PPARα and Sirt1 Mediate Erythropoietin Action in Increasing Metabolic Activity and Browning of White Adipocytes to Protect Against Obesity and Metabolic Disorders

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Erythropoietin (EPO) has shown beneficial effects in the regulation of obesity and metabolic syndrome; however, the detailed mechanism is still largely unknown. Here, we created mice with adipocyte-specific deletion of EPO receptor. These mice exhibited obesity and decreased glucose tolerance and insulin sensitivity, especially when fed a high-fat diet. Moreover, EPO increased oxidative metabolism, fatty acid oxidation, and key metabolic genes in adipocytes and in white adipose tissue from diet-induced obese wild-type mice. Increased metabolic activity by EPO is associated with induction of brown fat–like features in white adipocytes, as demonstrated by increases in brown fat gene expression, mitochondrial content, and uncoupled respiration. Peroxisome proliferator–activated receptor (PPAR)α was found to mediate EPO activity because a PPARα antagonist impaired EPO-mediated induction of brown fat–like gene expression and uncoupled respiration. PPARα also cooperates with Sirt1 activated by EPO through modulating the NAD+ level to regulate metabolic activity. PPARα targets, including PPARγ coactivator 1α, uncoupling protein 1, and carnitine palmityltransferase 1α, were increased by EPO but impaired by Sirt1 knockdown. Sirt1 knockdown also attenuated adipose response to EPO. Collectively, EPO, as a novel regulator of adipose energy homeostasis via these metabolism coregulators, provides a potential therapeutic strategy to protect against obesity and metabolic disorders. Diabetes 62:4122–4131, 2013

Obesity and its associated metabolic syndrome, including glucose intolerance and insulin resistance, are well-documented risk factors for cardiovascular disease, type 2 diabetes, and stroke. It is therefore essential to develop new strategies to treat metabolic syndrome and obesity. Recently, erythropoietin (EPO), the cytokine required for the erythrocyte production, has become attractive because of its important protective activity in the nonerythroid system. It is now recognized that EPO has protective effects in animal models of cardiac ischemia/reperfusion injury via stimulating endothelial cells to produce nitric oxide to regulate vascular tone and improve oxygen transport (1–5). EPO activity has also been reported for other nonhematopoietic tissue, including brain protection against ischemia, enhanced neural progenitor production, and anti-inflammatory effects (6–8). We demonstrated that disrupted EPO signaling in all nonerythroid tissues promotes obesity (9). However, the mechanism by which EPO functions as a metabolic regulator to cooperate with other coregulators to regulate energy homeostasis remains largely unknown. Compared with other nonhematopoietic tissue, we found that the EPO receptor (EpoR) is expressed at a high level in white adipose tissue (WAT) (6–20% of hematopoietic tissue), which is secondary to its primary expression site of EpoR, raising the possibility that endogenous EPO action in WAT may contribute importantly to protection against obesity and its associated metabolic disorders.

WAT adipocytes are specialized for the storage of excess energy such as triglycerides. It is now recognized, however, that WAT may also play a central role in energy homeostasis and systemic metabolism (10). Brown adipose tissue (BAT) adipocytes can dissipate calories as heat via uncoupled metabolism due to a pattern of gene expression that results in a high mitochondrial content and elevated cellular respiration (11). Observations that adult humans have functional and metabolically active BAT (12–15) suggest that a higher level of BAT may be protective against obesity and have stimulated interest concerning the therapeutic potential of augmenting brown fat to combat obesity and its associated metabolic disease. Brown adipocytes are found interspersed within the WAT under certain conditions such as cold exposure or after stimulation of the β3-adrenergic pathways (16,17). Advances have recently been achieved on identification of several transcriptional factors and coregulators that specifically promote development and acquisition of the BAT-specific gene expression profile, including uncoupling protein 1 (UCP1), PRDM16, peroxisome proliferator–activated receptor γ coactivator 1α (PGC-1α), and peroxisome proliferator–activated receptor α (PPARα) (18–20).

PGC-1α and PRDM16 are transcriptional coactivators involved in the control of energy metabolism, and ectopic expression of PGC-1α and PRDM16 in WAT induces acquisition of BAT features, including expression of mitochondrial and fatty acid oxidation and thermogenic genes to limit weight gain and improve glucose intolerance in
response to a high-fat diet (HFD) (20–23). PPARα plays an important role in lipid metabolism, and activation of PPARα in human WAT led to the appearance of brown fat gene expression, including UCP1, PGC-1α, and PRDM16 (19,24). PPARα has been considered a distinctive marker of BAT with respect to the WAT phenotype (25). Another metabolic sensor, Sirt1, an NAD+ dependent type III deacetylase sir-tuin, activates PGC-1α and various substrates, including PPARγ and PPARα, in skeletal muscle and adipocytes to contribute to energy expenditure and browning of WAT and resistance to dietary obesity (26–30).

Here, we show that EPO activity in WAT is responsible for energy homeostasis. We confirm that the development of a brown fat–like gene program and an increase in metabolic activity in WAT with EPO stimulation are associated with a rise in whole-body energy expenditure, a suppression of weight gain, and improvement of glucose intolerance and insulin resistance in response to an HFD. In contrast, the loss of EPO activity in WAT leads to opposite effects. Importantly, we demonstrate that PPARα is required for the EPO-stimulated increase in metabolic activity and induction of a thermogenic gene program in white adipocytes. The EPO-promoted increase in the NAD+ level activates Sirt1 activity, which cooperates with PPARα to mediate the adipocyte response to EPO. Our results clearly identify EPO as a critical mediator of energy homeostasis and provide a potential contribution to the protection against obesity and metabolic syndrome.

RESEARCH DESIGN AND METHODS

Animal studies. For analysis of body weight, body composition, and food intake, C57BL/6 mice (4 weeks; The Jackson Laboratory) were maintained under an 12-h light/dark cycle with free access to food and drinking water, except as indicated for pair-fed mice. Male mice were fed the HFD (60 kcal% fat/[high fat]) and separated into different groups. EPO treatment (3,000 units/kg; three times/week for 2.5 to 5 weeks as indicated) was administered by subcutaneous injection. The control and pair-fed mice were injected with PBS as the vehicle control. EpoRaP2KO mice were generated as described (31) to generate a fat-specific knockout (KO) of EpoR (EpoRfl/fl mice). EpoR expression was analyzed in multiple tissues, including spleen, subcutaneous WAT (S-WAT), and visceral WAT (V-WAT); SVF was analyzed from WAT, BAT, kidney, heart, liver, and muscle (Fig. 1A); and loss of EpoR expression in S-WAT and V-WAT was validated using Western blotting (Fig. 1B). Because ap2 is also expressed in macrophages, we isolated macrophages from WAT and detected EpoR expression in macrophages. The EpoR expression level in macrophages from EpoRfl/fl mice was not significantly different from wild-type (WT) mice (Supplementary Fig. 1A). EpoRap2KO mice fed normal chow showed a slight but significant increase in body weight and fat mass compared with littermate controls (Fig. 1C and D), and by 30 weeks, EpoRap2KO mice were 20% more massive, with a 65% increase in fat mass. Total activity, VO2, and total RER were also significantly lower in EpoRap2KO mice (Fig. 1E and F and Supplementary Fig. 1B). The difference in EpoRap2KO body weight was further accentuated when mice were challenged by the HFD (Fig. 1G); although food intake was comparable (Supplementary Fig. 1C). When 10-week-old, EpoRap2KO mice were fed the HFD for 6 weeks, they exhibited a 1.4-fold increase in fat mass compared with control mice (Fig. 1H), along with glucose intolerance and insulin resistance (Fig. 1I and J). Compared with the HFD control mice, these HFD EpoRap2KO mice exhibited higher fasting serum leptin and glucose levels, an increased insulin level (Fig. 1K), and a trend toward higher serum adiponectin (Supplementary Fig. 1D). Insulin-stimulated AKT (protein kinase B) activation (phosphorylation) contributes to glucose and energy homeostasis (32). Diet-induced obesity (DIO) reduced the insulin receptor substrate/phosphatidylinositol 3-kinase/AKT intracellular pathway in rats (33). We observed that EPO treatment increased AKT activity, as shown by increased AKT phosphorylation in S-WAT and in V-WAT in control mice. In contrast, EpoRap2KO mice showed decreased AKT phosphorylation, and AKT activity did not increase with EPO treatment in EpoRap2KO mice in contrast to control mice (Fig. 1L), suggesting EPO activity modulates AKT activation, which may have an effect on insulin signaling.

Mitochondrial DNA copy measurement. The amount of mitochondrial DNA (mtDNA) relative to nuclear DNA was determined by quantitative real-time PCR using primers for Ndi2 (NADH dehydrogenase subunit 2; mitochondrial genome) and Nune1 (nuclear genome).

RNA interference. For knockdown experiments, small interfering RNA (siRNA) specific for Sirt1 and negative control (Thermo Scientific Dharmacon) were transfected into 3T3-L1 adipocytes and primary mouse adipocytes (M-adipocytes) via the DharmaFECT transfection reagent.

Statistical analyses. Values are expressed as mean ± SEM. The unpaired two-tailed Student's t test was used to determine differences between vehicle- and EPO-treated groups. Results in multiple groups were compared by ANOVA. Significance was set at P < 0.05.

RESULTS

Targeted deletion of EpoR in adipocyte tissue leads to obesity, glucose intolerance, and insulin resistance. To determine the direct adipocyte EPO response, we mated chimeric EpoR floxed mice with mice containing a cre-recombinase gene controlled by the ap2 promoter (31) to generate a fat-specific knockout (KO) of EpoR (EpoRap2KO mice). EpoR gene expression was analyzed in multiple tissues, including spleen, subcutaneous WAT (S-WAT), and visceral WAT (V-WAT); SVF was analyzed from WAT, BAT, kidney, heart, liver, and muscle (Fig. 1A); and loss of EpoR expression in S-WAT and V-WAT was validated using Western blotting (Fig. 1B). Because ap2 is also expressed in macrophages, we isolated macrophages from WAT and detected EpoR expression in macrophages. The EpoR expression level in macrophages from EpoRap2KO mice was not significantly different from wild-type (WT) mice (Supplementary Fig. 1A). EpoRap2KO mice fed normal chow showed a slight but significant increase in body weight and fat mass compared with littermate controls (Fig. 1C and D), and by 30 weeks, EpoRap2KO mice were 20% more massive, with a 65% increase in fat mass. Total activity, VO2, and total RER were also significantly lower in EpoRap2KO mice (Fig. 1E and F and Supplementary Fig. 1B). The difference in EpoRap2KO body weight was further accentuated when mice were challenged by the HFD (Fig. 1G); although food intake was comparable (Supplementary Fig. 1C). When 10-week-old, EpoRap2KO mice were fed the HFD for 6 weeks, they exhibited a 1.4-fold increase in fat mass compared with control mice (Fig. 1H), along with glucose intolerance and insulin resistance (Fig. 1I and J). Compared with the HFD control mice, these HFD EpoRap2KO mice exhibited higher fasting serum leptin and glucose levels, an increased insulin level (Fig. 1K), and a trend toward higher serum adiponectin (Supplementary Fig. 1D). Insulin-stimulated AKT (protein kinase B) activation (phosphorylation) contributes to glucose and energy homeostasis (32). Diet-induced obesity (DIO) reduced the insulin receptor substrate/phosphatidylinositol 3-kinase/AKT intracellular pathway in rats (33). We observed that EPO treatment increased AKT activity, as shown by increased AKT phosphorylation in S-WAT and in V-WAT in control mice. In contrast, EpoRap2KO mice showed decreased AKT phosphorylation, and AKT activity did not increase with EPO treatment in EpoRap2KO mice in contrast to control mice (Fig. 1L), suggesting EPO activity modulates AKT activation, which may have an effect on insulin signaling.
To rule out a role for EpoR on other cell types, we treated EpoRaP2KO and WT mice with EPO for 3 weeks. EpoRaP2KO mice and WT mice showed an increase in hematocrit after EPO treatment (Supplementary Fig. 1). WT mice also exhibited the expected decrease in body weight of 15% at 3 weeks of EPO treatment (P < 0.01), and this decrease continued up to 7 weeks after EPO treatment (Fig. 1). The EpoRaP2KO mice, however, only exhibited a decreasing trend in body weight of 4% at 3 weeks of EPO treatment (Fig. 1N). These data strongly indicate that endogenous and exogenous EPO/EpoR in adipocytes plays an important role in the regulation of obesity and that EPO/EpoR activity in fat tissue is responsible for much of the EPO regulation of body weight, fat mass accumulation, glucose metabolism, and insulin sensitivity.

**EPO regulates mitochondrial genes in adipocytes and in WAT.** Increased mitochondrial biogenesis and related gene expression correlate with a reduction of DIO (34). We found that expression of mitochondrial biogenesis genes, including cytochrome c (CytC), isocitrate dehydrogenase 3α (Idh3a), and cytochrome c oxidase subunit 7A1 (Cox7a1) was decreased with the loss of EPO activity in S-WAT from EpoRaP2KO mice compared with littermate control mice (Fig. 2A). The key metabolic regulator, Pgc-1α, and fatty acid utilization gene carnitine palmitoyltransferase 1 (Cpt1) were also downregulated (Fig. 2A). To determine if the effects of EPO are independent of body weight, we performed paired-feeding studies in WT DIO mice without or with EPO treatment (3,000 units/kg, three times/week for 5 weeks). Importantly, although paired-fed mice exhibited a mild reduction in body fat content and fat pad weights, accompanied by modestly improved GTT and ITT compared with the vehicle-treated mice (Supplementary Fig. 2A and B), increased expression of CytC, Idh3a, Cpt1, Pgc-1α, and Cox7a1 in both S-WAT and V-WAT was only observed in EPO-treated DIO mice but not in vehicle-treated and paired-fed DIO mice (Fig. 2B and Supplementary Fig. 2C), suggesting the effects of EPO are independent of differences in body weight.

In vitro, mitochondrial genes and fatty acid oxidation gene were also upregulated by EPO in 3T3-L1 adipocytes and in H-adipocytes (Fig. 2C and D). CytC protein level
CS activity in S-WAT of EpoRaP2KO mice was decreased in vehicle-treated or paired-fed DIO mice (Fig. 3A). In vivo, we also observed that EPO treatment in DIO mice increased CS activity in S-WAT and V-WAT compared with control mice (Fig. 3B). In vitro, we determined the effect of EPO on the OCR as a measure of mitochondrial function (~related genes, PGClα and CPT1, without and with EPO treatment, in differentiated H-adipocytes was also determined. Cytochrome C (CytC) protein levels were determined by ELISA in 3T3-L1 adipocytes, M-adipocytes, and H-adipocytes, without and with EPO treatment (Fig. 3C). In S-WAT from the DIO mice treated for 5 weeks with EPO or PBS or paired-fed (Fig. 3D), and in 3T3-L1 adipocytes with PBS or EPO treatment (5 units/ml) (Fig. 3E). The EPO-stimulated increase in OCR of 3T3-L1 adipocytes under the basal condition and in the presence of oligomycin and FCCP under the hypoxic condition (Fig. 3F). EPO treatment also increased OCR in H-adipocytes (Fig. 3F). In vivo, adipocytes isolated from S-WAT after 5 weeks of EPO treatment in DIO mice also showed an increase in OCR (Fig. 3G). In comparison, adipocytes isolated from S-WAT of EpoR −/− mice exhibited reduced OCR compared with control mice (Fig. 3H). Taken together, these findings highlight a previously unrecognized role for EPO/EpoR activity in increasing cellular mitochondrial respiration and oxidative metabolism capacity beyond its effect of increased erythropoiesis and oxygen transport capacity, leading to increased oxygen utilization capacity and energy oxidative metabolism efficiency, and offset the adverse effect of obesity.

Mitochondrial dysfunction in adipose tissue is linked to obesity and type 2 diabetes in humans, as indicated by reduced oxidative phosphorylation capacity and reduced fatty acid β-oxidation in several tissues, including adipocytes (37,38). We treated adipocytes with palmitate, a substrate for fatty acid oxidation, and observed that EPO stimulation further enhanced the palmitate-stimulated increase in OCR (Fig. 3I–K), suggesting that EPO increases fatty acid oxidation in adipocytes. These data support the view that EPO increases energy expenditure in WAT and also demonstrate that EPO enhances the ability of adipocytes to metabolize fatty acid, which may limit the storage of excess lipid to protect against obesity.

EPO increases mitochondrial activity, cellular respiration capacity, and fatty acid utilization. As the key first pacemaking component of the tricarboxylic acid cycle, citrate synthase (CS) activity is an indicator of mitochondrial function. EPO stimulation increased CS activity in 3T3-L1 adipocytes, M-adipocytes, and H-adipocytes (Fig. 3A). In vivo, CS activity in 3T3-L1 adipocytes (37,38). We treated adipocytes with palmitate, a substrate for fatty acid oxidation, and observed that EPO stimulation further enhanced the palmitate-stimulated increase in OCR (Fig. 3I–K), suggesting that EPO increases fatty acid oxidation in adipocytes. These data support the view that EPO increases energy expenditure in WAT and also demonstrate that EPO enhances the ability of adipocytes to metabolize fatty acid, which may limit the storage of excess lipid to protect against obesity.
with BMI (12,13). However, EpoR expression in BAT is an order of magnitude lower (Fig. 1A), and expression of BAT-associated factors is unchanged in BAT isolated from EPO-treated mice (Supplementary Fig. 2E). We therefore hypothesized that the EPO-mediated antiobesity activity and associated metabolic improvement observed above may be related to the promotion of brown fat–associated markers in white adipocytes by EPO. We isolated S-WAT after 2.5 weeks of EPO treatment in young DIO mice before the readily detected change in body weight to rule out the effect of body weight difference. As we expected, these mice exhibited similar body weight but improved GTT and ITfcT compared with vehicle-treated mice (Supplementary Fig. 3A and B). With EPO treatment, we observed an increase in the expression of Ucp3 and Ppara in V-WAT (Fig. 4A), whereas S-WAT exhibited a larger scale and more dramatic increase of BAT-associated genes, including Cidea, Prdm16, Ucp3, Ppara, and Pgc-1α in EPO-treated DIO mice (Fig. 4B). We also isolated primary adipocytes from brown interscapular, white inguinal subcutaneous, and white visceral epididymal adipocytes from WT and EpoRaP2KO mice without and with EPO treatment. The expression of BAT-associated factors at the mRNA and protein level was unchanged in brown primary adipocytes (Supplementary Fig. 2E). In inguinal subcutaneous white primary adipocytes, cells harvested from EPO-treated WT mice showed increased expression of Cidea, Prdm16, Ucp1, Ucp3, Ppara, and Pgc-1α compared with untreated WT mice, whereas the expression of these genes was decreased in EpoRaP2KO mice adipocytes and was not different from EPO treated EpoRaP2KO mice (Fig. 4C). In visceral epididymal white adipocytes, cells from EPO-treated WT mice only showed increased expression of Ucp3, Ppara, and Pgc-1α, and the expression of these genes was reduced in adipocytes from EpoRaP2KO mice and remained unchanged in EpoRaP2KO mice treated with EPO (Fig. 4D). Western blotting confirmed that protein expression levels were consistent with the gene expression pattern (Fig. 4E and F). All other statistics were performed using the Student t test. Bar graphs are mean ± SEM. In vitro data are means of three independent experiments. *P < 0.05; **P < 0.01; ***P < 0.001.

![Figure 3. EPO increases adipocyte mitochondrial activity and VO2 capacity.](http://diabetesjournals.org/diabetes/article-pdf/62/12/4122/571758/4122.pdf)
treatment did not show any effect (Fig. 4G). The Resistin protein level was also decreased by EPO treatment in WT mice but not in EpoRaP2KO mice (Fig. 4H). These data suggest suppression of a WAT-associated program of gene expression by EPO.

We then analyzed CS activity as an indicator of mitochondrial function and bioenergetics marker. We did not observe any change of CS activity in brown primary adipocytes from WT mice treated with EPO or from EpoRaP2KO mice without or with EPO treatment compared with WT mice (Fig. 5A). This is consistent with the gene expression and protein expression analysis in brown primary adipocytes. However, in subcutaneous and visceral primary adipocytes isolated from WT mice treated with EPO and from EpoRaP2KO mice, CS activity was affected by EPO/EpoR signaling, increasing in WT mice treated with EPO and decreasing in EpoRaP2KO mice with or without EPO treatment compared with WT control mice (Fig. 5A). These data provide further evidence that BAT is not EPO responsive in contrast to S-WAT and V-WAT, with S-WAT exhibiting greater EPO induction of brown fat–associated gene expression. We also performed hematoxylin and eosin staining. The size of inguinal adipocytes was slightly reduced in the EPO-treated WT mice but not in the EpoRaP2KO mice (Fig. 5B). UCP1 immunohistochemistry staining further supports the shift in S-WAT toward brown fat–like characteristics with EPO treatment (Fig. 5C). UCP1 displayed stronger staining in S-WAT from EPO-treated WT control mice but decreased staining in EpoRaP2KO mice without or with EPO treatment compared with untreated WT control mice (Fig. 5C). These results indicated that the EPO promoted WAT to a BAT-like appearance, especially in S-WAT.

In culture, EPO stimulation in primary M-adipocytes also increased expression of BAT-associated genes (Fig. 5D), elevated mitochondrial gene expression (Fig. 5E), and mitochondrial density reflected by increased mitochondrial staining and mtDNA (Fig. 5F and 2G). Interestingly, in addition to increased total OCR, a higher uncoupled OCR was observed in EPO-treated M-adipocytes after injection of oligomycin, an index of uncoupled respiration and a characteristic of brown adipocytes (Fig. 5G), providing functional evidence for EPO promotion of BAT-like features in WAT. These findings collectively imply that EPO

FIG. 4. EPO promotes brown fat–associated gene and protein expression in WAT. Expression of BAT-associated genes in V-WAT (A) and S-WAT (B) in DIO mice treated without (PBS) and with EPO for 2.5 weeks was determined (n = 6). Expression of BAT-associated genes in primary adipocytes isolated from S-WAT (C) and V-WAT (D) in EpoRaP2KO (KO) and WT control mice without (PBS) and with EPO treatment were determined and compared (n = 4). The protein level of BAT-associated factors in S-WAT (E) and V-WAT (F) in EpoRaP2KO (KO) and littermate control mice without (PBS) and with EPO treatment (n = 4). Expression of mRNA (G) and protein (H) of white fat–associated genes (G) and Resistin in primary adipocytes isolated from S-WAT and V-WAT in EpoRaP2KO (KO) and WT control mice without (PBS) and with EPO treatment were determined (n = 4). One-way ANOVA was used in C, D, and G. All other statistics were performed using the Student t test. Bar graphs are mean ± SEM. In vitro data are means of three independent experiments. *P < 0.05; **P < 0.01.
stimulation in WAT promotes expression of BAT-associated genes and represses WAT-associated genes that may be responsible for the increased mitochondrial biogenesis and oxidative metabolism. Collectively, these effects may partly explain the manner in which EPO counteracts obesity and its associated metabolic disorders during DIO. However, we cannot exclude the possibility of systemic effects of EPO in promoting BAT-like characteristics in WAT in vivo, even if mice do not show any difference in body mass compared with control mice (Supplementary Fig. 3A).

**PPARα cooperates with Sirt1 to mediate the adipocyte response to EPO.** Next, we examined the mechanism by which EPO promotes brown features in WAT and increases metabolic activity. Our data showed that EPO increased PPARα expression and its downstream target PGC-1α (Fig. 4). PPARα was reported to promote the expression of the PRDM16 and UCP1, the two regulators determining brown adipocyte lineage. Activation of PPARα in human white adipocytes led to the appearance of a brown adipocyte pattern of gene expression, including induction of PGC-1α and PRDM16 (19). We treated M-adipocytes with the PPARα agonist GW6471 and found GW6471 abrogated EPO-mediated increases in BAT-enriched genes such as Prdm16, Ucp1, Ucp3, and PGC-1α (Fig. 6A). GW6471 also attenuated EPO-induced basal and uncoupled OCR in M-adipocytes (Fig. 6B), suggesting PPARα is required for EPO activity in increasing mitochondrial activity and promotion of brown features in WAT.

As a downstream target of PPARα, PGC-1α is also activated by its deacetylation via Sirt1, an NAD-dependent deacetylase and a member of sirtuins that is responsible for PGC-1α deacetylation (39). In EPO-stimulated 3T3-L1 adipocytes, PGC-1α was increased in EPO-treated mice and the decreased NAD+ level with the loss of EPO activity in EpoRaP2KO mice (Fig. 6C). PGC-1α acetylation in EPO-treated mice was decreased (Fig. 6D) further implying EPO modulates the intracellular NAD+ level to regulate PGC-1α activity. The increased intracellular NAD+ level in EPO-stimulated M-adipocytes and S-WAT from EPO-treated mice and the decreased NAD+ level with the loss of EPO activity in EpoRaP2KO mice (Fig. 6D) further imply EPO modulates the intracellular NAD+ level to activate Sirt1 activity. To elucidate further the role of Sirt1 in PPARα signaling in adipocytes, we treated primary adipocytes with a PPARα agonist GW7647, without or with Sirt1 knockdown. As shown in Fig. 5E, GW7647 treatment induced the expression of mitochondrial genes and BAT fat–enriched genes in M-adipocytes (Fig. 6E). However, the induction of these genes, including PPARα targets Pgc1a, Ucp1, Ucp3, and Cpt1a, was significantly lower when Sirt1 was knocked down using siRNA (Supplementary Fig. 3D) in adipocytes (Fig. 6E). Furthermore, Sirt1 knockdown attenuated the EPO-stimulated increase in BAT-enriched factors and mitochondrial genes (Fig. 6F and G) and in
basal OCR and uncoupled OCR and fatty acid oxidation (Fig. 6H and I), indicating Sirt1 mediates EPO activity in adipocyte metabolism. Together, these data suggest Sirt1 cooperates with PPARα to respond to EPO to increase metabolic activity and promote brown features in WAT, leading to protection against DIO and its associated metabolic syndrome.

**DISCUSSION**

Beyond the erythroid-specific effect in the erythrocyte production, EPO has gained interest because of its various nonhematopoietic activities (40,41). These EPO activities were attributed to EPO response in multiple tissues, including pancreatic β-cells, skeletal muscle, and brain. Here, we engineered mice with fat tissue–specific deletion of EpoR to exclude the multiple tissue influence of EPO on energy metabolism and demonstrated that endogenous EPO activity in WAT contributes importantly to protection from obesity and associated metabolic syndromes. Metabolic disturbance due to the imbalance between fat storage and energy expenditure causes the body to store more fat and burn less energy. We demonstrated that targeted deletion of EpoR in adipocyte tissue shows a disproportionate increase in body fat and decreases in whole-body VO2 and physical activity, with no change in food intake. This phenotype illustrates the importance of WAT in the endogenous metabolic response to EPO compared with other nonhematopoietic tissues. Furthermore, the increase in body weight gain in male adipocyte-specific EpoR KO is comparable to that observed in mice with EpoR deleted in nonhematopoietic tissue (9), suggesting that the loss of EPO signaling in WAT contributes to much of the increase in fat mass observed with loss of EpoR in nonhematopoietic tissue. In contrast, WT mice treated with EPO become leaner, with a decrease in accumulated body fat and an increase in activity. Although EPO treatment can also reduce food intake in WT mice, our paired-feeding experiments demonstrated that reduction in food intake is not sufficient to account for the increase in metabolic activity and the oxidative metabolism observed in WAT with EPO treatment. Collectively, these findings reveal insights into EPO effects on the molecular mechanisms of obesity and related metabolic diseases and point to the therapeutic potential of EPO beyond its known protective effect from tissue ischemia.

Energy metabolism imbalance in multiple tissues, including fat, has been recognized as an inducer of obesity and its associated metabolic disorders (42,43). Mitochondrial dysfunction and the resultant decreased lipid metabolism in WAT and excessive influx of fatty acids and
triglycerides in other tissues lead to the excessive fat accumulation and development of obesity-induced insulin resistance and glucose intolerance preceding type 2 diabetes (34,37,38). However, reduced mitochondrial number and activity in WAT in the obese state can be restored to reduce obesity in animal models (44,45). Increased expression of brown adipocyte–enriched factors, such as PRDM6 and PGC-1α in WAT, can increase mitochondrial biogenesis and metabolic activity to prevent obesity and metabolic diseases. Therefore, increasing these BAT-enriched factors may be a plausible strategy for the treatment of obesity. In our study, EPO increased mitochondrial metabolic activity and promoted expression of brown fat–associated genes, such as PRDM16, PGC-1α, and UCPs, possibly through activation of PPARα, especially in inguinal WAT. Our observations suggest a mechanism by which EPO activates metabolic coregulators, PPARα, PGC-1α, and PRDM16, and other factors to contribute to the appearance of brown features, including increased mitochondrial content, oxidative respiration, and fatty acid oxidation in WAT. The transcriptional network by which EPO regulates PPARα, PRDM16, and PGC-1α remains to be determined. We should note that a differential EPO response was found among the various fat depots to EPO treatment in WT mice, along with corresponding opposing changes in adipocyte-specific EpoR KO mice, that reflects the response to endogenous EPO. In contrast to inguinal WAT, epididymal WAT showed an EPO-induced increase only in the expression of UCP3, PPARα, and PGC-1α, whereas BAT exhibited no significant change in BAT-associated gene expression. Among the other fat depots, perianal WAT showed an expression pattern analogous to inguinal WAT, whereas mesenteric WAT showed changes that more closely resembled epididymal WAT, possibly indicating various origins of these fat depots and different physiological function.

Activation of Sirt1 can improve metabolic dysregulation, insulin resistance, and glucose intolerance by enhanced β-oxidation of free fatty acids and promotes mitochondrial biogenesis in obese mice in an NAD+ dependent way (28,29,46,47). NAD+ and NADH metabolism is important in oxidative metabolism and energy homeostasis, has been linked to protection against dietary obesity, and is a therapeutic target for associated metabolic diseases (48–50). Here, we reveal that EPO elevated the intracellular NAD+ level in white adipocytes. In contrast, the loss of EpoR in WAT leads to the opposite effect. This increase in NAD+ leads to changes in PGC-1α acetylation, likely due to an increase in Sirt1 activity. Furthermore, we show that EPO increases oxidative respiration and fatty acid oxidation in a Sirt1-dependent way in adipocytes, which may improve energy homeostasis and reduce lipid content in adipocytes in a model of obesity induced by an HFD. On the other hand, fatty acid synthesis or export may be altered because Sirt1 has been shown to regulate both processes (51,52). Our results also suggest that Sirt1 may be required for PPARα activity in the adipocyte response to EPO. The mechanism remains to be elucidated. It is likely that EPO may activate other sirtuins via the increased NAD+ level. However, we clearly show that EPO promoted brown remodeling of WAT and stimulated metabolic activity dependent on Sirt1.

Note that involvement of indirect EPO effects during DIO cannot be entirely excluded and that further investigation is warranted. Our study demonstrates for the first time that the loss of EPO/EpoR signaling in adipose tissue is sufficient to develop metabolic syndrome phenotype, including obesity, glucose intolerance, and insulin resistance, and supports the idea that EPO may contribute to protection against obesity and its associated metabolic syndromes.

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L.W. designed and conducted the experiments and wrote the manuscript. R.T., L.D., and H.R. conducted experiments and data analyses. H.W. analyzed data and provided EpoRlox/lox mice. J.B.K. assisted with the VO2 and fatty acid oxidation experiments. C.T.N. conceived the project, designed the experiments, analyzed data, and wrote the manuscript. C.T.N. is the guarantor of this work and, as such, had full access to all the data in the study and takes responsibility for the integrity of the data and the accuracy of the data analysis.

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REFERENCES


