Research Article

Cancer Immunology Research

Fine-tuning Tumor Immunity with Integrin Trans-regulation

Joseph M. Cantor, David M. Rose, Marina Slepak, and Mark H. Ginsberg

Abstract

Inefficient T-cell homing to tissues limits adoptive T-cell immunotherapy of solid tumors. αLβ2 and α4β1 integrins mediate trafficking of T cells into tissues via engagement of ICAM-1 and VCAM-1, respectively. Inhibiting protein kinase A (PKA)–mediated phosphorylation of α4 integrin in cells results in an increase in αLβ2-mediated migration on mixed ICAM-1–VCAM-1 substrates *in vitro*, a phenomenon termed "integrin trans-regulation." Here, we created an α4(S988A)-bearing mouse, which precludes PKA-mediated α4 phosphorylation, to examine the effect of integrin trans-regulation *in vivo*. The α4(S988A) mouse exhibited a dramatic and selective increase in migration of lymphocytes, but not myeloid cells, to sites of inflammation. Importantly, we found that the α4(S988A) mice exhibited a marked increase in T-cell entry into and reduced growth of B16 melanomas, consistent with antitumor roles of infiltrating T cells and progrowth functions of tumor-associated macrophages. Thus, increased α4 trans-regulation of αLβ2 integrin function biases leukocyte emigration toward lymphocytes relative to myeloid cells and enhances tumor immunity. *Cancer Immunol Res; 3(6); 661–7. ©2015 AACR.*

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Introduction

Different classes of leukocytes have opposing effects on the growth of tumors. Lymphocytes, particularly T cells, are critical components of the host defense that limit tumorigenesis (1). In sharp contrast, myeloid cells contribute cytokines that promote both tumor growth and angiogenesis (2, 3). α4 integrins play an important role in lymphocyte entry into sites of tissue injury (4, 5) in part because they markedly potentiate cell migration via signaling mediated by binding of paxillin to the α4 cytoplasmic tail (6). Protein kinase A (PKA)–mediated phosphorylation of the α4 integrin tail at Ser^{988} inhibits paxillin binding in migrating cells (7); consequently, the α4(S988A) mutation stabilizes the α4–paxillin interaction and increases α4 integrin signaling. The increased signaling downstream of α4(S988A) enhances integrin $\alpha_L\beta2$(LFA-1)–mediated migration of cells, a phenomenon termed integrin trans-regulation (8).

Here, we examined the biologic consequences of blockade of α4 phosphorylation by generating α4(S988A) mutant mice and found that these mice manifest a dramatic increase in recruitment of lymphocytes, but not myeloid cells, to an inflammatory site. We found that the α4(S988A) mutation markedly increased the abundance of T cells, but not myeloid cells, in heterotopic B16 melanomas. Increased lymphocyte homing is needed for efforts to develop adoptive immunotherapies for solid tumors (9–14). The potential therapeutic value of this form of integrin trans-regulation was established by the finding that subcutaneously implanted melanomas grew more slowly in α4(S988A) mice associated with markedly enhanced recruitment of T cells, but not myeloid cells, to the tumors. Thus, increasing integrin trans-regulation by blocking α4 integrin phosphorylation selectively recruits lymphocytes, but not myeloid cells, thereby reducing tumor growth. This finding suggests that modulation of integrin trans-regulation may be useful in overcoming a limitation of adoptive cellular immunotherapy of solid tumors.

Materials and Methods

Mice

α4(S988A) mice were generated as described in Supplementary Material of ref. 16. These mice were bred to create homozygous germline knock-ins and backcrossed to BL6 for >8 generations for all experiments, unless otherwise noted. For assessment of T-cell cytotoxic function, α4(S988A) mice were crossed with the OT-1 strain. *Rag1*$^{-/-}$ mice were used as recipients for *in vivo* competitive migration experiments. All mice were housed at the University of California San Diego animal facility, and all experiments were approved by the Institutional Animal Care and Use Committee.

Hematologic analysis

Blood from adult (10–20-week-old) α4(S988A) and α4[wild-type(wt)] mice was collected into tubes containing EDTA. Cell counts were obtained using an MS9-automated cell counter by the University of California San Diego Animal Care Program Diagnostic Laboratory, whose staff also manually performed differential counts on DiffQuick-stained smears.

Peritonitis model

Adult (10–20-week-old) α4(S988A) and control α4(wt) mice were injected i.p. with 4% (weight/vol) thioglycollate (Sigma-Aldrich) and sacrificed at various time points for peritoneal lavages. Cells (1×10^5) were adhered to glass slides with a

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Note: Supplementary data for this article are available at Cancer Immunology Research Online (http://cancerimmunolres.aacrjournals.org/).

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Cytospin4 instrument (ThermoShandon) and stained with Diff-Quick to differentiate cell types by light microscopy. The percentages of T, B1, and B2 cells were assessed by flow cytometry using fluorochrome-conjugated anti-CD3, anti-B220, and anti–Mac-1 antibodies. For competitive peritonitis assay, peritonitis was elicited as described above, but in *Rag1*$^{-/-}$ mice. Twenty-four hours after thioglycollate injection, mice received an i.v. injection of a mixture of splenocytes from adult α4(S988A) or control Ly5.1 α4(wt). Splenocyte suspensions and peritoneal lavages were stained using antibodies against CD3, CD8, and Ly5.1. The ratio of α4(S988A) to Ly5.1$^+$ α4(wt) splenocytes was compared between spleen and peritoneum as a measure of enrichment for α4(S988A) cells in the inflammatory site.

Lymphoid compartments

Bone marrow cell suspensions were prepared by flushing femur and tibia bones from adult α4(S988A) and α4(wt) mice. Single-cell suspensions from bone marrow, spleen, and thymus were treated with ACK lysis buffer, counted, and stained with fluorochrome-conjugated antibodies (eBioscience) against mouse B220 (RA3-6B2), IgM (II/41), IgD (11-26c), CD21/35 (7G6), CD23 (B3B4), CD3 (145-2C11), CD4 (GK1.5), CD8 (53-6.7) at optimized concentrations. Cells were washed and analyzed by flow cytometry. For integrin expression analysis, blood was collected by tail bleed, treated with ACK lysis buffer, and stained with anti-CD3, anti-α4, anti–αLβ2 integrin, or isotype control fluorochrome-conjugated antibodies before flow cytometry.

Humoral immune response

For antigen-specific antibody responses, α4(S988A) and α4(wt) mice (F5 backcross to BL6) were injected i.p. with 100 μg trinitrophenol-keyhole limpet hemocyanin (TNP-KLH; Biosearch) emulsified in 250 μL complete Freund's adjuvant (T-cell–dependent antigen). Blood serum was collected by centrifugation of tail vein bleed (100–200 μL with 1–2 mmol/L EDTA solution as an anticoagulant) before (preimmune) and at 1, 2, and 3 weeks after immunization. TNP-specific antibody concentrations in blood sera were assessed by direct ELISA with trinitrophenol-ovalbumin (TNP-OVA) as the coating antigen and alkaline phosphatase–conjugated polyclonal anti-mouse IgG (Sigma) as the detection antibody.

Migration assay

Resting B or T cells were purified from spleens of adult α4 (S988A) and control α4(wt) mice by negative depletion. Macrophages were differentiated from α4(S988A) or control α4(wt) bone marrow by culture in 30% L292 supernatant for one week. *In vitro* migration was assessed following a modified Boyden Chamber assay (8). Briefly, transwells (Costar) with 3.0-μm polycarbonate membrane inserts were coated with VCAM-1 and/or ICAM-1 Fc fusion proteins in carbonate buffer, pH 8.0. Transwell membranes were blocked in PBS, 2% BSA 30 minutes at room temperature. Cells (2.0×10^5) were added to the top chamber in complete medium (10% FBS). Complete medium containing 15 ng/mL stromal-derived factor-1α (SDF-1α; R&D Systems) was added to the lower chamber. To observe macrophage migration, 20 ng/mL of both SDF-1 and MCP-1 was necessary. After a 4-hour incubation at 37°C (overnight for macrophages), cells in the lower chamber and underside of transwell were harvested and counted by hemacytometer.

B16 melanoma model

B16 (f1 subclone) or Lewis lung carcinoma (LLC) cells were expanded in culture in complete medium (DMEM supplemented with 10% FBS, L-glutamine, βME, and pen/strep antibiotics). B16 cells (3×10^5) or LLC cells (1×10^6) were injected s.c. into the right hind flanks of adult α4(S988A) or control α4(wt) mice. When tumors became visible, length and width were measured daily with calipers. Tumors were assumed to be ellipsoid, and volume was calculated using the formula: (length × width^2)/2. Mice were sacrificed on day 15, and tumors were excised and weighed. To analyze tumor-infiltrating leukocytes (TIL), tumors were digested with collagenase (Sigma) for 20 to 30 minutes at 37°C and further processed to a single-cell suspension using a 7-mL tissue grinder (Kontes) and counted. Fluorochrome-conjugated antibodies were used to stain for tumor-infiltrating CD45$^+$ leukocytes and identify CD4$^+$ and CD8$^+$ T cells (CD3$^+$), as well as CD4$^+$Foxp3$^+$ regulatory T cell (Treg) and NK1.1$^+$ natural killer (NK) cells. Total subset numbers were calculated by multiplying the total cell number with percentage of CD45$^+$ and percentage of each subset. For lymphoid cell depletion, we injected anti-CD8 antibody (53-6.7, 100 μg) or a combination of anti-CD4, anti-CD8, and anti-NK1.1 antibodies (150 μg each), compared with isotype control antibody i.p. 2 days before and 5 days after B16 tumor-cell inoculation. Splenocytes harvested on day 15 were stained with antibodies specific for CD3, CD4, and CD8 to determine efficiency of T-cell depletion.

Analysis of clonal expansion *in vivo*

To analyze clonal expansion on a polyclonal T-cell receptor (TCR) background, α4(S988A) and wt mice were immunized with a combination of 100 μg poly I:C (Invivogen), 50 μg anti-CD40 antibody (Biolegend), and 500 μg ovalbumin protein (Sigma) in PBS i.p. Six days later, mice were sacrificed and spleen cells were stained using anti-CD8$^+$ antibody and an H-2K^b-SIINFEKL(PE) tetramer (Coulter).

Assessment of CD8 T-cell cytotoxic function

To generate functional CTLs, splenocytes from α4(S988A) and wt–OT-1 mice were cultured with 1 μg/mL SIINFEKL and 100 U/mL IL2 (NCI) for 6 days. Degranulation as a measure of cytotoxicity was measured as exposure of CD107a (LAMP-1) on the outer cell membrane. On day 6 following SIINFEKL stimulation, CTLs were harvested, counted, and cultured with SIINFEKL-pulsed (1 μg/mL for 1–2 hours) or -unpulsed splenocytes as targets in the presence of anti–LAMP-1-PE antibodies (eBioscience) for 2.5 hours at 37°C. Effector:target cultures were stained with anti-CD8 antibodies and analyzed by flow cytometry. For target lysis *in vitro*, CTLs were generated as above and cultured with an approximately 50/50 mixture of peptide-pulsed (CFSElo) and -unpulsed (CFSEhi) splenocyte targets overnight. Specific lysis is the percentage decrease in the percentage of the peptide-pulsed peak between CTL-containing and no-CTL control cultures.

Statistical analysis

The two-tailed *t* test was used for statistical comparison between groups in all experiments, except where otherwise noted. A value of $P < 0.05$ was considered statistically significant.

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Results and Discussion

The α4(S988A) mutation selectively increases lymphocyte migration to an inflammatory site

We used homologous recombination to generate α4(S988A) mice (Supplementary Fig. S1) and compared them to wt C57BL/6 mice as controls. We observed no statistically significant differences in formed elements of the blood (Supplementary Fig. S2A) or in lymphocyte numbers (with the possible exception of increased Pro-B cells in the bone marrow) in primary or secondary lymphoid tissue (Supplementary Fig. S2B) in α4(S988A) mice. Humoral immune responses to a T-dependent antigen were also similar between α4(S988A) and wt mice (Supplementary Fig. S2C).

Because the α4(S988A) mutation inhibits migration on substrates containing purified α4 integrin ligands (7), we hypothesized that α4(S988A) mice might exhibit a similar defect in leukocyte migration *in vivo*. We therefore used a thioglycollate-induced peritonitis model to test the effect of this mutation on leukocyte entry into an inflammatory site. To our great surprise, we observed a sharp increase in the number of lymphocytes infiltrating the peritoneum (Fig. 1), whereas myeloid cell infiltration was unaffected. We stained the peritoneal lavage with antibodies to identify B1, B2, and T lymphocytes and found that both B2 and T cells exhibited significantly (>3–4-fold) greater numbers in the α4(S988A) mice compared with that in the controls (Fig. 1). In contrast, the numbers of B1 cells, a resident peritoneal population (15), showed no statistically significant differences from controls, indicating that the α4(S988A) mutation specifically affects influx of lymphocytes. An *in vivo* competitive migration assay (Supplementary Fig. S3) using a mix of wt (congenically marked) and α4(S988A) splenocytes confirmed that the increased migration of α4(S988A) T cells is intrinsic to leukocytes and not merely due to effects of α4 phosphorylation in endothelial cells (16) or other changes in the inflammatory environment (e.g., production of chemokines by resident cells) in these mice. Therefore, interfering with α4 integrin phosphorylation selectively increased homing of lymphocytes to a site of inflammation but had no obvious effect on myeloid cells.

Integrin trans-regulation explains increased migration of α4(S988A) lymphocytes

Our initial expectation that the α4(S988A) mutation would decrease homing was based on results from *in vitro* migration on purified α4 integrin ligands. *In vivo* migration to an inflammatory site is complex and involves several classes of integrins (17). We previously reported that the presence of small amounts of α4 integrin ligands (e.g., VCAM-1) increases the migration of Jurkat T cells on αLβ2 integrin substrates (e.g., ICAM-1) *in vitro* (8). Paxillin binding to α4 is required for this effect, and enforced paxillin–α4 association enhanced the trans-regulation of αLβ2 by α4 integrins through increasing the activation of FAK and Pyk2 kinases (8, 18). The α4(S988A) mutation that blocks PKA phosphorylation of the α4 cytoplasmic tail could therefore enhance αLβ2-dependent migration. The *in vivo* peritonitis experiment requires migration on mixed substrates and is dependent on both α4 and β2 integrins (17), conditions in which the α4(S988A) mutation could increase migration of lymphocytes (8). To explore this possibility in a controlled setting, we purified B and T cells from α4(S988A) mice and measured their ability to migrate *in vitro* on purified α4 ligand (VCAM-1), purified β2 ligand (ICAM-1), or mixed substrates (VCAM-1 + ICAM-1) in response to the chemokine SDF-1α. Using a modified Boyden chamber assay, we confirmed that α4(S988A) lymphocytes exhibited reduced migration on a purified α4 integrin ligand: VCAM-1 (Fig. 2A). In contrast, when plated on substrates containing predominantly ICAM-1 and small amounts of VCAM-1, both B and T cells from α4(S988A) mice displayed enhanced migration that was dependent on both α4 and β2 integrins (Fig. 2B; Supplementary Fig. S4). This observation cannot be explained by differences in surface integrin expression levels, as T cells from α4(S988A) and

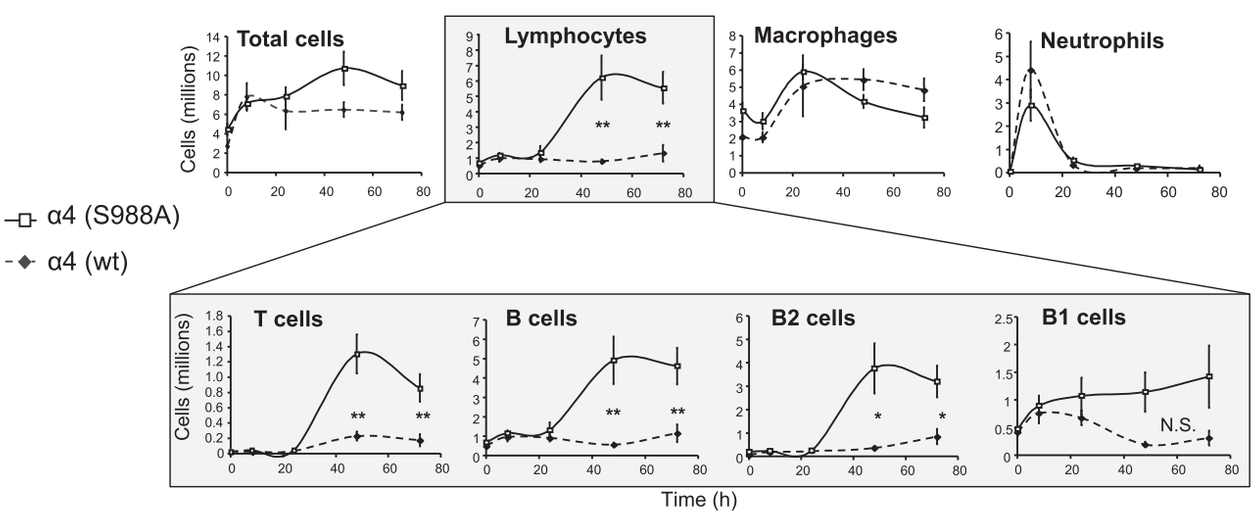

Figure 1.
Leukocyte migration in an α4(S988A) mouse. Adult α4(S988A) or wt mice were injected with 1 mL of thioglycollate medium i.p. Mice were sacrificed at various time points, and leukocytes from peritoneal lavages were enumerated and identified by cytospin and DiffQuick staining. Peritoneal lavages were also stained with antibodies to identify T cells ($CD3^+$), total B cells ($B220^+$), B2 cells ($B220^+CD11b^-$), and B1 cells ($B220^+CD11b^+$). Error bars, SEM of 5 mice for each group. *, $P < 0.03$; **, $P < 0.02$; N.S., not statistically significant.

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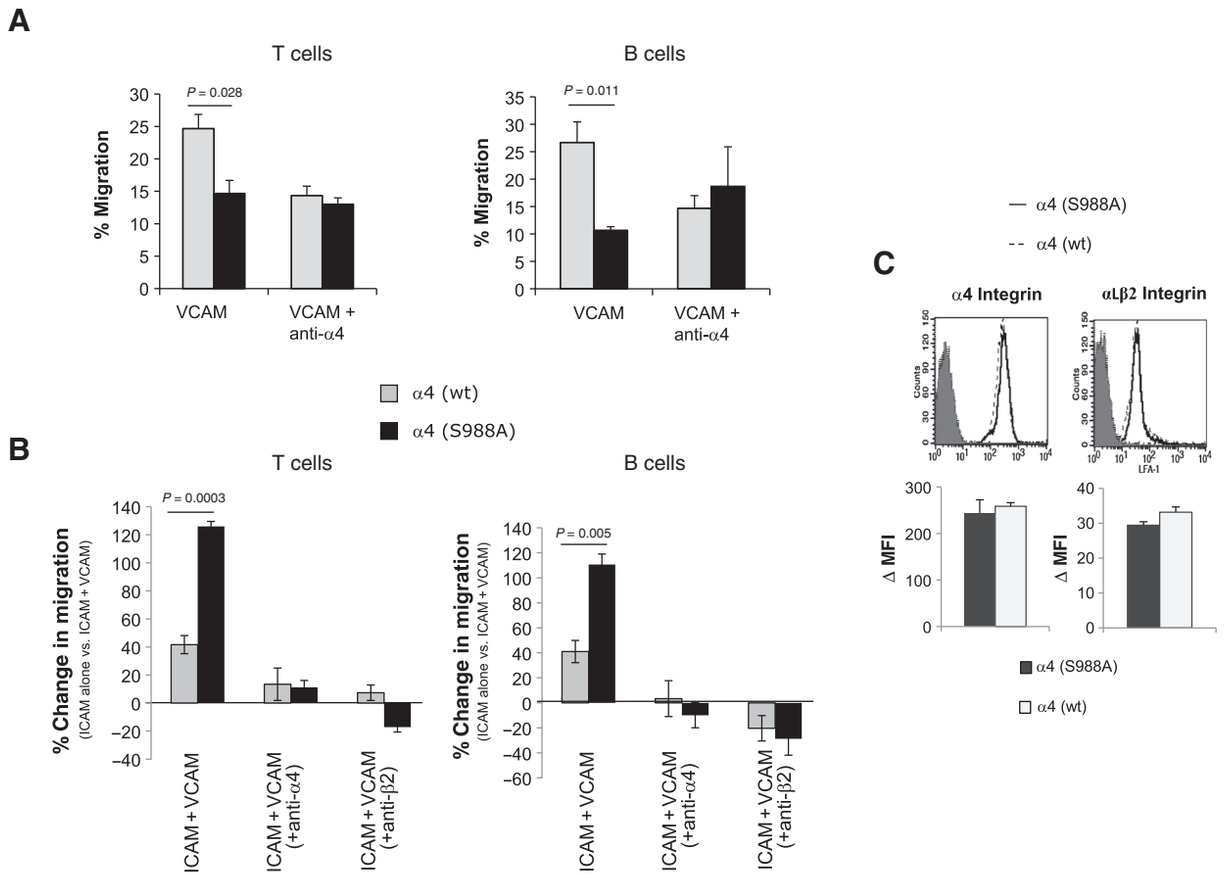

Figure 2.
Increased integrin trans-regulation in α4(S988A) lymphocytes. B and T cells were purified from spleens of α4(S988A) or control α4(wt) mice. Chemotactic migration toward SDF-1α (15 ng/mL) was assessed using a modified Boyden Chamber assay in wells coated with (A) VCAM-1 alone (2 μg/mL) or (B) ICAM-1 (5 μg/mL) ± VCAM-1 (0.02 μg/mL). For anti-integrin antibody-blocking studies, cells were treated with 10 μg/mL of either anti-α4 or anti–β2 integrin before the assay. B, percentage of increase in migration on ICAM-1 + VCAM-1 compared with ICAM-1 alone 100 × [(ICAM + VCAM migration − ICAM migration)/ICAM migration]. Error bars, SEM of 4 mice for each group. C, surface integrin expression levels on circulating α4(S988A) T cells. Blood leukocytes from α4(S988A) and α4(wt) control adult C57BL/6 mice ($n = 3$ per group) were stained with antibodies specific for T cells (CD3), α4 integrins (CD49d), and αLβ2 integrin heterodimer and analyzed by flow cytometry. Representative histograms show α4 or αLβ2 staining (soild and dotted lines) compared with nonspecific isotype control staining (filled peak). Bar graphs summarize staining from 3 mice per group; no statistically significant differences were observed. Δ MFI, change in mean fluorescence intensity.

α4(wt) mice show no difference in staining for α4 or αLβ2 integrins (Fig. 2C). These data indicate that the α4(S988A) mutation provides an increase in β2 integrin–dependent migration, i.e., integrin trans-regulation in primary lymphocytes.

Increased integrin trans-regulation reduces tumor growth by increasing T-cell homing

Specifically increasing lymphocyte entry into an inflammatory site might offer therapeutic benefit during an immune response to a tumor. T-cell migration to solid tumors is important for tumor immunity (1), whereas tumor infiltration by macrophages may promote tumor growth through increased angiogenesis and suppressed immunity (2, 3). In adoptive immunotherapy (11–14, 19), naïve T cells are modified and activated *in vitro*; however, T-cell homing is a critical limiting variable in solid tumor adoptive immunotherapy (9, 10). Based on our results with the peritonitis model, we hypothesized that α4(S988A) mice may have increased ability to resist tumors due to selective migration of lymphocytes to the tumor site. We tested this idea using a melanoma model (20), in which B16 melanoma cells are injected s.c., and tumor size is measured by weighing excised tumors 15 days later. α4 (S988A) mice had approximately 5-fold smaller tumors than wt BL6 controls (Fig. 3A), indicating that blocking α4 integrin phosphorylation on Ser988 increased tumor protection. This observation was not unique to the B16 melanoma, as α4 (S988A) mice also displayed increased resistance to the growth of Lewis lung carcinoma (Supplementary Fig. S5).

We next asked whether the α4(S988A) resistance to tumors was associated with increased T-cell homing. Indeed, B16 tumors in α4(S988A) mice had greater concentrations of T cells than those grown in wt mice (Fig. 3B, left). This finding was in striking contrast with a similar number of macrophages found in tumors from mice of both genotypes, supporting the idea that the decreased tumor growth is a result of selective homing of lymphocytes versus macrophages in the α4(S988A) mice. Among

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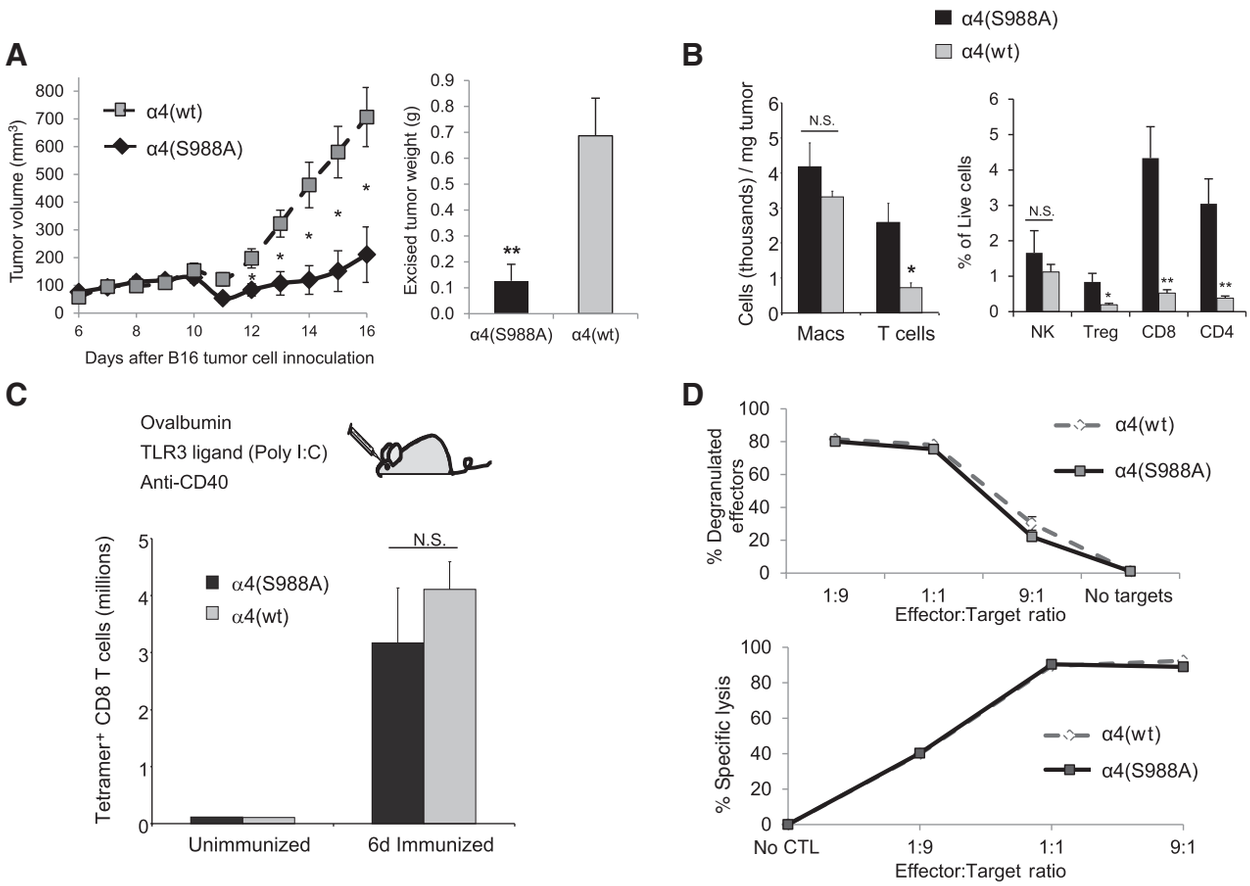

Figure 3.

Blocking α4 integrin phosphorylation increases tumor-infiltrating T cells and reduces tumor growth in mice. A, Reduced tumor growth in α4 (S988A) mice. B16 melanoma cells (3×10^5) were injected into α4(S988A) or control α4(wt) mice. Tumor area was measured daily and converted to an ellipsoid volume: (length × width^2)/2. Fifteen days later, tumors were excised and weighed. B, α4(S988A) T cells migrate more efficiently to a tumor site. B16 tumors were grown as in A. On day 15, mice were sacrificed, and excised tumors were weighed, digested with collagenase, and stained for (CD45$^+$) CD11b$^+$ macrophages, CD3$^+$ T cells, CD4$^+$ T cells, CD8$^+$ T cells, Foxp3$^+$(CD4$^+$) Treg, and NK1.1$^+$ NK cells. Error bars, SEM of ≥10 mice per group. *, $P < 0.015$; **, $P < 0.002$. C, T-cell clonal expansion. α4(S988A) and α4(wt) control littermate mice were immunized with ovalbumin protein, anti-CD40, and Poly I:C, i.p. in PBS. Six days later, splenocytes were isolated and stained with anti-CD8 antibody and an H-2K^b-SIINFEKL tetramer. D, T-cell cytotoxic function. Splenocytes of 8- to 12-week-old *OT-I*$^+$ α4(S988A) or OT-1$^+$ α4(wt) control littermate mice were differentiated to CTLs *in vitro* with SIINFEKL (1 μg/mL) and IL2 for 5 days and cultured for 2 hours at the indicated effector-to-target ratios (E:T) in the presence of anti–LAMP-1 antibody, followed by staining for CD8 and flow cytometric analysis. For target lysis, CTLs were cultured with SIINFEKL-pulsed or -unpulsed targets that had been differentially labeled with carboxyfluorescein diacetate succinimidyl ester. Specific killing was detected by flow cytometry as the decrease in the percentage of specific targets. No statistically significant differences were noted, with the exception of the 9:1 E:T ratio, at which the α4(S988A)-specific killing was 3.7% lower than that for α4(wt) CTLs ($P < 0.005$). $n = 3$ mice per group. Experiment was performed twice.

lymphoid cells, tumors in α4(S988A) mice had significantly greater numbers of CD4$^+$, CD8$^+$, and regulatory T cells; NK-cell abundance showed a modest, but statistically insignificant, increase compared with that of controls (Fig. 3B, right). The increased density of tumor-infiltrating T cells seen in α4(S988A) mice could be due to subtle, unrelated immunologic changes in this strain such as greater clonal expansion of tumor-specific T cells; however, expansion of antigen-specific T cells (SIINFEKL-tetramer$^+$) was similar between α4(S988A) and α4(wt) mice in response to immunization with ovalbumin (Fig. 3C). Furthermore, the cytotoxic function of α4(S988A) CD8$^+$ T cells was nearly identical to that of wt (Fig. 3D) cells as measured by degranulation of α4(S988A) OT-1 CD8$^+$ T cells and specific lysis of SIINFEKL-pulsed target cells. Thus, we concluded that the reduction in tumor growth observed in α4(S988A) mice is largely ascribable to increased lymphoid-cell homing. As an additional test of this conclusion, we used antibodies to deplete the majority of CD4 T, CD8 T, and NK cells. Depleting these multiple lymphoid subsets reversed the reduction in tumor growth in α4(S988A) relative to wt mice (Supplementary Fig. S6A). In contrast, partial depletion of CD8 T cells did not abolish the reduced tumor growth in α4(S988A) mice (Supplementary Fig. S6B). Thus, the capacity of the α4(S988A) mutation to increase the recruitment of multiple lymphocyte subsets is responsible for its ability to enhance tumor resistance.

The differential requirement of β2 integrins for migration of lymphocytes or myeloid cells *in vivo* can account for the remarkable leukocyte specificity of this form of trans-regulation. Whereas αLβ2 plays a major primary role in the migration of T cells to inflammatory sites (17, 21–23), macrophage migration to

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inflamed peritoneum is not dependent on β2 integrins and is reported to be solely dependent on α4β1 (24, 25). Thus, trans-regulation of migration would be absent in monocytes and macrophages because β2 integrins are not utilized. Indeed, we did not observe increased migration of macrophages from α4(S988A) or α4(wt) mice (Supplementary Fig. S7) on mixed ICAM-1–VCAM-1 substrates.

The finding that α4β1 trans-regulation of αLβ2 integrin–mediated migration promotes tumor immunity has important therapeutic implications. Inhibiting focal adhesion kinase (FAK) can suppress such trans-regulation (8), a finding that sounds a cautionary note in the use of FAK inhibitors in tumor therapy (26–28) and suggests that the effect of these agents on lymphocyte trafficking to tumors should be evaluated. Homing of infused lymphocytes is currently a rate-limiting step in applying T-cell immunotherapy to solid tumor cancers (9). Because lymphocytes are modified *ex vivo* before adoptive transfer for immunotherapy (11–14), the opportunity exists to simultaneously optimize their homing capacity by increasing integrin trans-regulation. α4 phosphorylation is type I-PKA–dependent (29); thus, increased trans-regulation could be induced by introduction of a dominant α4(S988A) integrin subunit, a cell permeating type I–specific A-kinase anchor protein (AKAP) peptide, or genetically encoded type I–specific PKA inhibitor (29, 30). Increased integrin trans-regulation might also have utilities beyond tumor immunotherapy. Migration and survival of long-lived plasma cells in the bone marrow appear to be dependent on α4 and αLβ2 integrins (31–35) and could involve integrin trans-regulation. Thus, enhanced integrin trans-regulation can offer a new approach to selectively potentiate β2 integrin–mediated homing of lymphocytes and plasma cells.

Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed.

Authors' Contributions

Conception and design: J.M. Cantor, D.M. Rose, M.H. Ginsberg
Development of methodology: D.M. Rose, M.H. Ginsberg
Acquisition of data (provided animals, acquired and managed patients, provided facilities, etc.): J.M. Cantor, D.M. Rose, M. Slepak
Analysis and interpretation of data (e.g., statistical analysis, biostatistics, computational analysis): J.M. Cantor, D.M. Rose, M.H. Ginsberg
Writing, review, and/or revision of the manuscript: J.M. Cantor, D.M. Rose, M.H. Ginsberg
Administrative, technical, or material support (i.e., reporting or organizing data, constructing databases): M. Slepak
Study supervision: M.H. Ginsberg

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