> _{trend}			.41				.51				.22	
<i>P</i> ‡			.17				.31				.07	
Adulthood												
Low	339	1453	1.00		191	813	1.00		148	942	1.00	
Low-mid	266	1492	0.80§	0.64-1.00	142	777	0.82	0.60-1.12	124	715	0.77	0.56-1.06
Mid-high	260	1425	0.76§	0.61-0.95	147	798	0.73§	0.55-0.97	113	627	0.82	0.58-1.15
High	289	1633	0.87	0.70-1.08	176	942	0.94	0.71-1.24	113	691	0.81	0.58-1.14
<i>P</i> _{trend}			.51				.72				.35	
<i>P</i> ‡			.43				.34				.22	
History of sunburn												
No	356	1548	1.00		210	801	1.00		146	747	1.00	
Yes	714	3681	0.77§	0.63-0.95	390	2077	0.77	0.57-1.03	324	1604	0.79	0.59-1.06
<i>P</i> ‡			.36				.26				.95	
History of sunlamp use												
No	588	3428	1.00		391	2095	1.00		197	1333	1.00	
Yes	633	2501	0.81§	0.69-0.96	285	1139	0.89	0.65-1.23	348	1362	0.81	0.63-1.04
<i>P</i> ‡			.37				.37				.45	

*Education for Epilymph and SCALE; current socioeconomic status for Engela, and skin pigmentation were included as confounders in the final models.

†*P*_{trend} in ORs.

‡*P* for heterogeneity among studies.

§*P* < .05.

Table 4. Exposure to UVR and risk of HL by histologic subtype and by tumor EBV status

UVR exposure	Histology						Tumor EBV status					
	Nodular sclerosis			Mixed cellularity			Positive			Negative		
	Cases (n = 928)	OR	95% CI	Cases (n = 242)	OR	95% CI	Cases (n = 208)	OR	95% CI	Cases (n = 526)	OR	95% CI
Overall*												
Low	182	1.00		55	1.00		56	1.00		121	1.00	
Low-mid	218	1.00	0.78-1.28	49	1.00	0.50-1.18	55	1.00	0.51-1.26	132	1.00	0.70-1.32
Mid-high	214	0.97	0.75-1.26	53	0.83	0.54-1.27	46	0.60†	0.36-0.98	135	0.93	0.67-1.28
High	202	0.88	0.68-1.14	65	0.85	0.56-1.29	51	0.56†	0.35-0.91	138	0.86	0.63-1.19
<i>P</i> _{trend} ‡		0.14			0.52			0.03			0.02	
<i>F</i> _S		0.40			0.28			0.09			0.36	
Childhood												
Low	186	1.00		42	1.00		50	1.00		109	1.00	
Low-mid	212	1.00	0.73†	50	0.91	0.63-1.31	52	0.60†	0.37-0.97	143	0.75	0.54-1.03
Mid-high	149	0.84	0.75-1.07	39	1.03	0.60-1.77	30	0.62	0.35-1.07	77	0.70	0.48-1.02
High	166	0.79	0.63-1.00	52	0.90	0.61-1.31	49	0.36	0.10-1.34	129	0.85	0.60-1.21
<i>P</i> _{trend}		0.28			0.59			0.06			0.55	
<i>F</i> _S		0.20			0.16			0.14			0.11	
Adult												
Low	231	1.00		57	1.00		58	1.00		107	1.00	
Low-mid	172	0.77	0.55-1.08	47	0.77	0.49-1.20	30	0.43†	0.26-0.73	93	0.69†	0.48-0.98
Mid-high	191	0.87	0.68-1.12	38	1.425	0.40-1.01	33	0.37†	0.21-0.65	101	0.90	0.63-1.27
High	168	0.75†	0.58-0.98	68	1.633	0.68-1.56	45	0.59†	0.36-0.96	104	0.92	0.65-1.29
<i>P</i> _{trend}		0.11			0.94			0.49			0.99	
<i>F</i> _S		0.34			0.44			0.10			0.53	
History of sunburn												
No	242	1.00		65	1.00		61	1.00		132	1.00	
Yes	488	0.67†	0.53-0.84	127	0.92	0.59-1.43	132	0.81	0.51-1.29	335	0.65†	0.48-0.89
<i>F</i> _S		0.56			0.98			0.27			0.44	
History of sunlamp use												
No	361	1.00		120	1.00		97	1.00		187	1.00	
Yes	455	0.85	0.70-1.03	103	0.80	0.58-1.10	111	0.69	0.47-1.02	338	0.88	0.68-1.13
<i>F</i> _S		0.32			0.72			0.10			0.98	

*Education for EpiLymph and SCALE, current socioeconomic status for Engela, and skin pigmentation were included as confounders in the final models.

†*P* < 0.05.

‡*P* for trend in ORs.

\$*P* for heterogeneity among studies.

This study has several major advantages. As an analysis of pooled data, it includes sufficient numbers of subjects to examine etiologic heterogeneity in risks across subtypes of HL, a relatively rare disease. Our detailed database allowed for adjustment for potential confounding factors across studies, evaluation of interactions, and a robust and complete assessment of selected HL subtypes, which were defined with established laboratory methods.^{25,26} Participation bias may have contributed to our observations if individuals with healthy lifestyles were overrepresented among the controls. However, greater participation of healthier controls might be expected to underestimate UVR exposure in the source population because UVR exposure is known to cause skin cancer, and such a bias would attenuate inverse associations with HL.

Study limitations include the loss of granularity resulting from the data harmonization required of pooled analyses. There was substantial variation across studies in the wording of questions about UVR exposure (except for dichotomized variables). Nevertheless, the information on individual UVR exposure was mostly collected as number of hours daily at a specific age, such that quantifying exposures into the broad categories of quartiles should accurately capture the gradient in UVR exposure. Furthermore, any exposure misclassification should not have been differential between cases and controls. We could neither distinguish occupational from recreational exposure nor evaluate cumulative lifetime UVR exposure or the use of UVR-protective clothing or lotion. However, the lack of clear differences in pooled risk estimates for UVR exposure before and after age 20 years suggests a potential biological effect of UVR on HL development independent of age at exposure, although our data do not rule out a cumulative effect of UVR. Given the large difference in age distributions between cases and controls, we cannot rule out residual confounding by age, although our models were adjusted for age as a continuous variable. However, age-stratified results, which are less likely to be confounded by age within strata, were similar to overall results, suggesting minimal, if any, residual confounding by age.

Given the known health risks for UVR exposure, recall bias might have led to underreporting of UVR exposure among cases. However, this possibility was potentially reduced by the studies' including many questions on other exposures and not emphasizing UVR exposure in the questionnaires, as well as by the lack of a previously known association between UVR exposure and HL risk.

Reverse causality, with an HL diagnosis leading to more time spent indoors, may also have contributed to spurious inverse associations with adulthood UVR exposure. However, the inverse associations with both childhood and adulthood UVR exposure, heterogeneity between risks of EBV-negative and EBV-positive HL (and consistent results from the case-case analysis), which would be expected to have an equivalent impact on reverse causality, and an observed dose-response relationship for EBV-positive HL make this possibility unlikely.

Given the inverse associations of UVR exposure with both HL and NHL development, several biological hypotheses can be considered for our study findings. Until recently, the strongest evidence came from the observation that the induction of vitamin D₃ synthesis in the skin by UVR²⁷ could have a protective effect against lymphoma development. This hypothetical mechanism is supported by the observation that vitamin D₃ promotes differentiation and inhibits proliferation of lymphoma cells *in vitro*,²⁸ and by the strong expression of the vitamin D receptor on HL tumor cells.²⁹ However, a pooled analysis of 10 cohort studies found no association between prediagnostic serum 25-hydroxyvitamin D levels and risk of NHL or its major histologic subtypes.³⁰

Two other plausible pathways could explain a protective effect of UVR on HL. First, the immune system could be modulated by

UVR induction of regulatory T cells,³¹ which are critical to inhibiting inflammation.^{32,33} In HL, the tumor microenvironment, which represents 99% of the tumor, comprises reactive cells that help maintain an inflammatory milieu,³⁴ and individual traits affect the tumor microenvironment in HL.³⁵ UVR appears to induce regulatory T cells through antigen presentation by UVR-damaged Langerhans cells in the lymph nodes³⁶ leading to an immunosuppressed state that may be protective, as suggested by the inverse association observed between the use of aspirin and HL risk.³⁷⁻³⁹ Second, the cellular DNA damage response activated by viral oncogenes such as those associated with EBV⁴⁰ could cause tumor suppression by effector T cells when enhanced by UVR exposure,⁴¹ which is known to induce multiple defense mechanisms to counterbalance its potential mutagenic and cytotoxic effects.⁴² The DNA damage response acts as an innate barrier to tumorigenesis,⁴³ whereas a disordered response may lead to lymphomas, including EBV-associated lymphoma.⁴⁴ Further research is clearly needed to explore these speculative mechanisms.

In conclusion, this analysis of HL risk across contemporaneous European case-control studies found UVR exposure to be inversely associated with HL risk overall. The stronger associations for EBV-positive HL, with a clear dose-response relationship, further support etiologic heterogeneity in HL. A clearer understanding of biological pathways related to UVR exposure could illuminate the pathogenesis of HL and facilitate the development of new targets to enhance the DNA damage response or regulatory T-cell activity for the purpose of reducing the occurrence of HL and other malignancies.

Acknowledgments

We thank Rita Leung and Sarah Shema from the Cancer Prevention Institute of California for their role in the data management and analysis of this study. We also thank Klaus Rostgaard from Statens Serum Institut for his assistance with the data analysis.

This work was supported in part by Grant No. R03 CA137828-01 from the National Cancer Institute, National Institutes of Health (E.T.C. and S.L.G.) and by a travel grant from the Fondation de France (A.M.). The Scandinavian Lymphoma Etiology study (Sweden) was supported by the Swedish Cancer Society (2009/659), the Stockholm County Council (20110209), and the Strategic Research Program in Epidemiology at Karolinska Institute (K.E.S.). The Epidemiology and Cancer Statistics Group Case-Control Study (United Kingdom) was supported by Leukaemia and Lymphoma Research (E.K.). The Epilymph study was partially supported by public grants (FIS PI11-01810, AGAUR, RTIC RD06/0020/0095, and CIBERESP) from the Instituto de Salud Carlos III (S.d.S.). The Epilymph study (Czech Republic study site) was supported by grants (RECAMO, CZ.1.00/2.1.00/03.0101) from The European Regional Development Fund and the State Budget of the Czech Republic (L.F.). The Engela study was supported by grants from the Association pour la Recherche contre le Cancer, the Fondation de France, and AFSSET, and by a donation from Faberge employees (J.C.).

Authorship

Contribution: E.T.C. designed the research; A.M. drafted the manuscript; C.W.S. performed the statistical analysis; A.M., K.E.S., S.d.S., E.K., M. Melbye, L.F., M. Maynadić, A.S., N.B., A.N., P. Boffetta, P.C., I.G., J.C., and H.H. contributed data; A.M., E.T.C., and S.L.G. interpreted

the results; E.T.C. and S.L.G. obtained funding for the research; and all the authors critically reviewed and revised the manuscript.

Conflict-of-interest disclosure: The authors declare no competing financial interests.

Correspondence: Alain Monnereau, Haematological Malignancies Registry of Gironde, Bergonie Institute, 229 Cours de l'Argonne, 33076 Bordeaux cedex, France; e-mail: a.monnerau@bordeaux.unicancer.fr.

Reference

- Mueller NE, Grufferman S, Chang ET. The epidemiology of Hodgkin's Lymphoma. In: Mauch PM, Armitage JO, Diehl V, Weiss LM, eds. *Hodgkin's Lymphoma*. Philadelphia, PA: Lippincott, Williams & Wilkins; 2008:7-23
- Sklar CA, Mertens AC, Mitby P, et al. Premature menopause in survivors of childhood cancer: a report from the childhood cancer survivor study. *J Natl Cancer Inst*. 2006;98(13):890-896.
- Metayer C, Lynch CF, Clarke EA, et al. Second cancers among long-term survivors of Hodgkin's disease diagnosed in childhood and adolescence. *J Clin Oncol*. 2000;18(12):2435-2443.
- Swerdlow AJ, Higgins CD, Smith P, et al. Myocardial infarction mortality risk after treatment for Hodgkin disease: a collaborative British cohort study. *J Natl Cancer Inst*. 2007;99(3):206-214.
- Hodgson DC. Late effects in the era of modern therapy for Hodgkin lymphoma. *Hematology (Am Soc Hematol Educ Program)*. 2011;2011:323-329.
- MacMahon B. Epidemiology of Hodgkin's disease. *Cancer Res*. 1966;26(6):1189-1201.
- Glaser SL, Lin RJ, Stewart SL, et al. Epstein-Barr virus-associated Hodgkin's disease: epidemiologic characteristics in international data. *Int J Cancer*. 1997;70(4):375-382.
- Cozen W, Katz J, Mack TM. Risk patterns of Hodgkin's disease in Los Angeles vary by cell type. *Cancer Epidemiol Biomarkers Prev*. 1992;1(4):261-268.
- Hjalgrim H, Smedby KE, Rostgaard K, et al. Infectious mononucleosis, childhood social environment, and risk of Hodgkin lymphoma. *Cancer Res*. 2007;67(5):2382-2388.
- Chang ET, Zheng T, Lennette ET, et al. Heterogeneity of risk factors and antibody profiles in Epstein-Barr virus genome-positive and -negative Hodgkin lymphoma. *J Infect Dis*. 2004;189(12):2271-2281.
- Smedby KE, Hjalgrim H, Melbye M, et al. Ultraviolet radiation exposure and risk of malignant lymphomas. *J Natl Cancer Inst*. 2005;97(3):199-209.
- Petridou ET, Dikaloti SK, Skalkidou A, Andrieu E, Dessypris N, Trichopoulos D; Childhood Hematology-Oncology Group. Sun exposure, birth weight, and childhood lymphomas: a case control study in Greece. *Cancer Causes Control*. 2007;18(9):1031-1037.
- Grandin L, Orsi L, Troussard X, et al. UV radiation exposure, skin type and lymphoid malignancies: results of a French case-control study. *Cancer Causes Control*. 2008;19(3):305-315.
- Boffetta P, van der Hel O, Kricker A, et al. Exposure to ultraviolet radiation and risk of malignant lymphoma and multiple myeloma—a multicentre European case-control study. *Int J Epidemiol*. 2008;37(5):1080-1094.
- Chang ET, Canchola AJ, Cockburn M, et al. Adulthood residential ultraviolet radiation, sun sensitivity, dietary vitamin D, and risk of lymphoid malignancies in the California Teachers Study. *Blood*. 2011;118(6):1591-1599.
- Wong KY, Tai BC, Chia SE, et al. Sun exposure and risk of lymphoid neoplasms in Singapore. *Cancer Causes Control*. 2012;23(7):1055-1064.
- Falk M. Differences in sun exposure habits between self-reported skin type and ultraviolet sensitivity measured by phototest. *Photodermatol Photoimmunol Photomed*. 2011;27(4):190-195.
- Willett EV, O'Connor S, Smith AG, Roman E. Does smoking or alcohol modify the risk of Epstein-Barr virus-positive or -negative Hodgkin lymphoma? *Epidemiology*. 2007;18(1):130-136.
- Jaffe ES, Harris NL, Stein H, Vardiman JW. *Pathology and Genetics of Tumours of Haematopoietic and Lymphoid Tissues*. Lyon, France: IARC Press; 2001
- Fritz A, Percy C, Jack A, et al. *International Classification of Diseases for Oncology*, 3rd ed. Geneva, Switzerland: World Health Organization. 2000
- Coups EJ, Phillips LA. A more systematic review of correlates of indoor tanning. *J Eur Acad Dermatol Venereol*. 2011;25(5):610-616.
- Stukel TA, Demidenko E, Dykes J, Karagas MR. Two-stage methods for the analysis of pooled data. *Stat Med*. 2001;20(14):2115-2130.
- Rothman K, Greenland S, Lash T. *Modern Epidemiology*. Philadelphia, PA: Lippincott, Williams & Wilkins; 2008
- World Health Organization. The Global Solar UV Index—a practical guide. WHO/SDE/OEH/02.2. <http://www.who.int/uv/publications/en/UVIGuide.pdf>. World Health Organization. 2012. Accessed August 22, 2013.
- Clarke CA, Glaser SL, Dorfman RF, Bracci PM, Eberle E, Holly EA. Expert review of non-Hodgkin's lymphomas in a population-based cancer registry: reliability of diagnosis and subtype classifications. *Cancer Epidemiol Biomarkers Prev*. 2004;13(1):138-143.
- Glaser SL, Gulley ML, Borowitz MJ, et al. Inter- and intra-observer reliability of Epstein-Barr virus detection in Hodgkin lymphoma using histochemical procedures. *Leuk Lymphoma*. 2004;45(3):489-497.
- Guyton KZ, Kensler TW, Posner GH. Vitamin D and vitamin D analogs as cancer chemopreventive agents. *Nutr Rev*. 2003;61(7):227-238.
- Hickish T, Cunningham D, Colston K, et al. The effect of 1,25-dihydroxyvitamin D3 on lymphoma cell lines and expression of vitamin D receptor in lymphoma. *Br J Cancer*. 1993;68(4):668-672.
- Renné C, Benz AH, Hansmann ML. Vitamin D3 receptor is highly expressed in Hodgkin's lymphoma. *BMC Cancer*. 2012;12:215.
- Purdue MP, Freedman DM, Gapstur SM, et al. Circulating 25-hydroxyvitamin D and risk of non-hodgkin lymphoma: Cohort Consortium Vitamin D Pooling Project of Rarer Cancers. *Am J Epidemiol*. 2010;172(1):58-69.
- Norval M, McLoone P, Lesiak A, Narbutt J. The effect of chronic ultraviolet radiation on the human immune system. *Photochem Photobiol*. 2008;84(1):19-28.
- Erdman SE, Poutahidis T. Cancer inflammation and regulatory T cells. *Int J Cancer*. 2010;127(4):768-779.
- Norval M, Halliday GM. The consequences of UV-induced immunosuppression for human health. *Photochem Photobiol*. 2011;87(5):965-977.
- Steidl C, Connors JM, Gascoyne RD. Molecular pathogenesis of Hodgkin's lymphoma: increasing evidence of the importance of the microenvironment. *J Clin Oncol*. 2011;29(14):1812-1826.
- Glimelius I, Rubin J, Rostgaard K, et al. Predictors of histology, tissue eosinophilia and mast cell infiltration in Hodgkin's lymphoma—a population-based study. *Eur J Haematol*. 2011;87(3):208-216.
- Schwarz T. 25 years of UV-induced immunosuppression mediated by T cells—from disregarded T suppressor cells to highly respected regulatory T cells. *Photochem Photobiol*. 2008;84(1):10-18.
- Chang ET, Zheng T, Weir EG, Borowitz M, Mann RB, Spiegelman D, Mueller NE. Aspirin and the risk of Hodgkin's lymphoma in a population-based case-control study. *J Natl Cancer Inst*. 2004;96(4):305-315.
- Chang ET, Cronin-Fenton DP, Friis S, Hjalgrim H, Sørensen HT, Pedersen L. Aspirin and other nonsteroidal anti-inflammatory drugs in relation to Hodgkin lymphoma risk in northern Denmark. *Cancer Epidemiol Biomarkers Prev*. 2010;19(1):59-64.
- Chang ET, Frøsvlev T, Sørensen HT, Pedersen L. A nationwide study of aspirin, other non-steroidal anti-inflammatory drugs, and Hodgkin lymphoma risk in Denmark. *Br J Cancer*. 2011;105(11):1776-1782.
- Nikitin PA, Yan CM, Forte E, et al. An ATM/Chk2-mediated DNA damage-responsive signaling pathway suppresses Epstein-Barr virus transformation of primary human B cells. *Cell Host Microbe*. 2010;8(6):510-522.
- Nikitin PA, Luftig MA. The DNA damage response in viral-induced cellular transformation. *Br J Cancer*. 2012;106(3):429-435.
- Bergink S, Jaspers NG, Vermeulen W. Regulation of UV-induced DNA damage response by ubiquitylation. *DNA Repair (Amst)*. 2007;6(9):1231-1242.
- Nikitin PA, Luftig MA. At a crossroads: human DNA tumor viruses and the host DNA damage response. *Future Virol*. 2011;6(7):813-830.
- Ciccia A, Elledge SJ. The DNA damage response: making it safe to play with knives. *Mol Cell*. 2010;40(2):179-204.