



Autoimmune Disease Research Solutions

- Comprehensive Support for Early Diagnosis and Drug Discovery
- High-quality Reagents for Nearly 50 Diseases
- Covering Immune Cell, Cytokine, and Kinase Targets

Learn More!

The Journal of Immunology

RESEARCH ARTICLE | APRIL 01 2003

IL-10 Mediates Sigma₁ Receptor-Dependent Suppression of Antitumor Immunity¹ **FREE**

Li X. Zhu; ... et. al

J Immunol (2003) 170 (7): 3585–3591.

<https://doi.org/10.4049/jimmunol.170.7.3585>

IL-10 Mediates Sigma₁ Receptor-Dependent Suppression of Antitumor Immunity¹

Li X. Zhu,[†] Sherven Sharma,^{†‡} Brian Gardner,^{†‡} Brian Escudro,[†] Kimberly Atianzar,[†] Donald P. Tashkin,^{*†} and Steven M. Dubinett^{2*†‡}

Sigma receptors are unique endoplasmic reticulum proteins that mediate signaling for a variety of drugs. We determined the effect of sigma₁ receptor agonists on immune responses in a syngeneic lung cancer model. Sigma₁ receptor agonists, including cocaine, up-regulated splenocyte IL-10 mRNA and protein production in vitro in a sigma receptor-dependent, pertussis toxin-sensitive manner. In vivo, sigma₁ receptor agonists promoted tumor growth and induced IL-10 at the tumor site. Increased tumor growth was prevented by administration of specific Abs to IL-10 or by administration of specific sigma₁ receptor antagonists. We report that sigma₁ receptor ligands, including cocaine, augment tumor growth through an IL-10 dependent mechanism. *The Journal of Immunology*, 2003, 170: 3585–3591.

Sigma receptors, widely distributed in neuronal and non-neuronal tissues, are unique drug-binding proteins (1). These receptors interact with a variety of compounds including cocaine, and dextromethorphan and benzomorphans such as pentazocine (2, 3). At least two subtypes of sigma receptors, classified as sigma₁ and sigma₂, are distinguishable by physiologic function and pharmacologic response.

Sigma ligands have potent immunoregulatory properties including the induction of IL-10 (4) and the suppression of IFN- γ and GM-CSF (5). In murine studies, treatment with sigma ligands prevented both graft-vs-host disease and delayed-type hypersensitivity granuloma formation (5). These studies indicate that sigma receptor-dependent signaling plays a role in immune-mediated responses. Cocaine, a sigma₁ receptor ligand, is also known to modulate immune function in vivo and in vitro (6, 7). Because immunocompetent animal models of tumorigenicity and tumor progression can serve as sensitive indicators of immune dysfunction, in the present studies we evaluated the capacity for sigma ligands to limit antitumor immune responses. Based on previous reports suggesting sigma receptor-dependent cytokine modulation (4, 5), we speculated that sigma receptor ligands would impact host antitumor immunity. We report for the first time that sigma₁ receptor-dependent signaling suppresses antitumor immune reactivity by up-regulating IL-10, resulting in augmentation of tumor growth.

Materials and Methods

Reagents

PRE-084 (2-(4-morpholino)ethyl-1-phenylcyclohexane-1-carboxylate) and SKF 10047 (sigma₁ receptor agonists), 4-IBP (high-affinity agonist for sigma₁ and moderate-affinity agonist for sigma₂ receptors), and BD1047 (sigma₁ receptor antagonist) were purchased from TOCRIS Cookson (Ellisville, MO). Cocaine hydrochloride (5–20 mg/ml in saline) was obtained from the National Institute of Drug Abuse. Reagents for PGE₂ assays were obtained from Cayman Chemical (San Diego, CA). Recombinant cytokines IL-10, IFN- γ , and TGF- β , as well as the corresponding Abs for these cytokine ELISAs were purchased from BD PharMingen (San Diego, CA). Recombinant TGF- β used for in vitro assays was purchased from R&D Systems (Minneapolis, MN). Staphylococcal enterotoxin B (SEB)³ and pertussis toxin (PT) were purchased from Sigma-Aldrich (St. Louis, MO). IL-10 mAb (JES-2A5) and control mAb (GL-113 5E) were provided by K. Moore (DNAX, Palo Alto, CA).

Mice

Pathogen-free male BALB/c mice (H-2^d) (8–12 wk of age) were obtained from Harlan Sprague Dawley (Indianapolis, IN) or Simonsen Laboratories (Gilroy, CA). Mice were maintained in the West Los Angeles Healthcare Center Animal Research Facility. All studies were approved by the institutional animal review committee.

Cell culture

The murine line 1 alveolar cell carcinoma (L1C2, H-2^d) cell line was used to establish the in vivo tumor model. L1C2 is a well-characterized line that is weakly immunogenic and shows progressive tumor growth in vivo (8). The cells were cultured in complete medium consisting of RPMI (Irvine Scientific, Santa Ana, CA) supplemented with 10% FCS (Gemini Bio-Products, Calabasas, CA) and antibiotics (penicillin and streptomycin; Gemini Bio-Products) and kept at 37°C in a humidified atmosphere containing 5% CO₂.

Evaluation of tumorigenicity

To determine the effect of sigma₁ receptor ligands on tumorigenicity in vivo, mice were pretreated for 2 wk with i.p. injections of PRE-084 (20 mg/kg), cocaine (5 mg/kg), or diluent control (saline) five times per week. For experiments with the sigma₁ receptor antagonist, BD1047 was given 30 min in advance of the agonist (i.e., cocaine or PRE-084) administration. Fourteen days following the initiation of PRE-084, cocaine, or diluent injections, 10⁵ L1C2 cells were implanted s.c. in the suprascapular area. To identify the immunoregulatory role of IL-10 following sigma₁ receptor agonist administration in vivo, Ab blocking studies were performed for 2 wk in BALB/c mice that were pretreated with PRE-084, cocaine, or diluent

*University of California Los Angeles Lung Cancer Research Program, Jonsson Comprehensive Cancer Center, Los Angeles, CA 90095; [†]Division of Pulmonary and Critical Care Medicine, David Geffen School of Medicine, University of California, Los Angeles, CA 90095; and [‡]Division of Pulmonary and Critical Care Medicine, Veterans Administration Greater Los Angeles Healthcare System, Los Angeles, CA 90073

Received for publication July 12, 2002. Accepted for publication January 28, 2003.

The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked *advertisement* in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

¹ This work is supported by National Institutes of Health Grants R01 DA08254, R01 CA71818, P50 90388 and Merit Review Research Funds from the Department of Veterans Affairs.

² Address correspondence and reprint requests to Dr. Steven M. Dubinett, University of California Los Angeles Lung Cancer Research Program, David Geffen School of Medicine, 37-131 Center for Health Sciences, 10833 Le Conte Avenue, Los Angeles, CA 90095-1690. E-mail address: sdubinett@mednet.ucla.edu

³ Abbreviations used in this paper: SEB, staphylococcal enterotoxin B; PT, pertussis toxin.

control as previously described and then were inoculated with 1×10^5 L1C2 cells by s.c. injection in the right suprascapular region. One day before tumor inoculation, mice received anti-IL-10 mAb (5 mg/kg) or control Ab (5 mg/kg) by i.p. injections three times a week for the duration of the experiment. Following implantation of tumor cells, mice continued to receive PRE-084, cocaine, antagonist BD1047, or diluent injections five times per week. Tumor growth was assessed as previously described (8). Two bisecting diameters of each tumor were measured with calipers and the volume was calculated using the formula $0.4 \times ab^2$, where a represents the larger diameter and b the smaller diameter.

Assessment of cytokine concentrations by ELISA

Non-necrotic tumors were isolated from the mice in the various treatment groups previously described for cytokine evaluation. IL-10, TGF- β , and IFN- γ levels were measured in the supernatant of tumor homogenates by cytokine-specific ELISA as previously described (9). To evaluate the sigma receptor-mediated production of cytokines independent of the influence of the tumor-bearing state, we assessed splenocyte cytokine production following administration of PRE-084 or cocaine in non-tumor-bearing mice. Mice received injections of the sigma receptor ligands 5 days per week for 2 wk. IL-10, TGF- β , and IFN- γ levels were measured in cultured splenocytes by cytokine-specific ELISA. Non-tumor-bearing BALB/c mice were pretreated with sigma receptor agonists and antagonists as previously described, and SEB (50 μ g/mouse) was administered via a lateral tail vein injection. Two hours following SEB injection, blood samples were obtained by retro-orbital puncture, and IL-10 in the sera quantified by ELISA. To determine the specificity of sigma $_1$ receptor agonist-induced lymphocyte IL-10 production, BALB/c splenocytes were cultured with IL-2 (100 U/ml) for 3 days and then stimulated with agonists PRE-084 (0.3 μ M), SKF 10047 (0.3 μ M), or cocaine (0.3 μ M) plus BD1047 (0.3 μ M) for 24 h. To determine whether the sigma $_1$ agonist-induced IL-10 production could be inhibited by PT, splenocytes were cultured as previously described and PT (10 ng/ml) was added 24 h before treatment with the agonists. Following an additional 24 h, IL-10 secreted in the culture supernatants was quantified by ELISA.

PGE $_2$ determinations by enzyme immunoassay

All reagents for the PGE $_2$ assay were obtained from Cayman Chemical (Ann Arbor, MI). PGE $_2$ ELISA was performed according to the manufacturer's instructions as routinely performed in our laboratory (10). The plate was read at 405 nm with Molecular Devices plate reader (Sunnyvale, CA).

Total RNA preparation, cDNA synthesis, and real-time PCR

To determine whether sigma $_1$ receptor ligands regulate IL-10 mRNA expression, total splenocyte RNA was isolated using methods previously described (11). The reverse transcriptase reaction was performed with 500 ng total RNA with a kit from Life Technologies (Carlsbad, CA). For IL-10 mRNA determination, quantitative real-time PCR was performed using Amplifluor murine IL-10 direct gene system kit (Intergen, Purchase, NY) in an iCycler (Bio-Rad, Hercules, CA). The primers for the IL-10 PCR were: Amplifluor forward primer; 5'-CAT ACT GCT AAC CGA CTC CT-3' and reverse primer; 5'-CTG GGG CAT CAC TTC TAC-3'. IL-10 cDNA provided in the kit was utilized to set up a standard curve. The PCR parameters were as follows: 94°C for 5 min (94°C for 15 s, 55°C for 40 s, 72°C for 1 min) \times 45 cycles, 72°C for 3 min (final temperature, 25°C). The β -actin housekeeping gene was amplified to correct for RNA amounts using Amplifluor murine β -actin direct gene system (Intergen). The RT-PCR amplifications were examined by 2% agarose gel to rule out secondary bands. Only single bands were visible at the expected positions after EtBr staining of gels. Data analyses were performed using SigmaPlot (SPSS, Chicago, IL).

Lymphocyte transfer

Lymphocytes were isolated from spleens of the following groups of mice: tumor bearers, PRE-084-treated mice, tumor bearers receiving PRE-084, or diluent-treated BALB/c mice. Lymphocytes were isolated by using MACS microbeads in accordance with the manufacturer's protocol (Miltenyi Biotec, Auburn, CA). A total of 5×10^7 lymphocytes per injection were transferred to normal mice by lateral tail vein injection 1 day before and 7 days following inoculation of 10^5 L1C2 tumor cells. As a control, the same amount of L1C2 tumor cells were implanted in naive BALB/c mice without lymphocyte transfer. Tumor volumes were assessed three times per week.

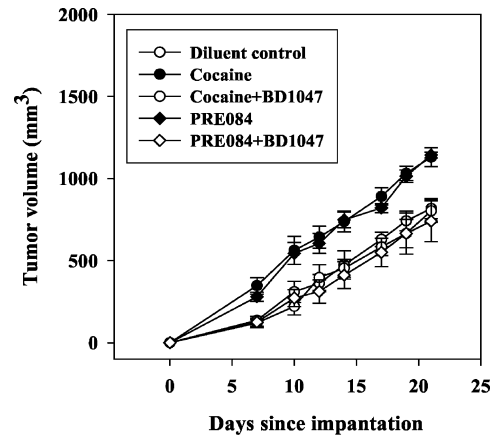


FIGURE 1. Sigma $_1$ receptor mediated promotion of tumor growth in vivo. BALB/c mice receiving i.p. injections of sigma receptor agonists cocaine (5 mg/kg), PRE-084 (20 mg/kg) or the agonists plus antagonists BD 1047 (20 mg/kg) for 2 wk were challenged with a s.c. injection of 10^5 L1C2 cells. Mice continued to receive the treatments for the duration of the experiment. There was a significant enhancement of L1C2 tumor growth in PRE-084 as well as in cocaine-treated mice ($p < 0.05$). The sigma $_1$ receptor agonist-mediated enhancement in tumor growth was ablated by treatment with BD1047 ($p < 0.05$). BD1047 treatment alone did not affect the tumor growth (data not shown) ($n = 8-12$ mice per group).

Results

Sigma $_1$ receptor agonists promote tumorigenicity in immunocompetent mice

To determine how sigma $_1$ receptor agonists modulate antitumor immune responses, we evaluated their effects on the growth of L1C2 tumors in vivo. L1C2 tumor cells were inoculated s.c. in PRE-084 or cocaine-treated BALB/c mice, and tumor growth was compared with that in diluent-treated control mice. As shown in Fig. 1, there was significant enhancement of L1C2 tumor growth in PRE-084 or cocaine-treated mice compared with control mice. These agonists acted in a sigma receptor-dependent manner as demonstrated by reversal of their tumor growth-promoting activity by BD1047, a sigma receptor antagonist. BD1047 treatment alone did not alter tumor growth (data not shown).

Sigma $_1$ receptor ligands modify tumor cytokine profiles in vivo

Based on previous studies demonstrating sigma $_1$ receptor modulation of cell-mediated immune responses (4, 5, 12, 13), we speculated that sigma $_1$ receptor ligands could be enhancing tumor growth by augmenting immune suppressive cytokine profiles in vivo. Non-necrotic tumors were isolated from L1C2 tumor-bearing BALB/c mice that had been treated with PRE-084 or cocaine as previously described and evaluated for IL-10, TGF- β , IFN- γ , and PGE $_2$ production. As shown in Table I, tumor homogenates from PRE-084- or cocaine-treated mice produced significantly more IL-10, TGF- β , and PGE $_2$ but less IFN- γ than did diluent-treated controls. Thus, sigma $_1$ ligand administration leads to a profile of increased immunosuppressive cytokines with enhanced IL-10, PGE $_2$, and TGF- β production but decreased IFN- γ release.

To determine the lymphocyte phenotype responsible for the production of IL-10 and TGF- β , we assessed the percentage of CD4 and CD8 T cells producing these cytokines at the tumor site. There was an increase in the percentage of both CD4 and CD8 T cells producing IL-10 in response to PRE-084 treatment. In contrast, there was no increase in CD4 and CD8 T cells producing TGF- β . However, the overall percentage of TGF- β -producing cells was increased in response to PRE-084. These findings suggest that

Table I. The σ_1 receptor agonist PRE084 and cocaine modulate cytokine production at the tumor site^a

	TGF- β (ng/ml/500 mg of tumor)	IL-10 (pg/ml/500 mg of tumor)	IFN- γ (pg/ml/500 mg of tumor)	PGE ₂ (ng/ml/500 mg of tumor)
Diluent control	6.1 \pm 1.6	58.0 \pm 7.3	155.3 \pm 16.8	11.9 \pm 0.7
PRE 084 (20 mg/kg)	10.4 \pm 0.7	435.0 \pm 7.3	62.7 \pm 6.8	25.7 \pm 3.8
<i>p</i> value	<0.05	<0.01	<0.01	<0.01
Diluent control	2.9 \pm 0.4	73.2 \pm 7.1	86.4 \pm 10.9	19.6 \pm 0.6
Cocaine (5 mg/kg)	4.3 \pm 0.3	128.8 \pm 19.5	41.2 \pm 2.5	33.1 \pm 4.4
<i>p</i> value	<0.05	<0.05	<0.05	<0.05

^a Non-necrotic tumors were isolated from the L1C2 tumor-bearing mice. Mice were treated with PRE 084, cocaine, or diluent control using the dosing and injection schedule as described above (*n* = 6 per group).

σ_1 receptor agonists may induce TGF- β production from tumor cells or other nonlymphocyte host-cell populations *in vivo*.

Sigma₁ receptor-dependent modulation of cytokine production in vivo in non-tumor-bearing mice

To evaluate the sigma receptor-mediated production of cytokines independent of the influence of the tumor-bearing state, we assessed cytokine production following administration of PRE-084 or cocaine in non-tumor-bearing mice. Mice receiving injections of these sigma ligands were found to have significant induction of splenocyte IL-10 production and decreased IFN- γ release (Table II). Thus, there was a sigma receptor-mediated production of cytokines independent of the influence of the tumor-bearing state. However, splenocyte TGF- β production was not significantly altered by these treatments. To assess the capacity of sigma ligands to alter the type 1 cytokine profile in response to superantigens, cytokines were measured following SEB administration in BALB/c mice. Superantigens including SEB have been found to polyclonally activate T cells *in vivo* leading to secretion of cytokines, including IL-2 and IFN- γ (14). In contrast, tolerance to superantigen-induced cytokine production is mediated by IL-10 and TGF- β (15). Sigma ligands can potently modulate SEB-induced cytokine production in mice (16). Based on these previous findings, we hypothesized that sigma receptor ligation would induce a cytokine pattern consistent with tolerance to superantigen and therefore result in high level IL-10 production. In keeping with this postulate, following SEB administration *in vivo*, the level of IL-10 was significantly increased in cocaine-treated mice (Fig. 2). A specific σ_1 receptor antagonist blocked this effect, thus documenting the σ_1 receptor dependence. Thus cocaine mediates a sigma receptor-dependent induction of IL-10 that is maintained in the setting of a potent type 1 cytokine stimulus.

Table II. *Sigma₁ receptor-dependent modulation of cytokine production in splenocytes from nontumor-bearing mice^a*

	IL-10 (pg/ml/2 \times 10 ⁶ cells)	IFN- γ (pg/ml/2 \times 10 ⁶ cells)	TGF- β (ng/ml/2 \times 10 ⁶ cells)
Diluent control	330 \pm 29	658 \pm 13	4 \pm 0
PRE 084 (20 mg/kg, daily, 2 wk)	609 \pm 50	244 \pm 51	4 \pm 0
<i>p</i> value	<0.01	<0.01	0.1
Cocaine (5 mg/kg, daily, 2 wk)	520 \pm 29	342 \pm 65	4 \pm 0
<i>p</i> value	<0.05	<0.01	0.2

^a To evaluate the sigma receptor-mediated production of cytokines independent of the influence of the tumor-bearing state, we assessed splenocyte cytokine production following administration of PRE 084 or cocaine in nontumor-bearing mice. Mice received injections of the sigma receptor ligands 5 days per week for 2 wk. IL-10, TGF- β , and IFN- γ levels were evaluated in cultured splenocytes by cytokine-specific ELISA (*n* = 6 per group).

Sigma receptor agonists induce IL-10 production in vitro

To determine whether sigma₁ receptor ligation directly up-regulates IL-10, BALB/c splenocytes were exposed to sigma₁-specific agonists *in vitro*. Consistent with the *in vivo* findings, we found that the sigma₁ receptor agonists PRE-084, SKF 10047, and cocaine induced IL-10 mRNA and protein production in splenocytes *in vitro*. In the presence of the sigma₁ receptor-specific antagonist BD1047, each of the agonists had significantly diminished IL-10 induction capacity (Fig. 3). BD1047 alone did not affect IL-10 levels (data not shown). Thus, sigma agonists significantly induce IL-10 *in vitro* and *in vivo* in a receptor-dependent manner. Consistent with G protein-coupled receptor signaling, the SKF 10047-, 4-IBP-, or cocaine-induced IL-10 production was significantly inhibited by PT (10 ng/ml). PT alone did not affect IL-10 production (Fig. 4). These findings indicate that sigma receptor agonists stimulate IL-10 production in a PT-sensitive manner.

Administration of anti-IL-10 mAb prevents the sigma₁ receptor agonist-mediated increase in tumor growth in vivo

IL-10 has been found to potently inhibit host immunity (17, 18) and may act at several points to interfere with either the generation or maintenance of antitumor immune responses. Based on the previously documented detrimental effects of IL-10, we speculated that sigma₁ receptor-mediated induction of IL-10 could be responsible for enhanced tumor growth *in vivo*. To determine the contribution of the heightened IL-10 production to the increased rate

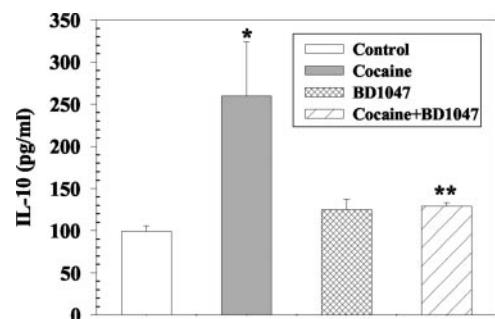
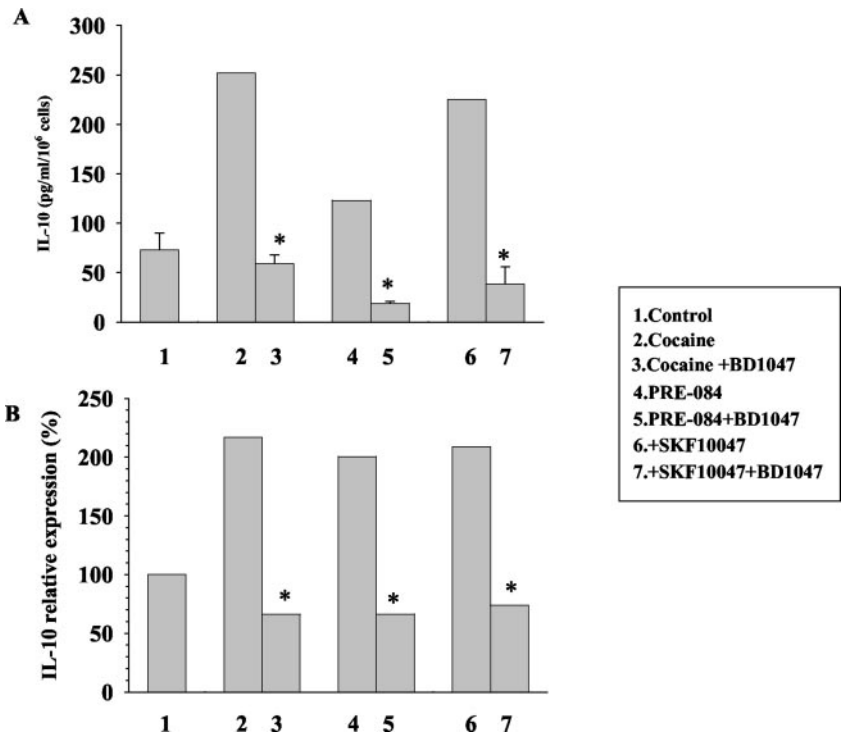


FIGURE 2. Sigma₁ receptor-dependent induction of IL-10 *in vivo* in response to SEB. BALB/c mice were pretreated five times per week for 2 wk with i.p. injections of cocaine (5 mg/kg), BD1047 (20 mg/kg), cocaine plus BD1047, or diluent control. The sigma₁ receptor antagonist, BD1047, was given 30 min in advance of agonist administration. SEB (50 μ g/mouse) was administered via a lateral tail vein injection. Two hours following SEB injection, blood samples were obtained by retro-orbital puncture and IL-10 in the sera was quantified by ELISA. BD1047 significantly inhibited the capacity of cocaine to induce IL-10 (*n* = 6 mice per group). *, *p* < 0.05 compared with diluent control; **, *p* < 0.05 compared with cocaine.

FIGURE 3. Sigma₁ receptor agonists induce splenocyte IL-10 mRNA and protein in vitro. *A*, BALB/c splenocytes were cultured with IL-2 (100 U/ml) for 3 days and then stimulated with sigma₁ agonists PRE-084 (0.3 μM), SKF 10047 (0.3 μM), or cocaine (0.3 μM) in the presence or absence of the sigma₁ receptor antagonist, BD1047 (0.3 μM), for 24 h. Sigma₁ receptor agonists induced splenic IL-10 production that could be completely blocked in the presence of the sigma₁ receptor antagonist BD1047 ($p < 0.05$). BD1047 treatment alone did not affect IL-10 production (data not shown). *B*, Total RNA was isolated from splenocytes following exposure to agonists and antagonists as in *A*. IL-10 mRNA expression was determined by quantitative real-time PCR. Sigma₁ receptor agonists, cocaine, PRE-084, and SKF 10047 potently induced splenic IL-10 mRNA that was blocked by the sigma₁ receptor antagonist BD1047 ($p < 0.05$).



of tumor growth in PRE-084- or cocaine-treated mice, anti-IL-10 mAb was administered to mice receiving PRE-084 or cocaine. Anti-IL-10 mAb, but not control Ab, prevented the sigma₁ receptor agonist-induced increase in tumor growth (Fig. 5, *A* and *B*).

Transfer of lymphocytes from sigma₁ receptor agonist-treated mice augments tumorigenicity in normal mice.

Because it appeared that sigma₁ receptor agonist mediated its immunosuppressive effects by up-regulating production of IL-10 from immune cells, we assessed the capacity of lymphocytes from sigma₁ receptor agonist-treated mice to transfer the immune deficit to normal control mice. Fifty million splenic lymphocytes from PRE-084-treated mice were transferred to normal mice by i.v. tail vein injection 1 day before and 7 days following s.c. inoculation of 10⁵ L1C2 tumor cells. Following transfer of lymphocytes from PRE-084-treated mice to normal controls, the L1C2 tumor growth was augmented revealing a similar pattern to that demonstrated for transfer of lymphocytes from tumor-bearing mice. Tumor growth in mice receiving lymphocytes from PRE-084-treated tumor-bearing mice showed an even greater enhancement in tumor growth (Fig. 6). In contrast, transfer of lymphocytes from diluent-treated control mice to tumor-bearing control mice did not alter tumor growth.

Discussion

Sigma receptors have been identified as important high-affinity binding sites for a variety of drugs (1). Although initially identified in neuronal tissue, these receptors have also been found at other sites including cells of the immune system (19). The fact that sigma receptor expression has been noted in a variety of tissue sites has led to the suggestion that they may serve a broader biological role in mediating signaling well beyond their original description as neurotransmitter receptors (1). Previous studies indicate that sigma ligands can profoundly affect immune function (4, 5, 12, 20–23). In the present study we find that sigma ligands, including cocaine, can promote tumor growth by inducing the production of IL-10 in an immunocompetent murine model of lung cancer. Specific sigma₁ receptor antagonists abrogated the tumor growth promoting activities of cocaine, as well as its capacity to induce IL-10 in vitro.

Cocaine was used as a model sigma receptor agonist in our studies because, at clinically relevant concentrations, it has been previously documented to interact with sigma₁ receptors (24). Furthermore, specific sigma₁ receptor antagonists or antisense oligonucleotides have been found to block cocaine-induced behaviors (25–28). Relevant to our current investigations of sigma receptor-mediated modulation of

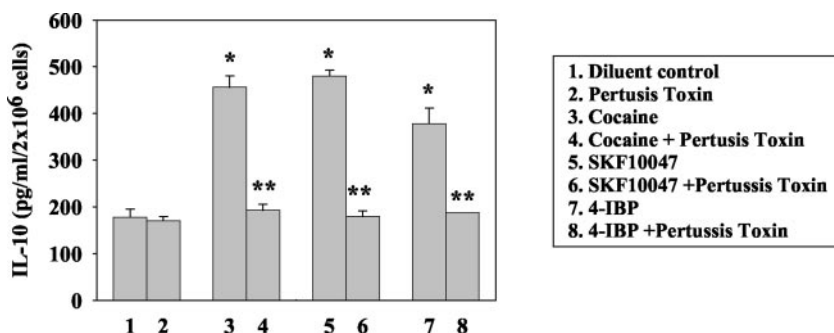


FIGURE 4. Sigma₁ receptor agonists induce IL-10 production in a PT-sensitive manner. BALB/c splenocytes were stimulated with IL-2 and treated with sigma₁ receptor agonists (0.3 μM) the presence or absence of PT (10 ng/ml, added 24 h before agonist treatment). PT consistently abrogated sigma₁ receptor agonist-mediated induction of IL-10 production. *, $p < 0.05$ compared with diluent control; **, $p < 0.05$ compared with cocaine, SKF 10047, or 4-IBP alone.

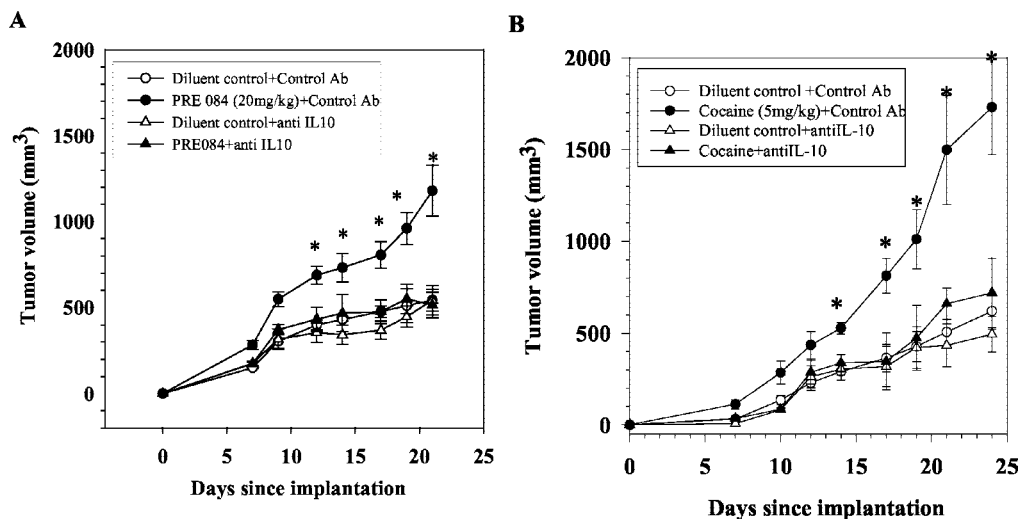


FIGURE 5. Administration of anti-IL-10 mAb prevents the σ_1 receptor agonist or cocaine-mediated increase in tumor growth in vivo. Following treatment with σ_1 receptor agonists, BALB/c mice were injected with 10^5 L1C2 cells s.c. Anti-IL-10 Ab, JES-2A5, (5 mg/kg) or control Ab were administered i.p. 1 day before tumor inoculation and then three times per week for the duration of the experiment. Mice continued to receive the σ_1 receptor agonists for the duration of the experiment. Anti-IL-10 mAb but not control Ab, prevented the PRE-084 or cocaine-induced increase in tumor growth ($p < 0.05$). The PRE-084 or cocaine-mediated increased tumor growth was not altered by control Ab treatment (data not shown) ($n = 8-12$ mice per group).

immune responses, cocaine has been found to interfere with host immunity (6, 29, 30). Cocaine has been shown to modulate cytokine production (31–33), alter the activity of CTLs and NK cells, and limit the B cell response to LPS (29). However, relatively little is known about the effects of cocaine on integrated immune responses such as the development of Ag-specific immunity or protection of the host against the challenge posed by infectious pathogens or tumor cells. In the current study, therefore, we sought to define the role of σ_1 receptor ligation in mediating immunosuppression in the context of tumor growth in vivo.

σ_1 receptor ligation led to augmentation of IL-10 in a PT-sensitive manner suggesting that this network involves G-coupled protein signaling. However, based on reports in the literature, the precise relationship between σ_1 receptors and PT-sensitive G protein coupled signaling is at present unclear. Because almost all G protein-associated receptors contain seven transmembrane regions, it has been suggested that σ_1 receptors, which have only one transmembrane domain, may not be directly linked to G proteins (34). σ_1 receptors may, however, be associated with G proteins via indirect mechanisms (34). σ_1 receptor protein has been localized at both the endoplasmic reticulum as well as the nuclear envelope (35, 36), and the complex events leading to its documented activities will require further investigation.

The possibility that immune independent events play a role in σ_1 ligand regulation of tumor growth has been suggested by previous studies. Vilner et al. (37) have shown that both σ_1 receptor subtypes are highly expressed in tumor cell lines from various tissues, and Crawford and Bowen (38) reported that σ_2 receptor agonists could activate apoptosis in breast tumor cells. In our present studies, σ_1 receptor agonists did not affect murine L1C2 tumor cell proliferation or apoptosis in vitro at concentrations of 0.03–3 μ M (data not shown). The disparity between our results and those of Crawford and Bowen (38) may be due to the difference in the cell lines evaluated or the distinct difference in the activities mediated by σ_1 vs σ_2 receptor ligation. Further studies will be necessary to define the importance of σ_1 receptor expression by tumor cells.

Previous studies suggest that populations of T cells in the tumor-bearing host may develop suppressor activities through the induc-

tion of IL-10 gene expression (17, 39). In accord with these studies documenting the importance of lymphocyte-derived IL-10 in the generation of tumor-induced tolerance, we speculated that σ_1 receptor agonists might act to enhance tumor growth by promoting the induction of lymphocyte IL-10 in vivo. The fact that transfer of lymphocytes from PRE-084-treated mice had the capacity to transfer the immune deficit that led to enhanced tumor growth in normal control mice strongly supports this hypothesis.

σ_1 receptor agonists mediated an increase in tumorigenicity via up-regulation of IL-10. The ability of IL-10 to promote tumor growth is consistent with the known activities of this cytokine.

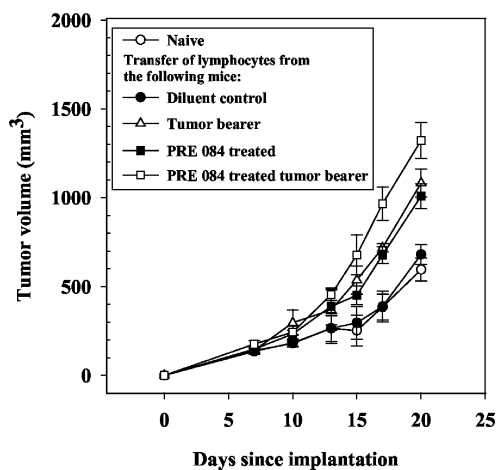


FIGURE 6. Transfer of lymphocytes from σ_1 receptor agonist-treated mice augments L1C2 tumorigenicity in normal mice. BALB/c mice were pretreated for 2 wk with i.p. injections of PRE-084 (20 mg/kg) or diluent control. Lymphocytes were isolated from spleens of the following groups of mice: tumor bearers, PRE-084-treated mice, tumor bearers receiving PRE-084 or diluent-treated BALB/c mice. A total of 5×10^7 lymphocytes/injection were transferred to normal mice by a lateral tail vein injection 1 day before and 7 days following inoculation of 10^5 L1C2 tumor cells. As a control, naive BALB/c mice were implanted with the same amount of L1C2 tumor cells without lymphocyte transfer. Tumor volumes were assessed three times per week. ($n = 6$ mice per group).

IL-10 inhibits a broad array of immune parameters that include Ag presentation, Ag-specific T cell proliferation (40, 41), and type 1 cytokine production (42, 43). Pretreatment of tumor targets with IL-10 renders the tumor cells more resistant to CTL-mediated lysis (44, 45). We have previously found that production of IL-10 by cutaneous carcinomas provides a mechanism for evasion of the local T cell immune response (46). We also found that transgenic mice overexpressing IL-10 under the control of the IL-2 promoter were unable to limit the growth of immunogenic tumors (47). Administration of blocking IL-10 mAbs restored in vivo antitumor responses in these transgenic mice. These findings suggest that lymphocyte-derived and/or tumor-derived IL-10 production antagonizes antitumor immunity (17, 18, 48). The capacity for effector cells to generate cytokines is a critical element in the generation of effective immunity. Although the tumor-bearing state is accompanied by up-regulation of immune suppressive cytokines (49, 50), we speculated that sigma₁ receptor agonists could further augment the production of these deleterious cytokines. The host-immune response against tumors has been documented to be down-regulated by soluble mediators in the tumor environment (51). Tumors may either directly release factors or orchestrate immune suppressive networks by inducing host-immune cell production of inhibitory cytokines. IL-10 is an important immune inhibitory cytokine produced or induced by tumors causing limitations in immune reactivity against the tumor (51–53). In addition to IL-10, TGF- β , PGE₂, and IFN- γ are important mediators of immune responses that alter tumor growth. Elevated TGF- β leads to enhanced tumor growth by antagonizing CTL generation (54) and macrophage activities (55, 56). Increased PGE₂ can enhance tumor progression by limiting apoptosis, while increasing angiogenesis and invasiveness (57). IFN- γ is a type 1 cytokine that promotes cell-mediated immunity and may therefore limit tumor growth. We found that sigma₁ receptor agonists increased the release of IL-10, TGF- β , and PGE₂, while decreasing IFN- γ at the tumor site, suggesting these agonists exaggerated tumor-induced immune suppression by regulating cytokine production. However, it appears that the increased TGF- β observed at the tumor site may be due to increased production by the tumor or tumor-induced cytokine production in host cells, because in the absence of tumor, sigma₁ receptor agonists did not increase the production of this cytokine in vivo. The fact that abrogation of IL-10 reversed the detrimental effects of sigma₁ receptor ligands strongly suggests that elevated IL-10 plays a significant role in mediating the sigma₁ receptor agonist-induced suppression of antitumor immunity. This is the first report of sigma₁ receptor-dependent alteration of antitumor immunity.

These findings may have important clinical implications for both medically prescribed drugs as well as those used in abuse. For example cocaine smokers have increased histopathologic abnormalities such as hyperplasia and squamous cell metaplasia compared with nonsmokers (58). In addition, cocaine smokers have elevated expression of molecular markers that have been associated with increased cancer risk such as Ki-67 and epidermal growth factor receptor (59). Thus cocaine smoking, like tobacco, has been implicated in field cancerization of bronchial epithelial cells (59). This leads to the speculation that the combination of these genetic alterations and sigma ligand-induced immune suppression may promote tumorigenesis. Further studies will be required to determine whether smoking cocaine causes cancer (60). Additional studies will also be necessary in tumor models and human cancer to define precisely both the details of the signaling events and the importance of these observations.

References

- Bowen, W. D. 2000. Sigma receptors: recent advances and new clinical potentials. *Pharm. Acta Helv.* 74:211.
- Walker, J. M., W. D. Bowen, F. O. Walker, R. R. Matsumoto, B. De Costa, and K. C. Rice. 1990. Sigma receptors: biology and function. *Pharmacol. Rev.* 42:355.
- Itzhak, Y., ed. 1994. *The Sigma Receptors: Neuroscience Perspective Series*, 1st Ed. Academic Press Inc., Orlando.
- Bourrie, B., M. Bouaboula, J. M. Benoit, J. M. Derocq, M. Esclangon, G. Le Fur, and P. Casellas. 1995. Enhancement of endotoxin-induced interleukin-10 production by SR 31747A, a sigma ligand. *Eur. J. Immunol.* 25:2882.
- Carayon, P., M. Bouaboula, J. F. Loubet, B. Bourrie, G. Petitpretre, G. Le Fur, and P. Casellas. 1995. The sigma ligand SR 31747 prevents the development of acute graft-versus-host disease in mice by blocking IFN- γ and GM-CSF mRNA expression. *Int. J. Immunopharmacol.* 17:753.
- Xu, W., T. Flick, J. Mitchel, C. Knowles, and K. Ault. 1999. Cocaine effects on immunocompetent cells: an observation of in vitro cocaine exposure. *Int. J. Immunopharmacol.* 21:463.
- Pellegrino, T., and B. M. Bayer. 1998. In vivo effects of cocaine on immune cell function. *J. Neuroimmunol.* 83:139.
- Miller, P. W., S. Sharma, M. Stolina, L. H. Butterfield, J. Luo, Y. Lin, M. Dohadwala, R. K. Batra, L. Wu, J. S. Economou, and S. M. Dubinett. 2000. Intratumoral administration of adenoviral interleukin 7 gene-modified dendritic cells augments specific antitumor immunity and achieves tumor eradication. *Hum. Gene Ther.* 11:53.
- Sharma, S., P. Miller, M. Stolina, L. Zhu, M. Huang, R. Paul, and S. Dubinett. 1997. Multi-component gene therapy vaccines for lung cancer: effective eradication of established murine tumors in vivo with interleukin 7/herpes simplex thymidine kinase-transduced autologous tumor and ex vivo-activated dendritic cells. *Gene Ther.* 4:1361.
- Stolina, M., S. Sharma, Y. Lin, M. Dohadwala, B. Gardner, J. Luo, L. Zhu, M. Kronenberg, P. W. Miller, J. Portanova, J. C. Lee, and S. M. Dubinett. 2000. Specific inhibition of cyclooxygenase 2 restores antitumor reactivity by altering the balance of IL-10 and IL-12 synthesis. *J. Immunol.* 164:361.
- Gardner, B., L. X. Zu, S. Sharma, Q. Liu, A. Makriyannis, D. P. Tashkin, and S. M. Dubinett. 2002. Autocrine and paracrine regulation of lymphocyte CB2 receptor expression by TGF- β . *Biochem. Biophys. Acta* 290:91.
- Liu, Y., B. B. Whitlock, J. A. Pultz, and S. A. Wolfe, Jr. 1995. Sigma-1 receptors modulate functional activity of rat splenocytes. *J. Neuroimmunol.* 59:143.
- Wang, Y., D. S. Huang, and R. R. Watson. 1994. In vivo and in vitro cocaine modulation on production of cytokines in C57BL/6 mice. *Life Sci.* 54:401.
- Proft, T., and J. Fraser. 1998. Superantigens: just like peptides only different. *J. Exp. Med.* 187:819.
- Miller, C., J. A. Ragheb, and R. H. Schwartz. 1999. Anergy and cytokine-mediated suppression as distinct superantigen-induced tolerance mechanisms in vivo. *J. Exp. Med.* 190:53.
- Bourrie, B., J. M. Benoit, J. M. Derocq, M. Esclangon, C. Thomas, G. Le Fur, and P. Casellas. 1996. A sigma ligand, SR 31747A, potentially modulates Staphylococcal enterotoxin B-induced cytokine production in mice. *Immunology* 88:389.
- Rohrer, J. W., and J. H. Coggin, Jr. 1995. CD8 T cell clones inhibit antitumor T cell function by secreting IL-10. *J. Immunol.* 155:5719.
- Sharma, S., M. Stolina, Y. Lin, B. Gardner, P. W. Miller, M. Kronenberg, and S. M. Dubinett. 1999. T cell-derived IL-10 promotes lung cancer growth by suppressing both T cell and APC function. *J. Immunol.* 163:5020.
- Ganapathy, M. E., P. D. Prasad, W. Huang, P. Seth, F. H. Leibach, and V. Ganapathy. 1999. Molecular and ligand-binding characterization of the sigma-receptor in the Jurkat human T lymphocyte cell line. *J. Pharmacol. Exp. Ther.* 289:251.
- Carr, D. J., S. Mayo, T. W. Woolley, and B. R. DeCosta. 1992. Immunoregulatory properties of ⁺-pentazocine and sigma ligands. *Immunology* 77:527.
- Casellas, P., B. Bourrie, X. Canat, P. Carayon, I. Buisson, R. Paul, J. Breliere, and G. Le Fur. 1994. Immunopharmacological profile of SR 31747: in vitro and in vivo studies on humoral and cellular responses. *J. Neuroimmunol.* 52:193.
- Garza, H. H., Jr., S. Mayo, W. D. Bowen, B. R. DeCosta, and D. J. Carr. 1993. Characterization of a ⁺-azidophenazocine-sensitive sigma receptor on splenic lymphocytes. *J. Immunol.* 151:4672.
- Su, T. P., E. D. London, and J. H. Jaffe. 1988. Steroid binding at sigma receptors suggests a link between endocrine, nervous, and immune systems. *Science* 240:219.
- Sharkey, J., K. A. Glen, S. Wolfe, and M. J. Kuhar. 1988. Cocaine binding at sigma receptors. *Eur. J. Pharmacol.* 149:171.
- Matsumoto, R. R., K. A. McCracken, M. J. Friedman, B. Pouw, B. R. De Costa, and W. D. Bowen. 2001. Conformationally restricted analogs of BD1008 and an antisense oligodeoxynucleotide targeting sigma₁ receptors produce anti-cocaine effects in mice. *Eur. J. Pharmacol.* 419:163.
- Ujike, H., S. Kuroda, and S. Otsuki. 1996. Sigma receptor antagonists block the development of sensitization to cocaine. *Eur. J. Pharmacol.* 296:123.
- Romieu, P., R. Martin-Fardon, and T. Maurice. 2000. Involvement of the sigma₁ receptor in the cocaine-induced conditioned place preference. *NeuroReport* 11:2885.
- McCracken, K. A., W. D. Bowen, and R. R. Matsumoto. 1999. Novel sigma receptor ligands attenuate the locomotor stimulatory effects of cocaine. *Eur. J. Pharmacol.* 365:35.
- Watzl, B., and R. R. Watson. 1990. Immunomodulation by cocaine: a neuroendocrine mediated response. *Life Sci.* 46:1319.

30. Colombo, L., M. Lopez, G. Chen, and R. Watson. 1999. Effect of short-term cocaine administration on the immune system of young and old C57BL/6 female mice. *Immunopharmacol. Immunotoxicol.* 21:755.
31. Baldwin, G. C., D. M. Buckley, M. D. Roth, E. C. Kleerup, and D. P. Tashkin. 1997. Acute activation of circulating polymorphonuclear neutrophils following in vivo administration of cocaine: a potential etiology for pulmonary injury. *Chest* 111:698.
32. Mao, J. T., M. Huang, J. Wang, S. Sharma, D. P. Tashkin, and S. M. Dubinett. 1996. Cocaine down-regulates IL-2-induced peripheral blood lymphocyte IL-8 and IFN- γ production. *Cell. Immunol.* 172:217.
33. Mao, J., L. Zhu, S. Sharma, K. Chen, M. Huang, S. Santiago, J. Gulsurd, D. Tashkin, and S. Dubinett. 1997. Cocaine inhibits human endothelial cell IL-8 production: the role of transforming growth factor- β . *Cell. Immunol.* 181:38.
34. Hayashi, T., T. Maurice, and T. P. Su. 2000. Ca²⁺ signaling via sigma₁-receptors: novel regulatory mechanism affecting intracellular Ca²⁺ concentration. *J. Pharmacol. Exp. Ther.* 293:788.
35. Dussosoy, D., P. Carayon, S. Belugou, D. Feraut, A. Bord, C. Goubet, C. Roque, H. Vidal, T. Combes, G. Loison, and P. Casellas. 1999. Colocalization of sterol isomerase and sigma₁ receptor at endoplasmic reticulum and nuclear envelope level. *Eur. J. Biochem.* 263:377.
36. Su, T. P., and T. Hayashi. 2001. Cocaine affects the dynamics of cytoskeletal proteins via sigma₁ receptors. *Trends Pharmacol. Sci.* 22:456.
37. Vilner, B. J., C. S. John, and W. D. Bowen. 1995. Sigma-1 and sigma-2 receptors are expressed in a wide variety of human and rodent tumor cell lines. *Cancer Res.* 55:408.
38. Crawford, K. W., and W. D. Bowen. 2002. Sigma-2 receptor agonists activate a novel apoptotic pathway and potentiate antineoplastic drugs in breast tumor cell lines. *Cancer Res.* 62:313.
39. Huang, M., S. Sharma, J. T. Mao, and S. M. Dubinett. 1996. Non-small cell lung cancer-derived soluble mediators and prostaglandin E₂ enhance peripheral blood lymphocyte IL-10 transcription and protein production. *J. Immunol.* 157:5512.
40. de Waal-Malefyt, R., J. Haanen, H. Spits, M. G. Roncarolo, A. Te Velde, C. Fidgor, K. Johnson, R. Kastelein, H. Yssel, and J. E. De Vries. 1991. Interleukin-10 (IL-10) and viral IL-10 strongly reduce antigen-specific human T-cell proliferation by diminishing the antigen-presenting capacity of monocytes via down-regulation of class-II major histocompatibility complex expression. *J. Exp. Med.* 174:915.
41. Taga, K., and G. Tosato. 1992. IL-10 inhibits human T cell proliferation and IL-2 production. *J. Immunol.* 148:1143.
42. Fiorentino, D. F., A. Zlotnik, T. R. Mosmann, M. Howard, and A. Ogarra. 1991. IL-10 inhibits cytokine production by activated macrophages. *J. Immunol.* 147:3815.
43. Mosmann, T., and K. W. Moore. 1991. The role of IL-10 in cross-regulation of TH1 and TH2 responses. *Immunol. Today* 12:A49.
44. Matsuda, M., F. Salazar, M. Petersson, G. Masucci, J. Hansson, P. Pisa, Q. Zhang, M. G. Masucci, and R. Kiessling. 1994. Interleukin 10 pretreatment protects target cells from tumor- and allo-specific cytotoxic T cells and down-regulates HLA class I expression. *J. Exp. Med.* 180:2371.
45. Salazar-Onfray, F., M. Petersson, L. Franksson, M. Matsuda, T. Blankenstein, K. Kärre, and R. Kiessling. 1995. IL-10 converts mouse lymphoma cells to a CTL-resistant, NK-sensitive phenotype with low but peptide-inducible MHC class I expression. *J. Immunol.* 154:6291.
46. Kim, J., R. L. Modlin, R. L. Moy, S. M. Dubinett, T. McHugh, B. J. Nickoloff, and K. Uyemura. 1995. IL-10 production in cutaneous basal and squamous cell carcinomas: a mechanism for evading the local T cell immune response. *J. Immunol.* 155:2240.
47. Hagenbaugh, A., S. Sharma, S. Dubinett, S. H.-Y. Wei, R. Aranda, H. Cheroutre, D. Fowell, S. Binder, B. Tsao, R. Locksley, K. Moore, and M. Kronenberg. 1997. Altered immune responses in IL-10 transgenic mice. *J. Exp. Med.* 185:2101.
48. Halak, B. K., H. C. Maguire, Jr., and E. C. Lattime. 1999. Tumor-induced interleukin-10 inhibits type 1 immune responses directed at a tumor antigen as well as a non-tumor antigen present at the tumor site. *Cancer Res.* 59:911.
49. Alleva, D. G., C. J. Burger, and K. D. Elgert. 1994. Tumor-induced regulation of suppressor macrophage nitric oxide and TNF- α production: role of tumor-derived IL-10, TGF- β and prostaglandin E₂. *J. Immunol.* 153:1674.
50. Maeda, H., and A. Shiraishi. 1996. TGF- β contributes to the shift toward Th2-type responses through direct and IL-10 mediated pathways in tumor-bearing mice. *J. Immunol.* 156:73.
51. Huang, M., M. Stolina, S. Sharma, J. Mao, L. Zhu, P. Miller, J. Wollman, H. Herschman, and S. Dubinett. 1998. Non-small cell lung cancer cyclooxygenase-2-dependent regulation of cytokine balance in lymphocytes and macrophages: up-regulation of interleukin 10 and down-regulation of interleukin 12 production. *Cancer Res.* 58:1208.
52. Seo, N., Y. Tokura, M. Takigawa, and K. Egawa. 1999. Depletion of IL-10- and TGF- β -producing regulatory $\gamma\delta$ T cells by administering a daunomycin-conjugated specific monoclonal antibody in early tumor lesions augments the activity of CTLs and NK cells. *J. Immunol.* 163:242.
53. de Visser, K. E., and W. M. Kast. 1999. Effects of TGF- β on the immune system: implications for cancer immunotherapy. *Leukemia* 13:1188.
54. Pardoux, C., C. Asselin-Paturel, J. Chehimi, F. Gay, F. Mami-Chouaib, and S. Chouaib. 1997. Functional interaction between TGF- β and IL-12 in human primary allogeneic cytotoxicity and proliferative response. *J. Immunol.* 158:136.
55. Fargeas, C., C. Y. Wu, T. Nakajima, D. Cox, T. Nutman, and G. Delespesse. 1992. Differential effect of transforming growth factor β on the synthesis of Th1- and Th2-like lymphokines by human T lymphocytes. *Eur. J. Immunol.* 22:2173.
56. Bogdan, C., and C. Nathan. 1993. Modulation of macrophage function by transforming growth factor β , interleukin-4, and interleukin-10. *Ann. NY Acad. Sci.* 685:713.
57. Dohadwala, M., R. K. Batra, J. Luo, Y. Lin, K. Krysan, M. Pold, S. Sharma, and S. M. Dubinett. 2002. Autocrine/paracrine prostaglandin E₂ production by non-small cell lung cancer cells regulates matrix metalloproteinase-2 and CD44 in cyclooxygenase-2-dependent invasion. *J. Biol. Chem.* 277:50828.
58. Fligiel, S. E., M. D. Roth, E. C. Kleerup, S. H. Barsky, M. S. Simmons, and D. P. Tashkin. 1997. Tracheobronchial histopathology in habitual smokers of cocaine, marijuana, and/or tobacco. *Chest* 112:319.
59. Barsky, S., M. Roth, E. Kleerup, M. Simmons, and D. Tashkin. 1998. Histopathologic and molecular alterations in bronchial epithelium in habitual smokers of marijuana, cocaine, and/or tobacco. *J. Natl. Cancer Inst.* 90:1198.
60. Mao, L., and Y. Oh. 1998. Does marijuana or crack cocaine cause cancer? *J. Natl. Cancer Inst.* 90:1182.