

Impact of Body Mass Index on the Risk of Colorectal Adenoma in a Metabolically Healthy Population

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Abstract

Metabolically healthy obese (MHO) states exist that seem to be protected from cardiovascular risks. Although obesity is a risk factor for colorectal adenoma (CRA), there has yet to be any study of the risks of CRA in MHO individuals. In this study, we compared CRA prevalence in MHO individuals versus metabolically healthy individuals who were normal in weight. This cross-sectional study involved 18,085 Korean adults (39.1 ± 6.7 years) who had a health checkup including a colonoscopy. High-risk CRA was defined as any adenoma over 1 cm, 3 or more adenomas, adenoma with a villous component, or high-grade dysplasia. Multinomial logistic regression models were used to measure the associations between body mass index (BMI) and the risk of low-risk and high-risk CRA. Low-risk and high-risk CRA were present in 9.3% and 1.4% of the study population, respectively. After adjusting for age, sex, smoking, drinking, exercise, family history of colorectal cancer, education, and use of analgesic and aspirin, compared with normal healthy individuals, the prevalence of low-risk and high-risk CRA was increased in MHO individuals [OR = 1.44; 95% confidence interval (CI), 1.23–1.69 and OR = 1.62; 95% CI, 1.09–2.41, respectively]. In fully adjusted models, the prevalence of low-risk and high-risk CRA was associated with increasing categories of BMI in a dose–response manner (*P* for trend < 0.001 and 0.01, respectively). Thus, excess body weight, even in the absence of a metabolic unhealthy state, was found to be positively associated with increased presence of CRAs. *Cancer Res*; 73(13); 4020–7. ©2013 AACR.

Introduction

Excess body weight, expressed as body mass index (BMI), is an important risk factor for colorectal cancer (CRC) and adenoma (CRA) in a sex-specific manner (1, 2), with higher BMIs associated with higher risks. Although the mechanisms underlying these associations are not fully understood, insulin resistance and related metabolic disturbances are considered the most plausible explanations (3). However, the prevalence of obesity-related metabolic disturbances varies widely among obese individuals (4, 5). For example, increased BMI is commonly, but not always, accompanied by insulin resistance and related disturbances. Based on previous findings, it is not clear whether obesity *per se* or the presence of co-existing metabolic

risk factors (6), such as diabetes, metabolic syndrome, and insulin resistance, is associated with CRC and CRA.

Recently, metabolically healthy as well as metabolically unhealthy states have been recognized to exist among obese individuals (5, 7). A unique subset of these individuals, termed metabolically healthy obese (MHO) individuals, despite having excessive body fat, seem to have a favorable metabolic profile without obesity-related metabolic abnormalities including insulin resistance, proatherogenic lipoprotein profile, proinflammatory state, or hypertension (5, 7). Furthermore, previous studies showed that MHO individuals were not at an increased risk for cardiovascular diseases compared to normal weight subjects (5, 8). Similarly, these individuals might not be at an increased risk for CRA or CRC, but no study has tested this hypothesis. This study evaluated the associations between BMI and CRA, established precursor lesions for CRC (9), through screening colonoscopies conducted on individuals who are metabolically healthy determined by a range of anthropometric and biochemical measures.

Materials and Methods

Study population

The study population consisted of examinees who underwent a colonoscopy as part of a comprehensive health screening program at Kangbuk Samsung Hospital, Seoul, Korea, from 2010 to 2011 (*N* = 62,171). The purpose of the screening program was to promote health through early detection of chronic diseases and their risk factors. Such programs are popular in Korea (10). In addition, in Korea, the Industrial

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Note: Supplementary data for this article are available at Cancer Research Online (<http://cancerres.aacrjournals.org/>).

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doi: 10.1158/0008-5472.CAN-12-3477

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Safety and Health Law requires employees to participate in annual or biennial health examinations. About 60% of the participants were employees of various companies and local governmental organizations and their spouses with the remaining participants registering individually for the program.

For this analysis, we excluded participants with missing anthropometric measures and incomplete colonoscopies (Fig. 1). Metabolically healthy participants were defined as those not having any metabolic abnormalities as follows (7, 11): (i) fasting blood glucose (FBG) ≥ 100 mg/dL or current use of blood glucose-lowering agents (12); (ii) blood pressure $\geq 130/85$ mm Hg or current use of blood pressure-lowering agents (12); (iii) elevated triglyceride levels ≥ 150 mg/dL or current use of lipid lowering agents (12); (iv) low high-density lipoprotein-cholesterol (HDL-C < 40 mg/dL in men or < 50 mg/dL in women; ref. 12); and (v) insulin resistance as homeostasis model assessment of insulin resistance (HOMA-IR) ≥ 2.5 (13). We further excluded subjects with history of inflammatory bowel disease and factors that could affect the association between BMI and CRA. As some individuals met more than one criterion for exclusion, the total number of eligible subjects for the study was 18,085 (Fig. 1).

This study was approved by the Institutional Review Board of Kangbuk Samsung Hospital, which exempted the requirement for informed consent as we only accessed data retrospectively that were de-identified.

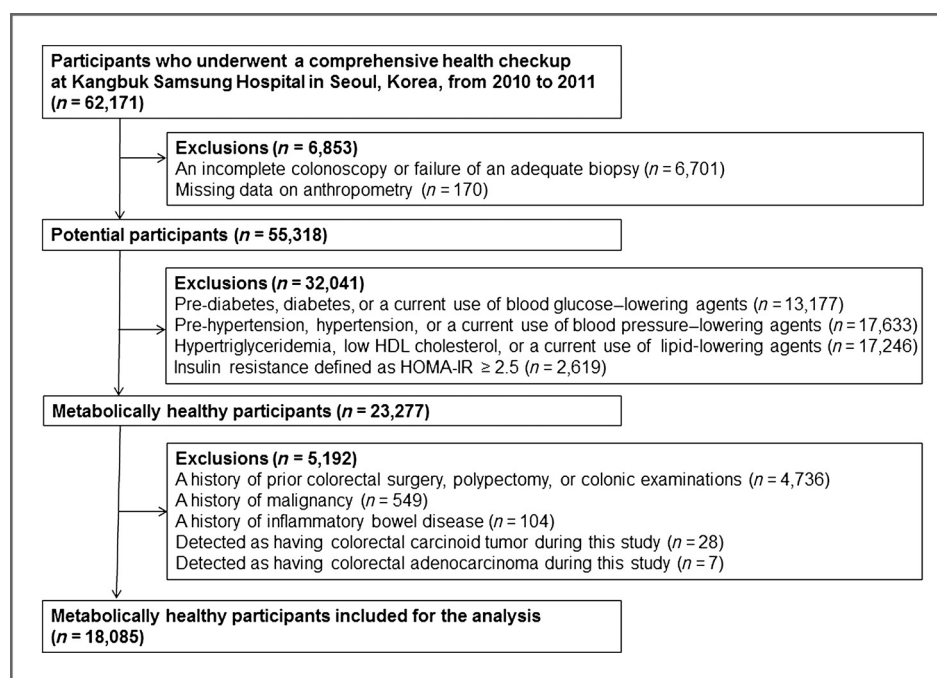
Measurements

Data on medical history, medication use, and health-related behaviors were collected through a self-administered questionnaire, whereas the physical measurements and serum biochemical parameters were measured by trained staff, all collected during the health examinations. Details regarding

alcohol use included the frequency of intake per week and the average amount of intake per episode. Current smokers were identified and the weekly frequency of moderate- or vigorous-intensity physical activity assessed. Family history of CRC was defined as CRC in one or more first-degree relatives at any age. Self-reported use of aspirin and analgesics of any type over the past month were assessed. Body weight was measured in light clothing and no shoes to the nearest 0.1 kg using a digital scale. Height was measured to the nearest 0.1 cm. BMI was calculated as weight in kilograms divided by height in meters squared. Trained nurses measured sitting blood pressure with standard mercury sphygmomanometers.

Blood samples were taken from the antecubital vein after at least a 10-hour fast. Serum levels of uric acid, total cholesterol, and triglyceride were determined using an enzymatic colorimetric assay; low-density lipoprotein-cholesterol and HDL-C levels were determined using a homogeneous enzymatic colorimetric assay; and alanine aminotransferase (ALT) and aspartate aminotransferase levels were determined by photometry using a Modular Analytics D2400 (Roche Diagnostics). Serum high-sensitivity C-reactive protein (hsCRP) level was determined using a particle-enhanced immunoturbidimetric assay on the Modular Analytics P800 apparatus (Roche Diagnostics). Serum insulin level was measured using electrochemiluminescence immunoassay on the Modular Analytics E170 apparatus (Roche Diagnostics) and serum fasting glucose level was measured using the hexokinase method on the Cobas Integra 800 apparatus (Roche Diagnostics). Insulin resistance was assessed with HOMA-IR according to the following equation: fasting blood insulin ($\mu\text{U}/\text{mL}$) \times FBG (mmol/L)/22.5. The Laboratory Medicine Department at Kangbuk Samsung Hospital in Seoul, Korea, has been accredited by the Korean Society of Laboratory Medicine and the Korean Association of

Figure 1. Flow diagram for the selection of study subjects.



Quality Assurance for Clinical Laboratories. The laboratory participates in the Collage of American Pathologists Survey Proficiency Testing.

Colonoscopy and histologic examination

Following careful bowel preparation with 4 L of polyethylene glycol–electrolyte oral lavage solution (Taejoon Pharm), colonoscopy was done on each subject by 1 of 13 experienced gastroenterologists using the EVIS LUCERA CV-260 colonoscope (Olympus) from the rectum to the cecum. All polypoid lesions were biopsied or removed and histologically assessed by experienced pathologists. Polyps were classified by number, size, and histologic characteristics [tubular, tubulovillous, or villous adenoma; hyperplastic polyp; sessile serrated polyp (also known as sessile serrated adenoma) or traditional serrated adenoma]. Hyperplastic polyps or other findings including diverticuli, hemorrhoids, anal fissure, and angiodysplasias were classified as normal colonoscopic findings in this study. The grade of dysplasia was classified as low or high grade. High-risk adenoma was defined as any adenoma larger than 1 cm, 3 or more adenomas, any adenoma with a villous component, or high-grade dysplasia (14). Subjects simultaneously diagnosed with high-risk and low-risk adenomas were classified as high-risk adenoma.

In a sensitivity analysis (Supplementary Tables S1 and S2), study subjects were categorized into 1 of 4 groups: (i) control group whose colonoscopy detected neither hyperplastic polyps nor adenomatous polyps, (ii) hyperplastic polyps only, (iii) low-risk adenoma, and (iv) high-risk adenoma. Subjects simultaneously diagnosed with hyperplastic polyps and adenomatous polyps were classified as either low-risk or high-risk adenoma according to their findings. For example, subjects with hyperplastic polyps and low-risk adenoma were classified as low-risk adenoma, whereas subjects with hyperplastic polyps and low-risk and high-risk adenomas were classified as high-risk adenoma.

Statistical analyses

Descriptive statistics were used to summarize the characteristics of participants by BMI categories in men and women. The BMI classification developed for Asian population was used (15): metabolically healthy underweight (MHU) = BMI < 18.5 kg/m²; metabolically healthy normal-weight (MHNW) = BMI of 18.5 to 23 kg/m²; metabolically healthy overweight (MHOW) = BMI of 23 to 25 kg/m²; and MHO = BMI ≥ 25 kg/m². The distribution of continuous variables was evaluated and appropriate transformations were conducted during analysis, as needed.

ORs were used to measure the association of BMI categories with the prevalence of one or more low-risk adenomas and one or more high-risk adenomas. In the main analyses, controls are subjects without any adenoma. Then in sensitivity analysis (Supplementary Tables S1 and S2), the controls are subjects without any adenoma or hyperplastic polyp. Multinomial logistic regression models were used to estimate ORs and 95% confidence intervals (95% CIs) after adjusting for potential confounders. The models were initially adjusted for age and sex, then for smoking, alcohol intake, exercise, educational

level, and family history of CRC. To determine linear trends of prevalence, the number of categories or quartiles was used as a continuous variable and tested on each model. Subgroup analyses were conducted according to gender, age group (<50 vs. ≥50 years of age), and lifestyle (current smoker vs. noncurrent smoker <20 vs. ≥20 g of alcohol per day, <1 time/week vs. ≥1 time/week of regular exercise); and interactions by subgroups were tested using the likelihood ratio tests (lrtest in STATA) comparing models with and without multiplicative interaction terms.

To compare the impact of BMI on CRA in the metabolically healthy population versus overall population, analyses were conducted not restricted to metabolically normal subjects (Supplementary Tables S3 and S4).

The statistical analysis was done using STATA version 11.2 (StataCorp LP). All reported *P* values are 2-tailed, and comparisons where *P* < 0.05 were considered statistically significant.

Results

With 7,179 women (39.7%) and 10,906 men (60.3%), the mean age and BMI of the 18,085 participants were 39.7 years (SD = 6.8) and 22.6 kg/m² (SD = 2.7, range 14.4–35.7), respectively. The baseline characteristics of the study participants in relation to BMI categories are outlined in Tables 1 and 2. Only for women was age positively associated with the BMI categories. For both men and women, current alcohol use, exercise, systolic and diastolic blood pressure, FBG, total cholesterol, triglycerides, uric acid, ALT, hsCRP, and HOMA-IR were positively associated with the BMI categories, whereas HDL-C was inversely associated with the BMI categories. The proportion of current smokers was the highest in MHU for men and in MHU and MHNW for women.

Table 3 shows the results of the multinomial logistic analyses with OR corresponding to 95% CIs of BMI categories for the prevalence of low-risk and high-risk adenoma. Of the 18,085 subjects, 1,674 subjects (9.3%) had low-risk adenomas and 248 subjects (1.4%) had high-risk adenomas. We first analyzed the relationships between the baseline BMI categories and each outcome adjusting for age and sex. Then, we adjusted for age, sex, smoking, drinking, regular exercise, family history of colon cancer, education level, aspirin use, and analgesic use. In the multivariate models, the prevalence of low-risk and high-risk CRA was associated with increasing categories of BMI categories in a dose-response manner (*P* for trend < 0.001 and 0.01, respectively). Compared to the MHNW group, the MHO group was at a significantly increased prevalence of both low-risk adenomas (OR = 1.44; 95% CI, 1.23–1.69) and high-risk adenomas (OR = 1.62; 95% CI, 1.09–2.41). To explore whether the association between BMI categories and CRA was mediated by HOMA-IR or hsCRP levels, we conducted additional analysis by adjusting for HOMA-IR and hsCRP levels. The results did not change. Further adjusting for glucose, SBP and DBP did not materially alter the estimates (data not shown).

The associations between the BMI categories and the prevalence of low-risk and high-risk adenomas were also examined by gender (Table 4). For men, an increase across BMI categories

Table 1. Baseline characteristics of metabolically healthy participants by BMI category among 10,906 men at health checkup center of Kangbuk Samsung Hospital in 2010 to 2011

Characteristics	Overall	BMI categories (kg/m ²)			
		MHU (<18.5)	MHNW (18.5–22.9)	MHOW (23.0–24.9)	MHO (≥25.0)
Men	N = 10,906	N = 215	N = 4,494	N = 3,445	N = 2,752
Age (years) ^a	39.1 (6.7)	39.4 (7.7)	39.0 (6.7)	39.4 (6.7)	39.0 (6.4)
Current smoke (%)	35.4	49.3	35.3	33.4	37.0
Alcohol intake (%) ^c	31.5	27.1	29.5	31.4	35.1
Regular exercise (%) ^d	58.3	43.8	54.9	60.5	62.2
Aspirin use (%)	1.1	0	1.1	1.2	1.0
Analgesics use (%)	2.7	2.3	2.6	2.8	2.9
Systolic blood pressure (mm Hg) ^a	110.9 (8.7)	106.9 (9.1)	109.7 (8.8)	111.5 (8.5)	112.7 (8.3)
Diastolic blood pressure (mm Hg) ^a	69.6 (6.5)	68.0 (7.3)	68.8 (6.6)	69.8 (6.4)	70.5 (6.3)
Glucose (mg/dL) ^a	88.7 (6.6)	85.9 (8.2)	87.9 (6.9)	89.2 (6.3)	89.6 (6.2)
Total cholesterol (mg/dL) ^a	196.5 (31.3)	182.0 (28.6)	191.7 (30.7)	198.4 (30.9)	203.2 (31.2)
HDL-C (mg/dL) ^a	56.7 (11.3)	66.0 (13.3)	59.3 (11.9)	55.6 (10.5)	52.9 (9.5)
Triglycerides (mg/dL) ^b	84.0 (65.0–107.0)	68.0 (55.0–86.0)	76.0 (60.0–97.0)	86.0 (67.0–108.0)	95.0 (74.0–118.0)
ALT (U/L) ^b	23.0 (18.0–32.0)	18.0 (14.0–23.0)	21.0 (16.0–28.0)	23.0 (18.0–33.0)	28.0 (21.0–39.0)
hsCRP (mg/L) ^b	0.50 (0.30–0.90)	0.30 (0.20–0.40)	0.40 (0.30–0.70)	0.50 (0.30–0.90)	0.60 (0.40–1.20)
HOMA-IR ^b	0.63 (0.43–0.93)	0.34 (0.21–0.50)	0.52 (0.35–0.73)	0.68 (0.47–0.96)	0.87 (0.59–1.19)

^aData are means (standard deviation).^bData are medians (interquartile range), or percentages.^c≥20 g of ethanol per day.^d≥1 time per week.

was positively associated with the prevalence of high-risk adenomas, as well as, low-risk adenomas in a dose-response manner (P for trend < 0.001 and 0.02, respectively). For women, the BMI categories were not statistically significantly related to low-risk adenomas, and the associations between the BMI categories and the prevalence of high-risk adenomas seemed to be nonlinear. The MHOW was positively associated with increased prevalence of high-risk adenomas (OR = 2.64; 95% CI, 1.30–5.36) whereas MHO was not statistically positively associated with high-risk adenoma. The overall interaction between gender and BMI categories for both low-risk adenoma and high-risk adenoma were not significant (P for interaction = 0.48).

The associations between the BMI categories and the prevalence of low-risk and high-risk adenomas were similar across the subgroups of the study participants with no significant interactions according to age group (<50 vs. ≥50 years of age), and lifestyle (current smoker vs. noncurrent smoker, <20 vs. ≥20 g of alcohol per day, <1 time/week vs. ≥1 time/week of regular exercise; data not shown).

In a sensitivity analysis, we examined the association of BMI categories with the prevalence of low-risk and high-risk adenomas with the control group consisting of subjects without hyperplastic polyps or adenomas. These analyses did not change any of the adenoma associations qualitatively (Supplementary Tables S1 and S2).

Supplementary Tables S3 and S4 show the impact of BMI on CRA in the population not restricted to the metabolically

healthy population. Overall, the associations between obesity and CRA were similar to that in the metabolically healthy population.

Discussion

This study shows that MHO individuals had a higher prevalence of high-risk CRA as well as low-risk CRA compared to MHNW individuals.

Previous studies have suggested that a higher BMI is associated with an increased risk of low-risk and high-risk CRA (16, 17). An important finding of our study is that this association exists even in metabolically healthy subjects selected exclusively for not having any metabolic syndrome components and insulin resistance. This suggests that increased BMI, even in the absence of a metabolic unhealthy state, can be an important risk factor for both low-risk and high-risk CRA. Another study showed that increased BMI and total body fat percentage, as a promoting factor, seems to increase adenoma growth (18). Our study supports that excess fat *per se*, even without metabolic abnormalities, is an independent risk factor for the development of CRA and its progression.

To our knowledge, this is the first study to address the hypothesis that MHO phenotype is a risk factor for precancerous lesions. Our findings raise the possibility that MHO individuals, a subset of obese individuals, might be protected from the cardiovascular risk standpoint but not protected from the cancer risk standpoint. Further studies are needed to address MHO phenotype as a possible risk for other cancers

Table 2. Baseline characteristics of metabolically healthy participants by BMI category among 7,179 women at health checkup center of Kangbuk Samsung Hospital in 2010 and 2011

Characteristics	Overall	BMI categories (kg/m ²)			
		MHU (<18.5)	MHNW (18.5–22.9)	MHOW (23.0–24.9)	MHO (≥25.0)
Women	N = 7,179	N = 700	N = 4,843	N = 1,056	N = 580
Age (years) ^a	40.7 (6.9)	37.6 (5.9)	40.3 (6.6)	42.8 (7.3)	43.5 (7.7)
Current smoke (%)	2.8	2.9	2.9	2.4	2.1
Alcohol intake (%) ^c	6.6	4.9	6.5	7.8	10.3
Regular exercise (%) ^d	47.7	39.5	49.0	47.6	46.4
Aspirin use (%)	1.3	1.4	1.3	1.6	0.7
Analgesics use (%)	4.9	5.6	4.8	4.7	5.7
Systolic blood pressure (mm Hg) ^a	104.4 (9.6)	101.4 (9.2)	103.9 (9.5)	106.6 (9.1)	108.9 (9.6)
Diastolic blood pressure (mm Hg) ^a	65.4 (6.6)	64.0 (6.4)	65.0 (6.5)	66.5 (6.4)	68.2 (6.7)
Glucose (mg/dL) ^a	86.8 (7.3)	84.6 (8.3)	86.7 (7.3)	87.9 (6.5)	88.8 (5.9)
Total cholesterol (mg/dL) ^a	189.8 (31.3)	182.0 (29.2)	188.4 (30.7)	194.8 (31.8)	202.3 (32.7)
HDL-C (mg/dL) ^a	67.9 (12.3)	71.8 (13.4)	68.5 (12.3)	65.4 (11.5)	62.8 (10.3)
Triglycerides (mg/dL) ^b	62.0 (51.0–79.0)	59.0 (49.0–70.0)	61.0 (50.0–76.0)	68.0 (55.0–87.0)	75.0 (60.0–96.0)
ALT (U/L) ^b	15.0 (12.0–20.0)	15.0 (12.0–19.0)	15.0 (12.0–20.0)	16.0 (13.0–21.0)	19.0 (14.0–24.0)
hsCRP (mg/L) ^b	0.30 (0.30–0.60)	0.30 (0.20–0.30)	0.30 (0.20–0.50)	0.40 (0.30–0.70)	0.60 (0.30–1.30)
HOMA-IR ^b	0.65 (0.43–0.95)	0.49 (0.31–0.72)	0.62 (0.40–0.88)	0.80 (0.54–1.10)	0.94 (0.68–1.31)

^aData are means (standard deviation).

^bData are medians (interquartile range), or percentages.

^c≥20 g of ethanol per day.

^d≥1 time per week.

where obesity is a known risk factor such as kidney, prostate, pancreas, breast, and endometrial cancers (19, 20).

With regard to the relationship between BMI, CRA, and CRC by sex, existing evidence suggests that the associations between obesity, CRA, and CRC are more consistent and stronger among men than women (1, 2, 21–23). Our study finding showed that higher BMI categories were significantly associated with increased prevalence of both low-risk and high-risk CRA in men but were inconsistent in women, showing a significantly increased prevalence of high-risk CRA only in women with MHOW: however, given the wide confidence limits for high-risk adenomas in women, it is difficult to exclude the effects of chance and characterize the occurrence pattern in any way. Previous studies reported weight change in addition to BMI was also independent factor for CRA or CRC (23, 24), especially in women (24). Also for women, the female hormonal status can affect the association between BMI and CRA (25). In this study, data on weight change, oral contraceptive use, or hormone therapy were not available for analysis. In addition, the inconclusive result for women can be explained by the small number of obese women, which may be insufficient to establish a relationship and lead to imprecise estimates.

In this study, the prevalence of MHO phenotype was 23.9% in obese individuals, lower than that in previous studies, with estimate as high as 30% in obese individuals (4, 5). Similarly, compared with previous reports (26), the prevalence of CRA was lower in this study, which is not unexpected due to our

stringent criteria for MHO. One challenge in evaluating CRA risk in the MHO phenotypes is the lack of a uniform definition for MHO. In this study, we defined metabolically healthy phenotype as subjects without any metabolic syndrome components and insulin resistance, which are commonly used to define MHO individuals (7).

The mechanisms that link increased BMI and CRA are not fully understood, although several possibilities have been raised (3, 27, 28). Insulin resistance is considered a key mechanism underpinning the obesity-colon cancer link (3, 27). In this metabolically healthy population, the positive association between increased BMI and CRA remained significant even after adjusting for metabolic parameters including HOMA-IR. CRA might be not necessarily related to insulin resistance in obese individuals (29). Another potential mechanism for colon carcinogenesis can be obesity-related inflammation. In our study, a significant association between increased BMI and CRA remained even after adjusting for CRP levels. A recent cross-sectional study reported a significant positive relationship between circulating levels of interleukin-6 (IL-6) and TNF- α and CRA, but a weaker, nonsignificant association between CRP and CRA (30). TNF- α and IL-6, inflammatory cytokines secreted by adipose tissue, were found to be involved in the early development of colorectal neoplasia (30, 31). In our study, IL-6 and TNF- α levels were not available, so we could not exclude obesity-related inflammation as a possible mediator between obesity and CRA.

Table 3. Associations between BMI and colorectal adenoma in 18,085 metabolically healthy participants among health checkup examinees at Kangbuk Samsung Hospital in 2010 to 2011.

BMI (kg/m ²) category	Person	Prevalent case	Age–sex adjusted OR (95% CI)	Multivariate OR ^a (95% CI)	
				Model 1	Model 2
Low-risk adenoma					
MHU (<18.5)	915	57	0.96 (0.72–1.28)	0.90 (0.61–1.33)	0.90 (0.61–1.33)
MHNW (18.5–22.9)	9,337	754	1.00 (reference)	1.00 (reference)	1.00 (reference)
MHOW (23.0–24.9)	4,501	470	1.11 (0.98–1.26)	1.17 (1.01–1.37)	1.17 (1.01–1.37)
MHO (≥25.0)	3,332	393	1.29 (1.12–1.47)	1.44 (1.23–1.69)	1.44 (1.21–1.71)
<i>P</i> for trend			<0.001	<0.001	<0.001
High-risk adenoma					
MHU (<18.5)	915	7	0.85 (0.39–1.85)	0.69 (0.24–1.96)	0.70 (0.25–2.00)
MHNW (18.5–22.9)	9,337	105	1.00 (reference)	1.00 (reference)	1.00 (reference)
MHOW (23.0–24.9)	4,501	78	1.35 (0.99–1.83)	1.50 (1.04–2.16)	1.48 (1.02–2.15)
MHO (≥25.0)	3,332	58	1.44 (1.02–2.01)	1.62 (1.09–2.41)	1.59 (1.04–2.43)
<i>P</i> for trend			0.02	0.01	0.01

^aModel 1: adjusted for age and sex, smoking status, alcohol intake, regular exercise, family history of colon cancer, educational level, analgesic and aspirin use; model 2: model 1 plus adjusted for HOMA-IR and hsCRP.

An association between excess fat *per se* and CRA in MHO phenotypes can be explained by altered adipocytokines. Adipose tissue is an important endocrine organ secreting numerous adipokines consisting of hormones, cytokines, and other signaling molecules that play roles in energy balance, inflammation, insulin sensitivity, and angiogenesis (32–34). Altered adipokine secretion from the adipose tissue, such as leptin and adiponectin, is considered a potential mediator for obesity-related colon cancer (35–38). Colon epithelial cells express adiponectin and leptin receptors,

supporting the potential of adiponectin and leptin to influence regulation of cellular processes within the colon (39). A recent study showed that adiponectin directly inhibits colon cancer cell proliferation (40). Leptin regulates proliferation in CRC by activating mitogenic and antiapoptotic signaling pathways (41, 42). Moreover, there is evidence for interactive effects of adiponectin and leptin in the early stage of colorectal tumorigenesis, distinct from their involvement in insulin resistance (43). Therefore, studies that further assess markers specifically indicating increased adipose

Table 4. Associations between BMI and colorectal adenoma by gender in 18,085 metabolically healthy participants among health checkup examinees at Kangbuk Samsung Hospital in 2010 to 2011

	BMI categories (kg/m ²)				<i>P</i> value for trend
	MHU (<18.5)	MHNW (18.5–22.9)	MHOW (23.0–24.9)	MHO (≥25.0)	
Men					
Number = 10,906	215	4,494	3,445	2,752	
Prevalent case of low-risk adenoma (%)	9.3	9.8	11.4	12.3	
Prevalent case of high-risk adenoma (%)	1.4	1.3	1.6	1.8	
aOR ^a (95% CI) for low-risk adenoma	0.92 (0.52–1.60)	1.00 (reference)	1.21 (1.02–1.43)	1.47 (1.24–1.75)	<0.001
aOR ^a (95% CI) for high-risk adenoma	0.85 (0.24–3.03)	1.00 (reference)	1.31 (0.86–1.98)	1.66 (1.08–2.56)	0.02
Women					
Number = 7,179	700	4,843	1,056	580	
Prevalent case of low-risk adenoma (%)	5.3	6.5	7.2	9.7	
Prevalent case of high-risk adenoma (%)	0.6	1.0	2.1	1.6	
aOR ^a (95% CI) for low-risk adenoma	0.87 (0.51–1.49)	1.00 (reference)	1.00 (0.67–1.49)	1.34 (0.84–2.14)	0.23
aOR ^a (95% CI) for high-risk adenoma	0.38 (0.05–2.85)	1.00 (reference)	2.64 (1.30–5.36)	1.14 (0.33–3.94)	0.06

NOTE: *P* = 0.48 for the overall interaction between gender and BMI categories for low-risk adenoma and high-risk adenoma (adjusted model).

^aAdjusted for age and sex, smoking status, alcohol intake, regular exercise, family history of colon cancer, educational level, analgesic, and aspirin use.

tissue will be helpful for establishing MHO as a risk factor for CRA or CRC.

There are several limitations to this study. First, the definition of insulin resistance used in this study is based on HOMA-IR and not on euglycemic insulin clamp, a reference method for assessing insulin resistance (44), which is invasive and not feasible in large populations. A second limitation is the use of BMI as a measure of obesity as it cannot distinguish between fat tissue and lean tissue. If the MHO group in this study has higher lean tissue than fat mass, the association between higher BMI categories and CRA could be attenuated. Third, we were unable to include dietary information, which could be a possible confounder for CRA (45). Another limitation is that a cross-sectional design precludes the determination of causality; however, the strength of our study design is that individuals having a first-time screening colonoscopy were included, minimizing the possibility of reverse causation. Finally, our findings cannot be simply extrapolated to other populations.

In conclusion, excess body weight, even in the absence of a metabolic unhealthy state, was associated with increased presence of both low-risk and high-risk CRA, established precursor lesions for CRC, possibly suggesting that the mechanisms linking excess body weight and CRA risk may go beyond insulin resistance. Further studies are needed to

address MHO as a possible risk factor for obesity-related cancers.

Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed.

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Acknowledgments

The authors thank T.S. Choi (Kangbuk Samsung Hospital, Information System, Seoul, Korea) for his help with technical support in gathering data, and also Dr. L. Kim (Edmonton, Alberta, Canada) for her help with revising this paper.

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Received September 3, 2012; revised March 22, 2013; accepted March 31, 2013; published OnlineFirst May 16, 2013.

References

1. Renehan AG, Tyson M, Egger M, Heller RF, Zwahlen M. Body-mass index and incidence of cancer: a systematic review and meta-analysis of prospective observational studies. *Lancet* 2008;371:569–78.
2. Ben Q, An W, Jiang Y, Zhan X, Du Y, Cai QC, et al. Body mass index increases risk for colorectal adenomas based on meta-analysis. *Gastroenterology* 2012;142:762–72.
3. Renehan AG, Frystyk J, Flyvbjerg A. Obesity and cancer risk: the role of the insulin-IGF axis. *Trends Endocrinol Metab* 2006;17:328–36.
4. Ogorodnikova AD, Kim M, McGinn AP, Muntner P, Khan U, Wildman RP. Incident cardiovascular disease events in metabolically benign obese individuals. *Obesity* 2012;20:651–9.
5. Stefan N, Kantartzis K, Machann J, Schick F, Thamer C, Rittig K, et al. Identification and characterization of metabolically benign obesity in humans. *Arch Intern Med* 2008;168:1609–16.
6. Giovannucci E. Metabolic syndrome, hyperinsulinemia, and colon cancer: a review. *Am J Clin Nutr* 2007;86:s836–42.
7. Primeau V, Coderre L, Karelis AD, Brochu M, Lavoie ME, Messier V, et al. Characterizing the profile of obese patients who are metabolically healthy. *Int J Obes (Lond)* 2011;35:971–81.
8. Meigs JB, Wilson PW, Fox CS, Vasan RS, Nathan DM, Sullivan LM, et al. Body mass index, metabolic syndrome, and risk of type 2 diabetes or cardiovascular disease. *J Clin Endocrinol Metab* 2006;91:2906–12.
9. Fearon ER, Vogelstein B. A genetic model for colorectal tumorigenesis. *Cell* 1990;61:759–67.
10. Park HW, Byeon JS, Yang SK, Kim HS, Kim WH, Kim TI, et al. Colorectal neoplasm in asymptomatic average-risk Koreans: the KASID prospective multicenter colonoscopy survey. *Gut Liver* 2009;3:35–40.
11. Kuk JL, Ardern CI. Are metabolically normal but obese individuals at lower risk for all-cause mortality? *Diabetes Care* 2009;32:2297–9.
12. Alberti KG, Eckel RH, Grundy SM, Zimmet PZ, Cleeman JI, Donato KA, et al. Harmonizing the metabolic syndrome: a joint interim statement of the International Diabetes Federation Task Force on Epidemiology and Prevention; National Heart, Lung, and Blood Institute; American Heart Association; World Heart Federation; International Atherosclerosis Society; and International Association for the Study of Obesity. *Circulation* 2009;120:1640–5.
13. Matthews DR, Hosker JP, Rudenski AS, Naylor BA, Treacher DF, Turner RC. Homeostasis model assessment: insulin resistance and beta-cell function from fasting plasma glucose and insulin concentrations in man. *Diabetologia* 1985;28:412–9.
14. Lieberman DA, Rex DK, Winawer SJ, Giardiello FM, Johnson DA, Levin TR. Guidelines for colonoscopy surveillance after screening and polypectomy: a consensus update by the US Multi-Society Task Force on Colorectal Cancer. *Gastroenterology* 2012;143:844–57.
15. WHO Western Pacific Region Ial. The Asia-pacific perspective: redefining obesity and its treatment. Health Communications Australia Pty Limit: Sydney, Australia; 2000.
16. Tsilidis KK, Brancati FL, Pollak MN, Rifai N, Clipp SL, Hoffman-Bolton J, et al. Metabolic syndrome components and colorectal adenoma in the CLUE II cohort. *Cancer Causes Control* 2010;21:1–10.
17. Okabayashi K, Ashrafian H, Hasegawa H, Yoo JH, Patel VM, Harling L, et al. Body mass index category as a risk factor for colorectal adenomas: a systematic review and meta-analysis. *Am J Gastroenterol* 2012;107:1175–85.
18. Almendingen K, Hofstad B, Vatn MH. Does high body fatness increase the risk of presence and growth of colorectal adenomas followed up *in situ* for 3 years? *Am J Gastroenterol* 2001;96:2238–46.
19. Calle EE, Rodriguez C, Walker-Thurmond K, Thun MJ. Overweight, obesity, and mortality from cancer in a prospectively studied cohort of U.S. adults. *N Engl J Med* 2003;348:1625–38.
20. Parr CL, Batty GD, Lam TH, Barzi F, Fang X, Ho SC, et al. Body-mass index and cancer mortality in the Asia-Pacific Cohort Studies Collaboration: pooled analyses of 424,519 participants. *Lancet Oncol* 2010;11:741–52.
21. Anderson JC, Messina CR, Dakhllalah F, Abraham B, Alpern Z, Martin C, et al. Body mass index: a marker for significant colorectal neoplasia in a screening population. *J Clin Gastroenterol* 2007;41:285–90.
22. Morimoto LM, Newcomb PA, Ulrich CM, Bostick RM, Lais CJ, Potter JD. Risk factors for hyperplastic and adenomatous polyps: evidence

- for malignant potential? *Cancer Epidemiol Biomarkers Prev* 2002;11:1012–8.
23. Sedjo RL, Byers T, Levin TR, Haffner SM, Saad MF, Tooze JA, et al. Change in body size and the risk of colorectal adenomas. *Cancer Epidemiol Biomarkers Prev* 2007;16:526–31.
 24. Nock NL, Thompson CL, Tucker TC, Berger NA, Li L. Associations between obesity and changes in adult BMI over time and colon cancer risk. *Obesity (Silver Spring)* 2008;16:1099–104.
 25. Chen J, Iverson D. Estrogen in obesity-associated colon cancer: friend or foe? Protecting postmenopausal women but promoting late-stage colon cancer. *Cancer Causes Control* 2012;23:1767–73.
 26. Rundle AG, Lebwohl B, Vogel R, Levine S, Neugut AI. Colonoscopic screening in average-risk individuals ages 40 to 49 vs. 50 to 59 years. *Gastroenterology* 2008;134:1311–5.
 27. Khandekar MJ, Cohen P, Spiegelman BM. Molecular mechanisms of cancer development in obesity. *Nat Rev Cancer* 2011;11:886–95.
 28. Yehuda-Shnaidman E, Schwartz B. Mechanisms linking obesity, inflammation and altered metabolism to colon carcinogenesis. *Obesity Rev* 2012;13:1083–95.
 29. Sedjo RL, D'Agostino RB Jr, Ahnen D, Levin TR, Haffner SM, Tooze JA, et al. Lack of association between insulin sensitivity and colorectal adenoma risk. *Nutr Cancer* 2011;63:6–11.
 30. Kim S, Keku TO, Martin C, Galanko J, Woosley JT, Schroeder JC, et al. Circulating levels of inflammatory cytokines and risk of colorectal adenomas. *Cancer Res* 2008;68:323–8.
 31. Balkwill F, Charles KA, Mantovani A. Smoldering and polarized inflammation in the initiation and promotion of malignant disease. *Cancer Cell* 2005;7:211–7.
 32. Renehan AG, Roberts DL, Dive C. Obesity and cancer: pathophysiological and biological mechanisms. *Arch Physiol Biochem* 2008;114:71–83.
 33. Chen J. Multiple signal pathways in obesity-associated cancer. *Obes Rev* 2011;12:1063–70.
 34. Huang XF, Chen JZ. Obesity, the PI3K/Akt signal pathway and colon cancer. *Obes Rev* 2009;10:610–6.
 35. Kelesidis I, Kelesidis T, Mantzoros CS. Adiponectin and cancer: a systematic review. *Br J Cancer* 2006;94:1221–5.
 36. Drew JE. Molecular mechanisms linking adipokines to obesity-related colon cancer: focus on leptin. *Proc Nutr Soc* 2012;71:175–80.
 37. McTiernan A. Obesity and cancer: the risks, science, and potential management strategies. *Oncology (Williston Park)* 2005;19:871–81; discussion 81–2, 85–6.
 38. Ho GY, Wang T, Gunter MJ, Strickler HD, Cushman M, Kaplan RC, et al. Adipokines linking obesity with colorectal cancer risk in postmenopausal women. *Cancer Res* 2012;72:3029–37.
 39. Drew JE, Farquharson AJ, Padidar S, Duthie GG, Mercer JG, Arthur JR, et al. Insulin, leptin, and adiponectin receptors in colon: regulation relative to differing body adiposity independent of diet and in response to dimethylhydrazine. *Am J Physiol Gastrointest Liver Physiol* 2007;293:G682–91.
 40. Kim AY, Lee YS, Kim KH, Lee JH, Lee HK, Jang SH, et al. Adiponectin represses colon cancer cell proliferation via AdipoR1- and -R2-mediated AMPK activation. *Mol Endocrinol* 2010;24:1441–52.
 41. Wang D, Chen J, Chen H, Duan Z, Xu Q, Wei M, et al. Leptin regulates proliferation and apoptosis of colorectal carcinoma through PI3K/Akt/mTOR signalling pathway. *J Biosci* 2012;37:91–101.
 42. Amemori S, Ootani A, Aoki S, Fujise T, Shimoda R, Kakimoto T, et al. Adipocytes and preadipocytes promote the proliferation of colon cancer cells *in vitro*. *Am J Physiol Gastrointest Liver Physiol* 2007;292:G923–9.
 43. Yamaji T, Iwasaki M, Sasazuki S, Tsugane S. Interaction between adiponectin and leptin influences the risk of colorectal adenoma. *Cancer Res* 2010;70:5430–7.
 44. DeFronzo RA, Tobin JD, Andres R. Glucose clamp technique: a method for quantifying insulin secretion and resistance. *Am J Physiol* 1979;237:E214–23.
 45. Cross AJ, Ferrucci LM, Risch A, Graubard BI, Ward MH, Park Y, et al. A large prospective study of meat consumption and colorectal cancer risk: an investigation of potential mechanisms underlying this association. *Cancer Res* 2010;70:2406–14.