Leaf gas exchange performance and the lethal water potential of five European species during drought

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Establishing physiological thresholds to drought-induced mortality in a range of plant species is crucial in understanding how plants respond to severe drought. Here, five common European tree species were selected (Acer campestre L., Acer pseudoplatanus L., Carpinus betulus L., Corylus avellana L. and Fraxinus excelsior L.) to study their hydraulic thresholds to mortality. Photosynthetic parameters during desiccation and the recovery of leaf gas exchange after rewatering were measured. Stem vulnerability curves and leaf pressure–volume curves were investigated to understand the hydraulic coordination of stem and leaf tissue traits. Stem and root samples from well-watered and severely drought-stressed plants of two species were observed using transmission electron microscopy to visualize mortality of cambial cells. The lethal water potential (ψ_lethal) correlated with stem P₉⁹ (i.e., the xylem water potential at 99% loss of hydraulic conductivity, PLC). However, several plants that were stressed beyond the water potential at 100% PLC showed complete recovery during the next spring, which suggests that the ψ_lethal values were underestimated. Moreover, we observed a 1 : 1 relationship between the xylem water potential at the onset of embolism and stomatal closure, confirming hydraulic coordination between leaf and stem tissues. Finally, ultrastructural changes in the cytoplasm of cambium tissue and mortality of cambial cells are proposed to provide an alternative approach to investigate the point of no return associated with plant death.

Keywords: cambium vitality, embolism resistance, hydraulic failure, leaf turgor, photosynthesis, plant death, xylem water potential.

Introduction

Xylem sap in transpiring plants is typically under tension according to the cohesion–tension theory (Dixon and Joly 1895, Tyree 2003a). The cohesive force of the water column in xylem conduits is quantified in vulnerability curves, which show the relationship between xylem water potential (ψₓ, MPa) and the percentage loss of hydraulic conductivity due to embolism formation (PLC %; Tyree et al. 2003, Cochard et al. 2008, 2013). One could expect that with more severe and frequent drought events ψₓ may become more negative, resulting in higher amounts of embolized conduits and higher levels of PLC than under well-watered conditions. A decline in hydraulic conductivity would have potential effects on reducing photosynthesis, transpiration and stomatal conductance (Sperry 2000, Hubbard et al. 2001, Miyashita et al. 2005), therefore affecting plant growth and productivity (Tyree 2003b, Brodribb 2009). A continued decline of hydraulic conductivity could eventually lead to hydraulic failure of the water transport system, with the propagation of xylem embolism reaching a point of no return from which the plant is unable to recover (Nardini et al. 2013). During severe drought, hydraulic failure can result in complete desiccation of plant tissues and organs (Hoffmann et al. 2011), and induce partial or complete plant mortality (Anderegg 2015).

Considerable attention has been focused on the resistance of plants to xylem hydraulic failure (Hartmann et al. 2015), mostly including hydraulic parameters at the stem level, such as xylem embolism resistance (Lopez et al. 2005), xylem hydraulic safety margins (Choat et al. 2012), sapwood capacitance (Scholz et al. 2014) and sapwood water storage (Pineda-Garcia et al. 2013). Stem and root samples from well-watered and severely drought-stressed plants of two species were observed using transmission electron microscopy to visualize mortality of cambial cells. The lethal water potential (ψ_lethal) correlated with stem P₉⁹ (i.e., the xylem water potential at 99% loss of hydraulic conductivity, PLC). However, several plants that were stressed beyond the water potential at 100% PLC showed complete recovery during the next spring, which suggests that the ψ_lethal values were underestimated. Moreover, we observed a 1 : 1 relationship between the xylem water potential at the onset of embolism and stomatal closure, confirming hydraulic coordination between leaf and stem tissues. Finally, ultrastructural changes in the cytoplasm of cambium tissue and mortality of cambial cells are proposed to provide an alternative approach to investigate the point of no return associated with plant death.
However, as a direct indicator of hydraulic resistance to plant death, the lethal water potential ($\psi_m$), i.e., the water potential corresponding to the point of no recovery, has been investigated in a small number of species thus far. Lethal water potential of five angiosperm species corresponded to $P_{88}$ ($\psi_m$ at 88% stem PLC; Urdi et al. 2013), and $M_{\psi_m}$ of poplar and beech seedlings were related to $P_{50}$ ($\psi_m$ at 50% stem PLC; Barigah et al. 2013a). In contrast, $\psi_{\text{ lethal}}$ of four gymnosperm species were associated with $P_{50}$ ($\psi_m$ at 50% stem PLC; Brodribb and Cochr 2009). This divergence between angiosperm and gymnosperm tree species is associated with xylem anatomical differences, and various contrasting traits, such as phenology, growth allotropes, and competition sensitivity (Carnicer et al. 2013).

At the leaf level, stomatal closure is a crucial protective strategy against xylem hydraulic failure (Brodribb and Holbrook 2004, Choat et al. 2007, Chen et al. 2010). In general, plants can be separated into two major groups based on their stomatal control strategy during drought stress: isohydrastic plants maintain a relatively stable daytime leaf water potential ($\psi_l$, MPa) under mild and moderate drought conditions by closing their stomata gradually to reduce gas exchange and water loss (Kumagai and Porporato 2012, Sade et al. 2012). In contrast, anisohydrastic plants tolerate a decline in $\psi_l$ by keeping stomata open to enable continuous gas exchange within certain levels of water stress (Tardieu and Simonneau 1998). Nevertheless, evidence suggests that stomatal closure for both isohydrastic and anisohydrastic plants is coordinated with xylem embolism (Sperry and Pockman 1993, Salleo et al. 2000, Meinzer et al. 2009; Manzoni et al. 2014), although this coordination varies to some degree across species (Bond and Kavanagh 1999). For instance, most studies illustrate that $\psi_l$ at stomatal closure ($M_{\psi_l}$ MPa) scales with $\psi_x$ at the onset of xylem embolism ($P_{50}$ MPa; Nardini et al. 2001, Cochard et al. 2002). Some studies, however, suggest a much closer relationship between $M_{\psi_l}$ and $P_{50}$ than $P_e$ (Sparks and Black 1999, Martorell et al. 2014).

Stomatal closure is generally assumed to be the first protective mechanism against drought stress (Tyree et al. 1998). However, the structural threshold to drought-induced tree mortality is not clear. At the tissue level, meristematic tissue such as cambium determines tree growth by producing new layers of xylem and phloem (Liphschitz and Lev-Yadun 1986, Rossi et al. 2013), which allows woody plants to replace dysfunctional cells and to facilitate secondary growth. Previous work showed that the cambium is highly resistant to leaf defoliation in Abies balsamea seedlings (Rossi et al. 2009b), and is the last resilient tissue to dehydration in Populus nigra seedlings (Barigah et al. 2013b). These results suggest that cambium vitality can be a useful indicator of tree growth and also mortality (Gričar et al. 2014). Nevertheless, the majority of studies on cambium focused on seasonal changes of cambium activity (Farrar and Evert 1997, Fuchs et al. 2010), or quantification of cambium cells with different irrigation regimes (Abe et al. 2003, Rossi et al. 2009a, de Luis et al. 2011, Balducci et al. 2013). Research on the ultrastructure of cambial cells in relation to plant death remains surprisingly limited (Thomas 2013).

In this study, we aimed to estimate $\psi_{\text{ lethal}}$ of five angiosperm tree species and expected that these values would be closely related to stem $P_{88}$ values. We applied drought-stress rehydration experiments on seedlings of the species studied and quantified the dynamics of photosynthesis and transpiration during the drought and rewatering period to estimate $\psi_{\text{ lethal}}$. We defined a moderate stress level as $\psi_m$ (midday leaf water potential) less negative than $P_{50}$, while severe water stress was considered to have $\psi_m$ between $P_{50}$ and $P_{88}$. Death was predicted to occur at $\psi_m$ values that were more negative than $P_{88}$. We also hypothesized that there is a coordination of $M_{\psi_l}$ and $P_e$ for the five tree species selected. We conducted stomatal response curves, leaf pressure–volume (PV) curves and stem vulnerability curves to test the hydraulic coordination between leaf and stem organs. In addition, we hypothesized that cambium would suffer cellular failure at the time of plant death in both stem and root organs.

**Materials and methods**

**Plant material**

The following five European angiosperm species were selected: *Acer campestre* L., *Acer pseudoplatanus* L., *Carpinus betulus* L., *Corylus avellana* L. and *Fraxinus excelsior* L. Seedlings of these species were used for a drought experiment. These seedlings were 2–4 years old and 30–90 cm tall. In November 2013, samples belonging to the same population of plants were bought from a local nursery (Kordes Jungpflanzen Handels GmbH, Bilsen, Germany), and transported to the Botanical Garden of Ulm University (48°25’9.84” N, 9°57’59.759” E). The seedlings were transferred to 3-l pots, with a composition of 50% organic soil, 20% sand, 20% loam and 10% turf. In spring and early summer 2014, all seedlings were well-watered on a daily basis and pesticides were applied twice to avoid insect herbivory.

Branches from adult trees grown in the Botanical Garden of Ulm University were used for stem vulnerability curves and leaf PV curves. All branches were ~2–5 years old and from the same five species selected for the drought experiment. The adult trees at Ulm University were from the same population as the plants from the local nursery. Samples were taken from sun-exposed branches at a height of 2–3 m during early morning in the growing season of 2013 and 2014.

**Experimental design**

In June 2014, 11–14 healthy seedlings of each species were placed under a rainout shelter (8 × 4 m), which was constructed with a special UV transparent foil (Lumisol clear AF; Folitec, Westerburg, Germany), having a light transmittance of 90%. The ground was covered with a special UV translucent foil (Lumisol clear AF; Folitec, Westerburg, Germany), having a light transmittance of 70%. The seedlings were related to a UV-B transmittance of 70%. The ground was covered with a special UV translucent foil (Lumisol clear AF; Folitec, Westerburg, Germany), having a light transmittance of 70%.
water or rain. In addition, this foil reduced the growth of weeds. The soil was also given a slight inclination to facilitate water run-off. The temperature and humidity was measured with a data logger (EL-USB-2, Lascar electronics Inc., Salisbury, UK). The average temperature and relative humidity were 17.9 ± 6.5 °C and 75 ± 20.6%, respectively, over the entire duration of the experiment, i.e., from 10 July 2014 to 26 September 2014.

Before starting the drought experiment, plants were well-watered on a daily basis. Overall, the approach applied was based on Brodribb and Cochard (2009) and Urli et al. (2013). For a list of the physiological parameters described below, acronyms and their definitions, see Table 1. A Li-Cor 6400 XT portable photosynthesis system (LI-COR, Lincoln, NE, USA) was used to measure the maximum CO₂ assimilation rate (A max, mmol CO₂ m⁻² s⁻¹), transpiration rate (E max, mmol H₂O m⁻² s⁻¹) and stomatal conductance (g s max, mol H₂O m⁻² s⁻¹). The LED red and blue light source was set to a fixed intensity of 1200 μmol m⁻² s⁻¹. Other parameters included a 400 μmol mol⁻¹ reference CO₂ concentration, 20 °C block temperature and 500 μmol s⁻¹ pump flow rate. The A max, E max and g s max were measured on three mature 3- to 5-month-old leaves for each plant. These leaves were labelled to repeat similar measurements over time. Photos were taken from leaves smaller than the Li-Cor leaf chamber to estimate the leaf area measured. Five individuals from each species were randomly selected to measure ψ s, with three leaves being measured on a single plant. Leaves were bagged with aluminium bags for at least 1 h before midday to reach equilibrium between ψ s with ψ i, and ψ s was measured using a Scholander pressure chamber (PMS Instrument Company, Albany, OR, USA).

Drought was induced by withholding water from the plants over a variable number of days, ranging from 7 to 56 days. The plants were water-stressed to different levels based on the ψ s measurements obtained. In general, we aimed for moderate and severe levels of water stress, as well as lethal doses of drought. Midday leaf water potential was measured on one to three leaves from each plant with the Scholander pressure chamber every 1–3 days. For each plant, measurements of A, E and g s

### Table 1. Acronyms and units of the parameters measured in this study with their corresponding definitions.

<table>
<thead>
<tr>
<th>Acronyms</th>
<th>Units</th>
<th>Definitions</th>
</tr>
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<tbody>
<tr>
<td>A</td>
<td>μmol CO₂ m⁻² s⁻¹</td>
<td>CO₂ assimilation rate</td>
</tr>
<tr>
<td>A max</td>
<td>μmol CO₂ m⁻² s⁻¹</td>
<td>Maximum CO₂ assimilation rate</td>
</tr>
<tr>
<td>E</td>
<td>mmol H₂O m⁻² s⁻¹</td>
<td>Transpiration rate</td>
</tr>
<tr>
<td>E max</td>
<td>mmol H₂O m⁻² s⁻¹</td>
<td>Maximum transpiration rate</td>
</tr>
<tr>
<td>g s</td>
<td>mol H₂O m⁻² s⁻¹</td>
<td>Stomatal conductance rate</td>
</tr>
<tr>
<td>g s max</td>
<td>mol H₂O m⁻² s⁻¹</td>
<td>Maximum stomatal conductance rate</td>
</tr>
<tr>
<td>PLC</td>
<td>%</td>
<td>Percentage loss of hydraulic conductivity</td>
</tr>
<tr>
<td>PLCₐ</td>
<td>%</td>
<td>PLC corresponding to ψ lethalₐ</td>
</tr>
<tr>
<td>PLCₜ</td>
<td>%</td>
<td>PLC corresponding to ψ lethalₜ</td>
</tr>
<tr>
<td>PLCₙ</td>
<td>%</td>
<td>PLC corresponding to ψ lethalₙ</td>
</tr>
<tr>
<td>Pₐ</td>
<td>MPa</td>
<td>ψ s at the onset of xylem embolism</td>
</tr>
<tr>
<td>P₅₀</td>
<td>MPa</td>
<td>ψ s at 50% PLC</td>
</tr>
<tr>
<td>P₈₈</td>
<td>MPa</td>
<td>ψ s at 88% PLC</td>
</tr>
<tr>
<td>RWCₜ</td>
<td>%</td>
<td>Relative water content at turgor loss point</td>
</tr>
<tr>
<td>Sₐ₅₀</td>
<td></td>
<td>Slope of the tangent drawn through the midpoint of the A response curve</td>
</tr>
<tr>
<td>Sₐₜₜ</td>
<td></td>
<td>Slope of the tangent drawn through the midpoint of the E response curve</td>
</tr>
<tr>
<td>Sₙ₉₀ₐ₄</td>
<td></td>
<td>Slope of the tangent drawn through the midpoint of the g s response curve</td>
</tr>
<tr>
<td>Sₚ₉₀ₜ</td>
<td></td>
<td>Slope of the tangent drawn through the midpoint of the stem vulnerability curve</td>
</tr>
<tr>
<td>t₁/₂</td>
<td>days</td>
<td>Number of days for A, E and g s to recover to more than half of the maximum values</td>
</tr>
<tr>
<td>ψₐ</td>
<td>MPa</td>
<td>ψ s at the cessation of photosynthesis</td>
</tr>
<tr>
<td>ψₜ</td>
<td>MPa</td>
<td>ψ s at the cessation of transpiration</td>
</tr>
<tr>
<td>ψₙ</td>
<td>MPa</td>
<td>ψ s at stomatal closure</td>
</tr>
<tr>
<td>ψₙ₉₀ₐ₄</td>
<td>MPa</td>
<td>ψ s at 50% of maximum stomatal conductance</td>
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<tr>
<td>ψ</td>
<td>MPa</td>
<td>Leaf water potential</td>
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<tr>
<td>ψ lethal</td>
<td>MPa</td>
<td>Xylem lethal water potential</td>
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<tr>
<td>ψ lethalₐ</td>
<td>MPa</td>
<td>Xylem lethal water potential calculated based on A</td>
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<tr>
<td>ψ lethalₜ</td>
<td>MPa</td>
<td>Xylem lethal water potential calculated based on E</td>
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<td>ψ lethalₙ</td>
<td>MPa</td>
<td>Xylem lethal water potential calculated based on g s</td>
</tr>
<tr>
<td>ψₙ</td>
<td>MPa</td>
<td>Midday leaf water potential/midday xylem water potential</td>
</tr>
<tr>
<td>ψₙ₉₈</td>
<td>MPa</td>
<td>Xylem water potential</td>
</tr>
<tr>
<td>ψₚ</td>
<td>MPa</td>
<td>ψ s at turgor loss point</td>
</tr>
<tr>
<td>ϵ</td>
<td>MPa</td>
<td>Modulus of elasticity</td>
</tr>
<tr>
<td>ε</td>
<td>MPa</td>
<td>Osmotic potential at full turgor</td>
</tr>
</tbody>
</table>
were based on the three leaves that were labelled and measured previously. All Li-Cor measurements were conducted from 9 am to 1 pm and were repeated every 2–3 days. Given considerable fluctuation in light intensity and temperature during the 4 h of measurements, all plants were measured in a fixed order, which means that temporal differences for individual plants were kept as minimal as possible. In case leaves selected for our Li-cor measurements dropped off, additional, fully developed leaves were used to take measurements.

Once the $A$, $E$ and $g_s$ values were lower than 50% of $A_{\text{max}}$, $E_{\text{max}}$ and $g_{s_{\text{max}}}$ for each specific plant and the required $\psi_m$ was targeted, plants were rewatered daily during the late afternoon and early evening until the soil was fully saturated. Photosynthetic recovery was then measured in the morning on a daily basis until values were higher than 50% of the maximum. In case a particular plant showed slow recovery, Li-Cor measurements were not conducted daily, but every 2–3 days. The number of days for $A$, $E$ and $g_s$ to recover to more than half of the maximum values ($t_{1/2}$, days$^{-1}$) were counted. For some plants, $t_{1/2}$ was estimated by applying a linear regression of the data, especially when the exact recovery day was missed. Lethal water potential was calculated by plotting $\psi_m$ against $1/t_{1/2}$ (Brodribb and Cochard 2009, Brodribb et al. 2010), which was done for $A$, $E$ and $g_s$ (Urli et al. 2013). After the experiment, all plants were watered on a daily basis, including the plants that showed no photosynthetic recovery. We then determined survival of the severely desiccated plants in April 2015 by noting whether or not there was leaf flushing.

**Embolism resistance of stem xylem**

In late May 2013 and 2014, four to eight branches from at least five specimens per species that were ~5 years old were collected in the morning. Branches were immediately wrapped in wet towels and dark plastic foil to prevent dehydration. The samples were then transported to Göttingen University and embolism resistance was measured within 5 days using the Cavitron technique (Cochard et al. 2005). Stems were connected to a Xylem apparatus (Bronkhorst, Montigny les Cormeilles, France) for determining the maximum hydraulic conductivity at 6 kPa, interrupted by three 10-min flushes at 120 kPa to remove potential embolism. Subsequently, branch samples of 27.5 cm length were mounted in a custom-built rotor chamber of the Cavitron, which uses a commercially available centrifuge as basis (Sorvall RC-5C, Thermo Fisher Scientific, Waltham, MA, USA), and spun at defined velocities as recorded with the CaviSoft programme (version 2.1, University of Bordeaux, Bordeaux, France) to generate stem vulnerability curves. For all hydraulic measurements we used demineralized, filtered (0.2 μm) and degassed water, to which 10 mM KCl and 1 mM CaCl$_2$ were added.

For *F. excelsior*, the maximum vessel length based on air injection was ~1.0–1.2 m, which exceeded the sample length of the Cavitron (Cochard et al. 2005). Therefore, the bench dehydration technique was used to construct stem vulnerability curves for this species. Eight 1.5–2.0 m long sun-exposed branches were collected from five adult trees in the Botanical Garden of Ulm University, immediately brought to the laboratory, re-cut under water, and then dehydrated in the laboratory with different time intervals ranging from 1 to 24 h (Wang et al. 2014). After bagging up the plant material for 1 h to obtain equilibrium between $\psi_l$ and stem $\psi_s$, three leaves attached to the current year stem were measured for $\psi_l$. Branches were then rehydrated in water for half an hour to eliminate cutting artefacts, and were then recut under water several times into 7–10 cm long current year stem segments (Wheeler et al. 2013, Torres-Ruiz et al. 2015). These final segments were used to take PLC measurements with a Sperry apparatus:

$$\frac{\text{PLC}}{\text{TOD}} = \frac{K_{\text{max}} - K_{\text{nl}}}{K_{\text{max}}}$$  \hspace{1cm} (1)$$

with $K_{\text{nl}}$ (kgs$^{-1}$ m MPa$^{-1}$) representing the hydraulic conductivity of the stems under different water potential before being flushed with degassed and distilled water, and $K_{\text{max}}$ (kgs$^{-1}$ m MPa$^{-1}$) representing the hydraulic conductivity of the same stems after being flushed.

**PV curves**

In August 2013, three branches that were 2–5 years old were collected in the early morning. The samples were cut off at a height of 2–3 m from three different adult trees per species growing in the Botanical Garden of Ulm University. They were put into bags filled with wet tissue and immediately brought to the laboratory. Three mature leaves per species were selected to generate leaf PV curves. Briefly, the initial $\psi_l$ was measured using a Scholander pressure chamber and the fresh weight was measured using a balance. Then, the leaves were bench dried for several hours while $\psi_l$ and the corresponding drop in weight were measured until $\psi_l$ had dropped to about ~3 MPa. Finally, the leaves were put into the oven at >70 °C for at least 2 days and their dry mass was measured. The leaf PV curves obtained were then used to calculate the osmotic potential at full turgor ($\pi_o$), $\psi_l$ at the turgor loss point ($\psi_{tlp}$), the modulus of elasticity ($E$) and the relative water content at the turgor loss point ($\text{RWC}_{tlp}$) (Sack and Pasquet-Kok 2011).

**Transmission electron microscopy**

Stem and root material from *C. avellana* and *C. betulus* was tested for cambium vitality of both organs. We selected these two species as they belong to the same family and are both diffuse porous species, with *C. betulus* being more resistant to embolism while *C. avellana* was more vulnerable to embolism ($P_{50} = −3.79$ and $−2.09$ MPa, respectively). For both species, we selected a well-watered control plant and a plant that was exposed to drought stress for ~1 month. We assumed that the drought-stressed plants, which had all their leaves wilted and $\psi_m$ around

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...ψ\textsubscript{lethal} were dead. The drought-stressed plant of *C. avellana* was stressed for ~10 days longer than the time required to reach ψ\textsubscript{lethal} for this species. The *C. betulus* seedling, however, was stressed for a shorter period than the average period needed to reach ψ\textsubscript{lethal}. Stem and root cambium samples (~2 mm\(^3\)) from well-watered and severely wilted plants were collected on 9 September 2014 from the main stem (~3 cm above the soil) and the main root (~2 cm below ground). The samples were immediately fixed in a standard solution (2.5% glutaraldehyde, 0.1 mol phosphate, 1% saccharose, pH 7.3). Further transmission electron microscopy (TEM) preparation followed a standard protocol (Scholz et al. 2013). Ultra-thin transverse sections were observed with a JEOL 1400 TEM (JEOL Ltd., Tokyo, Japan) and ImageJ (version 1.48v, National Institutes of Health, Bethesda, MD, USA) was applied to measure the cambial cell wall thickness.

**Statistics**

A sigmoidal function was used in Matlab (version R2013b, MathWorks, Natick, MA, USA) to fit curves for *A, E* and *gs* data versus ψ\textsubscript{m} (Brodribb and Cochard 2009). For each species, the average values of *A\textsubscript{max}, E\textsubscript{max} and *gs\textsubscript{max} based on the measurements from all well-watered individual plants were used to manually fix the starting points of the curves near the *y*-intercept:

\[
y = \frac{a}{1 + \exp(-(x - x_0)/b)},
\]

with *x* (MPa) representing the absolute value of ψ\textsubscript{m}, *y* representing the photosynthetic parameters (*A, E* and *gs*), and *a*, *b*, *x_0* being constants.

### Table 2. Summary of physiological parameters measured in five angiosperm species.

Table 2. Summary of physiological parameters measured in five angiosperm species. Material from seedlings was used for measurements of *A, E* and *gs* while stem vulnerability curves and PV curves were conducted on tree branches with a similar age as the seedlings. Standard deviations (SD) were given for the parameters extracted from three PV curves, but were not available for the other parameters as the data collected were combined into a single curve. Acronyms and units follow the definitions shown in Table 1.

<table>
<thead>
<tr>
<th>Measurements</th>
<th>Parameters</th>
<th><em>A. campestrae</em></th>
<th><em>A. pseudoplatanus</em></th>
<th><em>C. avellana</em></th>
<th><em>C. betulus</em></th>
<th><em>F. excelsior</em></th>
</tr>
</thead>
<tbody>
<tr>
<td>Vulnerability curve</td>
<td><em>P_50</em></td>
<td>−4.80</td>
<td>−2.99</td>
<td>−2.09</td>
<td>−3.79</td>
<td>−2.72</td>
</tr>
<tr>
<td></td>
<td><em>P_88</em></td>
<td>−5.33</td>
<td>−4.13</td>
<td>−2.77</td>
<td>−4.43</td>
<td>−3.75</td>
</tr>
<tr>
<td></td>
<td><em>Ee</em></td>
<td>−4.15</td>
<td>−1.75</td>
<td>−1.33</td>
<td>−3.02</td>
<td>−1.60</td>
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<tr>
<td></td>
<td><em>S\textsubscript{PS50}</em></td>
<td>0.77</td>
<td>0.40</td>
<td>0.66</td>
<td>0.65</td>
<td>0.44</td>
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<tr>
<td><em>A</em></td>
<td>*A\textsubscript{max}</td>
<td>12.97</td>
<td>10.62</td>
<td>7.85</td>
<td>11.12</td>
<td>12.26</td>
</tr>
<tr>
<td></td>
<td>*ψ\textsubscript{fz}</td>
<td>−3.37</td>
<td>−1.44</td>
<td>−1.67</td>
<td>−3.27</td>
<td>−2.94</td>
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<tr>
<td></td>
<td>*S\textsubscript{PS50}</td>
<td>−4.45</td>
<td>−13.48</td>
<td>−12.13</td>
<td>−4.06</td>
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<tr>
<td><em>gs</em></td>
<td>*g\textsubscript{s max}</td>
<td>0.22</td>
<td>0.17</td>
<td>0.14</td>
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<td>*S\textsubscript{Ps50}</td>
<td>−1.89</td>
<td>−0.97</td>
<td>−1.24</td>
<td>−2.18</td>
<td>−1.55</td>
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<tr>
<td></td>
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<td>−0.27</td>
<td>−0.65</td>
<td>−0.06</td>
<td>−0.10</td>
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<tr>
<td></td>
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<td>−3.39</td>
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<tr>
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<td>−6.84</td>
<td>−0.71</td>
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<td>PV curve</td>
<td>*π\textsubscript{Ip}</td>
<td>−1.87 ± 0.17</td>
<td>−1.75 ± 0.14</td>
<td>−1.85 ± 0.65</td>
<td>−2.33 ± 0.07</td>
<td>−2.08 ± 0.18</td>
</tr>
<tr>
<td></td>
<td><em>ε</em></td>
<td>7.43 ± 0.16</td>
<td>8.02 ± 2.14</td>
<td>4.57 ± 2.11</td>
<td>17.02 ± 7.81</td>
<td>9.51 ± 2.78</td>
</tr>
<tr>
<td></td>
<td>*π\textsubscript{e}</td>
<td>−1.55 ± 0.07</td>
<td>−1.47 ± 0.11</td>
<td>−1.39 ± 0.46</td>
<td>−1.87 ± 0.17</td>
<td>−1.74 ± 0.11</td>
</tr>
<tr>
<td></td>
<td>RWC\textsubscript{Ip}</td>
<td>78.19 ± 1.26</td>
<td>77.97 ± 6.35</td>
<td>61.68 ± 15.54</td>
<td>86.91 ± 4.99</td>
<td>78.81 ± 9.37</td>
</tr>
</tbody>
</table>

*A*, *E* and *gs* were calculated as the *x*-intercepts of tangents drawn through the midpoint of the fitted curves, and referred to ψ\textsubscript{s} at the cessation of photosynthesis, transpiration and stomatal closure, respectively. The slopes of the tangents for these curves (S\textsubscript{A50}, S\textsubscript{E50} and S\textsubscript{gs50}, Table 1) were also derived from this function.

The following Weibull function (Cai et al. 2014) was used to fit the stem vulnerability curves in Matlab, to determine *P_50*, *P_88* and *S\textsubscript{Ps50}.* *E* was calculated as the *x*-intercept of a tangent drawn through the midpoint of the following Weibull curve (Meinzer et al. 2009):

\[
\text{PLC} = 100 \times \left( 1 - e^{-\left( \frac{\psi}{T} \right)^c} \right)
\]

with *T* representing the tension of the Cavitron (MPa), and *b* and *c* being Weibull constants.

### Results

**Stem vulnerability curves and leaf photosynthetic dynamics during drought**

There was considerable variation in the stem vulnerability curves of the five species (Table 2; Figure 1). The most embolism vulnerable species was *C. avellana* (*P_50* = −2.1 MPa) and the most embolism resistant one was *A. campestrae* (*P_50* = −4.8 MPa). The stem vulnerability curves could be fitted with the Weibull function.
(root mean square error ≤ 5.4) and all curves were sigmoidal. The start of the stem vulnerability curve of *A. pseudoplatanus*, however, showed PLC values that were slightly higher than the fitted curve (Figure 1c).

Overall, *A*, *E* and *g*ₚ were significantly correlated with each other (*P* < 0.01). Variation in the photosynthetic response to different levels of drought was observed in the five species (Figure 1; Table 2), which was also clearly based on variation in *S*ₜ₅₀, *S*ₑ₅₀ and *S*ₜ₅₀ values (Table 2). Furthermore, embolism-resistant species such as *A. campestre* and *C. betulus* showed higher photosynthetic capacity, while more embolism vulnerable species (i.e., *C. avellana* and *A. pseudoplatanus*) showed lower

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**Figure 1.** Stem vulnerability curves and stomatal response curves under different midday leaf water potential (*ψₘ*) for the five species studied (a–e). For stem vulnerability curves, each PLC value (closed circles) is based on the mean value ± SD from four to eight stems, except for *F. excelsior* (d), where one point is collected from one stem segment. Each point (triangles) in the stomatal response curves is the mean value of three measurements from a single leaf. The regression between the xylem water potential at stomatal closure (*gₛₑ*) and the xylem water potential at the onset of embolism (*Pₑ*) is shown in (f). This regression line was forced to go through the origin and the grey line represents the 1 : 1 line. *Ac*, *Acer campestre*; *Ap*, *Acer pseudoplatanus*; *Ca*, *Corylus avellana*; *Cb*, *Carpinus betulus* and *Fe*, *Fraxinus excelsior*. 

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photosynthetic levels. However, statistical analysis showed no significant linear regression between $A_{max}$ and $P_{50}$ ($R^2 = 0.57$, $P = 0.14$), $E_{max}$ and $P_{50}$ ($R^2 = 0.01$, $P = 0.86$) and $g_{s \max}$ and $P_{50}$ ($R^2 = 0.33$, $P = 0.31$).

**Stomatal response to drought and leaf turgor**

*Carpinus betulus* and *C. avellana* were considered to be isohydric as $g_s$ decreased dramatically within a small range of $\psi_m$, which was also shown by more negative $S_{g50}$ values compared with the other species (Table 2). *Fraxinus excelsior*, *A. campestre* and *C. betulus* were anisohydric. They were found to tolerate a declining $\psi_m$ with a slow reduction of gas exchange (Figure 1). The anisohydric stomatal behaviour of these species was also reflected in their high (i.e., close to zero) $S_{g50}$ values.

*Acer pseudoplatanus*, *C. avellana* and *F. excelsior* had less negative $P_e$ values than *A. campestre* and *C. betulus*. The former three species also had less negative $A_e$, $E_e$ and $\psi_{gse}$ values than the latter two. There was a weak and non-significant 1 : 1 correlation between $\psi_{gse}$ and $P_e$ across the five species ($R^2 = 0.57$, $P = 0.14$; Figure 1f). A similar but significant relationship was found between $A_e$ and $P_e$ ($R^2 = 0.55$, $P = 0.002$). There was also no significant correlation between $g_{s \max}$ and $P_{50}$ ($R^2 = 0.2$, $P = 0.15$), $\psi_{g50}$ and $P_{50}$ ($R^2 = 0.45$, $P = 0.22$) and between $E_e$ and $P_e$ ($R^2 = 0.57$, $P = 0.14$).

Moreover, there was a linear, non-significant relationship between $\psi_{gse}$ and $\pi_{o50}$ ($R^2 = 0.56$, $P = 0.14$; Figure 2), which was different from the 1 : 1 relationship. A positive relationship between $\psi_{g50}$ ($\psi_m$ corresponding to 50% stomatal closure) and $\pi_{o50}$ was found ($R^2 = 0.63$, $P = 0.11$). Both relationships suggested that stomatal closure and leaf turgor were coordinated.

There was no relationship between $P_{50}$ and $\pi_o$ (osmotic potential at full turgor; $R^2 = 0.04$, $P = 0.74$).

**Lethal water potential**

In general, the seedlings of all species showed relatively quick recovery (<9 days) under moderate water stress, and relatively slow recovery (up to 18 days) or recovery failure after severe water stress. Similar recovery trends were found for $E$, $A$ and $g_s$, although there were minor differences in the recovery time $t_{1/2}$. The relationship between $1/2tlp$ and $\psi_m$ was based on $E$ had the best fit compared with $A$ and $g_s$. Therefore, $E$ was the best indicator to estimate $\psi_{ lethal}$ for the five species studied.

The relationship between $t_{1/2}$ as based on $E$ values and $\psi_m$ could be fitted with a linear regression for all species. Both *C. avellana* and *F. excelsior* had low fitting probability ($R^2 = 0.29$, $P = 0.08$ and $R^2 = 0.36$, $P = 0.04$, respectively; Figure 3a–e). There were intraspecific differences in $t_{1/2}$ among the plants stressed to similar $\psi_m$. For example, $t_{1/2}$ of two *C. avellana* seedlings with $\psi_m$ at $-2.9$ and $-3$ MPa were 11 and 7 days, respectively. All five species were able to recover to half of the $E_{max}$ overnight, except for *C. betulus*, which required at least 2 days (Figure 3b). However, a few *F. excelsior* seedlings showed quick recovery of $E$ overnight for $\psi_m$ values from $-1.75$ to $-3.91$ MPa.

The average $\psi_{ lethal}$ of the five tree species ranged from $-4.39 \pm 0.26$ to $-6.06 \pm 0.47$ MPa, which corresponded to PLC values between 87 and 100%, with minor differences only for $A$, $E$ and $g_s$ (Table 3). Lethal water potential calculated based on $A$ ($\psi_{ lethalA}$), $E$ ($\psi_{ lethalE}$) and $g_s$ ($\psi_{ lethalg}$) were positively correlated with embolism resistance of the stem xylem. Specifically, $\psi_{ lethal}$ had a linear but non-significant relationship with $P_{50}$, $P_{88}$ and $P_{99}$ ($R^2 = 0.20$, 0.17 and 0.13, and $P = 0.45$, 0.49 and 0.58, respectively). Nevertheless, $\psi_{ lethal}$ was closest to $P_{99}$ and the linear regression between both parameters was close to a 1 : 1 relationship when we included data from six additional species from literature ($y = 0.93x + 0.65$, $R^2 = 0.51$, $P = 0.02$).

Two seedlings of *C. avellana* that were water-stressed during 2 weeks showed complete leaf mortality. However, new leaves flushed at the bottom of the stem within a few weeks after rewatering. Moreover, leaf flushing was observed after winter in April 2015 for a total of six seedlings from all five species that were water-stressed beyond $\psi_{ lethal}$ (Figure S1 available as Supplementary Data at [Tree Physiology Online](http://www.treephys.oxfordjournals.org)).

**Cambium vitality**

The fusiform stem cambium cells of the well-watered *C. avellana* and *C. betulus* showed a maximum thickness of 1.59 and 2.23 μm, respectively, and many lipid droplets in a dense cytoplasm (Figure 4a and c). The cell membrane of the cambium cells was closely attached to the cell walls. The cambium cells in a stem sample of *C. avellana* included numerous small vacuoles, protein bodies, an electron dense nucleus and in some cases nucleolus. In *C. betulus*, the cambium showed vacuoles and a
smooth and dense endoplasmic reticulum. No clear ultrastructural differences were noticed between the stem and the root cambium in the well-watered control plants.

The plasmalemma of stem cambium cells in the water-stressed plants was shrunken or ruptured, showing clear leakage and breakdown of cellular organelles (Figure 4b and d). In C. avellana, all fusiform cambium cells were distinctly damaged, showing various empty vesicles, but little or no intact cell organelles. Also, nuclei and nucleoli were not observed in these cells. About 90% of the fusiform cambium cells in C. betulus were

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**Figure 3.** Linear regression between midday leaf water potential (\(\psi_m\)) and the reciprocal of the time (1/\(t_{1/2}\)) required to recover to more than half of the maximum transpiration rate (\(E_{\text{max}}\)) in five species (a–e). The more negative the plants were water-stressed, the higher the 1/\(t_{1/2}\) values, except for a few plants from F. excelsior. The lethal water potential based on \(E (\psi_{\text{lethal}})\) was calculated as the intercept value of the linear regression on the x axis, ranging from \(-4.4\) to \(-6\) MPa. A similar approach was applied to \(A\) and \(g_s\) to estimate \(\psi_{\text{lethalA}}\) and \(\psi_{\text{lethalgs}}\). The linear regression (f) between the mean lethal water potential (\(\psi_{\text{lethal}}\), MPa) and \(P_{99}\) was based on five species from this study (closed circles) and six species from the literature (triangles; Barigah et al. 2013a, Urli et al. 2013). Ac, Acer campestre; Ap, Acer pseudoplatanus;Cb, Carpinus betulus; Ca, Corylus avellana; Fs, Fagus sylvatica; Fe, Fraxinus excelsior; Pd, Populus deltoides × Populus nigra; Pt, Populus tremula; Qi, Quercus ilex; Qp, Quercus petraea; Qr, Quercus robur.
Table 3. Lethal water potential ($\psi_{lethal}$) of the five species studied based on photosynthesis rate ($A$), transpiration rate ($E$) and stomatal conductance ($g_s$), together with the corresponding PLC values, as well as the mean lethal water potential and PLC values ($\pm$SD). Standard deviation values are not available for $\psi_{lethal}$ and the corresponding PLC, which are calculated from single linear regressions shown in Figure 3.

<table>
<thead>
<tr>
<th>Parameters</th>
<th>A. campestre</th>
<th>A. pseudoplatanus</th>
<th>C. avellana</th>
<th>C. betulus</th>
<th>F. excelsior</th>
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<tr>
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<td>$-6.09$</td>
<td>$-4.61$</td>
<td>$-4.86$</td>
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<tr>
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<td>$-4.46$</td>
<td>$-4.77$</td>
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<td>$-5.62$</td>
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<td>$-5.91$</td>
<td>$-4.09$</td>
<td>$-4.85$</td>
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<tr>
<td>$\psi_{lethal}$</td>
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<td>$-4.39 \pm 0.26$</td>
<td>$-4.83 \pm 0.05$</td>
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<tr>
<td>PLC</td>
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<td>$92.14 \pm 4.35$</td>
<td>$100.00 \pm 0.00$</td>
<td>$95.52 \pm 7.51$</td>
<td>$99.99 \pm 0.01$</td>
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Discussion

Seedlings that were drought-stressed to $\psi_s$ values close to 99% stem PLC failed to recover after the drought experiment in 2014 (Table 3). This confirms that angiosperms are more resistant to high levels of PLC than gymnosperms (Brodribb and Cochard 2009). However, plants that were drought-stressed beyond the estimated $\psi_{lethal}$ showed leaf flushing during next spring, which indicates that $\psi_{lethal}$ values based on photosynthetic recovery time are underestimated, and that true values of $\psi_{lethal}$ are likely to be at $\psi_s$ beyond 100% PLC. This finding suggests that lethal levels of drought might be higher than values previously reported (Tyree and Sperry 1988, Tyree et al. 2002, Kursar et al. 2009, Barigha et al. 2013a, Urli et al. 2013), although differences could be due to intraspecific variation, the experimental set up, the age of the plant material, the pot size (Poorter et al. 2012) and micro-environmental conditions. Flushing of new leaves was also reported for Populus tremula and Quercus robur by Urli et al. (2013). According to the vulnerability segmentation hypothesis (Tyree and Ewers 1991), distal parts of plants such as leaves could be more easily sacrificed than stems and roots. Therefore, leaf mortality and leaf abscession does not equal plant death per se (Lu et al. 2010). Instead, it is a protective mechanism for deciduous plants to cope with severe drought, and to preserve water and nutrients in tissues that remain alive (Griffiths et al. 2014).

The recovery performance of xylem hydraulic conductivity was negatively correlated with embolism resistance under moderate water stress levels in seven temperate tree species (Ogasa et al. 2013). We did not find a significant correlation between photosynthetic recovery and xylem embolism resistance. However, given the positive correlation between xylem embolism resistance and $\psi_{lethal}$ as well as a functional link between plant micromorphology and embolism resistance (Lens et al. 2011, Markestein et al. 2011), we speculate that $\psi_{lethal}$ is linked with structural features. Acer pseudoplatanus and C. avellana have relatively large leaves (leaf area was 116.80 ± 40 and 59.48 ± 23.47 cm², respectively) compared with the other species investigated, which also results in their lower Huber values. These characters could be associated with the less negative $\psi_{lethal}$ values in both species. Nevertheless, the lack of a significant correlation between $\psi_{lethal}$ and $\psi_s$ suggests that either additional factors may affect $\psi_{lethal}$ or that our experimental approach was not sufficiently accurate.

In general, the technique applied to estimate $\psi_{lethal}$ provides a laborious and time-consuming approach, which has several shortcomings (Figure 3). One of the main problems concerns the intraspecific variation in the recovery rates measured, which could be explained by differences in the microclimate during recovery. For instance, one seedling of F. excelsior that was stressed to −2.65 MPa recovered to more than half of the $g_s$ overnight upon rewatering under a midday temperature of 20.5 °C and 73% relative air humidity, while another seedling that was stressed to −2.6 MPa required ~4 days to recover under warmer midday temperatures (on average 28.25 °C) and drier relative air humidity (on average 46%). This kind of variation could be avoided by working under climate controlled conditions. However, even plants with a similar $\psi_s$ that were recovering under similar vapour pressure deficit levels showed considerable differences in recovery rates. For example, one plant of C. avellana that was water-stressed to −2.92 MPa recovered after 11 days to half of the $g_s$ while another plant water-stressed to −3 MPa required only 7 days to recover under similar temperature and relative humidity. Additional shortcomings could be that photosynthetic parameters decrease considerably with leaf ageing (Mason et al. 2013, Locke and Ort 2014), which could result in overestimation of $\psi_{lethal}$. Moreover, it is possible that water potential measurements of $\psi_s$ and $\psi_s$ become uncoupled during the gradual process of leaf wilting.

Photosynthetic recovery and xylem embolism resistance. However, given the positive correlation between xylem embolism resistance and $\psi_{lethal}$ as well as a functional link between plant micromorphology and embolism resistance (Lens et al. 2011, Markestein et al. 2011), we speculate that $\psi_{lethal}$ is linked with structural features. Acer pseudoplatanus and C. avellana have relatively large leaves (leaf area was 116.80 ± 40 and 59.48 ± 23.47 cm², respectively) compared with the other species investigated, which also results in their lower Huber values. These characters could be associated with the less negative $\psi_{lethal}$ values in both species. Nevertheless, the lack of a significant correlation between $\psi_{lethal}$ and $\psi_s$ suggests that either additional factors may affect $\psi_{lethal}$ or that our experimental approach was not sufficiently accurate.

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Figure 4. Fusiform cambium cells in transverse stem sections from well-watered (a and c) and severely wilted (b and d) seedlings of *C. avellana* (a and b) and *C. betulus* (c and d). Cambium ray initials in stem (e) and root (f) of severely stressed seedlings of *C. betulus*. CR, cambium ray initial; C, fusiform cambium; ER, endoplasmic reticulum; FW, fibre wall; LD, lipid droplet; N, nucleus; Nu, nucleolus; Pl, plasmalemma; Ve, vesicle; V, vacuole.
and die back, even after bagging up leaves to take water potential measurements.

Several seedlings of *F. excelsior* showed a surprisingly fast recovery after severe water stress levels (Figure 3d). An explanation for the quick recovery is that this species has a relatively large pith (accounting for ~20% of the stem surface area), which could function as hydraulic capacitance tissue during drought stress. We did not find positive root pressure in this species (Cochard et al. 1997), which could also promote fast recovery by refilling embolized conduits. Moreover, *F. excelsior* is the only ring-porous species among the five tree species, which has long and large early wood vessels (maximum vessel diameter = 71 μm). These long and wide vessels might contribute to its high ability to absorb water even under dry soil conditions (Käch et al. 2009). In addition, active osmotic adjustment of leaf tissue in *F. excelsior* may facilitate the large recovery of gas exchange upon rewatering (Guicherd et al. 1993).

*Acer pseudoplatanus* and *C. avellana* can be regarded as isohydric plants, while the other three species are studied more anisohydric. However, this classification might be arbitrary because no clear divergences emerged across 70 woody species due to a continuum in leaf gas exchange response to drought. Accordingly, the water potential at 50% of the maximum stomatal conductance (ψ_{gs50}) was proposed as a quantitative indicator between species (Klein 2014). Indeed, both ψ_{gs50} and S_{gs50} values among the five species studied show a continuum in stomatal behaviour, with isohydric and anisohydric as two extremes.

Ishohydric plants are found to be more vulnerable to xylem embolism, as well as being more commonly distributed in moist habitats, compared with anisohydric plants (Vogt 2001, Meinzer and McCulloch 2013). As embolism resistance is a criterion to evaluate the drought resistance of plants, a close and strong relationship between drought resistance and stomatal control behaviour is likely to occur. Our results are consistent with this hypothesis, as the two isohydric species were the most vulnerable species to xylem embolism, and also are more vulnerable to drought, i.e., it took *A. pseudoplatanus* and *C. avellana* ~14 days of drought stress to reach their lethal water potential. The anisohydric species, however, showed more negative *P*_{50} and ψ_{etm} values, and were more resistant to water stress: ψ_{etm} values in *F. excelsior*, *A. campestre* and *C. betul*us required 25, 47 and 53 days of drought stress, respectively.

A 1 : 1 relationship between stomatal closure and stem embolism onset was observed among the five species (Figure 1f), which demonstrates the physiological integration between stems and leaves (Cochard et al. 2002, Ennajeh et al. 2008). In addition, it reflects the sophisticated stomatal regulation strategy, which is also affected by many other factors, such as ABA concentration (Brodribb et al. 2014, Mcadam and Brodribb 2015) and vapour pressure deficit (Will et al. 2013). Although leaf turgor appears to be an important parameter in plant drought tolerance and plasticity at a global scale (Bartlett et al. 2012, 2014), we did not find a relationship between ψ_{etm} and *P*_{50} for the five species studied. The positive but non-significant correlation between ψ_{etm} and ψ_{gs50} and between *P*_{50} and ψ_{gs50} (Figure 2; Brodribb et al. 2003) is obvious considering that stomatal guard cells need to maintain turgor for stomatal opening.

The cambium ultrastructure of the water-stressed plants can be described as necrosis (van Doorn et al. 2011), with rupture of the plasma membrane and the lack of a vacuole and protoplasm (Figure 4b and d) as typical characters. We observed total fusiform cambium failure at the cell level in *C. avellana*, while 90% cellular failure occurred in the more drought resistant *C. betul*us, suggesting that the ~1 month of drought stress did not result in a similar level of death. Similar cellular patterns were described at four different stages of mortality for the root apical meristem in *Arabidopsis* (Duan et al. 2010). Since cambium is one of the most resistant tissues of the plant to drought stress (Barigah et al. 2013b), visualization of cambium breakdown can be used as a method to determine plant death (Figure 4). We also suggest that the gradual changes in cambial cell death can be used to characterize different stages of plant death (Duan et al. 2010).

Ray cells are responsible for transport of water and nutrients in a radial direction, connecting phloem with xylem parenchyma via ray initial cells (Samuels et al. 2006, Plavcová and Jansen 2015). The higher resistance of the ray initial cells to drought-induced necrosis in roots of *C. betul*us compared with the fusiform cambium cells (Figure 4e and f) could be explained by cytoplasmic connection of the ray initial cells with xylem and phloem ray cells. It is known that this symplastic pathway is important for water and carbon reserves (Chaffey and Barlow 2001, Sokolowska and Zagórska-Marek 2012, Spicer 2014, Pfautsch et al. 2015). Therefore, vitality and connectivity of ray parenchyma cells in the wood could play a role in plant survival (Morris et al. 2015). Nevertheless, data on ray cell death are rare. Nakaba et al. (2006) reported that xylem ray parenchyma remained living between 3 and 10 years in *Abies sachalinensis*, with the upper and lower radial cells showing less symplastic connectivity and faster death than the central ray cells.

In conclusion, our results demonstrated that in the five angiosperm tree species studied, ψ_{etm} based on photosynthetic recovery time was close to stem ψ_{99}. During the next spring, however, recovery of plants that passed the estimated ψ_{etm} suggests that the method applied is probably underestimating the true ψ_{etm} values. This finding also suggests that 100% PLC may not necessarily equal plant death in angiosperm seedlings. Further research on the drought tolerance of meristematic tissues such as cambium could provide an alternative method to predict and define more precisely the point of no recovery and ψ_{etm}. 

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Supplementary data

Supplementary data for this article are available at Tree Physiology Online.

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Conflict of interest

None declared.

References


