

CLINICAL TRIALS AND OBSERVATIONS

Phase 2 study of azacytidine plus sorafenib in patients with acute myeloid leukemia and *FLT-3* internal tandem duplication mutation

Farhad Ravandi,¹ Mona Lisa Alattar,¹ Michael R. Grunwald,² Michelle A. Rudek,³ Trivikram Rajkhowa,² Mary Ann Richie,¹ Sherry Pierce,¹ Naval Daver,¹ Guillermo Garcia-Manero,¹ Stefan Faderl,¹ Aziz Nazha,¹ Marina Konopleva,¹ Gautam Borthakur,¹ Jan Burger,¹ Tapan Kadia,¹ Sara Deltasala,¹ Michael Andreeff,¹ Jorge Cortes,¹ Hagop Kantarjian,¹ and Mark Levis²

¹Department of Leukemia, University of Texas MD Anderson Cancer Center, Houston, TX; and ²Division of Hematological Malignancies, ³Division of Chemical Therapeutics, Johns Hopkins Sidney Kimmel Cancer Center, Baltimore, MD

Key Points

- Azacytidine and sorafenib are effective in patients with relapsed and refractory *FLT3*-mutated AML.

Patients received 5-azacytidine (AZA) 75 mg/m² intravenously daily for 7 days and sorafenib 400 mg orally twice daily continuously; cycles were repeated at ~1-month intervals. Forty-three acute myeloid leukemia (AML) patients with a median age of 64 years (range, 24-87 years) were enrolled; 37 were evaluable for response. *FMS*-like tyrosine kinase-3 (*FLT3*)-internal tandem duplication (ITD) mutation was detected in 40 (93%) patients, with a median allelic ratio of 0.32 (range, 0.009-0.93). They had received a median of 2 prior treatment regimens (range, 0-7); 9 had failed prior therapy with a *FLT3* kinase inhibitor. The response rate was 46%, including 10 (27%) complete response with incomplete count recovery (CRi), 6 (16%) complete responses (CR), and 1 (3%) partial response. The median time to achieve CR/CRi was 2 cycles (range, 1-4), and the median duration of CR/CRi was 2.3 months (range, 1-14.3 months). Sixty-four percent of patients achieved adequate (defined as >85%) *FLT3* inhibition during their first cycle of therapy. The degree of *FLT3* inhibition correlated with plasma sorafenib concentrations. *FLT3* ligand levels did not rise to levels seen in prior studies of patients receiving cytotoxic chemotherapy. The combination of AZA and sorafenib is effective for patients with relapsed AML and *FLT3*-ITD. This trial was registered at clinicaltrials.gov as #NCT01254890. (*Blood*. 2013;121(23):4655-4662)

Introduction

Internal tandem duplication (ITD) mutations in the juxtamembrane domain of the *FMS*-like tyrosine kinase-3 (*FLT3*) gene have been detected in ~25% of patients with acute myeloid leukemia (AML) in several large, retrospective studies.¹ These mutations are associated with leukocytosis, higher marrow blast percentage, and poor outcomes.²⁻⁴ Their presence is associated with an increased risk of relapse and a shorter overall survival (OS).^{2,5} This negative impact also exists in the setting of relapse; patients harboring the *FLT3*-ITD mutation have a significantly worse outcome in first relapse compared with those without the mutation.⁶⁻⁸

Several novel agents targeting the *FLT3* kinase have shown promising activity in patients with AML and mutated *FLT3*.⁹⁻¹¹ These small molecule tyrosine kinase inhibitors (TKIs), such as lestaurtinib (CEP-701), midostaurin (PKC412), and tandutinib (MLN518), have been evaluated either as monotherapy or in combination with cytotoxic chemotherapy.^{11,12} These agents block the autophosphorylation of the *FLT3* kinase, leading to inhibition of cell proliferation and induction of apoptosis. *FLT3* inhibitors have been particularly active in patients with *FLT3*-ITD mutations.¹³⁻¹⁵ Sorafenib (Nexavar), another small molecule *FLT3* TKI, induces pronounced apoptosis in vitro in blast cells from AML patients.¹² This promotion of programmed cell death is accompanied by

extracellular signal-regulated kinase (ERK)1/2 inactivation and caspase-independent downregulation of MCL-1.¹² Sorafenib is an orally active multikinase inhibitor with potent activity against *FLT3* and the Raf/ERK/mitogen-activated protein kinase pathway.¹⁶ In phase 1 clinical trials, sorafenib significantly reduced the number of leukemia blasts in the peripheral blood (PB) and bone marrow (BM) in patients with the *FLT3*-ITD mutation.^{12,17,18} Melzelder and coauthors¹⁹ have also reported that sorafenib induced remission and facilitated allogeneic stem cell transplant in patients with refractory AML with the *FLT3*-ITD mutation.

In vitro studies have demonstrated that *FLT3* kinase inhibitors are synergistic with cytotoxic agents when used simultaneously with or immediately after chemotherapy.²⁰ A number of clinical trials combining *FLT3* TKIs with traditional chemotherapy have been reported.^{14,21} In a phase 1/2 study of idarubicin, high-dose cytarabine, and sorafenib in younger patients (median age of 53 years) with AML, all patients with mutated *FLT3* achieved complete response (CR) or CR with incomplete platelet recovery (CRp).²¹ Mutant *FLT3* was suppressed in all 10 patients evaluated, with fivefold greater suppression of mutant *FLT3* compared with wild-type *FLT3* on plasma inhibitory assays.²¹ This study demonstrated that sorafenib could be safely administered with chemotherapy and result in

Submitted January 24, 2013; accepted April 8, 2013. Prepublished online as *Blood* First Edition paper, April 23, 2013; DOI 10.1182/blood-2013-01-480228.

F.R. and M.L.A. contributed equally to this study.

The online version of this article contains a data supplement.

The publication costs of this article were defrayed in part by page charge payment. Therefore, and solely to indicate this fact, this article is hereby marked "advertisement" in accordance with 18 USC section 1734.

© 2013 by The American Society of Hematology

potent inhibition of FLT3 signaling. However, a German multicenter, randomized, placebo-controlled, double-blind trial of chemotherapy with or without sorafenib in older (>60 years) patients with AML did not show an improvement in CR rate, event-free survival, or OS for the patients receiving sorafenib, and sorafenib was associated with increased toxicity.²²

It has been suggested that elevated FLT3 ligand (FL) levels related to aplasia induced by chemotherapy regimens can block the action of TKIs such as sorafenib on the FLT3 kinase or could promote the survival of FLT3-ITD blasts by augmenting signaling from the mutant receptor.^{11,23,24} We hypothesized that the combination of 5-azacytidine (AZA) with sorafenib may be associated with less resistance by promoting lower levels of FL than traditional chemotherapy regimens. The objectives of this study were to determine the feasibility, safety and efficacy of combining sorafenib with AZA and to determine whether reduced levels of FL associated with AZA will translate to higher responses.

Materials and methods

Study design and eligibility

This phase 1/2 single-arm study was conducted in patients with refractory or relapsed AML from January 2011 to September 2012. The study was approved by the University of Texas–MD Anderson Cancer Center Institutional Review Board, and all participating patients signed an informed consent document in accordance with the Declaration of Helsinki. Patients were ≥ 18 years of age, had a diagnosis of AML, and needed to have failed prior induction therapy or relapsed after achieving a response to prior therapy; patients >60 years of age who refused standard induction therapy or were deemed unfit for it were also eligible to participate. Other requirements for study entry included an Eastern Cooperative Oncology Group (ECOG) performance status ≤ 2 , adequate hepatic (serum total bilirubin $\leq 1.5 \times$ upper limit of normal [ULN]), alanine aminotransferase, and aspartate aminotransferase $\leq 2.5 \times$ ULN), renal (serum creatinine $\leq 1.5 \times$ ULN), and pancreatic (amylase and lipase $\leq 2 \times$ ULN) function. Exclusion criteria included patients with a diagnosis of acute promyelocytic leukemia, known HIV infection, or active viral hepatitis (B or C); evidence of a bleeding diathesis or coagulopathy; and a history of solid organ transplant. Nursing and pregnant females were excluded. In addition, patients were excluded if there was a known history of congestive heart failure greater than class II New York Heart Association, uncontrolled hypertension, malignant disease of the central nervous system, or advanced malignant hepatic tumors. Presence of the FLT3-ITD mutation was not a necessary inclusion criteria but such patients were actively sought for inclusion.

Treatment regimen

Initially, to assess the safety of the combination, patients were treated with a low dose of sorafenib (200 mg orally twice daily) in combination with AZA. After 3 patients were evaluated without any significant toxicity observed, all patients were enrolled in the phase 2 part of the study and received sorafenib 400 mg orally twice daily. They received AZA 75 mg/m²/day subcutaneously or intravenously daily for 7 days per cycle without any interruptions; in addition, sorafenib was given orally twice daily continuously every cycle starting on day 1 of AZA. Cycles were repeated every 4 to 8 weeks at the discretion of the treating physician. In general, and in the absence of significant toxicity, sorafenib was continued without interruption even if the AZA administration was delayed (eg, due to cytopenias). A minimum of 1 full cycle (defined as AZA for 7 days and sorafenib for 28 days) was required for a patient to be eligible for evaluation of efficacy. Any patient who received ≥ 1 dose of either drug was considered evaluable for toxicity. After the first 3 cycles of therapy, subsequent cycles of AZA were administered only if the absolute neutrophil count was $\geq 1 \times 10^9/L$, and platelets were $\geq 30 \times 10^9/L$ in the absence of residual leukemia in the BM.

If prolonged myelosuppression (>60 days) with evidence of a hypocellular marrow (marrow cellularity <5% without evidence of leukemia) was observed, lower doses of AZA could be administered if approved by the principal investigator. Concomitant use of strong CYP3A4 inducers within 7 days of initiating dosing was prohibited. However, the use of azoles such as voriconazole and posaconazole was not prohibited. Inpatient dose escalation with shortening the interval between AZA courses was allowed if it was felt to be in the best interest of the patient (eg, if there was a perceived or real rise in peripheral blasts). All patients received antimicrobials, supportive care, and transfusions of blood products according to the institutional guidelines.

Laboratory correlative studies

Whole blood at designated time points was collected into heparinized vacuum tubes, and after centrifugation, the plasma was stored frozen. In vivo FLT3 inhibition was determined using a surrogate ex vivo assay, the plasma inhibitory activity (PIA) assay, as previously described, using the Molm-14 cell line (human AML with a naturally occurring FLT3-ITD mutation).²⁵ First, the frozen plasma samples were thawed and clarified by centrifugation. For each sample, $\sim 3 \times 10^6$ Molm-14 cells were suspended in 1 mL of patient plasma. The suspension was incubated for 1 hour at 37°C. The cells were subsequently washed twice with ice-cold phosphate-buffered saline and lysed, and immunoprecipitation and phospho-FLT3 (P-FLT3) immunoblotting was carried out as described. Densitometry was used to calculate the relative level of P-FLT3 in each plasma sample. The density of each band was expressed as a percentage of the density of the baseline plasma band for each patient or the density of the control plasma band.

FL concentrations in plasma samples were determined using an enzyme-linked immunosorbent assay kit obtained from R & D Systems (Minneapolis, MN) as previously described.²³

Sorafenib and sorafenib *N*-oxide concentrations were determined using a validated liquid chromatography/tandem mass spectrometry method, as previously described.²⁶

FLT3-ITD mutation status was determined in DNA from initial, follow-up, and relapsed unsorted BM aspirate samples by a polymerase chain reaction–based method with an analytical sensitivity of 1% to 2% mutation-bearing cells. FLT3 allele burden was determined by ratio of the area under the mutated and unmutated polymerase chain reaction amplicon peaks detected following capillary electrophoresis on 3100 or 3130 Genetic Analyzers (Applied Biosystems, Foster City, CA). Manual 400-cell differential performed on smears and multicolor flow cytometry on aspirate samples was used to track the levels of residual leukemia blasts. When leukemic blasts were detected, FLT3 mutant ratios²³ were normalized to blast count.

Response assessment

CR was defined by the presence of <5% blasts in the BM, with $>1 \times 10^9/L$ neutrophils and $>100 \times 10^9/L$ platelets in the PB with no detectable extramedullary disease.²⁷ Patients who met the above criteria but had neutrophil or platelet counts less than the stated values were considered to have achieved CRi (CR with incomplete recovery of PB counts). Partial response (PR) required all of the hematologic values for a CR but with a decrease of $\geq 50\%$ in the percentage of blasts to 5% to 25% in the BM aspirate. CR duration was calculated from the time of achieving CR until relapse. Relapse was defined by the recurrence of >5% blasts in BM aspirate not related to count recovery or the development of extramedullary disease. OS was calculated from the time of diagnosis until death. Patients were censored at the time of last contact with healthcare professionals at our institution.

Statistical analysis

Survival curves were plotted by the Kaplan-Meier method and compared using the log-rank test. Differences in subgroups by different covariates were evaluated using the χ^2 test for nominal values and the Mann-Whitney U and Fischer's exact test for continuous variables. In the PIA analysis, the log-rank (Mantel-Cox) test was used to evaluate the survival curves for statistical significance.

Table 1. Patient characteristics

Characteristics	No. of patients (%)
Total enrolled	43
Median age in years [range]	64 [24-87]
Male: female	22 (51):21 (49)
Median WBC × 10 ⁹ /L [range]	8.9 [0.4-107.8]
Median percentage blasts in PB [range]	51 [0-99]
Median percentage blasts in BM [range]	66 [2-98]
Cytogenetics	
Diploid	19 (44)
Chromosome 5 or 7/complex	11 (26)
Miscellaneous	13 (30)
Median no. prior therapy [range]	2 [0-7]

WBC, white blood cell count.

Results

Patient characteristics

Between January 2011 and September 2012, a total of 43 patients with AML meeting the eligibility criteria were enrolled. Six patients were inevaluable as they discontinued therapy before response assessment at 1 month. Pretreatment characteristics of the evaluable patients are summarized in Table 1. Their median age was 64 years (range, 24-87 years). Karyotype included 19 (44%) patients with diploid cytogenetics, 11 (26%) with chromosome 5/7 or complex cytogenetic abnormalities, and 13 (30%) with other miscellaneous chromosomal abnormalities. *FLT3*-ITD was detected prior to the initiation of treatment in 40 of 43 (93%) patients with a median allelic burden of 0.32 (range, 0.009-0.93) in those with *FLT3*-ITD. Patients had received a median of 2 prior treatments (range, 0-7) including 16 (37%) patients who had received ≥3 prior regimens. Among the 37 evaluable patients, 6 had no prior therapy, 12 were primarily refractory to their previous induction regimen, and 19 had relapsed after prior therapy. Nine patients had failed prior therapy with *FLT3* kinase inhibitors (5 with AC220, 1 with PKC412, and 6 with sorafenib, either as monotherapy or together with chemotherapy or plerixafor); 3 had failed 2 prior *FLT3* inhibitors (Figure 1). Seven patients had received a prior allogeneic stem cell transplant.

Response and outcomes

The overall composite response rate among the evaluable patients was 46% (Table 2), including 6 (16%) with CR, 10 (27%) with CRi, and 1 (3%) with PR (in this patient, BM blasts declined from 66% to 6% with normalization of blood counts). Overall, patients received a median of 3 (range, 1-18) treatment cycles. Among the responders, the median number of cycles to response was 2 (range, 1-4), and the median time to achieving response was 2 months (range, 1-4.6 months). Response rate was higher in previously untreated patients (4 of 6 [67%] patients including 1 with CR, 2 with CRi, and 1 with PR) compared with primary refractory patients (7 of 12 [58%] patients including 3 CR and 4 CRi) and compared with relapsed patients (6 of 19 [32%] patients including 2 CR and 4 CRi; *P* = not significant). The median duration of response was 2.3 months (range, 1-14.3 months; Figure 2A). Six patients have proceeded to allogeneic stem cell transplant, including 2 non-responders. With a median follow-up of 5.5 months (range, 1-16 months), the median OS for all evaluable patients is 6.2 months (Figure 2B) with a longer OS of 7.8 months for responders vs 6.0 months for nonresponders (*P* = .01). Of the 16 responding patients, 3 relapsed after 1 month in remission, an additional 6 relapsed after 3 months in remission, 4 underwent stem cell transplant, and 7 remain in CR at last follow-up. The median event free survival was 3.8 months (range, 1.0-16.4 months; Figure 2C). There was no statistical difference for the duration of response or OS between previously untreated, primary refractory, or relapsed patients (data not shown).

Among the patients who had received prior therapy with *FLT3* kinase inhibitors, 3 with prior sorafenib therapy (including 1 patient who had also received prior PKC412 and 1 patient who had also received prior AC220) achieved CR (1) or CRi (2), with a CR/CRi duration of 1.7, 5.4, and 5.6+ months. One died in CRi at 1.7 months and 1 relapsed after 5.4 months, with the third patient continuing in CR. Six other patients with prior *FLT3* kinase therapy did not respond.

Safety and toxicity

The majority (53%) of patients experienced grade <3 adverse effects attributable to sorafenib (Table 3). The most common grade ≥3

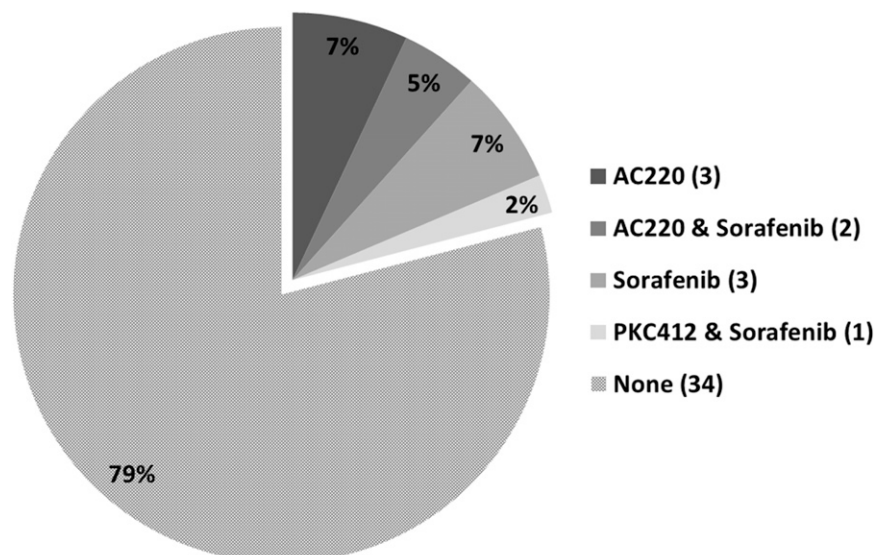


Figure 1. Previous *FLT3* kinase inhibitor exposure.

Table 2. Overall response

Parameter	N = 37 (%)
Median no. cycles received [range]	3 [1-18]
Median follow-up in weeks [range]	24 [5-71]
Response	
CR	6 (16)
Cri	10 (27)
PR	1* (3)
NR	18 (49)
Died	2 (5)
Median no. cycles to response [range]	2 [1-7]
4-wk mortality	1 (3)
8-wk mortality	1 (3)

*Patient's BM blast percentage fell from 66% to 6% with normalization of peripheral counts.

adverse events were thrombocytopenia, neutropenia, anemia, and neutropenia with fever or infection. Only 4 of 18 (22%) of patients with grade ≥ 3 hematologic adverse effects had normal PB counts prior to initiation of the study. Only 1 patient discontinued treatment because of a grade 4 nonischemic (proven by cardiac catheterization) cardiomyopathy. Although hepatotoxicity was seen, most instances were grade 1 or 2. Fatigue was the most common adverse event overall (47%) but was most often grade 1 (29%) in severity. Overall, the 4-week mortality was 9% (4 of 43 patients enrolled), and the 8-week mortality was 16% (7 of 43). Among

patients evaluable for response, the 4-week mortality was 3% (1 of 37), and the 8-week mortality was 3% (1 of 37), with all 6 deaths occurring during the study treatment resulting from infection. Reduction of the dose and/or duration of AZA (to 75 mg/m² daily for 5 days [n = 3] or 50 mg/m² daily for 5 days [n = 1]) occurred in 4 patients due to gastrointestinal toxicity or myelosuppression that was deemed to be related to therapy.

Correlation of response with FLT3 kinase inhibition and pharmacokinetic studies

Among the patients included in the clinical analysis, there were 22 patients for whom both clinical outcome data and plasma samples spanning ≥ 1 cycle of therapy were available. These 22 patients were included in our analysis of FLT3 inhibition and FL concentrations. As predicted, the relatively low-intensity therapy with AZA resulted in minimal or no increase in plasma FL levels (Figure 3A). The levels of FL observed following intensive chemotherapy (data from the Cephalon 204 trial of chemotherapy followed by lestaurtinib²⁴) are shown for comparison. Mean FL concentrations at baseline and at the end of cycles 1, 2, and 3 were 8, 35, 149, and 65 ng/mL, respectively. Median FL levels at the same time points were 0, 16, 24, and 46 ng/mL, respectively. In vivo FLT3 inhibition, as determined by the PIA assay (Figure 3B), was highly variable. Sorafenib concentrations, determined in a limited subset of these samples, showed a relatively tight correlation with the degree of in

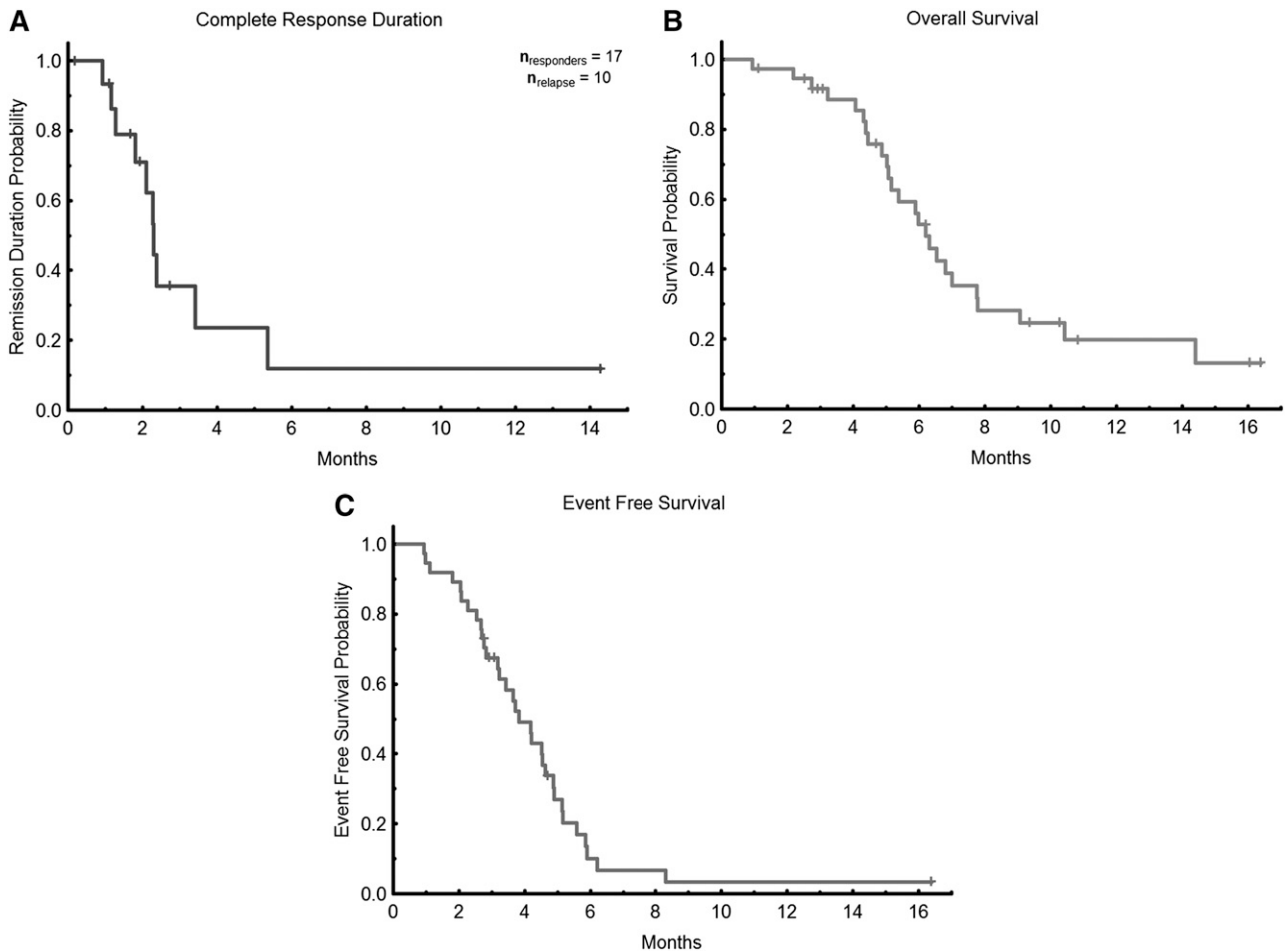
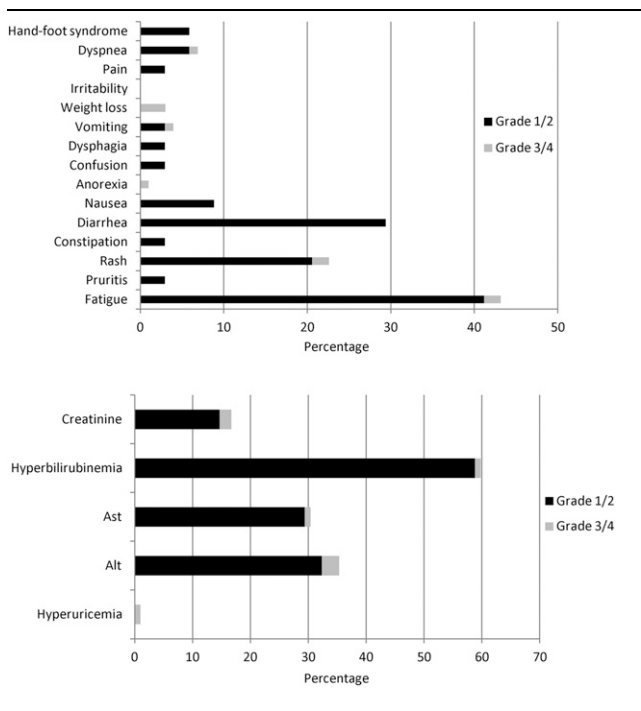


Figure 2. Patient outcomes. (A) Remission duration in responders. (B) OS. (C) Event-free survival.

Table 3. Adverse events and chemical abnormalities



Adverse events (top) and chemical abnormalities (bottom).
 *Hypertension in 1 patient (3%), typhlitis in 1 patient (3%), and pleural effusion in 2 patients (6%).

vivo *FLT3* inhibition (Figure 4). This suggests that the variability in *FLT3* inhibition in vivo was primarily due to variable sorafenib levels. In vitro studies of other *FLT3* inhibitors suggest that *FLT3* inhibition to <15% of baseline activity is necessary for optimal cytotoxicity. Therefore, this level was used as the threshold for adequate *FLT3* inhibition in the current trial. In this study, 64% of patients achieved *FLT3* inhibition to >15% of baseline at some point during their first cycle of therapy. Median survival in patients who achieved this degree of inhibition was 238 days. Median survival in patients who did not reach this level was 154 days. The difference in survival did not attain statistical significance ($P = .13$; Figure 5).

Correlation of response with *FLT3*-ITD allele burden

The *FLT3*-ITD burden prior to therapy was not significantly different among the responders and nonresponders. Similarly, at 1 to 3 months after therapy, the levels of *FLT3*-ITD allele burden were similar between responders and nonresponders (Figure 6).

Discussion

The treatment of patients with primary refractory or relapsed AML remains unsatisfactory.²⁸ *FLT3*-ITD mutations are associated with an inferior response to salvage therapy and a poor outcome at relapse.^{6,7,29} A number of *FLT3* kinase inhibitors are in development and have been evaluated in patients with *FLT3* mutated AML. A large randomized trial combining lestaurtinib with chemotherapy did not improve the outcome of patients with *FLT3* mutated AML in first relapse.²⁴ However, the investigators noted that patients who achieved adequate target inhibition were more likely to respond

and benefit from the addition of lestaurtinib. Furthermore, the authors suggested that elevated levels of FL may be associated with resistance to lestaurtinib.^{23,24}

Sorafenib has demonstrated activity in patients with relapsed and refractory AML.¹⁹ However, responses are of limited duration, and a number of mechanisms of resistance to *FLT3* inhibition have been demonstrated in preclinical and clinical studies.^{23,30,31} We have previously demonstrated a high response rate to the combination of sorafenib with cytarabine and anthracycline-based induction therapy in formerly untreated younger patients with AML.²¹ The responses were of limited duration, with more than half of patients with the *FLT3*-ITD mutation relapsing at a median follow-up of 9 months. The median CR duration was 8.5 months.³² We hypothesized that a regimen combining sorafenib with AZA was less likely to be associated with elevated FL levels, and hence, less likely to be associated with development of resistance to *FLT3* inhibition.

Furthermore, we hypothesized that the combination may be associated with differentiation of the leukemic cells. Normal myeloid differentiation involves CCAAT/enhancer-binding protein α (C/EBP α) and PU.1 expression.^{33,34} *FLT3*-ITD mutations lead to differentiation arrest by inhibiting expression of these transcription factors compared with wild-type *FLT3* (*FLT3*-WT).^{35,36} Targeting of transcription factors by *FLT3*-ITD signaling has been confirmed by microarray expression profiling.³⁷ Recent mouse models have

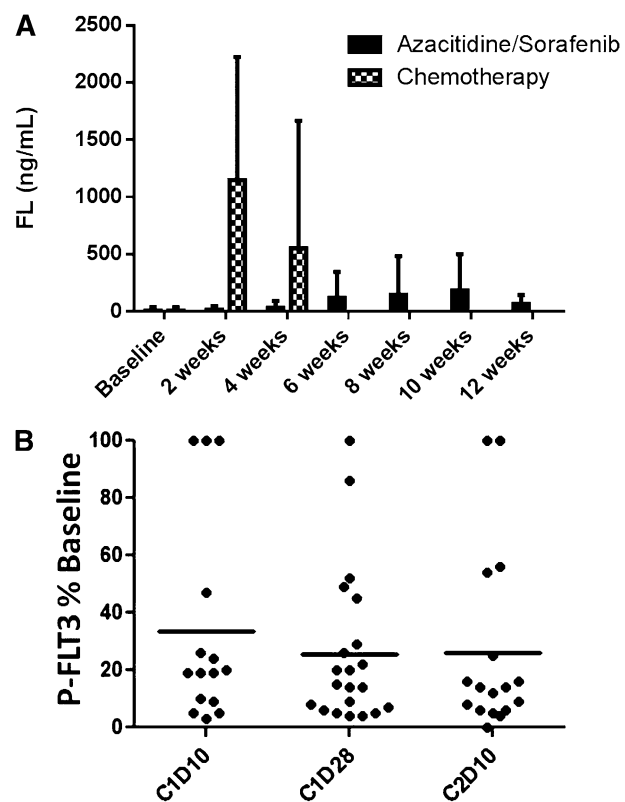


Figure 3. FL levels and in vivo *FLT3* inhibition. (A) Plasma FL levels determined by enzyme-linked immunosorbent assay using samples collected from the current trial of 5-azacitidine/sorafenib (solid columns). For comparison (hatched columns) is shown FL levels from plasma samples collected from relapsed/refractory *FLT3*-ITD AML patients on the Cephalon 204 trial²⁴ 2 or 4 weeks after starting salvage chemotherapy (mitoxantrone/etoposide/cytarabine). (B) *FLT3* inhibition grouped according to treatment time point. PIA assays (see Materials and methods) were performed on individual plasma samples collected during therapy. Densitometry measurements from individual *FLT3* PIA assays are plotted against time point. The solid lines indicate the mean level of P-*FLT3* for the patient group at each time point. C, cycle; D, day.

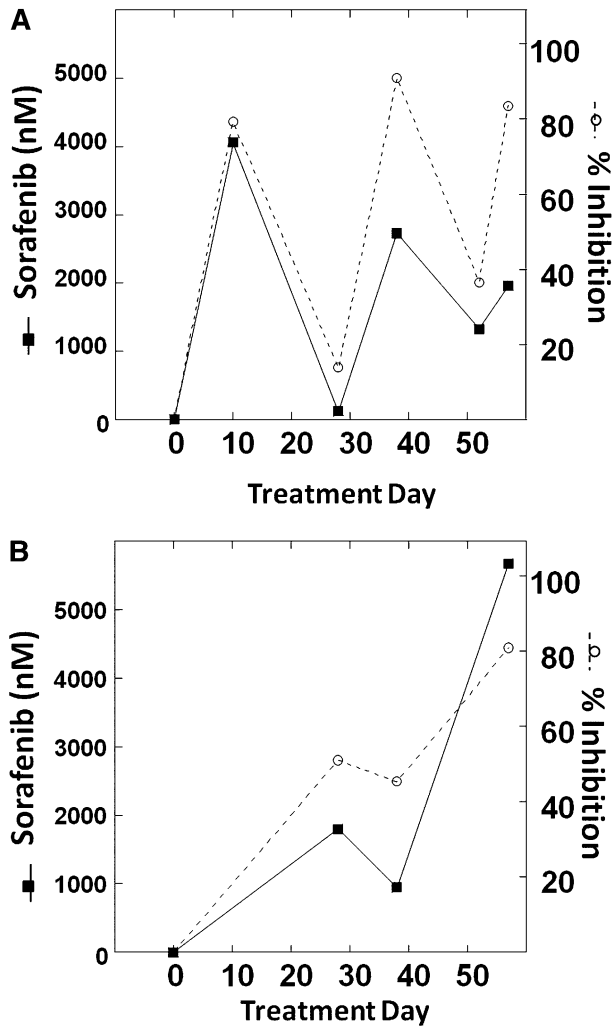


Figure 4. Plasma levels of sorafenib versus in vivo FLT3 inhibition. Plasma levels of sorafenib at individual time points from 2 different patients are compared with the results of PIA assays from those same time points. The PIA results are expressed inversely, as percent inhibition. (A) and (B) represent results from 2 different patients.

shown that FLT3 inhibitors like lestaurtinib induce differentiation in *FLT3* mutated AML cells through suppression of C/EBP α .³⁶ In vitro models of FLT3 inhibition have demonstrated that in the BM microenvironment, FLT3 inhibition does not induce immediate apoptosis. Rather, FLT3 inhibition promotes differentiation, and this differentiation is dependent on the expression of functional C/EBP α .³⁸ Others have demonstrated that activating mutations of *FLT3* block C/EBP α function by ERK1/2 mediated phosphorylation of its serine 21 residue, leading to the differentiation block in the leukemia cells.³⁹ The demethylating agents decitabine and AZA also have clearly demonstrable differentiation-inducing properties.⁴⁰⁻⁴² AZA, as well as histone deacetylase inhibitors, has been shown not only to maintain normal hematopoietic stem cell renewal but also to induce terminal differentiation in AML cells.^{41,43} We hypothesized that the combination of FLT3 inhibition with hypomethylation therapy might be synergistic in inducing differentiation in vivo. We saw clinical evidence of differentiation in several patients during the course of therapy as demonstrated by a single case provided in the supplemental Materials on the *Blood* website.

Several mechanisms of resistance to FLT3-TKIs have been proposed. Most prominently, the acquisition of mutations in the

ATP-binding pocket of FLT3 inducing clinical resistance to FLT3 inhibitors has now been reported.^{11,30} We did not systemically evaluate the patients at relapse for the development of such mutations. Other mechanisms of resistance include activation of compensatory survival pathways (eg, through activating *NRAS* mutations) that render leukemic cells independent of FLT3,⁴⁴ as well as enhanced activation of signal transducer and activator of transcription pathways and overexpression of survivin.⁴⁵ Last, autocrine and/or paracrine FLT3 receptor stimulation via FL can lead to resistance to FLT3 inhibitors. Increased FL expression has been demonstrated in response to standard myelosuppressive chemotherapy.²³

In prior studies, FL levels have been demonstrated to rise markedly during the aplasia caused by chemotherapy.^{23,46,47} In this study of AZA and sorafenib, FL levels did not rise to the levels seen in prior studies of FL levels in patients receiving cytotoxic chemotherapy. High FL levels have been shown to impede the efficacy of FLT3 inhibitors including sorafenib.²³ Therefore, it has been postulated that treatment regimens that do not cause abrupt increases in FL levels may create a more favorable environment for FLT3 inhibition.²³ In the current study, it appears that low FL levels are not sufficient to foster optimal activity of the FLT3 inhibitor sorafenib. As noted above, other factors may have been responsible for suboptimal FLT3 inhibition in some patients.

In the current study, FLT3 inhibition was variable. There was a sizable subset of patients who achieved excellent FLT3 inhibition and sustained this inhibition. While not statistically significant, our survival curve shows a trend toward improved survival in patients with adequate FLT3 inhibition during cycle 1. The small number of patients with evaluable clinical outcomes and sufficient plasma samples may have been an important limitation to this survival analysis. On the other hand, 36% of patients did not meet the threshold for FLT3 inhibition during the first month of therapy. Moreover, when FLT3 inhibition occurred, it was not always sustained. There are several possible explanations for the failures to achieve adequate FLT3 inhibition in some cases. First, sorafenib concentrations can be variable in patients receiving this medication, possibly due to age, gender, and effects of concomitant medications (chemotherapy or CYP3A4 inhibitors) on the metabolism of the drug.^{25,26,48} Variability in FLT3 inhibition as assessed by the PIA assay has previously been noted with sorafenib.²⁶ Second, some

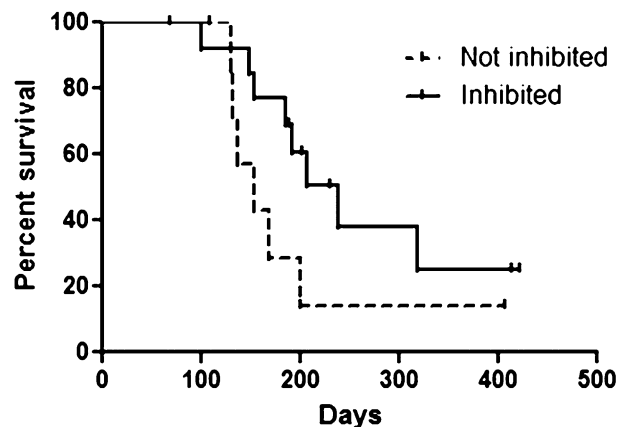


Figure 5. Survival grouped by PIA assay results. The Inhibited group consisted of 14 patients whose P-FLT3 levels were <15% of baseline at ≥ 1 point during the first cycle of therapy. The Not inhibited group consisted of 8 patients whose P-FLT3 levels were never less than 15% of baseline during the first cycle of therapy. Although there was a trend toward improved survival in the Inhibited group, this finding did not reach statistical significance.

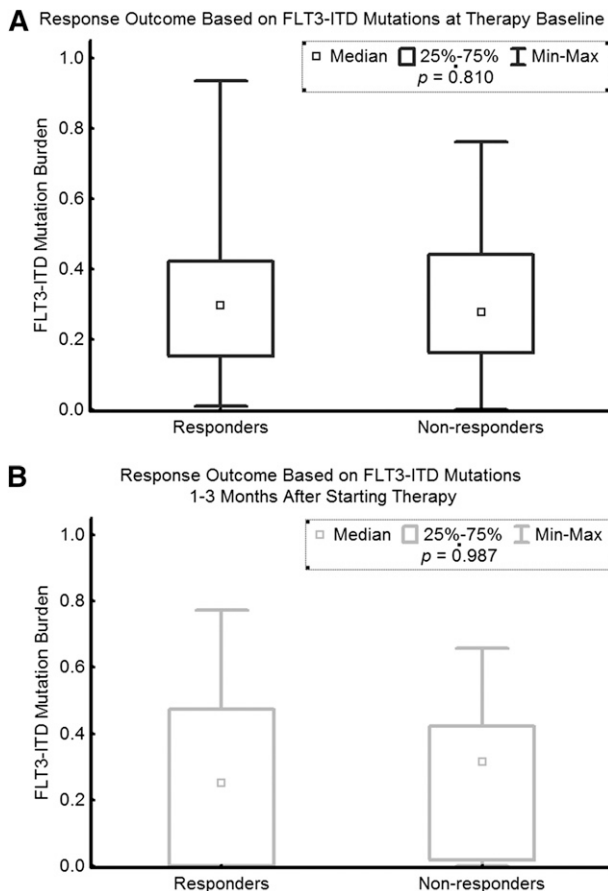


Figure 6. Outcomes by FLT3-ITD allele burden. (A) Baseline FLT3-ITD mutation burden of responders vs nonresponders. (B) FLT3-ITD mutation burden of responders versus nonresponders at 1 to 3 months after treatment initiation.

patients in the current study were administered reduced doses of sorafenib due to toxicity. Finally, the plasma concentration of sorafenib required to inhibit FLT3 is significantly higher than the inhibitory concentration of newer agents such as quizartinib.⁴⁹

Previous studies have reported the feasibility of combining sorafenib and chemotherapy.^{21,37} Serve et al²² randomized 197 elderly patients with AML to receive either placebo or sorafenib with the standard “3+7” induction chemotherapy. The addition of sorafenib did not improve event-free survival or OS and was associated with increased toxicity. Here we demonstrated that the combination of AZA plus sorafenib is effective and well tolerated, producing a high number of responses in patients with relapsed/refractory FLT3-ITD AML, a condition for which

treatment alternatives are limited and associated with poor outcomes. Future studies should evaluate this combination and other combinations of demethylating agents with FLT3 kinase inhibitors both in the relapse and frontline settings.

Acknowledgments

This work was supported by a research grant from Celgene and in part by National Cancer Institute (NCI) Leukemia Specialized Program of Research Excellence grant P50 CA100632. This work was also supported by grants from the NCI (NCI Leukemia Special Program of Research and Education P50 CA100632-06, R01 CA128864) and the American Society of Clinical Oncology (M.L.). M.L. is a Clinical Scholar of the Leukemia and Lymphoma Society. The project described was supported by the Analytical Pharmacology Core of the Sidney Kimmel Comprehensive Cancer Center at Johns Hopkins (National Institutes of Health, National Cancer Institute grants P30 CA006973 and UL1 RR025005 and Shared Instrument grant 1S10RR026824-01).

The content is solely the responsibility of the authors and does not necessarily represent the official views of the National Cancer Institute or the National Institutes of Health.

Authorship

Contribution: M.L.A. collected and analyzed data and wrote the manuscript; M.L., M.R.G., T.R., and M. A. Rudek conducted in vitro experiments, analyzed data, and wrote the manuscript; H.K., N.D., G.G.-M., S.F., M.K., G.B., J.B., A.N., M.A., T.K., and J.C. treated patients on the trial and critically reviewed the manuscript; M. A. Richie, S.P., and S.D. collected and analyzed clinical data; and F.R. designed the research, collected and analyzed the data, and wrote the manuscript.

Conflict-of-interest disclosure: F.R. received research funding and honoraria from Onyx/Bayer and Celgene. G.G.-M. received research funding and honoraria from Celgene. J.C. received research funding from Celgene. M. A. Rudek received research funding from Celgene. The remaining authors declare no competing financial interests.

Correspondence: Farhad Ravandi, Department of Leukemia, Unit 428, The University of Texas MD Anderson Cancer Center, 1515 Holcombe Blvd, Houston, TX 77030; e-mail: fravandi@mdanderson.org.

References

- Nakao M, Yokota S, Iwai T, et al. Internal tandem duplication of the flt3 gene found in acute myeloid leukemia. *Leukemia*. 1996;10(12):1911-1918.
- Kottaridis PD, Gale RE, Frew ME, et al. The presence of a FLT3 internal tandem duplication in patients with acute myeloid leukemia (AML) adds important prognostic information to cytogenetic risk group and response to the first cycle of chemotherapy: analysis of 854 patients from the United Kingdom Medical Research Council AML 10 and 12 trials. *Blood*. 2001;98(6):1752-1759.
- Fröhling S, Schlenk RF, Breitnick J, et al; AML Study Group Ulm. Acute myeloid leukemia. Prognostic significance of activating FLT3 mutations in younger adults (16 to 60 years) with acute myeloid leukemia and normal cytogenetics: a study of the AML Study Group Ulm. *Blood*. 2002;100(13):4372-4380.
- Thiede C, Steudel C, Mohr B, et al. Analysis of FLT3-activating mutations in 979 patients with acute myelogenous leukemia: association with FAB subtypes and identification of subgroups with poor prognosis. *Blood*. 2002;99(12):4326-4335.
- Abu-Duhier FM, Goodeve AC, Wilson GA, et al. FLT3 internal tandem duplication mutations in adult acute myeloid leukaemia define a high-risk group. *Br J Haematol*. 2000;111(1):190-195.
- Ravandi F, Kantarjian H, Faderl S, et al. Outcome of patients with FLT3-mutated acute myeloid leukemia in first relapse. *Leuk Res*. 2010;34(6):752-756.
- Chevallier P, Labopin M, Turlure P, et al. A new Leukemia Prognostic Scoring System for refractory/relapsed adult acute myelogenous leukaemia patients: a GOELAMS study. *Leukemia*. 2011;25(6):939-944.
- Kurosawa S, Yamaguchi T, Miyawaki S, et al. Prognostic factors and outcomes of adult patients with acute myeloid leukemia after first relapse. *Haematologica*. 2010;95(11):1857-1864.

9. Gilliland DG, Griffin JD. The roles of FLT3 in hematopoiesis and leukemia. *Blood*. 2002;100(5):1532-1542.
10. Small D. FLT3 mutations: biology and treatment. *Hematology (Am Soc Hematol Educ Program)*. 2006;2006(1):178-184.
11. Kindler T, Lipka DB, Fischer T. FLT3 as a therapeutic target in AML: still challenging after all these years. *Blood*. 2010;116(24):5089-5102.
12. Zhang W, Konopleva M, Shi YX, et al. Mutant FLT3: a direct target of sorafenib in acute myelogenous leukemia. *J Natl Cancer Inst*. 2008;100(3):184-198.
13. Smith BD, Levis M, Beran M, et al. Single-agent CEP-701, a novel FLT3 inhibitor, shows biologic and clinical activity in patients with relapsed or refractory acute myeloid leukemia. *Blood*. 2004;103(10):3669-3676.
14. Stone RM, DeAngelo DJ, Klimek V, et al. Patients with acute myeloid leukemia and an activating mutation in FLT3 respond to a small-molecule FLT3 tyrosine kinase inhibitor, PKC412. *Blood*. 2005;105(1):54-60.
15. DeAngelo DJ, Stone RM, Heaney ML, et al. Phase 1 clinical results with tandutinib (MLN518), a novel FLT3 antagonist, in patients with acute myelogenous leukemia or high-risk myelodysplastic syndrome: safety, pharmacokinetics, and pharmacodynamics. *Blood*. 2006;108(12):3674-3681.
16. Liu L, Cao Y, Chen C, et al. Sorafenib blocks the RAF/MEK/ERK pathway, inhibits tumor angiogenesis, and induces tumor cell apoptosis in hepatocellular carcinoma model PLC/PRF/5. *Cancer Res*. 2006;66(24):11851-11858.
17. Zhang W, Konopleva M, Ruvolo VR, et al. Sorafenib induces apoptosis of AML cells via Bim-mediated activation of the intrinsic apoptotic pathway. *Leukemia*. 2008;22(4):808-818.
18. Borthakur G, Kantarjian H, Ravandi F, et al. Phase I study of sorafenib in patients with refractory or relapsed acute leukemias. *Haematologica*. 2011;96(1):62-68.
19. Metzelder S, Wang Y, Wollmer E, et al. Compassionate use of sorafenib in FLT3-ITD-positive acute myeloid leukemia: sustained regression before and after allogeneic stem cell transplantation. *Blood*. 2009;113(26):6567-6571.
20. Levis M, Pham R, Smith BD, Small D. In vitro studies of a FLT3 inhibitor combined with chemotherapy: sequence of administration is important to achieve synergistic cytotoxic effects. *Blood*. 2004;104(4):1145-1150.
21. Ravandi F, Cortes JE, Jones D, et al. Phase I/II study of combination therapy with sorafenib, idarubicin, and cytarabine in younger patients with acute myeloid leukemia. *J Clin Oncol*. 2010;28(11):1856-1862.
22. Serve H, Wagner R, Sauerland C, Brunnberg U, Krug U, Schaich M. Sorafenib in combination with standard induction and consolidation therapy in elderly AML patients: results from a randomized, placebo-controlled phase II trial. *Blood*. 2010;116(21). Abstract 333.
23. Sato T, Yang X, Knapper S, et al. FLT3 ligand impedes the efficacy of FLT3 inhibitors in vitro and in vivo. *Blood*. 2011;117(12):3286-3293.
24. Levis M, Ravandi F, Wang ES, et al. Results from a randomized trial of salvage chemotherapy followed by lestaurnitinib for patients with FLT3 mutant AML in first relapse. *Blood*. 2011;117(12):3294-3301.
25. Levis M, Brown P, Smith BD, et al. Plasma inhibitory activity (PIA): a pharmacodynamic assay reveals insights into the basis for cytotoxic response to FLT3 inhibitors. *Blood*. 2006;108(10):3477-3483.
26. Pratz KW, Cho E, Levis MJ, et al. A pharmacodynamic study of sorafenib in patients with relapsed and refractory acute leukemias. *Leukemia*. 2010;24(8):1437-1444.
27. Cheson BD, Bennett JM, Kopecky KJ, et al; International Working Group for Diagnosis, Standardization of Response Criteria, Treatment Outcomes, and Reporting Standards for Therapeutic Trials in Acute Myeloid Leukemia. Revised recommendations of the International Working Group for Diagnosis, Standardization of Response Criteria, Treatment Outcomes, and Reporting Standards for Therapeutic Trials in Acute Myeloid Leukemia. *J Clin Oncol*. 2003;21(24):4642-4649.
28. Breems DA, Van Putten WL, Huijgens PC, et al. Prognostic index for adult patients with acute myeloid leukemia in first relapse. *J Clin Oncol*. 2005;23(9):1969-1978.
29. Wagner K, Damm F, Thol F, et al. FLT3-internal tandem duplication and age are the major prognostic factors in patients with relapsed acute myeloid leukemia with normal karyotype. *Haematologica*. 2011;96(5):681-686.
30. Smith CC, Wang Q, Chin CS, et al. Validation of ITD mutations in FLT3 as a therapeutic target in human acute myeloid leukaemia. *Nature*. 2012;485(7397):260-263.
31. Man CH, Fung TK, Ho C, et al. Sorafenib treatment of FLT3-ITD(+) acute myeloid leukemia: favorable initial outcome and mechanisms of subsequent nonresponsiveness associated with the emergence of a D835 mutation. *Blood*. 2012;119(22):5133-5143.
32. Al-Kali A, Cortes J, Faderl S, et al. Patterns of molecular response to and relapse after combination of sorafenib, idarubicin, and cytarabine in patients with FLT3 mutant acute myeloid leukemia. *Clin Lymphoma Myeloma Leuk*. 2011;11(4):361-366.
33. Pabst T, Mueller BU, Zhang P, et al. Dominant-negative mutations of CEBPA, encoding CCAAT/enhancer binding protein-alpha (C/EBPalpha), in acute myeloid leukemia. *Nat Genet*. 2001;27(3):263-270.
34. Radomska HS, Huettner CS, Zhang P, Cheng T, Scadden DT, Tenen DG. CCAAT/enhancer binding protein alpha is a regulatory switch sufficient for induction of granulocytic development from bipotential myeloid progenitors. *Mol Cell Biol*. 1998;18(7):4301-4314.
35. Hayakawa F, Towatari M, Kiyoi H, Tanimoto M, Kitamura T, Saito H, Naoe T. Tandem-duplicated Flt3 constitutively activates STAT5 and MAP kinase and introduces autonomous cell growth in IL-3-dependent cell lines. *Oncogene*. 2000;19(5):624-631.
36. Zheng R, Friedman AD, Levis M, Li L, Weir EG, Small D. Internal tandem duplication mutation of FLT3 blocks myeloid differentiation through suppression of C/EBPalpha expression. *Blood*. 2004;103(5):1883-1890.
37. Mizuki M, Schwable J, Steur C, et al. Suppression of myeloid transcription factors and induction of STAT response genes by AML-specific Flt3 mutations. *Blood*. 2003;101(8):3164-3173.
38. Sexauer A, Perl A, Yang X, et al. Terminal myeloid differentiation in vivo is induced by FLT3 inhibition in FLT3/ITD AML. *Blood*. 2012;120(20):4205-4214.
39. Radomska HS, Bassères DS, Zheng R, et al. Block of C/EBP alpha function by phosphorylation in acute myeloid leukemia with FLT3 activating mutations. *J Exp Med*. 2006;203(2):371-381.
40. Ng KP, Ebrahim G, Negrotto S, et al. p53 independent epigenetic-differentiation treatment in xenotransplant models of acute myeloid leukemia. *Leukemia*. 2011;25(11):1739-1750.
41. Negrotto S, Ng KP, Jankowska AM, et al. CpG methylation patterns and decitabine treatment response in acute myeloid leukemia cells and normal hematopoietic precursors. *Leukemia*. 2012;26(2):244-254.
42. Curik N, Burda P, Vargova K, et al. 5-azacitidine in aggressive myelodysplastic syndromes regulates chromatin structure at PU.1 gene and cell differentiation capacity. *Leukemia*. 2012;26(8):1804-1811.
43. Milhem M, Mahmud N, Lavelle D, Araki H, DeSimone J, Saunthararajah Y, Hoffman R. Modification of hematopoietic stem cell fate by 5aza 2'deoxyctidine and trichostatin A. *Blood*. 2004;103(11):4102-4110.
44. Piloto O, Wright M, Brown P, Kim KT, Levis M, Small D. Prolonged exposure to FLT3 inhibitors leads to resistance via activation of parallel signaling pathways. *Blood*. 2007;109(4):1643-1652.
45. Zhou J, Bi C, Janakakumara JV, et al. Enhanced activation of STAT pathways and overexpression of survivin confer resistance to FLT3 inhibitors and could be therapeutic targets in AML. *Blood*. 2009;113(17):4052-4062.
46. Lyman SD, Seaberg M, Hanna R, et al. Plasma/serum levels of flt3 ligand are low in normal individuals and highly elevated in patients with Fanconi anemia and acquired aplastic anemia. *Blood*. 1995;86(11):4091-4096.
47. Wodnar-Filipowicz A, Lyman SD, Gratwohl A, Tichelli A, Speck B, Nissen C. Flt3 ligand level reflects hematopoietic progenitor cell function in aplastic anemia and chemotherapy-induced bone marrow aplasia. *Blood*. 1996;88(12):4493-4499.
48. Zimmerman EI, Roberts JL, Li L, et al. Ontogeny and sorafenib metabolism. *Clin Cancer Res*. 2012;18(20):5788-5795.
49. Zarrinkar PP, Gunawardane RN, Cramer MD, et al. AC220 is a uniquely potent and selective inhibitor of FLT3 for the treatment of acute myeloid leukemia (AML). *Blood*. 2009;114(14):2984-2992.