Cumulative information available about the organization of amplified chromosomal regions in human tumors suggests that the amplification repeat units, or amplicons, can be of a simple or complex nature. For the former, amplified regions generally retain their native chromosomal configuration and involve a single amplification target sequence. For complex amplicons, amplified DNAs usually undergo substantial reorganization relative to the normal chromosomal regions from which they evolve, and the regions subject to amplification may contain multiple target sequences. Previous efforts to characterize the 7p11.2 epidermal growth factor receptor (EGFR) amplicon in glioblastoma have relied primarily on the use of markers positioned by linkage analysis and/or radiation hybrid mapping, both of which are known to have the potential for being inaccurate when attempting to order loci over relatively short (<1 Mb) chromosomal regions. Due to the limited resolution of genetic maps that have been established through the use of these approaches, we have constructed a 2-Mb bacterial and P1-derived artificial chromosome (BAC-PAC) contig for the EGFR region and have applied markers positioned on its associated physical map to the analysis of 7p11.2 amplifications in a series of glioblastomas. Our data indicate that EGFR is the sole amplification target within the mapped region, although there are several additional 7p11.2 genes that can be coamplified and overexpressed with EGFR. Furthermore, these results are consistent with EGFR amplicons retaining the same organization as the native chromosome 7p11.2 region from which they are derived.

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3 Abbreviations used are as follows: BAC, bacterial artificial chromosome; EGFR, epidermal growth factor receptor; EST, expressed sequence tag; ORF, open reading frame; P1, P1-derived artificial chromosome; PCR, polymerase chain reaction; RT, reverse transcriptase; STS, sequence tagged site.

Gene amplification is common in most types of human solid tumors (Knuutila et al., 1998). Amplified DNA is manifested cytogenetically as homogeneously staining regions or double-minutes (Cowell, 1982), each of which contains multiple copies of an amplification repeat unit or amplicon (Amler and Schwab, 1989; Fakharzadeh et al., 1993). The size of the repeat unit for a specific amplicon varies between tumors, but nearly all studies attempting to address this issue report lengths within a range of a few hundred to a few thousand kilobases (Akiyama et al., 1994; Elkahloun et al., 1996; Hiemstra et al., 1994; Mostert et al., 1998; Matsumura et al., 2000; Reiter and Brodeur, 1996).

Amplicon lengths are sufficient to encompass more than 1 expressed sequence, and the common, well-characterized human chromosomal amplification regions have been shown to encompass multiple, distinct genes. However, it is generally considered that most distinct expressed sequences contained in a specific amplicon are not targeted for amplification, but rather are amplified because of their genomic proximity to a single gene...
whose amplification-associated overexpression confers tumor growth advantage (Fischer et al., 1996; Reiter and Brodeur 1998). Results associated with recent investigations of 20q13 (Albertson et al., 2000; Collins et al., 1998) and 17q23 (Monni et al., 2001; Orselli et al., 1999; Wu et al., 2001) amplifications in breast cancer may challenge the validity of this generalization.

Detailed physical and/or transcript maps have been developed for many amplification regions, including those containing MYCN (2p24: Amler et al., 1996; Reiter and Brodeur, 1996), CCND1 (11q13: Cooper et al., 1998; Courseaux et al., 1997), and CDK4 and MDM2 (12q13-15: Elkahloun et al., 1996). For the latter of these regions, amplicons have been shown to contain as many as 12 distinct genes (Fischer et al., 1996). There are, however, amplification regions that have yet to be described at a level of detail comparable with those indicated above.

Such is the case for the 7p11.2 localized EGFR region, which is amplified in approximately 40% of glioblastomas (Ekstrand et al., 1991; Libermann et al., 1985; Wong et al., 1987). Amplified EGFRs have been shown to undergo 5’- as well as 3’-end rearrangement in glioblastoma (Wong et al., 1992; Ekstrand et al., 1992), and although the genomic consequences of these rearrangements have been mapped in detail (Eley et al., 1998; Frederick et al., 2000), little is known about the consequence of EGFR amplifications on flanking genomic sequences.

With these issues in mind, the primary goal of the present study was to accurately determine the organization and expressed sequence content for 7p11.2 sequences in instances of 7p11.2 amplification in glioblastoma. Previous attempts toward achieving a similar objective have interpreted 7p11.2 amplifications using maps whose expressed (EST) and nonexpressed (STS) sequence markers were positioned by linkage, radiation hybrid, or YAC analysis (Liu et al., 1998, 2000; Wang et al., 1998a). For the present investigation, we developed a high-resolution BAC-PAC physical map for the 7p11.2 region subject to amplification, and applied information from the map to an analysis of a large series of glioblastoma DNAs.

Materials and Methods

Contig Development

7p11.2 contig construction was initiated by using PCR to screen the RPCI-11 BAC library (Research Genetics, Huntsville, Ala.), as well as RPCI-4/5 PAGs localized to 7p11.2 by the Genome Sequencing Center at Washington University (St. Louis, MO), with primers known to amplify EGFR genomic DNA from total human DNA (Eley et al., 1998; Frederick et al., 2000). The EGFR content of clones identified in this manner was verified by Southern analysis and/or sequencing, and their 7p11.2 localization was demonstrated by fluorescence in situ hybridization. Additional 7p11.2 genomic clones were identified by PCR with primers for EST and STS markers previously shown to be coamplified with EGFR (Wang et al., 1998a). The clones obtained through this process were located within the Washington University human BAC library fingerprint database, and predicted overlapping clones were identified by using the FingerPrinted Contig program (Sanger Center, Cambridge, U.K.).

Fingerprint and Restriction Analysis of Genomic Clones

To confirm the FingerPrinted Contig-predicted overlap of BACs and PACs, we analyzed genomic clones by HindIII and NotI restriction mapping. HindIII digests for fingerprint analysis were electrophoresed as described by Marra et al. (1997). For the NotI restriction digests, bacteria were grown in 1.5 ml Luria-Bertani (LB) broth overnight, and their recombinant DNAs were harvested by alkaline lysis. Harvested DNAs were resuspended in 20 ml water, and 10-µl samples were digested with NotI for 2 h at 37°C. Entire digest was run on a 1.0% pulsed field gel for 17 h at 14°C with a 120° field angle, linear ramping factor, and a switch time from 5 to 15 s.

Database-Assisted Identification of 7p11.2-Expressed Sequences

The Human GeneMap ’99 maintained by the National Center for Biotechnology Information (http://www.ncbi.nlm.nih.gov/genemap99/) was accessed to identify ESTs that had been assigned between the DNA markers D7S2422 and D7S494, which are indicated as flanking EGFR. Each of the 32 ESTs identified by this approach were analyzed by PCR and/or Southern analysis for possible inclusion within the BAC/PAC contig.

cDNA Selection for Identification of 7p11.2-Expressed Sequences

Equal quantities of mRNAs from fetal brain and placenta (Clontech, Palo Alto, Calif.) were mixed and converted to cDNA (cDNA Synthesis System; Roche Molecular Biochemicals, Indianapolis, Ind.). Linkers were incubated (2 h at 4°C) with 3 mg cDNA in a 10-µl reaction containing 2 mg phosphorylated OLIGO1 and OLIGO2 linkers (Integrated DNA Technologies, Coralville, Iowa), 3 units New England Biolabs T4 DNA ligase, and 3 ml of 10X New England Biolabs ligase buffer. The cDNA with ligated linkers was then denatured with 2 mg of C5-t DNA as well as 500 ng of ribosomal RNA and was allowed to partially renature for 4 h at 65°C with gentle shaking in an Eppendorf Thermomixer 5437 (Brinkmann, Westbury, NY).

One hundred nanograms each of BAC and PAC forming an approximate 1.3-Mb core region of the larger contig (Fig. 1B) was coamplified with Gibco BRL Bionick Kit (Life Technologies, Inc., Grand Island, N.Y.). The coamplified DNAs were precipitated with ethanol, resuspended in a volume of 5 ml, and denatured for 5 min at 95°C. The denatured, coamplified BAC DNAs were immediately added to 5 ml of 2X hybridization buffer (1.5 M NaCl, 40 mM sodium phosphate [pH 7.2], 10 mM EDTA [pH 8.0], 10X Denhardt solution, 1.2% sodium dodecyl sulfate) and 20 ml of partially renatured cDNA, and the resulting mixture was incubated for 34 h at 65°C with shaking in the thermomixer. The coamplified BAC clones with bound cDNAs were mixed with...
Fig. 1. EGFR-anchored 7p11.2 contig and 7p11.2 amplification patterns in glioblastoma. A. Genomic map showing locations of markers used in developing the contig and/or used in the Southern (*) analysis of tumor DNAs. Functional genes are indicated in bold italics, and the portion of the contig spanned by each gene is indicated by a solid line (arrowheads at the ends of solid lines indicate the direction of the associated ORF for 3 of the genes). Isolated arrowheads indicate locations of identified pseudogenes (RF directionality indicated by arrowhead orientation). A length/distance ruler is directly above the genomic map of the contig region (outer boundaries indicated by diagonal lines), and the locations of associated NotI restriction sites are indicated directly below. B. BACs and PACs constituting the minimal tile for the contig. Closed circles after library identification numbers indicate clones that have been sequenced. Numbers below each clone indicate extent of overlap (kb) with neighboring clones as determined by fingerprint analysis. Bracketed portion of the contig shows the clones used for cDNA hybrid selection. C. 7p11.2 sequence content determined for 40 tumors and 3 cell lines with 7p11.2 amplification; arrowheads terminate between adjacent markers showing amplicon boundaries. Diagonal lines represent outer boundaries for the region that was examined by Southern analysis. Number of samples showing each type of amplicon pattern is indicated at the left.

1 mg of streptavidin-coated iron beads (Dynabead M280, Dynal, Lake Success, N.Y.) and incubated for 15 min at room temperature. A magnetic particle concentrator was used to separate the beads from the solution, and these were subsequently washed 2 times for 15 min at room temperature in 1× saline–sodium citrate/0.1% sodium dodecyl sulfate, followed by 3 washes for 15 min at 65°C in 0.1× saline–sodium citrate/0.1% sodium dodecyl sulfate. After the final wash, bound cDNAs were eluted by rinsing with 50 ml of 150 mM NaOH for 10 min. The cDNAs were then purified (PCR Purification Kit, Qiagen, Valencia, Calif.) and amplified by PCR using...
the Expand HI-FI PCR amplification kit (Boehringer-Mannheim, Berkeley, Calif.) with OLIGO1 and 2 primers. The PCR products were again purified and subjected to a second round of hybrid selection to increase the fractional yield of cDNAs associated with 7p11.2 genes. A library of enriched, selected cDNAs was then generated by cloning the final PCR products into the EcoRI site of the pZERO-2 vector (Invitrogen, Calif.). The entire cDNA selection procedure was performed 4 times, and from each cDNA library produced, there were 200 clones selected for fluorescence sequencing of inserts.

Tumor Specimens, Cell Lines, and Nucleic Acid Extraction

All tumors in this study were obtained from patients undergoing surgical treatment at the Mayo Clinic. Three cell lines with EGFR amplification were also examined: MDA-468 and A431 cells were purchased from American Type Culture Collection (Rockville, MD), and SKMG-3 cells were obtained from Dr. Christopher Thomas, Mayo Clinic, Jacksonville, FL (Thomas et al., 2001). DNAs were isolated from snap-frozen tumor tissues or cell culture pellets and purified as described previously (James et al., 1988). RNAs were extracted from ~2 mm–square sections of frozen tumor tissue with Trizol (Life Technologies, Inc.).

Southern and Northern Analysis

Southern analysis of tumor DNAs and large insert genomic clones was performed as previously described (Eley et al., 1998). Briefly, HindIII- or NotI-treated DNAs were electrophoresed through agarose gels, transferred to nylon-reinforced nitrocellulose membranes (Nytran, Schleicher and Schuell, Keene, N.H.), and fixed to the membrane by baking in a vacuum oven at 80°C for 2 h. Membranes were hybridized overnight with probes labeled by random priming and were washed the next day before being exposed to X-ray film. Ambion Human Multiple Tissue Northern Blot II filters (Ambion, Austin, Tex.) were used to assess normal tissue expression of candidate expressed sequences. Conditions used for filter hybridization and rinsing have been previously described (Reiter et al., 2001).

RT-PCR Analysis of Gene Expression

Expression of LANCL2 and GASP in glioblastomas was assessed using multiplexed, competitive RT-PCR in which HPRT (hypoxanthine phosphoribosyltransferase) served as the reference transcript. Reaction mixtures consisted of 0.1 mg tumor RNA, 3 units RT, and 0.15 units AmpliTaq Gold (Perkin-Elmer, Foster City, Calif.) and were performed in solutions containing 200 mM of each dNTP, 1.5 mM MgCl2, 50 mM KCl, and 10 mM Tris-HCl (pH 8.3) by using a reaction profile of 94°C for 20 s, 55°C for 30 s, and 72°C for 60 s. Reactions were for 30 cycles. Primer sets used for RT-PCR were as follows: forward primer 5'-TGACCAAACCTACCTGCTCC-3' with reverse primer 5'-TACAGATAACCTGCCCCTTGCCC-3' for LANCL2 (cDNA product = 252 bp), forward primer 5'-TGGTGATGGAAGGAGTGGAGGC-3' with reverse primer 5'-TTTATGTCCCCCTGCTTGACTGC-3' for HPRT (cDNA product = 190 bp). Upstream and downstream primers for each target gene were known to be separated by noncoding intron sequence, as indicated by the sequences of BAC clones RP11-775L16 and RP11-792I05. Multiplexed PCR products were electrophoresed on a 1.5% agarose gel, and images of ethidium bromide-stained gels were evaluated for target/reference cDNA signal intensity ratios, as determined using Molecular Analyst software (Bio-Rad, Hercules, Calif.). The intensity ratio values were sorted into 2 groups (tumors with or without amplification of the corresponding gene) and analyzed using the Mann-Whitney U test.

Generation of Full-Length cDNAs for Coamplified Genes

Expressed sequences associated with Unigenes Hs.134342 and Hs.4750 that showed amplification, amplification-associated overexpression, and normal tissue expression were expanded to full-length cDNAs by electronic walking based on sequences in dbEST and by 5’ and 3’ RACE (SMART RACE cDNA Amplification Kit, Clontech, Palo Alto, Calif.) as previously described (Reiter et al., 2001; Wang et al., 1998b). Resulting full-length sequences for LANCL2 and GASP have been deposited in Genbank (Accession Nos. AF353942 and AF395824, respectively).

Results

The development of a contig for analyzing 7p11.2 amplicons was initiated by using PCR to screen an arrayed BAC library and selected PAC clones with primers specific for the synthesis of EGFR genomic fragments. Positive clones were further examined by PCR for the extent of their EGFR coding sequence content, and this analysis revealed that the entire EGFR coding sequence was contained within the combined genomic sequences of PAC RP5-1091E12 and BAC RP11-339F13. The genomic sequence from these clones showed that the EGFR ORF spans 193 kb (Accession Nos. AC006977 and AF288738) and lies in a telomere-to-centromere orientation (Reiter et al., 2001).

To build the contig outward from the EGFR-containing clones, primers for STS and EST markers previously determined to be coamplified with EGFR (Wang et al., 1998a), were used to screen the same BAC library. This approach identified several BACs, although none of these overlapped with the 2 EGFR clones. To fill in the gaps between EGFR-containing and EGFR-flanking clones, additional, overlapping BACs were identified by FingerPrinted Contig analysis of the Washington University fingerprint database and by again screening the BAC library using primers, the sequences of which were derived from end-sequencing of the 7p11.2 clones that had already been identified. The repeated application of these approaches led to the assembly of an ~2-Mb contig,
Table 1. Locations and amplification frequencies of 7p11.2 markers

<table>
<thead>
<tr>
<th>Locus</th>
<th>Distance from EGFR</th>
<th>Amplification frequency (%)</th>
<th>Contig clone(s)</th>
</tr>
</thead>
<tbody>
<tr>
<td>GRB10</td>
<td>&gt; 1 Mb</td>
<td>7.5</td>
<td>Outside of contig</td>
</tr>
<tr>
<td>Hs.138283&lt;sup&gt;b&lt;/sup&gt;</td>
<td>370 kb</td>
<td>40</td>
<td>CTD-2016H9</td>
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<tr>
<td>Hs.34772&lt;sup&gt;b&lt;/sup&gt;</td>
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<td>40</td>
<td>CTD-2016H9</td>
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<tr>
<td>Hs.21659&lt;sup&gt;b&lt;/sup&gt;</td>
<td>350 kb</td>
<td>40</td>
<td>GS-018A18</td>
</tr>
<tr>
<td>D7S2542&lt;sup&gt;c&lt;/sup&gt;</td>
<td>235 kb</td>
<td>57.5</td>
<td>GS-018A18</td>
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<tr>
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<td>175 kb</td>
<td>70</td>
<td>RP11-164O17</td>
</tr>
<tr>
<td>sWSS2822&lt;sup&gt;c&lt;/sup&gt;</td>
<td>130 kb</td>
<td>82.5</td>
<td>RP11-745C15</td>
</tr>
<tr>
<td>Pseudo-ARHGAP5</td>
<td>70 kb</td>
<td>100</td>
<td>RP4-791C19</td>
</tr>
<tr>
<td>EGFR intron sequences</td>
<td></td>
<td>100</td>
<td>RP5-1091E12</td>
</tr>
<tr>
<td>Pseudo-CALM1</td>
<td>50 kb</td>
<td>95</td>
<td>RP11-339F13</td>
</tr>
<tr>
<td>WI-16990&lt;sup&gt;b&lt;/sup&gt;</td>
<td>170 kb</td>
<td>70</td>
<td>RP11-775L16</td>
</tr>
<tr>
<td>LANCL2</td>
<td>260 kb</td>
<td>50</td>
<td>RP11-775L16</td>
</tr>
<tr>
<td>GASP</td>
<td>340 kb</td>
<td>35</td>
<td>RP11-792I05</td>
</tr>
<tr>
<td>SHGC-30962&lt;sup&gt;b&lt;/sup&gt;</td>
<td>420 kb</td>
<td>25</td>
<td>RP11-792I05</td>
</tr>
<tr>
<td>GBAS</td>
<td>700 kb</td>
<td>10</td>
<td>RP11-015K19</td>
</tr>
<tr>
<td>L-3-PS3</td>
<td>700 kb</td>
<td>10</td>
<td>RP11-015K19</td>
</tr>
<tr>
<td>FKBP9</td>
<td>&gt; 1 Mb</td>
<td>5</td>
<td>Outside of contig</td>
</tr>
<tr>
<td>HNRNP5</td>
<td>&gt; 1 Mb</td>
<td>5</td>
<td>Outside of contig</td>
</tr>
</tbody>
</table>

<sup>a</sup>Based on results from 40 glioblastomas with EGFR amplification. None of these markers was amplified in any of 78 tumors without amplified EGFR.

<sup>b</sup>Denotes unnamed Unigene or EST.

<sup>c</sup>Denotes STS probe for which there is no known expressed sequence association.
marker amplification is inversely proportional to its distance from the amplification target sequence, \( \text{EGFR} \). In all instances, the results for individual tumors suggested that the 7p11.2 DNA contained within an amplicon is contiguous, or uninterrupted relative to its normal chromosomal configuration. The only indication of reorganization of 7p11.2 sequences was made evident in tumor DNAs with intragenic rearrangements of \( \text{EGFR} \) that have been previously described (Eley et al., 1998; Frederick et al., 2000).

Probes representing each of the 14 putative expressed sequences outside of the \( \text{EGFR} \) coding region, as well as additional probes for trapped \( \text{EGFR} \) intron sequences, were used for the Northern analysis of normal tissue RNAs and for cell line RNAs established from tumors with \( \text{EGFR} \) amplification. Each of the \( \text{EGFR} \) intron probes as well as 5 of the 12 probes for sequences outside of \( \text{EGFR} \) (Hs.138283, Hs.34772, Hs.21659, WI-16990, and SHGC-30962) failed to show expression by this analysis: Similar results have been noted by others who have examined catalogued EST sequences for evidence of expression (Ewing and Green, 2000; Hillier et al., 1996). In addition, Northern results associated with the 3 cell lines having 7p11.2 amplification, combined with sequence data for genomic clones near the \( \text{EGFR} \) core region (Fig. 1), indicated that the \( \text{ARHGAP5} \) and \( \text{CALM1} \) cDNAs were trapped by pseudogenes flanking \( \text{EGFR} \) telomerically and centromerically, respectively (functional \( \text{ARHGAP5} \) and \( \text{CALM1} \) genes reside on 14q). An additional pseudogene for \( \text{RPL31} \) (ribosomal protein L31) was localized to the contig region through sequence analysis of BAC GS018A18.

Seven of the expressed sequence probes were amplified in tumor DNAs, expressed in normal tissue RNAs, and overexpressed in cell line RNAs in instances where their corresponding gene was amplified. Five of the 7 probes were associated with known genes: \( \text{GRB10} \), \( \text{GBAS} \), \( \text{L-3-PSP} \), \( \text{FKBP9} \), and \( \text{HNRNPU} \). For the 2 probes for which corresponding sequences were associated with unnamed Unigene entries, Hs.134342 and Hs.4750, the Northern results showed distinct transcript sizes for each (1.8 and 3.8 kb for Hs.134342 and Hs.4750, the Northern results showed distinct transcript sizes for each (1.8 and 3.8 kb for Hs.134342, and 3.6 and 4.2 kb for Hs.4750: Fig. 2), suggesting that each was associated with a unique gene. Furthermore, the probes for these Unigenes showed distinct normal tissue expression patterns relative to each other, as well as to \( \text{EGFR} \): Hs.134342 was most abundant in testes, Hs.4750 was most abundant in thymus and ovary, and \( \text{EGFR} \) was most abundant in placenta (Fig. 2).

Electronic walking, performed by aligning ESTs associated with each Unigene, was used to generate 1.5- and 1.2-kb cDNAs for the Hs.134342 and Hs.4750 transcripts, respectively. SMART RACE was then employed to extend the 1.5-kb sequence for Hs.134342 such that a longest ORF of 1353 nucleotides was identified and to extend the 1.2-kb transcript of Hs.4750 in order to reveal its longest ORF of 507 nucleotides. The gene with which Hs.134342 is associated has been named Lantibiotic Synthetase C-like-2 (\( \text{LANCL2} \), Accession No. AF353942) because of its homology with the bacterial Lantibiotic Synthetase C family of genes, and it represents the second such gene cloned from humans (Bauer et al., 2000). Extensive homology and pattern searches of the 507-bp ORF generated for Hs.4750 have yet to reveal any significant similarities with known genes. However, the cDNA encodes a protein that is generally hydrophilic and contains a leader peptide, suggesting that it is secreted. Due to the frequency of its amplification in glioblastoma, as well as the putative leader peptide and hydrophilic nature of its encoded protein, we have named the gene for Hs.4750 \( \text{GASP} \) (Glioblastoma Amplified Secreted Protein, Accession No. AF395824).

To determine whether \( \text{LANCL2} \) and \( \text{GASP} \) show transcript overexpression in association with gene amplification, competitive RT-PCR analyses were performed using a subset of glioblastoma RNAs for which the amplification status of each gene was known in the corresponding tumor (Fig. 3). The results from these analyses showed a significant association between amplification and increased transcript level for both \( \text{LANCL2} \) (Fig. 3) and \( \text{GASP} \): \( P = 0.024 \) and 0.016, respectively. Gene dosage is, therefore, a major determinant of \( \text{LANCL2} \) and \( \text{GASP} \) expression in glioblastoma, although the variable levels of \( \text{LANCL2} \) (Fig. 3) and \( \text{GASP} \) transcript that were indicated among tumors without amplification suggest the existence of additional factors that influence the expression of these genes.

![Fig. 2. Normal tissue expression patterns for \( \text{EGFR} \) and 2 novel genes identified in association with this study, as indicated by Northern analysis. Probes used for \( \text{LANCL2} \) and \( \text{GASP} \) were full-length cDNAs. The sizes of transcripts (kb) revealed by each probe are indicated to the left of their corresponding image.](https://academic.oup.com/neuro-oncology/article-abstract/4/2/86/1298291)
Fig. 3. Comparison of competitive RT-PCR results for LANCL2 expression in glioblastomas with (T1-T6) or without (T7-T9) LANCL2 amplification. RT-PCR reaction products were resolved in a 1.5% agarose gel, and a negative image of the ethidium bromide–stained cDNAs (upper band, LANCL2; lower band, HPRT) were examined for intensity using Molecular Analyst software (Bio-Rad). The results for the normalized tumor LANCL2:HPRT intensity ratios are indicated by the heights of the histogram bars, the unit definition for which is based on the signal ratio determined for normal brain (NB). Histogram bars are shown directly above the corresponding reaction products from which their values (heights) were determined. The association between elevated LANCL2 expression and LANCL2 amplification was determined to be significant ($P = 0.024$).

**Discussion**

The primary objective of this study was to develop a high resolution physical and transcript map of the genomic region surrounding EGFR to more precisely characterize the content of amplicons associated with chromosomal region 7p11.2. Our primary interest in developing such a map is related to the results of recent investigations that suggest the existence of multiple independent amplification targets in at least some chromosomal regions (Albertson et al., 2000; Orsetti et al., 1999). In the event that EGFR proved to be the sole amplification target in the 7p11.2 region, the application of a physical-transcript map to the analysis of EGFR amplicons was still expected to be informative about the identities of bystander genes that can be coamplified with EGFR and the frequencies of their coamplification.

The contig that was developed to address these issues spans approximately 2 Mb of genomic sequence. Application of the physical map derived from the contig, in combination with the Southern analysis of tumor DNAs, sequence analysis of contig clones, and the Northern and RT-PCR analysis of normal as well as tumor RNAs, indicates there are 5 functional genes and 3 pseudogenes within the contig region (Fig. 1); 3 additional genes, GRB10, HNRNPU, and FKBP9, lie outside of the contig region, but are nonetheless occasionally coamplified with EGFR.

L-3-PSP and GBAS, as well as the 2 novel expressed sequences identified in conjunction with this project, LANCL2 and GASP, are all proximal (centromeric) to EGFR. Of these, LANCL2 and GASP are nearest to EGFR and are within 250 kb from its centromeric, 3’ end, whereas GBAS and L-3-PSP are approximately 700 kb centromeric of EGFR. Due to their genomic proximity, LANCL2 and GASP, as well as GBAS and L-3-PSP, are contained on single BAC clones: RP11-775L16 and RP11-792I05 for the former pair, and RP11-015K19 for the latter.

Unexpectedly, our results suggest that there are no functional genes in a 900-kb region that is telomeric of EGFR. This finding is especially surprising given the localization of 3 independent Unigenes, and a NoI restriction site, within a region 450 to 480 kb from the 5’ end of EGFR (Fig. 1A). However, neither our multiple tissue Northern analysis, performed with portions of the Unigene sequences in question, nor the cDNA hybrid selection procedure yielded results that support the assignment of functional genes to this region.

The results of Southern analysis of 7p11.2 sequences in 118 tumors and 3 cell lines support the conclusion that EGFR is the only amplification target of the 7p11.2 region. The lack of EGFR-independent amplification for any of the contig-localized probes, as well as for probes associated with the 3 genes outside of the contig region, is consistent with such a conclusion. We had previously established this point based on the interpretation of EGFR-7p11.2 amplifications with a map, the markers of which were positioned by linkage and radiation hybrid analysis (Wang et al., 1998a). Although the change from radiation-hybrid to physical map has not affected our conclusion regarding the singular nature of the 7p11.2 amplification target, our current interpretation of EGFR-7p11.2 amplicon structure, using a 2-Mb physical map with correctly ordered markers, shows that 7p11.2 amplicons are continuous and not structurally complex as we and others have previously suggested (Liu et al., 1998, 2000; Wang et al., 1998a). The simple structure of 7p11.2 amplicons appears analogous to that for the MYCN-2p24 amplification region in neuroblastoma (Noguchi et al., 1996; Reiter and Brodeur, 1998) and contrasts with structurally complex amplicons such as those arising from 2q13 (Albertson et al., 2000; Collins et al., 1998) and 17q23 (Monni et al., 2001; Orsetti et al., 1997; Wu et al., 2001) in breast cancer.

As shown for MYCN amplicons (Manohar et al., 1995; Noguchi et al., 1996), EGFR amplicons frequently contain bystander genes, and the 7p11.2 bystanders can be coamplified and overexpressed with EGFR. Approximately one half of the tumors and cell lines that were examined for 7p11.2 amplicon content showed amplification of multiple 7p11.2 genes. However, only 2 of these genes were coamplified at significant frequencies: LANCL2 (50%) and GASP (35%). LANCL2 amplification was observed in 17% of all glioblastomas examined here, making it the second most commonly amplified gene in these tumors. Amplification of EGFR invariably results in increased expression of its corresponding mRNA and protein (Wong et al., 1987), and the results presented here sug-
gest this to be the case for GASP and LANCL2 as well. In
instances where GASP and/or LANCL2 were coamplified
with EGFR, competitive RT-PCR analysis showed
overexpression of their corresponding transcripts.
Whether there are biologic consequences associated
with the coamplification and overexpression of these
bystander genes is an issue that has yet to be addressed
and is one of considerable interest because of the preva-
ience of bystander gene amplification in cancer. Based on
the data shown here, however, it seems likely that
LANCL2 and GASP amplification minimally result in
the properties of these genes could provide useful
information for the development of innovative therapies.

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