Minimally Invasive Approaches to Optimize Block Grafting: A Case Report

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INTRODUCTION

A narrow alveolar ridge is a challenging situation that surgeons may face when considering implant oral rehabilitation. Many surgical approaches exist, and the most frequently used is bone grafting.1

The subperiosteal tunneling approach is a minimally invasive procedure that allows the surgeon to allocate the graft in a space that is obtained between the soft tissues and the underlying bone through an access represented by a single incision on the mesial limit of the bone defect.2,3 This approach ensures minimal discomfort for the patient after the surgery and mostly steady coverage of the graft during the healing time, with minimal risk of exposure, infection, and failure.4,5 The subperiosteal tunneling technique can be performed with autologous bone, which is still considered the gold standard for bone regeneration.6

On the other hand, autologous bone grafting requires donor-site surgery, which often means donor-site surgery-related complications, especially when larger amounts of bone are needed and extraoral sites are accessed for the harvest. Moreover, this technique needs general anesthesia.7,8 Homologous grafting (or allograft) refers to the use of bone from living human donors or from cadavers. The bone is processed and stored in tissue banks. Depending on the type of treatment, fresh frozen bone (FFB), freeze-dried bone, and demineralized freeze-dried bone can be obtained.4,9,10

The case presented herein combines the tunnel flap technique and the use of homologous bone to achieve the least invasive approach to bilateral block grafting of an upper maxillary affected by severe horizontal bone atrophy in order to allow a full-arch implant rehabilitation.

CASE PRESENTATION

A 55-year-old white woman was referred to the Dental School of the University Federico II of Naples. No significant medical data (diabetes, hypertension, allergies) were present in her clinical history; laboratory results were substantially normal.

The patient was partially edentulous and showed severe horizontal bone atrophy in the maxillary arch. The only teeth left in the arch were the canines and the second molars, which had already undergone several treatments.

Because of the periodontal and endodontic condition of the remaining teeth, implant therapy and a full arch prosthetic restoration were chosen for the patient. Horizontal bone augmentation was necessary, as the ridge showed no sufficient horizontal dimensions for the placement of implants in the right and left posterior sectors, and block grafting was a mandatory option because of the degree of atrophy and the noncontensive anatomy of the defect. There was no need for an additional vertical augmentation (Figure 1).

A minimally invasive approach was chosen for the exposure of the receiving sites. Instead of creating a traditional flap, a mucoperiosteal tunnel was created with a periosteal elevator inserted through a single incision line of moderate dimensions per hemiarch on the mesial edge of each of the defects (right and left) from the occlusal to the apical limit of the ridge on its vestibular side (Figure 2b).

Tissue bank ilium bicortical bone (FFB) was chosen as grafting material. After being defrosted in sterile saline solution, each block was split, obtaining cortical bone on one side and trabecular bone on the other. The cortical bone was to be positioned on the interface with the soft tissue, and the trabecular bone was to be placed on the interface with the host site (Figure 2c). The host site had already been drilled to facilitate bleeding, start the regional acceleratory process, and expose trabecular bone.

Three microscrews fixed on the mesial side of the graft allowed the complete fixation of the block, avoiding any movement or tilting (Figure 2d). Platelet-rich fibrin (PRF) was then placed on the graft.

Besides the block grafting on the right and left posterior regions, a flap was elevated in the frontal region, which showed good bone quality, quantity, and anatomy; two $4 \times 11.5$ mm submerged tapered implants were placed (Figure 2e). Polygalactin 910 (Coated Vicryl, Ethicon, Somerville, NJ) 4/0 resorbable sutures were used for flaps closure.

The 4 remaining teeth were not extracted in this phase so they could be used to support a fixed provisional prosthesis (Figure 2f). The patient was scheduled for regular follow-ups for the next 5 months.
After 5 months of healing, the computerized tomography follow-up and surgical re-entry were performed (Figure 3). After removal of the provisional prosthesis and infiltration of local anesthesia, mucoperiosteal flaps were raised to expose both grafted areas. The grafted areas showed minimal bone resorption: the bone was still at the level of the top of the microscrews, which were removed. The bleeding during the removal of the screws demonstrated the successful integration of the graft.

After a slight osteoplasty to give right conformation to the ridge, measurements were taken showing a horizontal gain between 2 and 4 mm. Three implants per side were placed—with the same dimensions for the frontal region (4 × 11.5 mm). No immediate prosthetical load was realized, and the provisional prosthesis was recemented on the remaining teeth.

After a 4-month healing period, the implants were uncovered and healing abutments were placed. The four remaining teeth were extracted and a removable provisional prosthesis was provided for use over the 2-week soft-tissue healing period. Afterward, a full arch resin restoration was loaded on the implants for 1 month, followed by a definitive metal ceramic prosthesis. The 16-month follow-up showed no marginal bone loss and no bone resorption, as well as soft-tissue stability on the grafted areas.

**Figure 1.** Computerized tomographic image showing severe horizontal atrophy.

**Figure 2.** (a) Clinical case of patient with severe bone atrophy. (b) A minimally invasive approach with a single incision realizing instead of a mucoperiosteal tunnel. (c) Positioning of tissue bank ilium bicortical bone (fresh frozen bone). (d) Complete fixation of the block with three microscrews. (e) Positioning of 2 submerged implants in the frontal region. (f) Fixed provisional prosthesis.
Barone et al.8 established success criteria for a bone graft: the absence of exposure and infection of the graft in the postoperative period, incorporation of the graft to the receptor bed, absence of radiolucent areas, bleeding of the grafted bone when the fixation screws are removed, and achievement of adequate volume and density to place the dental implants. In the case presented herein these criteria were all satisfied.

The tunnel technique provided primary intention closure of the surgical wound, thereby avoiding dehiscences and infections, and reducing the edema and the postoperative discomfort for the patient. Dehiscences are reported to be among the most frequent complications associated with bone grafting, so preventing them can result in a higher predictability of the graft.3,11,12

Clinical and histologic studies support the use of FFB,6,10,13 and it has been suggested that the results of a graft with homologous and autologous bone are comparable, even though the healing phase is more critical and longer for the allograft.14 The FFB is believed to have both osteoconductive properties—thanks to its solid structure and medullary spaces that allow blood vessel infiltration and bone ingrowth—and osteoinductive action; freezing does not affect the bone morphogenetic proteins contained in the bone.15 In addition, polymerase chain reaction serologic tests on the donor tissue ensure the safety of the graft: viral infections transmission by unprocessed deep-frozen, nonirradiated grafts from screened donors is currently less than 1:200 000 for hepatitis C virus and 1:1 million for human immunodeficiency virus. Such a risk is virtually nonexistent for processed bone grafts.

Homologous bone from tissue banks offers obvious advantages such as: readily available source of large volumes of material, extremely low antigenic potential, unblemished safety record in dentistry, and, of course, the absence of a donor-site surgery for the patient and its risk of surgical complications.7,18

The use of PRF can also increase the predictability of bone formation. Osteoblasts show high affinity for PRF membranes,19 and the constant release of growth factors guides the healing and remodeling of connective tissues.20,21 Dohan Ehrenfest et al have demonstrated that PRF, through a constant release of growth factors, is twice as effective at enhancing soft tissue healing, stimulating blood vessels ingrowth, cellular migration, and differentiation into bone-forming cellular lines.22 Furthermore, the presence of cytokines lowers the risk of infection within the graft by controlling inflammation.23

Although there is no evidence that a delayed implant placement in the grafted areas is more predictable, the data obtained so far,24,25 and the use of allograft that proved to have a longer and more critical healing phase26 were the reasons why we chose such protocol.

For the same reason, delayed prosthetic loading was chosen, even though primary stability of the implants was good. The possibility of using a provisional teeth-supported fixed prosthesis balanced the choice of a longer therapy.
**CONCLUSIONS**

When the surgical protocol is accurate and no complications occur during the healing phase, homologous bone grafting is a valid alternative to autologous grafting. The obvious advantage of this technique is sparing the patient a donor-site surgery. The combination of homologous graft with the tunnel technique reduces surgical trauma, postoperative discomfort, and the risk of exposure and failure of the graft during the healing phase. The use of PRF to cover the graft can enhance bone formation and soft-tissue healing.

In our experience, onlay grafting with frozen bone is a predictable technique for horizontal augmentation. In the case of a vertical augmentation, however, our advice is grafting with a sandwich interpositional technique.

**ABBREVIATIONS**

FFB: fresh frozen bone  
PRF: platelet-rich fibrin

**REFERENCES**


