The Phytoestrogen Daidzein Affects the Antioxidant Enzyme System of Rat Hepatoma H4IIE Cells

Elke Röhrdanz,2 Sandra Ohler, Quynh-Hoa Tran-Thi and Regine Kahl

Institute of Toxicology, Heinrich-Heine-Universität Düsseldorf, D-40225 Düsseldorf, Germany

ABSTRACT Phytoestrogens such as the soy isoflavonoid daidzein have potential health benefits. The antioxidant properties of phytoestrogens are considered to be responsible for their protective effects. The antioxidant enzyme (AOE) system plays an important role in the defense of cells against oxidative insults. To determine whether flavonoids can exert antioxidative effects not only directly but also indirectly by modulating the AOE system, we investigated the influence of the flavonoid daidzein on the expression of different AOE. Daidzein treatment of hepatoma H4IIE cells increased catalase mRNA expression two- to threefold. Expression levels of copper zinc superoxide dismutase (CuZnSOD) were not affected by exposure to daidzein. Manganese superoxide dismutase (MnSOD) mRNA expression levels decreased slightly and glutathione peroxidase (GPx) levels increased slightly after daidzein exposure. Changes in AOE mRNA expression levels were significant at 300 μmol/L daidzein. To elucidate the mechanisms underlying the strong increase in catalase mRNA, transfection experiments were performed. Transient transfection of hepatoma cells with reporter plasmids containing different parts of the upstream region of the catalase gene showed a significant one- to threefold increase in reporter gene activity after daidzein exposure. This indicates that daidzein can directly activate the rat catalase promoter region. Despite the increase in catalase mRNA, daidzein pretreatment of cells did not protect against oxidative stress resulting from H2O2 exposure. On the contrary, daidzein itself exerted a mild oxidative stress. In conclusion, the changes in the AOE system provoked by daidzein affected the oxidant rather than the antioxidant properties of daidzein. J. Nutr. 132: 370–375, 2002.

KEY WORDS: antioxidant enzymes • daidzein • expression • hepatoma cell • lipid peroxidation

Phytoestrogens are a group of naturally occurring diphenolic compounds present in legumes, whole grains, fruits and vegetables. These dietary flavonoids have drawn much attention because there are suggestions that they might benefit human health. High consumption of phytoestrogen-rich food has been linked to a reduced incidence of cancers at different sites including breast, prostate and colon (1–4). Flavonoids in regularly consumed foods may also reduce the risk of death from coronary heart disease (5,6).

Several in vitro and animal studies were undertaken to clarify the biological and physiologic processes that account for the cancer chemoprotective effects of flavonoids. The estrogenic activities of the flavonoids may play an important role in their health-enhancing properties. Flavonoids have been reported to bind to estrogen receptors (ER)3 and prevent cell growth in hormone-dependent cancer cells (1,7,8). Another proposed mechanism for cellular protection is the induction of phase II detoxification enzymes, leading to detoxification of proximate carcinogens generated by phase I metabolism. Indeed, some phytoestrogens were found to induce the phase II enzyme NADPH quinone reductase (9). In addition to these different modes of action, the protective effects of flavonoids have been attributed mainly to their antioxidant properties. Flavonoids scavenge free radicals, chelate redox-active metal ions and increase metallothionein expression (10–12). Metallothionein can protect cells from heavy-metal toxicity, but also exerts antioxidant activity.

In the defense against oxidative stress, the antioxidant enzyme (AOE) system of cells plays an important role. The antioxidant enzymes include the superoxide dismutases (SOD), catalase and glutathione peroxidases (GPx) (13–15). Catalase and GPx convert H2O2 to H2O and the SOD catalyze the dismutation of the superoxide radical anion. There are three forms of SOD in mammals, i.e., MnSOD located in mitochondria, CuZnSOD found mainly in the cytosol and an extracellular SOD localized in the extracellular fluid.

The expression of AOE can be regulated by oxidative stress (16–19). Induction of catalase mRNA expression could be elicited by H2O2 or hyperoxia in different mammalian cells (16–18,20). MnSOD expression was also found to be selectively induced by inflammatory mediators (21,22) as well as oxidative stress provoked either by H2O2 (17,23) or the redox-
cycling compound paraquat (24). However, little is known about the influence of flavonoids on the AOE system.

There are many reports relating to the reactivity of flavonoids with active oxygen species, thus emphasizing their antioxidant potential via a direct radical scavenging mechanism (10,25). On the other hand, flavonoids were found to induce the expression of enzymes such as metallothionein or NADPH quinone reductase, which may exert protective effects (9,12). We wanted to know whether flavonoids also can affect AOE expression, thus exerting indirect antioxidant activities. Therefore, we treated hepatoma cells with the flavonoid daidzein. We investigated the expression of different AOE and concurrently determined whether exposure of cells to daidzein protected against oxidative stress.

MATERIALS AND METHODS

Culture of hepatoma cells. Rat hepatoma H4IIE cells were grown in Dulbecco's modified Eagle's medium (PAA Laboratories, Colbe, Germany) and human hepatoma HepG2 cells in RPMI (PAA Laboratories) supplemented with 10% fetal calf serum, penicillin (100 U/L) and streptomycin (100 mg/L) in a humidified atmosphere of 5% CO2 in air at 37°C. Cells were seeded in 6-well plates. Unless stated otherwise, cells were treated for 24 h with different concentrations of daidzein, which was dissolved in dimethyl sulfoxide. The final concentration of the solvent never exceeded 0.7%. Control cells were treated with dimethyl sulfoxide. For the experiments involving H2O2 exposure, cells were pretreated for 24 h with daidzein in different concentrations. Then the medium was changed to one containing 2 mmol/L H2O2, and cells were incubated for another 6 h before malondialdehyde (MDA) measurement.

Viability assay. To determine the viability of daidzein-treated cells, mitochondrial dehydrogenase activity was measured in the MTT-assay as described elsewhere (26). Additionally, the Neutral Red assay was performed according to Bornfreund and Puerner (27).

Lipid peroxidation. Hepatoma H4IIE cells were pretreated with 200 or 300 μmol/L daidzein for 24 h followed by a 6-h treatment with 2 mmol/L H2O2. The respective controls received no treatment at all, were pretreated with medium and then with 2 mmol/L H2O2, or were given a daidzein pretreatment only. The amount of lipid peroxidation was determined by measuring the release of the breakdown product MDA into the medium. MDA was determined after reaction of cell culture supernatants with thiobarbituric acid and subsequent HPLC as previously described (24). Measurements were compared with untreated control cells. In further experiments, H4IIE cells were exposed to different concentrations of daidzein only for 24 h and the amount of MDA released into the medium was determined.

RNA isolation and RNA analysis. Total RNA was isolated from cells using Trizol-Reagent (GIBCO BRL, Eggenstein, Germany). For Northern blot analysis, 5 μg of total RNA was resolved by electrophoresis in a 1% agarose, 2.25 mmol/L formaldehyde gel in a running buffer containing 20 mmol/L MOPS, pH 7, 0.5 mmol/L sodium phosphoribosyltransferase (Roche Diagnostics GmbH, Mannheim, Germany) by random hexamer priming (Roche Diagnostics GmbH, Mannheim, Germany). Blots were prehybridized and hybridized with cDNAs for rat catalase and CuZnSOD mRNA, 602 bp for GPx mRNA and 450 bp for GAPDH mRNA. The conditions of PCR were set to be in the linear phase of amplification to allow for semiquantification of mRNA content. From each PCR reaction, 5 μL was electrophoresed in a 1% agarose gel in Tris-borate-EDTA buffer. The cDNA bands were visualized by UV illumination after the gels were stained with ethidium bromide. Gels were photographed and scanned densitometrically.

Transfection and reporter gene assays. To evaluate the transcriptional control of the catalase gene, deletion plasmids were constructed from a plasmid containing 1.1 kb of the rat catalase upstream sequence (~1072 bp to +54 bp of transcription initiation site) in front of a luciferase expression vector (18). The different promoter-reporter constructs of the catalase upstream region in front of the luciferase vector are shown in Figure 4.

HepG2 cells were transiently transfected at 50–70% confluency with a modified calcium phosphate precipitation method followed by glycerol shock as previously described (32). Transfection cells were given fresh media 2–3 h before. Each well was cotransfected with 4 μg of a plasmid containing the luciferase reporter-construct and a plasmid containing the β-galactosidase gene driven by the SV40 promoter (Promega, Mannheim, Germany). After 24 h, transfection cells were treated with 50 or 100 μmol/L daidzein for another 24 h. For harvesting, the cells were washed twice with 1 × PBS and then lysed in cell culture lysis buffer (Promega). Protein concentration of the cellular extract was determined according to Bradford (33), using bovine serum albumin as standard. Cellular protein (10 μg) was used for luciferase assays. Luciferase activity was determined with a Multi-Biolumat LB 9505 C (Berthold, Bad Wildbad, Germany) using luciferase assay reagent according to the manufacturer's protocol (Promega). With every transfection assay pGL3-basic and pGL3-control plasmid (Promega), containing SV 40 promoter and enhancer sequences, were transfected separately to additionally control for transfection efficiency. β-Galactosidase activity was determined using 0.1% 5-bromo-4-chloro-3-indolyl β-D-galactopyranoside as substrate (28). To minimize variability due to the difference in transfection efficiency among plates, the activity of luciferase and chloramphenicol acetyltransferase was corrected on the basis of the activity of β-galactosidase.

Statistical methods. Results are expressed as means ± SEM. Significance was evaluated by ANOVA followed by Fisher's Protected Least Significant Difference test. A probability of P < 0.05 was used as the criterion for significant difference.

RESULTS

Cytotoxicity. The cytotoxicity of daidzein was determined with two different assays (data not shown). In H4IIE cells, the MTT assay was more sensitive than the Neutral Red assay. According to the MTT test, cytotoxicity was apparent with 200 μmol/L daidzein. There was a significant reduction in viable cells of ~24%. This increased to 30% with 300 μmol/L daidzein and to 54% with 500 μmol/L daidzein. With the Neutral Red assay, cytotoxicity of daidzein became apparent (40% dead cells) and was significant when cells were exposed to 500 μmol/L daidzein.

Effect of daidzein on AOE expression in H4IIE cells. H4IIE cells were grown to confluency and then treated with different concentrations of daidzein for 24 h. Figure 1A shows the Northern blots for catalase and CuZnSOD mRNA expression. In Figure 1B the densitometric data relative to E18 S rRNA for catalase mRNA expression. In Figure 2A, the PCR for detection of MnSOD and GPx is depicted and in Figure 2B, the densitometric data relative to GAPDH mRNA expression. Daidzein treatment led to an increase of catalase mRNA expression (Fig 1B). With 300 μmol/L daidzein, a
threefold increase in catalase mRNA levels was detected. With 300 μmol/L daidzein, we found a 100% increase in catalase activity (data not shown, mean of two experiments). On the basis of the Neutral Red assay, daidzein was not cytotoxic in this concentration range, whereas the MTT assay indicated minor cytotoxicity of daidzein with 24–30% dead cells. CuZnSOD mRNA expression was not affected by daidzein exposure; thus densitometric data are not shown. There was a slight, but significant 20% decrease in MnSOD mRNA expression after exposure of cells to 300 μmol/L daidzein (Fig. 2B). An increase of ~40% in Gpx mRNA expression generally occurred at all concentrations of daidzein tested except 200 μmol/L daidzein (Fig. 2B). A threefold increase in catalase mRNA levels was detected. With 300 μmol/L daidzein, we found a 100% increase in catalase activity (data not shown, mean of two experiments).

Influence of daidzein on the transcriptional activity of the catalase reporter gene constructs. The promoter-reporter constructs for the catalase gene, as shown in Figure 3, were transiently transfected into Hep G2 cells. Cells were then treated with different concentrations of daidzein for 24 h. In all of the experiments, the nontreated promoter-reporter constructs showed basal activity compared with the pGL3-basic vector.

FIGURE 1 Catalase and copper zinc superoxide dismutase (CuZnSOD) mRNA expression in H4IIE cells after daidzein exposure. (A) Northern blot analysis of catalase and CuZnSOD mRNA. Cells were exposed to the indicated concentrations of daidzein for 24 h. Northern blot analysis was carried out using rat catalase and CuZnSOD cDNAs as probes, n = 3–6 individual experiments. (B) Quantitative densitometric data for catalase mRNA expression levels quantitated relative to 18 S RNA levels shown at the bottom of (A). The results are expressed as a percentage of control (control = 0.77 ± 0.41). Values are means ± SEM, n = 3–6 different experiments; *P < 0.05 vs. controls.

FIGURE 2 Manganese superoxide dismutase (MnSOD) and glutathione peroxidase (Gpx) mRNA expression in H4IIE cells after daidzein exposure. (A) Polymerase chain reaction (PCR) blot of Gpx, GAPDH and MnSOD mRNAs obtained by reverse transcriptase (RT)-PCR after exposure of H4IIE cells to the indicated concentrations of daidzein for 24 h. (B) Gpx/GAPDH and MnSOD/GAPDH quotient after densitometric scanning of PCR blots obtained after RT-PCR shown in (A). Values are expressed as a percentage of controls (Gpx/GAPDH control = 0.82 ± 0.17; MnSOD/GAPDH control = 4.07 ± 0.53). Values are means ± SEM, n = 3–4 different experiments; *P < 0.05 vs. controls.

FIGURE 3 Promoter-reporter constructs of the 5′-flanking region of the rat catalase gene and the luciferase plasmid. Putative silencer elements and possible binding sites of different transcription factors are also shown (53,54).
Twenty-four hours after transfection, cells were treated with 50 or 100 μmol/L daidzein for another 24 h. Luciferase assays were conducted in treated and untreated cells. Luciferase activity values are shown as a percentage of controls (pLuc1 control = 0.77 ± 0.29, pLuc(-)2 control = 0.17 ± 0.10, pLuc2 control = 0.23 ± 0.05). Values are means ± SEM, n = 4–6 different experiments; *P < 0.05 vs. controls, †P < 0.05 vs. pLuc1 treated with 50 μmol/L daidzein.

FIGURE 4 Promoter-activity of pLuc1, pLuc(-)2 and pLuc2 after exposure to daidzein. HepG2 cells were transfected as described. Twenty-four hours after transfection, cells were treated with 50 or 100 μmol/L daidzein for another 24 h. Luciferase assays were conducted in treated and untreated cells. Luciferase activity values are shown as a percentage of controls (pLuc1 control = 0.77 ± 0.29, pLuc(-)2 control = 0.17 ± 0.10, pLuc2 control = 0.23 ± 0.05). Values are means ± SEM, n = 4–6 different experiments; *P < 0.05 vs. controls, †P < 0.05 vs. pLuc1 treated with 50 μmol/L daidzein.

The cytotoxicity of daidzein was measured with two different assays. The two highest concentrations of daidzein used for the evaluation of AOE expression levels were slightly cytotoxic in the MTT assay but nontoxic in the Neutral Red assay. The Neutral Red assay determines membrane uptake, whereas the MTT assay shows mitochondrial dehydrogenase activity (26,27). Obviously mitochondrial functions were first impaired by daidzein treatment, before a marked damage of cell membranes and thus the destruction of the whole cell became evident.

Daidzein treatment of hepatoma cells dramatically increased catalase mRNA expression. Kameoka et al. (12) investigated catalase and CuZnSOD expression after daidzein treatment of CaCo2-cells and found no change in the expression pattern of either enzyme. This may have been due to the differences in cell lines because basal as well as inducible mRNA expression levels of AOE may differ depending on the cell type investigated (16,22,34). To further elucidate the mechanisms underlying the increase in catalase mRNA by daidzein, we performed transfection experiments. Transfection of hepatoma cells with reporter genes containing different parts of the rat catalase upstream region and subsequent exposure to daidzein always resulted in an increase in reporter gene activity of treated compared with nontreated cells. These results clearly indicate that the increase of catalase mRNA

**DISCUSSION**

The influence of phytoestrogens on the AOE system has not yet been investigated in detail. We treated rat liver hepatoma H4IIE cells with the isoflavonoid daidzein. H4IIE cells contained high levels of AOE and induction of AOE through oxidative stress has been shown previously in H4IIE cells (17); thus, they present a good model for these investigations.

Lipid peroxidation. To determine whether the increase in catalase mRNA expression levels by daidzein exposure may exert antioxidant effects, H4IIE cells were pretreated with different concentrations of daidzein followed by a treatment with 2 mmol/L H2O2. The extent of lipid peroxidation was determined by measuring the release of the breakdown product MDA into the medium (Fig. 5A). Treatment with 2 mmol/L H2O2 increased MDA levels only ~3 times compared with controls. Daidzein pretreated cells were not protected from H2O2. On the contrary, MDA levels increased ~4 times in these cells compared with controls and compared with the cells treated with H2O2 only. Treatment with 300 μmol/L daidzein alone increased MDA levels ~50% compared with controls. Further experiments (Fig. 5B) were performed to confirm this effect of daidzein on MDA levels. H4IIE cells were exposed to different concentrations of daidzein and the amount of MDA released into the medium was determined. An increase in MDA levels of ~50% was detected in H4IIE cell supernatants treated with 200 or 500 μmol/L daidzein.

FIGURE 5 Formation of malondialdehyde (MDA) in H4IIE cells after treatment with daidzein. (A) Cells were incubated with the indicated concentrations of daidzein for 24 h. The medium was changed to medium containing 2 mmol/L H2O2 and cells were incubated for another 6 h. The amount of MDA released into the medium was determined. The results are expressed as a percentage of controls exposed to medium only (control = 113 ± 19 pmol MDA/10⁶ cells). Values are means ± SEM, n = 4 different experiments; *P < 0.05 vs. controls with no treatment; †P < 0.05 vs. cells treated with 2 mmol/L H2O2 only. (B) Cells were incubated with the indicated concentrations of daidzein for 24 h and the amount of MDA released into the medium was determined. The results are expressed as a percentage of controls (control = 188 ± 30 pmol MDA/10⁶ cells). Values are means ± SEM, n = 4 different experiments; *P < 0.05 vs. controls.
after daidzein treatment is due to transcriptional activation of
the rat catalase promoter. Little is known about transcriptional
activation of the catalase gene. In general, post-transcriptional
processes were found to be important for regulation of catalase
gene expression. A redox-sensitive RNA binding protein was
described for rat lung, which acts by increasing RNA stability
(19). In previous experiments, we found catalase gene induc-
tion to be regulated transcriptionally in hepatoma cells,
whereas in hepatocytes, a translational mechanism was
assumed (17).

Isoflavonoids interact with ER. For daidzein, a stronger
interaction with the β- than the α-subtype of the ER was
shown (7,35). Similar in vitro interactions of phytoestrogens
with human and rat ERα and ERβ have been reported, indi-
cating that ER react similarly to phytoestrogens in the two
species (7,35,36). Azevedo et al. (37) even found regulation of
AOE to be dependent on sex steroids. Hepatoma cells such as
those used for our experiments, however, are ER-negative,
especially for later passages (38). Thus, a transcriptional acti-
vation of the catalase promoter via ER interactions can be
excluded.

To determine whether the increase in catalase expression
protects the cells from oxidative damage, cells were pretreated
with daidzein and then exposed to H2O2. Unexpectedly, in
daidzein-pretreated cells, the oxidative damage, as measured
by the lipid peroxidation product MDA, was more distinct
than in cells treated with H2O2 only. Changes in the AOE
mRNA expression pattern resulting from daidzein pretreat-
ment may have led to an imbalance in the concerted action of
the different AOE. This may have rendered the cells more
sensitive toward the oxidative stress provoked by hydrogen
doxygen treatment. Daidzein itself caused a mild oxidative
stress. The antioxidant or prooxidant potential of flavonoids is
a controversial topic in the literature (10,39). Increasing levels
of daidzein were found to inhibit LDL oxidation in vitro (40).
Daidzein also protected against lecithin peroxidation induced
by the superoxide anion generated through the xanthine-
organophosphate system (41). On the other hand, in rat liver
micromes, daidzein was not able to inhibit enzymatically
induced lipid peroxidation (42). Hodgson et al. (43) also
found no in vivo antioxidant activity of daidzein. A number of
flavonoids are able to produce reactive oxygen species by autooxidation and redox-cycling (44). Similar mechanisms may
be responsible for the oxidative stress we observed.

Oxidative stress was found to upregulate catalase mRNA
expression in different cell lines as well as in neonatal rats
under hyperoxic conditions (16,19). Different regulatory ele-
ments that play a role in the regulation of genes after oxidative
stress have been identified (45,46). No consensus sequences of
these well-described transcription factors can be found in the
catalase upstream region (see Fig. 4). A putative AP2-se-
quence and a sequence resembling the binding site of the
antioxidant responsive element are the only elements identi-
fied by sequence comparison so far. The truncated catalase-
reporter-constrasts showed a higher inducible reporter gene
activity after exposure to 50 μmol/L daidzein than did the
complete construct. Perhaps gene activity is suppressed by
silencer elements present in the whole construct; such possible
silencers were described by Takeuchi et al. (47).

In addition to catalase, we also found an increase in GPx
mRNA expression levels, which was not as distinct. In the
5'-flanking region of the human GPx gene, two cis-acting
elements were characterized, which are of importance for gene
regulation in an oxidative environment (48). These elements
were found to be active in human cardiomyocytes and HeLa
cell cultures. The rat GPx gene is likely regulated in H4IIE cells.

Surprisingly, there was a slight decrease in MnSOD mRNA
expression after daidzein exposure. Induction of the MnSOD
gene by tumor necrosis factor-α and oxidative stress has been
described in several studies (49,50). Daidzein is an efficient
inhibitor of casein kinase II, which phosphorylates nuclear
oncogene products and a number of proteins involved in gene
expression (31). It is possible that daidzein interferes with
signal transduction pathways important for transcription of
MnSOD.

In addition, CuZnSOD mRNA expression was not regu-
lated coordinately with catalase and GPx expression. After
daidzein exposure, no changes in CuZnSOD mRNA expres-
sion levels were detected. In previous studies, we (17) and
others (23,52) found that CuZnSOD often showed a stable
expression pattern despite changes in the other AOE after
exposure to different oxidants. This differential regulation of
AOE expression is dependent on daidzein exposure as well.

In conclusion, our experiments showed that daidzein af-
fected the AOE expression pattern. Interestingly, these
changes did not add to the antioxidant, but rather to the
oxidant properties of daidzein. The possible effect of phy-
toestrogens on human health may also be linked to the AOE
system; as such, our in vitro studies suggest possible mecha-
nisms of daidzein action. Further investigations will show
whether daidzein has similar effects on the AOE system in
vivo. Therefore, when evaluating the antioxidant capacity of
flavonoids their ability to modulate AOE expression should
also be considered.

LITERATURE CITED

1. Adicuretz, H., Mousavi, Y., Clark, J., Höckerstedt, K., Hämmäläinen, E.,
Wähälä, K., Mäkelä, T. & Hase, T. (1992) Dietary phytoestrogens and cancer:
1200.
(1989) A prospective study of dietary habits, diet and prostate cancer among
lignans on colon carcinogenesis and β-glucuronidase activity. Carcinogenesis
17: 1343–1348.
5. Hertog, M.G.L., Kromhout, D., Aravanis, C., Blackburn, H., Buzina, R.,
Fidanza, F., Giampaoli, S., Jansen, A., Menotti, A., Nedeljkovic, S., Pekkanen,
(1995) Flavonoid intake and long-term risk of coronary heart disease and cancer
flavone intake is associated with cardiovascular disease risk factors in postmeno-
7. Morito, K., Hirose, T., Kinjo, H., Hirakawa, T., Okawa, M., Nohara, T.,
NAPQI, quinone reductase by dietary phytoestrogens in colon Caco2/205 cells.
In vitro scavenger activity of some flavonoids and melatans against O2-
11. Morel, I., Lescoat, G., Cogrel, P., Sergent, O., Pasdeloup, N., Brisset, P.,
the flavonoids catechin, quercetin, and procyanidins on iron-loaded rat hepatec-
antioxidant proteins in human intestinal Caco-2 cells treated with dietary flava-
toxicity. In: Reviews in Biochemical Toxicology (Hodgson, E., Bend, J. R. &
15. Ursini, F., Maiorino, M., Brigelius-Flohe, R., Aumann, K. D., Roveri, A.,
Downloaded from https://academic.oup.com/jn/article-abstract/132/3/370/4687301 by guest on 13 October 2018
INFLUENCE OF DAIDZEIN ON THE AOE SYSTEM

375