

Elevated Platelet Count Appears to Be Causally Associated with Increased Risk of Lung Cancer: A Mendelian Randomization Analysis



Ying Zhu¹, Yongyue Wei^{1,2,3}, Ruyang Zhang^{1,2,3}, Xuesi Dong⁴, Sipeng Shen², Yang Zhao^{1,3}, Jianling Bai¹, Demetrius Albanes⁵, Neil E. Caporaso⁵, Maria Teresa Landi⁵, Bin Zhu⁵, Stephen J. Chanock⁵, Fangyi Gu⁶, Stephen Lam⁷, Ming-Sound Tsao⁸, Frances A. Shepherd⁸, Adonina Tardon⁹, Ana Fernández-Somoano⁹, Guillermo Fernandez-Tardon⁹, Chu Chen¹⁰, Matthew J. Barnett¹⁰, Jennifer Doherty¹⁰, Stig E. Bojesen^{11,12,13}, Mattias Johansson¹⁴, Paul Brennan¹⁴, James D. McKay¹⁴, Robert Carreras-Torres¹⁴, Thomas Muley^{15,16}, Angela Risch^{16,17,18}, Heunz-Erich Wichmann¹⁹, Heike Bickeboeller²⁰, Albert Rosenberger²⁰, Gad Rennert²¹, Walid Saliba²¹, Susanne M. Arnold²², John K. Field²³, Michael P.A. Davies²³, Michael W. Marcus²³, Xifeng Wu²⁴, Yuanqing Ye²⁴, Loic Le Marchand²⁵, Lynne R. Wilkens²⁵, Olle Melander²⁶, Jonas Manjer²⁶, Hans Brunnström²⁷, Rayjean J. Hung²⁸, Geoffrey Liu²⁸, Yonathan Brhane²⁸, Linda Kachuri²⁸, Angeline S. Andrew²⁹, Eric J. Duell³⁰, Lambertus A. Kiemeny³¹, Erik HFM van der Heijden³¹, Aage Haugen³², Shanbeh Zienolddiny³², Vidar Skaug³², Kjell Grankvist³³, Mikael Johansson³³, Penella J. Woll³⁴, Angela Cox³⁴, Fiona Taylor³⁴, Dawn M. Teare³⁵, Philip Lazarus³⁶, Matthew B. Schabath³⁷, Melinda C. Aldrich³⁸, Richard S. Houlston³⁹, John McLaughlin⁴⁰, Victoria L. Stevens⁴¹, Hongbing Shen⁴², Zhibin Hu⁴², Juncheng Dai⁴², Christopher I. Amos⁴³, Younghun Han⁴³, Dakai Zhu⁴³, Gary E. Goodman⁴⁴, Feng Chen^{1,3}, and David C. Christiani^{1,2,3}

¹Department of Biostatistics, School of Public Health, Nanjing Medical University, Nanjing, China. ²Department of Environmental Health, Harvard School of Public Health, Boston, Massachusetts. ³China International Cooperation Center (CICC) for Environment and Human Health, Nanjing Medical University, Nanjing, China. ⁴Department of Epidemiology and Biostatistics, School of Public Health, Southeast University, Nanjing, China. ⁵Division of Cancer Epidemiology and Genetics, NCI, NIH, Bethesda, Maryland. ⁶Department of Cancer Prevention and Control, Roswell Park Comprehensive Cancer Center, Buffalo, New York. ⁷British Columbia Cancer Agency, Vancouver, British Columbia, Canada. ⁸University Health Network, Princess Margaret Cancer Centre, Toronto, Ontario, Canada. ⁹University of Oviedo and CIBERESP, Faculty of Medicine, Oviedo, Spain. ¹⁰Program in Epidemiology, Fred Hutchinson Cancer Research Center, Seattle, Washington. ¹¹Department of Clinical Biochemistry, Herlev and Gentofte Hospital, Copenhagen University Hospital, Copenhagen, Denmark. ¹²Faculty of Health and Medical Sciences, University of Copenhagen, Copenhagen, Denmark. ¹³Copenhagen General Population Study, Herlev and Gentofte Hospital, Copenhagen, Denmark. ¹⁴International Agency for Research on Cancer, World Health Organization, Lyon, France. ¹⁵Thoraxklinik at University Hospital Heidelberg, Heidelberg, Germany. ¹⁶Translational Lung Research Center Heidelberg (TLRC-H), Heidelberg, Germany. ¹⁷German Center for Lung Research (DZL), Heidelberg, Germany. ¹⁸University of Salzburg and Cancer Cluster Salzburg, Salzburg, Austria. ¹⁹Research Unit of Molecular Epidemiology, Institute of Epidemiology II, Helmholtz Zentrum München, German Research Center for Environmental Health, Neuherberg, Germany. ²⁰Department of Genetic Epidemiology, University Medical Center, Georg-August-University Göttingen, Germany. ²¹Department of Community Medicine and Epidemiology, Clalit National Cancer Control Center at Carmel Medical Center and Technion Faculty of Medicine, Technion-Israel Institute of Technology, Haifa, Israel. ²²Markey Cancer Center, University of Kentucky, Lexington, Kentucky. ²³Institute of Translational Medicine, University of Liverpool, Liverpool, United Kingdom. ²⁴Department of Epidemiology, University of Texas MD Anderson Cancer Center, Houston, Texas. ²⁵Epidemiology Program, University of Hawaii Cancer Center, Honolulu, Hawaii. ²⁶Faculty of Medicine, Lund University, Lund, Sweden. ²⁷Department of Pathology, Lund University, Lund, Sweden. ²⁸Lunenfeld-Tanenbaum Research Institute, Sinai Health System, University of Toronto, Toronto, Ontario, Canada. ²⁹Department of Epidemiology, Geisel School of Medicine, Hanover, New Hampshire. ³⁰Unit of

Nutrition and Cancer, Catalan Institute of Oncology (ICO-IDIBELL), Barcelona, Spain. ³¹Radboud University Medical Center, Radboud Institute for Health Sciences, Nijmegen, the Netherlands. ³²National Institute of Occupational Health, Oslo, Norway. ³³Department of Medical Biosciences, Umeå University, Umeå, Sweden. ³⁴Department of Oncology and Metabolism, University of Sheffield, Sheffield, United Kingdom. ³⁵School of Health and Related Research, University of Sheffield, England, United Kingdom. ³⁶Department of Pharmaceutical Sciences, College of Pharmacy, Washington State University, Spokane, Washington. ³⁷Department of Cancer Epidemiology, H. Lee Moffitt Cancer Center and Research Institute, Tampa, Florida. ³⁸Department of Thoracic Surgery, Division of Epidemiology, Vanderbilt University Medical Center, Nashville, Tennessee. ³⁹The Institute of Cancer Research, London, England. ⁴⁰Public Health Ontario, Toronto, Ontario, Canada. ⁴¹American Cancer Society, Inc., Atlanta, Georgia. ⁴²Department of Epidemiology and Biostatistics, Jiangsu Key Lab of Cancer Biomarkers, Prevention and Treatment, Collaborative Innovation Center for Cancer Personalized Medicine, School of Public Health, Nanjing Medical University, Nanjing, China. ⁴³Biomedical Data Science, Geisel School of Medicine at Dartmouth, Hanover, New Hampshire. ⁴⁴Swedish Medical Group, Seattle, Washington.

Note: Supplementary data for this article are available at Cancer Epidemiology, Biomarkers & Prevention Online (<http://cebp.aacrjournals.org/>).

Y. Zhu, Y. Wei, and R. Zhang contributed equally to this article.

D.C. Christiani is the senior author of this article.

Corresponding Authors: Feng Chen, School of Public Health (SPH), Nanjing Medical University, 818 East Tianyuan Road, Jiangning District, Nanjing, 211166, China. Phone: 86-25-8686-8414; Fax: 86-25-8686-8435; E-mail: fengchen@njmu.edu.cn; and David C. Christiani, Harvard School of Public Health, Building I, Room 1401, 665 Huntington Avenue, Boston, MA 02115. E-mail: dchris@hsph.harvard.edu

doi: 10.1158/1055-9965.EPI-18-0356

©2019 American Association for Cancer Research.

Abstract

Background: Platelets are a critical element in coagulation and inflammation, and activated platelets are linked to cancer risk through diverse mechanisms. However, a causal relationship between platelets and risk of lung cancer remains unclear.

Methods: We performed single and combined multiple instrumental variable Mendelian randomization analysis by an inverse-weighted method, in addition to a series of sensitivity analyses. Summary data for associations between SNPs and platelet count are from a recent publication that included 48,666 Caucasian Europeans, and the International Lung Cancer Consortium and Transdisciplinary Research in Cancer of the Lung data consisting of 29,266 cases and 56,450 controls to analyze associations between candidate SNPs and lung cancer risk.

Results: Multiple instrumental variable analysis incorporating six SNPs showed a 62% increased risk of overall non-small cell lung cancer [NSCLC; OR, 1.62; 95% confidence interval (CI), 1.15–2.27; $P = 0.005$] and a 200% increased risk for small-cell lung cancer (OR, 3.00; 95% CI, 1.27–7.06; $P = 0.01$). Results showed only a trending association with NSCLC histologic subtypes, which may be due to insufficient sample size and/or weak effect size. A series of sensitivity analysis retained these findings.

Conclusions: Our findings suggest a causal relationship between elevated platelet count and increased risk of lung cancer and provide evidence of possible antiplatelet interventions for lung cancer prevention.

Impact: These findings provide a better understanding of lung cancer etiology and potential evidence for antiplatelet interventions for lung cancer prevention.

Introduction

Lung cancer, a highly invasive, rapidly metastasizing cancer, has been the leading cause of cancer-related deaths worldwide for decades, accounting for more than one million deaths each year (1). Smoking is a major risk factor for lung cancer and accounts for about 80% of male and 50% of female lung cancer cases (2). In addition, environmental–occupational exposures (3, 4), lifestyle, and genetic variants (5) have been broadly explored as risks/predisposing factors for lung cancer. However, aspects of lung cancer risk remain largely unexplained and thus warrant further study.

The lung was recently noted to play a major role in platelet biogenesis and act as an ideal bioreactor for production of mature platelets from megakaryocytes, which account for approximately 50% of total platelet production (6). Platelets are an important element in coagulation and inflammation, and diverse mechanisms link activated platelets to cancer progression (7, 8). It has been identified that several variants in those chromosomal regions associated with platelet count have associations with myocardial infarction and autoimmune and hematologic disorders. Tumor-educated blood platelets have emerged as promising biomarker sources for noninvasive detection of cancer, and it was demonstrated to discriminate patients with non-small cell lung cancer (NSCLC) from healthy individuals and patients with various noncancerous inflammatory conditions (9, 10). Indeed, high platelet count is associated with increased mortality in a variety of cancers, including malignant mesothelioma (11), gynecologic malignancies (12), and breast cancer (13). In addition, platelet-to-lymphocyte ratio and mean platelet volume also add value in early diagnosis of lung cancer (14) and prognosis prediction (15, 16). These findings, taken together, indicate that disordered platelet production may be connected to lung carcinogenesis. However, due to potential unmeasured confounders in observational studies, the association between platelet count and lung cancer risk remains unclear.

Mendelian randomization is based on the principle that an individual's genotype is randomized at conception (17) and utilizes genetic variants as instrumental variables for the association between phenotypic exposures and outcomes to eliminate bias due to unmeasured confounders. Genetic variants used as

instrumental variables should meet the following assumptions: (i) genetic variants are associated with exposure, (ii) genetic variants affect outcome only via the exposure, and (iii) genetic variants are not associated with any confounders of the exposure–outcome association (18). By finding a genetic marker that satisfies instrumental variable assumptions, Mendelian randomization analysis has been broadly used to estimate unconfounded associations between exposure and outcome (19), such as the effect of higher adult height on escalated cancer risk (20–24).

In this study, we performed summary data–based Mendelian randomization (25) analysis, which is the extension of two sample Mendelian randomization, using curated platelet count–related SNPs as instrumental variables to evaluate the association between platelet count and lung cancer risk by using summary statistics from recent large-scale genome-wide association studies (GWAS).

Materials and Methods

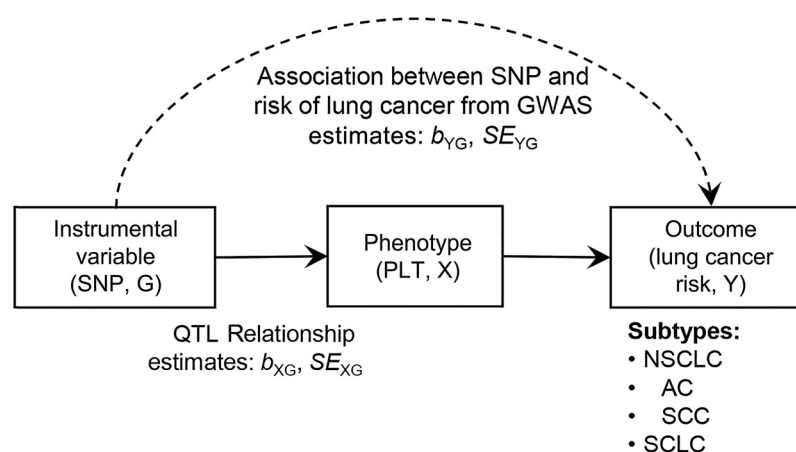
Data source and study population

Mendelian randomization analysis was conducted to estimate the effect of platelet count (X) on risk of lung cancer (Y) using genetic variants (G) as instrumental variables (26). According to the Mendelian randomization analysis diagram described in Fig. 1, we used coefficients of genetic variants on platelet count (b_{XC}) and their standard errors (SE_{XC}) from the recently published study of Gieger and colleagues, which pooled 23 studies and included approximately 48,666 individuals of European descent (27).

A total of 54 genetic variants were identified that were associated with platelet count (Supplementary Table S1). One of the key assumptions underlying Mendelian randomization is that the genetic variants (SNPs) used as instrumental variables are only related to the outcome of interest through the exposure variable under study. No pleiotropic pathways should exist from platelet-related SNPs to lung cancers through intermediates other than platelet count. Thus, six genetic variants (rs17030845, rs6141, rs3792366, rs210134, rs708382, and rs6065) were further selected as qualified instrumental variables that have prior functional knowledge supporting their association with platelets and no apparent link to cancer through intermediates other than

Figure 1.

Diagram of Mendelian randomization analysis. Mendelian randomization aims to estimate the unbiased causal relationship between platelet count (PLT) and lung cancer risk by incorporating genetic variants as instrumental variables (IVs). Dashed line represents the association between instrumental variable (SNP) and outcome (risk of lung cancer), denoted using b_{YG} in log(OR) scale and its standard error (SE_{YG}), which were obtained from GWAS. Estimates of quantitative trait loci relationship between SNP and phenotype (platelet count) were obtained from a recently published article and were described by b_{XG} and SE_{XG} . Lung cancer risk was assessed for NSCLC, adenocarcinoma (AC), SCC, and SCLC.



platelets. By the way, the SNP rs6141 in THPO narrowly misses the level required for nominal significance ($P < 5 \times 10^{-8}$) with $P = 6.18 \times 10^{-8}$ in Europeans, but shows genome-wide significance in Japanese (28). Therefore, it is still included serving as instrument variable for platelet count.

Coefficients (b_{YG}) and corresponding standard errors (SE_{YG}) of the association between genetic variants and lung cancer risk were obtained from meta-analysis of existing OncoArray and TRICL GWAS studies, which were detailed previously (29). Briefly, overall NSCLC samples were composed from OncoArray and TRICL GWASs, including 29,266 cases and 56,450 controls, and subgroup analyses were performed for 11,273 adenocarcinoma, 7,426 squamous cell carcinoma (SCC), and 2,664 small-cell lung cancer (SCLC) cases (Supplementary Table S2).

Mendelian randomization analysis

Mendelian randomization analysis with multiple instrumental variables was performed using an inverse-variance weighted (IVW) method combining the effect of genetic variants by weighted score. This score was used as an instrumental variable to estimate the effect of platelet count on lung cancer risk (26):

$$\hat{b}_{YX_{IVW}} = \frac{\sum_{i=1}^N \left(\frac{b_{XG_i} b_{YG_i}}{SE_{YG_i}^2} \right)}{\sum_{i=1}^N \left(\frac{b_{XG_i}}{SE_{YG_i}} \right)^2}, SE_{YX_{IVW}} = \sqrt{\frac{1}{\sum_{i=1}^N \left(\frac{b_{XG_i}}{SE_{YG_i}} \right)^2}}$$

In which $N = 6$ represents the number of instrumental variables included, and $b_{YX_{IVW}}$ and $SE_{YX_{IVW}}$ represent the effect of platelet count on lung cancer risk in log(OR) scale and its corresponding SE. Associations of platelet count on risk of overall NSCLC and individual subtypes were analyzed. Results

are presented as OR for lung cancer risk per $100 \times 10^9/L$ increment of platelet count.

In addition, penalized IVW, robust IVW, MR-Egger, penalized MR-Egger, and robust MR-Egger methods were used for sensitivity analyses to evaluate robustness of the findings (30). Step forward modeling was used to add an optimal instrumental variable each time from the left 48 SNPs, adding to the six curated SNPs for multiple instrumental variable analysis, until there was no improvement of statistical significance (P) for the test of causal effect. The modeling process was terminated when no added SNP increased $-\log_{10}(P)$ by 20% or 10%. Besides, Mendelian randomization analysis with a single-instrumental variable (one SNP at a time) was performed as supplementary. Effect of platelet count on lung cancer risk [b_{YX} in log(OR) scale] and its standard error (SE_{YX}) were estimated as follows (31):

$$\hat{b}_{YX} = \frac{b_{YG}}{b_{XG}}, SE_{YX} = \frac{SE_{YG}}{b_{XG}}$$

All analyses were performed using R Software Version 3.3.1 (The R Foundation). All tests were two-sided, and $P \leq 0.05$ was considered statistically significant unless stated otherwise.

Results

Among 48,666 Europeans, 54 SNPs were quantitatively associated with platelet count with $P \leq 5 \times 10^{-8}$ (Supplementary Table S1; ref. 27). Associations of those 54 SNPs with risk of lung cancer were analyzed among 29,266 cases and 56,450 controls from OncoArray and previous GWAS studies. Demographics and study descriptions were detailed previously (29) and are briefly listed in Supplementary Table S2 as well. Summarized association results of SNPs and lung cancer risk are listed in Supplementary

Table 1. SNPs of specific platelet-related genes

SNP	Chr:position (hg19)	Gene	Reference allele	Effect allele	EAF (%)	Function	b (95% CI)	P
rs17030845	2: 43687879	THADA	C	T	9.65	Intron	-3.58 (-4.67 to -2.49)	1.27×10^{-10}
rs6141	3: 184090266	THPO	T	C	47.39	3' UTR	-2.47 (-3.36 to -1.57)	6.18×10^{-8}
rs3792366	3: 122839876	PDIA5	A	G	38.68	Intron	2.153 (1.44-2.87)	3.60×10^{-9}
rs210134	6: 33540209	BAK1	G	A	29.29	500 bp Downstream	-4.96 (-5.73 to -4.18)	7.11×10^{-36}
rs708382	17: 42442344	FAM171A2-ITGA2B	T	C	39.66	2 kb Upstream	-2.44 (-3.28 to -1.59)	1.51×10^{-8}
rs6065	17: 4836381	GPIBA	C	T	8.53	Missense	4.19 (2.96-5.43)	2.92×10^{-11}

Abbreviations: CI, confidence interval; EAF, effect allele frequency; UTR, untranslated region.

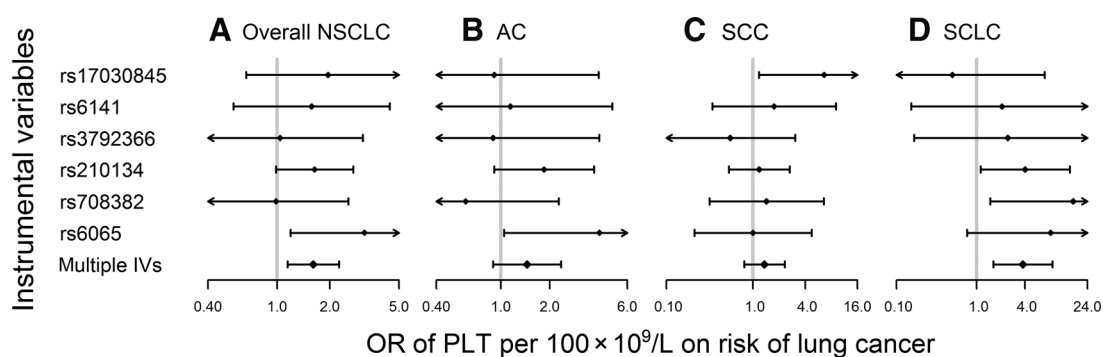


Figure 2. Causal associations between platelet count and lung cancer risk. Forest plots of causal associations between platelet count (PLT) and risk of lung cancer using Mendelian randomization analysis incorporating different genetic variants as instrumental variables (IVs). Associations of platelet count with risk of NSCLC (A), adenocarcinoma (AC; B), SCC (C), and SCLC (D) were analyzed based on single-instrumental variable or multiple instrumental variables using IVW analysis.

Table S3. According to instrumental variable assumptions that had evidence only related to platelets, 6 SNPs which are relatively independent and situated in different chromosomes were selected for Mendelian randomization analysis (Table 1), and 48 SNPs were excluded (Supplementary Table S4).

In multiple instrumental variable analysis combining all six relatively independent SNPs situated in different chromosomes, a significant association between platelet count and overall NSCLC risk is revealed, showing that each $100 \times 10^9/L$ increment of platelet count was associated with a 62% increase in NSCLC risk [95% confidence interval (CI), 1.15–2.27; $P = 0.005$; Figs. 2A and 3A]. In addition, five different methods of sensitivity analysis, including penalized IVW, robust IVW, MR-Egger, penalized MR-Egger, and robust MR-Egger, retained this association (Table 2). In NSCLC subtype analysis, it failed to detect significant associations between platelet count and the risk of lung adenocarcinoma (OR, 1.51; 95% CI, 0.92–2.48; $P = 0.11$; Figs. 2B and 3B) and SCC (OR, 1.59; 95% CI, 0.86–2.92; $P = 0.14$; Figs. 2C and 3C). On the

other hand, it is suggested that platelet count is significantly associated with the risk of SCLC (OR, 3.00; 95% CI, 1.27–7.06; $P = 0.01$; Figs. 2D and 3D). The results of single-instrumental variable are presented in Supplementary Table S5. No correction was conducted for them because a single-weak instrument will have lower power to reject the null hypothesis (32).

We also performed a step forward modeling strategy to include more instrumental SNPs in the multiple instrumental variable model. Including more SNPs as instrumental variables yielded similar, yet more significant, causal estimates (Supplementary Table S6; Supplementary Fig. S1).

Discussion

This Mendelian randomization study suggests that each $100 \times 10^9/L$ increment in platelets results in a 62% increased risk of NSCLC and, notably, a 200% increased risk of SCLC. However, this study failed to show evidence of a relationship between

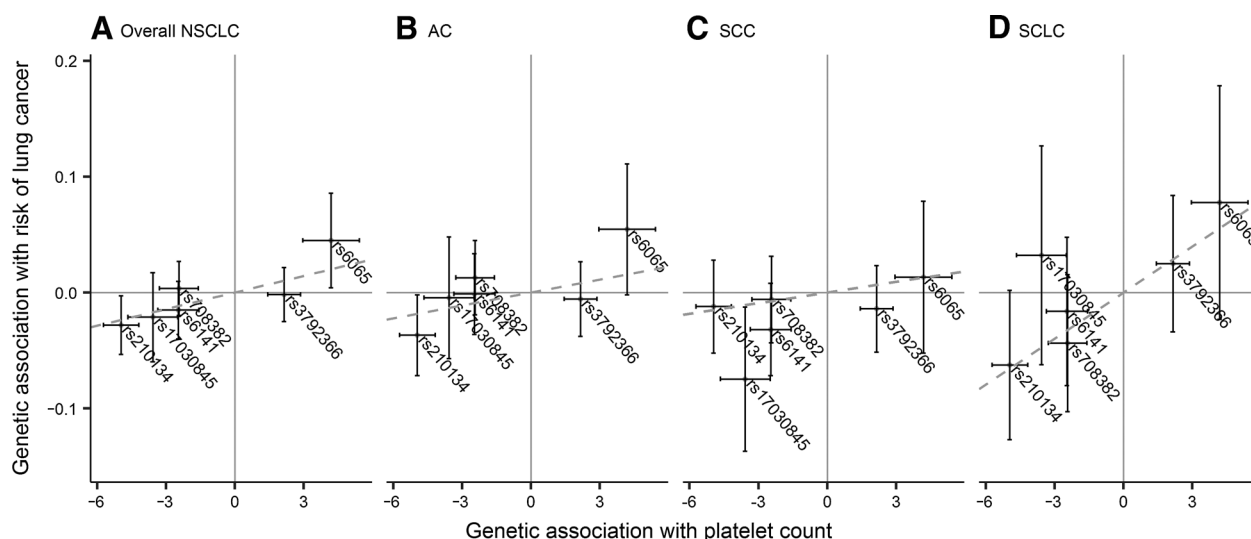


Figure 3. Associations between SNPs and lung cancer risk. Scatter plots displaying estimates of the association between each SNP and risk of lung cancer against quantitative relationship of each SNP on platelet count for NSCLC (A), adenocarcinoma (AC; B), SCC (C), and SCLC (D). Slope of the gray dashed line through the plot represents IVW regression estimate for the causal effect of platelet count on lung cancer risk.

Table 2. Association between platelet count and risk of lung cancer using multiple instrumental variable analysis

SNP	Overall NSCLC		Adenocarcinoma		SCC		SCLC	
	OR (95% CI)	P	OR (95% CI)	P	OR (95% CI)	P	OR (95% CI)	P
IVW	1.62 (1.15–2.27)	0.005	1.51 (0.92–2.48)	0.11	1.59 (0.86–2.92)	0.14	3.00 (1.27–7.06)	0.01
Penalized IVW	1.62 (1.15–2.27)	0.005	1.51 (0.92–2.48)	0.11	1.59 (0.86–2.92)	0.14	3.00 (1.27–7.06)	0.01
Robust IVW	1.63 (1.26–2.11)	<0.001	1.51 (0.90–2.53)	0.12	1.54 (0.93–2.56)	0.09	3.30 (1.52–7.15)	0.003
MR-Egger	3.25 (1.16–9.11)	0.03	6.06 (1.45–25.27)	0.01	1.75 (0.22–13.84)	0.59	3.29 (0.24–45.11)	0.37
Penalized MR-Egger	3.25 (1.16–9.11)	0.03	6.06 (1.45–25.27)	0.01	1.75 (0.22–13.84)	0.59	3.29 (0.24–45.11)	0.37
Robust MR-Egger	3.23 (1.80–5.78)	<0.001	5.88 (2.74–12.61)	<0.001	1.70 (0.48–6.08)	0.41	3.56 (1.25–10.14)	0.02

NOTE: OR of platelet count on lung cancer risk per $100 \times 10^9/L$ increment of platelet count.

platelet count and risk of adenocarcinoma and SCC, probably resulting from insufficient sample size. As comparing with SCLC, the effect size of platelet count on adenocarcinoma and SCC are weaker, larger sample size is needed (33).

Platelets have been studied for decades as an important regulator of inflammation and thrombosis (34), which are broadly interrelated with human carcinogenesis (13). Platelets are also recognized as a stimulator of proangiogenic factors (13) and a major source of VEGF (35), platelet-derived growth factor (36, 37), and basic fibroblast growth factor (37), which act as promoters of tumor growth in lung (38–44). New evidence suggests that platelets are relevant to defensive, physiologic immune responses of the lungs and to inflammatory lung diseases (45). Thus, higher platelet count has a potential biological connection to increased risk of lung cancer. Interestingly, p-selectin, an important adhesion molecule expressed on the surface of activated platelets, is more highly expressed in lung adenocarcinomas and SCC than in healthy populations (46). These results indicate a considerable role of platelets in lung carcinogenesis.

Intriguingly, a recent study indicates that cancer cells depend on platelets to avoid anoikis and succeed in metastasis (47). Platelets induce resistance to anoikis *in vitro* and are critical for metastasis *in vivo* by activating RhoA-MYPT1-PP1-mediated YAP1 dephosphorylation and promoting its nuclear translocation to inhibit apoptosis. However, the unknown underlying mechanism warrants future well-designed functional experiments to clarify the role of platelets in these cellular processes.

In addition, antiplatelet agents, such as purinergic antagonists, are used clinically because they affect inflammatory pathways (48). Recent publications demonstrate that platelets suppress T-cell responses against tumors through production and activation of immunosuppressive factors. These results suggest the use of a combination of immunotherapy and platelet inhibitors, such as aspirin (49, 50) and clopidogrel, as a therapeutic strategy against cancer (51, 52). Therefore, it is possible that antiplatelet therapy could reduce lung cancer risk.

However, we acknowledge some limitations in our study. First, some associations between genetic instrumental variables and phenotype (platelet count) were insufficient and thus may result in a "weak instrument" phenomenon (53). Second, in some scenarios, inconsistent results were observed between INW and MR-Egger (or regular and penalized/robust) models. This phenomenon indicates that genetic variants probably have horizontal pleiotropy, and thus Mendelian randomization assumptions are likely violated (54). Moreover, there is heterogeneity across results incorporating different SNP sets as instrumental variables, which indicates that the instrumental variable should be curated carefully before Mendelian randomization analysis. In this study, all platelet count-related SNPs were curated, and six were retained to better satisfy Mendelian

randomization assumptions. Third, a linear association was assumed between platelet count and lung cancer risk. However, the shape could be nonlinear and thus warrants further study incorporating individual-level data. Fourth, we only evaluated platelet count as a potential causal factor, whereas platelet function plays a comparable causal role in this pathway. More detailed platelet information should be measured in future studies, including immature platelet fractions and function. In addition, we assumed that study populations used for the genetic instrument for platelet count and for risk of lung cancer were representative of the same general Caucasian population, which may not be true. Therefore, additional functional studies are needed to further evaluate the mechanisms that underlie associations between platelets and lung cancer risk.

Nonetheless, our findings do suggest a role of platelet count in risk of lung cancer. The results provide a better understanding of lung cancer etiology and evidence for a possible role of antiplatelet interventions in lung cancer prevention.

Disclosure of Potential Conflicts of Interest

G. Liu has received speakers bureau honoraria from Pfizer, Astra Zeneca, Takeda, Roche, Novartis, BMS, and Merck. E.H.F.M. van der Heijden reports receiving other commercial research support from Philips Medical Systems and Astra Zeneca Oncology, and has received speakers bureau honoraria from Pentax Medical. No potential conflicts of interest were disclosed by the other authors.

Disclaimer

Sponsors had no role in the design of the study, collection and analysis of data, or preparation of the manuscript.

Authors' Contributions

Conception and design: Y. Wei, R. Zhang, P. Brennan, J.K. Field, J. McLaughlin, H. Shen, F. Chen, D.C. Christiani

Development of methodology: Y. Wei, R. Zhang, B. Zhu, P. Brennan, J. McLaughlin, D.C. Christiani

Acquisition of data (provided animals, acquired and managed patients, provided facilities, etc.): D. Albanes, N.E. Caporaso, M.T. Landi, B. Zhu, S.J. Chanock, S. Lam, M.-S. Tsao, F.A. Shepherd, A. Tardon, A. Fernández-Somoano, G. Fernandez-Tardon, C. Chen, J. Doherty, S.E. Bojesen, M. Johansson, P. Brennan, J.D. McKay, R. Carreras-Torres, T. Muley, A. Risch, H.-E. Wichmann, H. Bickeboeller, A. Rosenberger, G. Rennert, W. Saliba, S.M. Arnold, J.K. Field, M.P.A. Davies, M.W. Marcus, X. Wu, L. Le Marchand, L.R. Wilkens, O. Melander, J. Manjer, H. Brunnström, R.J. Hung, G. Liu, A.S. Andrew, E.J. Duell, L.A. Kiemeny, E.H.F.M. van der Heijden, A. Haugen, S. Zienolddiny, V. Skaug, K. Grankvist, M. Johansson, P.J. Woll, A. Cox, F. Taylor, D.M. Teare, P. Lazarus, M.B. Schabath, M.C. Aldrich, R.S. Houlston, J. McLaughlin, H. Shen, Z. Hu, J. Dai, C.I. Amos, G.E. Goodman, D.C. Christiani

Analysis and interpretation of data (e.g., statistical analysis, biostatistics, computational analysis): Y. Zhu, Y. Wei, R. Zhang, X. Dong, S. Shen, Y. Zhao, J. Bai, N.E. Caporaso, B. Zhu, P. Brennan, H. Bickeboeller, S.M. Arnold, M.W. Marcus, R.J. Hung, Y. Brhane, L. Kachuri, M.B. Schabath, Y. Han, G.E. Goodman, D.C. Christiani

Writing, review, and/or revision of the manuscript: Y. Zhu, Y. Wei, R. Zhang, Y. Zhao, J. Bai, D. Albanes, N.E. Caporaso, M.T. Landi, S.J. Chanock, F. Gu, S. Lam, M.-S. Tsao, F.A. Shepherd, A. Tardon, A. Fernández-Somoano, G. Fernandez-Tardon, C. Chen, J. Doherty, S.E. Bojesen, M. Johansson, P. Brennan, R. Carreras-Torres, T. Muley, A. Risch, H.-E. Wichmann, H. Bickeboeller, G. Rennert, W. Saliba, J.K. Field, M.P.A. Davies, M.W. Marcus, X. Wu, Y. Ye, L. Le Marchand, L.R. Wilkens, O. Melander, H. Brunnström, R.J. Hung, G. Liu, Y. Brhane, E.J. Duell, L.A. Kiemeny, E.H.F.M. van der Heijden, A. Haugen, V. Skaug, K. Grankvist, M. Johansson, P.J. Woll, A. Cox, D.M. Teare, P. Lazarus, M.B. Schabath, M.C. Aldrich, J. McLaughlin, V.L. Stevens, C.I. Amos, G.E. Goodman, F. Chen, D.C. Christiani

Administrative, technical, or material support (i.e., reporting or organizing data, constructing databases): Y. Zhao, M.J. Barnett, M. Johansson, P. Brennan, J.K. Field, M.P.A. Davies, Y. Ye, O. Melander, J. Manjer, G. Liu, L.A. Kiemeny, S. Zienolddiny, M.B. Schabath, C.I. Amos, D. Zhu, G.E. Goodman, F. Chen, D.C. Christiani

Study supervision: R. Zhang, P. Brennan, S.M. Arnold, R.J. Hung, L.A. Kiemeny, J. McLaughlin, H. Shen, F. Chen, D.C. Christiani

Other (final acceptance of manuscript): F.A. Shepherd

Acknowledgments

We thank the participants and staff for their important contributions to this study. This study was supported by the NIH (CA092824 and CA209414, to D.C. Christiani), National Natural Science Foundation of China (81530088 and 81473070, to F. Chen; 81373102, to Y. Zhao), State's Key Project of Research and Development Program (2016YFE0204900, to F. Chen), Key Project of Natural Science Foundation of Jiangsu, China (14JA31002, to F. Chen), A Project Funded by the Priority Academic Program Development of Jiangsu Higher Education Institutions (PAPD), and Outstanding Young Teachers Training Program of Nanjing Medical University (to Y.Y. Wei). Transdisciplinary Research for Cancer in Lung (TRICL) and Oncoarray funding sources are detailed as follows: Transdisciplinary Research for Cancer in Lung (TRICL) of the International Lung Cancer Consortium (ILCCO) was supported by grants U19-CA148127 and CA148127S1. ILCCO data harmonization was supported by the Cancer Care Ontario Research Chair of Population Studies (to R.J. Hung) and the Lunenfeld-Tanenbaum Research Institute, Sinai Health System. The CAPUA study was supported by FIS-FEDER/Spain grants FIS-01/310, FIS-PI03-0365, and FIS-07-BI060604; FICYT/Asturias grants FICYT PB02-67 and FICYT IB09-133; and the University Institute of Oncology (IUOPA) of the University of Oviedo and the Ciber de Epidemiologiay Salud Pública (CIBERESP), Spain. Work performed in the CARET study was supported by the NIH/NCI UM1 CA167462 (principal investigator: G.E. Goodman), NIH UO1-CA6367307 (principal investigators: Omen, G.E. Goodman), NIH R01 CA111703 (principal investigator: C. Chen), and NIH 5R01 CA151989-01A1 (principal investigator: J. Doherty). The Liverpool Lung project was supported by the Roy Castle Lung Cancer Foundation. The Harvard Lung Cancer Study was supported by the NIH/NCI grants CA092824, CA090578, and CA074386. The Multiethnic Cohort Study was partially supported by NIH grants CA164973, CA033619, CA63464, and CA148127. Work performed in the MSH-PMH study was supported by the Canadian Cancer Society Research Institute (020214), Ontario Institute of Cancer and Cancer Care Ontario Chair Award (to R.J. Hung and G. Liu), and Alan Brown Chair and Lusi Wong Programs at Princess Margaret Hospital Foundation. NJLCS was funded by the State Key Program of National Natural Science of China (81230067), National Key Basic Research Program (2011CB503805), and Major Program of the National Natural Science Foundation of China (81390543). The Norway study was supported by the Norwegian Cancer Society, Norwegian Research Council. The Shanghai Cohort Study (SCS) was supported by NIH R01 CA144034 (principal investigator: J.M. Yuan) and UM1 CA182876 (principal investigator: J.M. Yuan). The Singapore Chinese Health Study (SCHS) was supported by NIH R01 CA144034 (principal investigator: J.M. Yuan) and UM1 CA182876 (principal investigator: J.M. Yuan). Work in the TLC study has been supported in part by the James & Esther

King Biomedical Research Program (09KN-15), NIH Specialized Programs of Research Excellence (SPORE; P50 CA119997), and a Cancer Center Support Grant (CCSG) at the H. Lee Moffitt Cancer Center and Research Institute, an NCI designated Comprehensive Cancer Center (P30-CA76292). The Vanderbilt Lung Cancer Study – BioVU dataset used for the described analyses was obtained from Vanderbilt University Medical Center's BioVU, which is supported by institutional funding, the 1S10RR025141-01 instrumentation award, and the Vanderbilt Nature Genetics: doi:10.1038/ng.3892. The Clinical and Translational Science Awards (CTSA) grant UL1TR000445 was from the National Center for Advancing Translational Sciences (NCATS)/NIH. M.C. Aldrich was supported by NIH/NCI K07CA172294 (to principal investigator: M.C. Aldrich). The Copenhagen General Population Study (CGPS) was supported by the Chief Physician Johan Boserup and Lise Boserup Fund, the Danish Medical Research Council, and Herlev Hospital. The NELCS study was supported by grant P20RR018787 from the National Center for Research Resources (NCRR), a component of the NIH. The Kentucky Lung Cancer Research Initiative was supported by the Department of Defense (Congressionally Directed Medical Research Program, U.S. Army Medical Research, and Materiel Command Program) under award 10153006 (W81XWH-11-1-0781). This research was also supported by unrestricted infrastructure funds from the UK Center for Clinical and Translational Science, NIH grant UL1TR000117, and Markey Cancer Center NCI Cancer Center Support Grant (P30 CA177558), Shared Resource Facilities: Cancer Research Informatics, Biospecimen and Tissue Procurement, and Biostatistics and Bioinformatics. The M.D. Anderson Cancer Center study was supported in part by NIH grants P50 CA070907 and R01 CA176568 (to X. Wu), Cancer Prevention & Research Institute of Texas RP130502 (to X. Wu), and University of Texas MD Anderson Cancer Center institutional support for the Center for Translational and Public Health Genomics. The deCODE study of smoking and nicotine dependence was funded in part by grant R01-DA017932 from the National Institutes on Drug Abuse (NIDA). The Lodz center study was partially funded by Nofer Institute of Occupational Medicine under task NIOM 10.13: Predictors of mortality from non-small cell lung cancer field study. Genetic sharing analysis was funded by NIH grant CA194393. The ResoLuCENT study (Resource for the Study of Lung Cancer Epidemiology in North Trent) is funded by the Sheffield Hospitals Charity, Sheffield Experimental Cancer Medicine Centre, and Weston Park Hospital Cancer Charity. F. Taylor was supported by a Cancer Research UK/ Yorkshire Cancer Research Clinical Fellowship. B. Zhu's work was supported by the Intramural Research Program of the Division of Cancer Epidemiology and Genetics, NCI. The Environment And Genetics in Lung cancer Etiology (EAGLE) study (to principal investigator: M.T. Landi) was supported by the Intramural Research Program of NIH, NCI, Division of Cancer Epidemiology and Genetics. F. Gu was supported by Roswell Park Cancer Institute and Cancer Center Supporting Grant P30CA016056. L. Le Marchand was supported by a program project grant from the NCI, NIH: P01 CA168530 and grant U01 CA164973. The Toronto study (to principal investigator: J. McLaughlin) was supported by Canadian Cancer Society Research Institute (020214). The Canadian Urban Environmental Health Research Consortium is funded by the Canadian Institutes for Health Research (J. McLaughlin). S.J. Chanock was supported by the Intramural Research Program of the NIH NCI's Division of Cancer Epidemiology, and the American Cancer Society. This work was also supported by Cancer Research UK (C1298/A8362, to R.S. Houlston; C1298/A8780 and C1298/A8362, to J. McLaughlin; and C18281/A19169 to R. Carreras-Torres).

The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked *advertisement* in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

Received April 5, 2018; revised May 11, 2018; accepted January 17, 2019; published first January 30, 2019.

References

1. Siegel R, Ma J, Zou Z, Jemal A. Cancer statistics, 2014. *CA Cancer J Clin* 2014;64:9–29.
2. Jemal A, Bray F, Center MM, Ferlay J, Ward E, Forman D. Global cancer statistics. *CA Cancer J Clin* 2011;65:87–108.
3. Katanoda K, Sobue T, Satoh H, Tajima K, Suzuki T, Nakatsuka H, et al. An association between long-term exposure to ambient air pollution and mortality from lung cancer and respiratory diseases in Japan. *J Epidemiol* 2011;21:132–43.

4. Cohen BL. Testing a BEIR-VI suggestion for explaining the lung cancer vs. radon relationship for U.S. counties. *Health Phys* 2000; 78:522-7.
5. Kligerman S, White C. Epidemiology of lung cancer in women: risk factors, survival, and screening. *AJR Am J Roentgenol* 2011;196:287-95.
6. Lefrancais E, Ortiz-Munoz G, Caudrillier A, Mallavia B, Liu F, Sayah DM, et al. The lung is a site of platelet biogenesis and a reservoir for haematopoietic progenitors. *Nature* 2017;544:105-9.
7. Gasic GJ, Gasic TB, Stewart CC. Antimetastatic effects associated with platelet reduction. *Proc Natl Acad Sci U S A* 1968;61:46-52.
8. Ji Y, Sheng L, Du X, Qiu G, Su D. Elevated platelet count is a strong predictor of poor prognosis in stage I non-small cell lung cancer patients. *Platelets* 2015;26:138-42.
9. Best MG, Sol N, In 't Veld S, Vancura A, Muller M, Niemeijer AN, et al. Swarm intelligence-enhanced detection of non-small-cell lung cancer using tumor-educated platelets. *Cancer Cell* 2017;32: 238-52.e9.
10. Joosse SA, Pantel K. Tumor-educated platelets as liquid biopsy in cancer patients. *Cancer Cell* 2015;28:552-4.
11. Tural Onur S, Sokucu SN, Dalar L, Iliaz S, Kara K, Buyukkale S, et al. Are neutrophil/lymphocyte ratio and platelet/lymphocyte ratio reliable parameters as prognostic indicators in malignant mesothelioma? *Ther Clin Risk Manag* 2016;12:651-6.
12. Menczer J. Preoperative elevated platelet count and thrombocytosis in gynecologic malignancies. *Arch Gynecol Obstet* 2016;295:9-15.
13. Franco AT, Corken A, Ware J. Platelets at the interface of thrombosis, inflammation, and cancer. *Blood* 2015;126:582-8.
14. Nikolic I, Kukulj S, Samarzija M, Jelec V, Zarak M, Orehovec B, et al. Neutrophil-to-lymphocyte and platelet-to-lymphocyte ratio help identify patients with lung cancer, but do not differentiate between lung cancer subtypes. *Croat Med J* 2016;57:287-92.
15. Omar M, Tanriverdi O, Cokmert S, Oktay E, Yersal O, Pilanci KN, et al. Role of increased mean platelet volume (MPV) and decreased MPV/platelet count ratio as poor prognostic factors in lung cancer. *Clin Respir J* 2018;12: 922-9.
16. Oncel M, Kiyici A, Oncel M, Sunam GS, Sahin E, Adam B. Evaluation of platelet indices in lung cancer patients. *Asian Pac J Cancer Prev* 2015;16: 7599-602.
17. Palmer TM, Sterne JA, Harbord RM, Lawlor DA, Sheehan NA, Meng S, et al. Instrumental variable estimation of causal risk ratios and causal odds ratios in Mendelian randomization analyses. *Am J Epidemiol* 2010;173: 1392-403.
18. VanderWeele TJ, Tchetgen Tchetgen EJ, Cornelis M, Kraft P. Methodological challenges in mendelian randomization. *Epidemiology* 2014;25:427-35.
19. Didelez V, Sheehan N. Mendelian randomization as an instrumental variable approach to causal inference. *Stat Methods Med Res* 2007;16: 309-30.
20. Thrift AP, Risch HA, Onstad L, Shaheen NJ, Casson AG, Bernstein L, et al. Risk of esophageal adenocarcinoma decreases with height, based on consortium analysis and confirmed by Mendelian randomization. *Clin Gastroenterol Hepatol* 2014;12:1667-76.
21. Thrift AP, Gong J, Peters U, Chang-Claude J, Rudolph A, Slattery ML, et al. Mendelian randomization study of height and risk of colorectal cancer. *Int J Epidemiol* 2015;44:662-72.
22. Nuesch E, Dale C, Palmer TM, White J, Keating BJ, van Iperen EP, et al. Adult height, coronary heart disease and stroke: a multi-locus Mendelian randomization meta-analysis. *Int J Epidemiol* 2015;45: 1927-37.
23. Khankari NK, Shu XO, Wen W, Kraft P, Lindstrom S, Peters U, et al. Association between adult height and risk of colorectal, lung, and prostate cancer: results from meta-analyses of prospective studies and Mendelian randomization analyses. *PLoS Med* 2016;13:e1002118.
24. Davies NM, Gaunt TR, Lewis SJ, Holly J, Donovan JL, Hamdy FC, et al. The effects of height and BMI on prostate cancer incidence and mortality: a Mendelian randomization study in 20,848 cases and 20,214 controls from the PRACTICAL consortium. *Cancer Causes Control* 2015;26:1603-16.
25. Zhu Z, Zhang F, Hu H, Bakshi A, Robinson MR, Powell JE, et al. Integration of summary data from GWAS and eQTL studies predicts complex trait gene targets. *Nat Genet* 2016;48:481-7.
26. Burgess S, Butterworth A, Thompson SG. Mendelian randomization analysis with multiple genetic variants using summarized data. *Genet Epidemiol* 2013;37:658-65.
27. Gieger C, Radhakrishnan A, Cvejic A, Tang W, Porcu E, Pistis G, et al. New gene functions in megakaryopoiesis and platelet formation. *Nature* 2011; 480:201-8.
28. Kamatani Y, Matsuda K, Okada Y, Kubo M, Hosono N, Daigo Y, et al. Genome-wide association study of hematological and biochemical traits in a Japanese population. *Nat Genet* 2010;42:210-5.
29. McKay JD, Hung RJ, Han Y, Zong X, Carreras-Torres R, Christiani DC, et al. Large-scale association analysis identifies new lung cancer susceptibility loci and heterogeneity in genetic susceptibility across histological subtypes. *Nat Genet* 2017;49:1126-32.
30. Bowden J, Del Greco MF, Minelli C, Davey Smith G, Sheehan N, Thompson J. A framework for the investigation of pleiotropy in two-sample summary data Mendelian randomization. *Stat Med* 2017;36: 1783-802.
31. Burgess S, Small DS, Thompson SG. A review of instrumental variable estimators for Mendelian randomization. *Stat Methods Med Res* 2017;26: 2333-55.
32. Davies NM, von Hinke Kessler Scholder S, Farbmacher H, Burgess S, Windmeijer F, Smith GD. The many weak instruments problem and Mendelian randomization. *Stat Med* 2015;34:454-68.
33. Burgess S. Sample size and power calculations in Mendelian randomization with a single instrumental variable and a binary outcome. *Int J Epidemiol* 2014;43:922-9.
34. Thomas MR, Storey RF. The role of platelets in inflammation. *Thromb Haemost* 2015;114:449-58.
35. Verheul HM, Hoekman K, Luyck-de Bakker S, Eekman CA, Folman CC, Broxterman HJ, et al. Platelet: transporter of vascular endothelial growth factor. *Clin Cancer Res* 1997;3:2187-90.
36. Pinedo HM, Verheul HM, D'Amato RJ, Folkman J. Involvement of platelets in tumour angiogenesis? *Lancet* 1998;352:1775-7.
37. Cross MJ, Claesson-Welsh L. FGF and VEGF function in angiogenesis: signalling pathways, biological responses and therapeutic inhibition. *Trends Pharmacol Sci* 2010;22:201-7.
38. Ferrara N, Gerber HP, LeCouter J. The biology of VEGF and its receptors. *Nat Med* 2003;9:669-76.
39. Xiao XY, Lang XP. Correlation between MMP-7 and bFGF expressions in non-small cell lung cancer tissue and clinicopathologic features. *Cell Biochem Biophys* 2015;73:427-432.
40. Otaka Y, Rokudai S, Kaira K, Fujieda M, Horikoshi I, Iwakawa-Kawabata R, et al. SIXBP4 drives tumor growth and is associated with poor prognosis through PDGF receptor signaling in lung squamous cell carcinoma. *Clin Cancer Res* 2017;23:3442-52.
41. Naykoo NA, Dil A, Rasool R, Shah S, Ahangar AG, Bhat IA, et al. Single nucleotide polymorphisms, haplotype association and tumour expression of the vascular endothelial growth factor (VEGF) gene with lung carcinoma. *Gene* 2017;608:95-102.
42. Jahanban-Esfahlan R, Seidi K, Monfaredan A, Shafie-Irannejad V, Abbasi MM, Karimian A, et al. The herbal medicine *Melissa officinalis* extract effects on gene expression of p53, Bcl-2, Her2, VEGF-A and hTERT in human lung, breast and prostate cancer cell lines. *Gene* 2017; 613:14-9.
43. Hu M, Hu Y, He J, Li B. Prognostic value of basic fibroblast growth factor (bFGF) in lung cancer: a systematic review with meta-analysis. *PLoS One* 2016;11:e0147374.
44. Dadrich M, Nicolay NH, Flechsig P, Bickelhaupt S, Hoeltgen L, Roeder F, et al. Combined inhibition of TGFbeta and PDGF signaling attenuates radiation-induced pulmonary fibrosis. *Oncoimmunology* 2016;5: e1123366.
45. Middleton EA, Weyrich AS, Zimmerman GA. Platelets in pulmonary immune responses and inflammatory lung diseases. *Physiol Rev* 2016; 96:1211-59.
46. Gong L, Cai Y, Zhou X, Yang H. Activated platelets interact with lung cancer cells through P-selectin glycoprotein ligand-1. *Pathol Oncol Res* 2012;18: 989-96.
47. Haemmerle M, Taylor ML, Gutschner T, Pradeep S, Cho MS, Sheng J, et al. Platelets reduce anoikis and promote metastasis by activating YAP1 signaling. *Nat Commun* 2017;8:310.

48. Pitchford SC. Novel uses for anti-platelet agents as anti-inflammatory drugs. *Br J Pharmacol* 2007;152:987–1002.
49. Cao Y, Nishihara R, Wu K, Wang M, Ogino S, Willett WC, et al. Population-wide impact of long-term use of aspirin and the risk for cancer. *JAMA Oncol* 2016;2:762–9.
50. Oh SW, Myung SK, Park JY, Lee CM, Kwon HT. Aspirin use and risk for lung cancer: a meta-analysis. *Ann Oncol* 2011;22:2456–65.
51. Bordon Y. Tumour immunology: platelets - a new target in cancer immunotherapy? *Nat Rev Immunol* 2017;17:348.
52. Rachidi S, Metelli A, Riesenber B, Wu BX, Nelson MH, Wallace C, et al. Platelets subvert T cell immunity against cancer via GARP-TGF β axis. *Sci Immunol* 2017;2:eaai7911.
53. Burgess S, Thompson SG. Bias in causal estimates from Mendelian randomization studies with weak instruments. *Stat Med* 2011;30:1312–23.
54. Bowden J, Burgess S, Smith GD. Difficulties in testing the instrument strength independent of direct effect assumption in mendelian randomization. *JAMA Cardiol* 2017;2:929–30.