Skin denervation and cutaneous vasculitis in systemic lupus erythematosus

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To understand the clinical significance and mechanisms of cutaneous denervation in systemic lupus erythematosus (SLE), we assessed intraepidermal nerve fibre (IENF) density of the distal leg in 45 SLE patients (4 males and 41 females, aged 38.4 ± 13.6 years) and analysed its correlations with pathology, lupus activity, sensory thresholds and electrophysiological parameters. Compared with age- and gender-matched control subjects, SLE patients had lower IENF densities (3.08 ± 2.17 versus 11.27 ± 3.96 fibres/mm, P < 0.0001); IENF densities were reduced in 38 patients (82.2%). Pathologically, 11 patients (24.4%) were found to have definite cutaneous vasculitis; the severity and extent of cutaneous vasculitis were correlated with IENF densities. Patients with active lupus had even lower IENF densities than those with quiescent lupus (1.86 ± 1.37 versus 4.15 ± 2.20 fibres/mm, P = 0.0002). By linear regression analysis, IENF densities were negatively correlated with the SLE disease activity index (r = 0.527, P = 0.0002) and cumulative episodes of lupus flare-up within 2 years before the skin biopsy (r = 0.616, P = 0.0014). Clinically, skin denervation was present not only in the patients with sensory neuropathy but also in the patients with neuropsychiatric syndrome involving the CNS. SLE patients had significantly elevated warm threshold temperatures (P = 0.003) and reduced cold threshold temperatures (P = 0.048); elevated warm threshold temperatures were associated with the reduced IENF densities (P = 0.032). In conclusion, cutaneous vasculitis and lupus activities underlie skin denervation with associated elevation of thermal thresholds as a major manifestation of sensory nerve injury in SLE.

Keywords: neuropathy; skin biopsy; systemic lupus erythematosus; vasculitis; skin innervation

Abbreviations: anti-dsDNA antibodies = anti-double-stranded DNA antibodies; IENF = intraepidermal nerve fibre; NCS = nerve conduction studies; PGP 9.5 = protein gene product 9.5; SAP = sensory action potential; SLE = systemic lupus erythematosus


Introduction

Cutaneous tissues are frequently affected among the diverse involvement of organ systems in systemic lupus erythematosus (SLE) (Kotzin, 1996), and the skin is richly innervated by unmyelinated nerve terminals in the epidermis (Kennedy and Wendelschafer-Crabb, 1993). These observations raise the possibility that skin innervation may be susceptible to immunological derangements in SLE. Recently, skin biopsies with the quantification of intraepidermal nerve fibre (IENF) density have become a new approach to investigate the integrity of epidermal innervation (McCarthy et al., 1995; Herrmann et al., 1999; McArthur and Griffin, 2005). IENF densities are significantly reduced in neuropathies involving...
SLE patients were recruited from National Taiwan University Hospital (Taipei, Taiwan) (April 2002 to March 2004). Diagnosis of SLE was based on consensus criteria (Tan et al., 1982) and the neuropsychiatric syndromes were classified according to the standardized definitions (American College of Rheumatology, 1999). Four or more non-neuropsychiatric criteria were required to establish the diagnosis. Patients visited the rheumatologic clinic regularly (S.C.H. and K.L.L.) for laboratory testing, including complements (C3 and C4) and anti-double-stranded DNA (anti-dsDNA) antibodies. The diagnosis of various neuropsychiatric syndromes was based on a review of medical records and neurological examinations (M.T.T. and S.T.H.). To identify neuropsychiatric syndromes based on objective findings, we excluded mild neuropsychiatric symptoms, i.e. headaches, mild depression and anxiety (Ainiala et al., 2001a; Mitsikostas et al., 2004). Patients with clinically suspected involvement of the CNS underwent neuroimaging (CT or MRI), neurophysiological studies (evoked potentials and EEGs) and CSF examinations. In this report, these patients were defined as having CNS neuropsychiatric syndromes.

Peripheral neuropathy was defined according to the neuropathic symptoms or signs. The sensory signs were classified as the small-fibre type with impairment of at least one sensation to pinprick and temperature and the large-fibre type with impairment of at least one kinaesthetic sensation to joint positioning and vibration (Shun et al., 2004). To avoid confounding symptoms and signs, potential additional neuropathies were not defined in patients with CNS neuropsychiatric syndromes. Patients with conditions known to be associated with peripheral neuropathies were excluded. Initially, 63 consecutive SLE patients were evaluated and 18 of them were excluded: 8 with impaired renal functions, 5 with diabetes, 4 with administration of potentially neurotoxic agents and 1 with alcoholism.

At the time of the skin biopsy, patients were assigned to an active lupus group or a quiescent lupus group. Active lupus or lupus flare-up was defined based on the presence of at least three of the following seven criteria: (i) renal manifestations (proteinuria ≥500 mg/day or urinary casts); (ii) arthritis (≥2 peripheral joints involved); (iii) mucocutaneous manifestations (malar rash, discoid rash or oral ulcers); (iv) serositis (pericarditis, pleurisy or peritonitis); (v) haematological abnormalities (haemolytic anaemia, leukocytes <3500/mm³, lymphocytes <1500/mm³ or platelets <100 000/mm³); (vi) evolving changes in C3 (≥30% reduction in the C3 level); and (vii) evolving changes in anti-dsDNA antibodies (≥50% increase in anti-dsDNA antibodies). These organ syndromes (criteria i–v) had to be newly developed and were defined according to the scale of the European Consensus Lupus Activity Measurement (Vitali et al., 1992). Changes in serological markers (criteria vi and vii) were relative to the last examination. In regularly followed-up SLE patients, baseline values were those obtained at least 1 month before the skin biopsy (Vitali et al., 1992). In newly diagnosed SLE patients, baseline values were those obtained at least 1 month after the skin biopsy. Therefore, changes in serological markers were defined as follows: [(value obtained at the time of skin biopsy – baseline value)/(baseline value)] × 100%. At each visit, disease activity was measured using the commonly used SLE Disease Activity Index (Bombardier et al., 1992). This is based on the presence or absence of 24 most-important descriptors of disease activity, covering the most-frequently affected organ systems in SLE.

Age- and gender-matched control subjects were retrieved from the database, and were carefully evaluated by detailed questionnaires and neurological examinations to exclude anyone with neurological disorders (Chien et al., 2001; Chang et al., 2004; Lin et al., 2005).

**Skin biopsy**

A skin biopsy was taken following the established procedures after informed consent had been obtained (Chien et al., 2001). The skin surface was anaesthetized with 2% lidocaine, and a punch of 3 mm in diameter was taken from the lateral side of the distal leg, 10 cm above the lateral malleolus. IENF densities are region-dependent (Johansson et al., 1999; Chang et al., 2004). The selection of biopsy site was based on two criteria: (i) the absence of active lupus skin
lesions and (ii) the symptomatic side if sensory symptoms were present in only one extremity. All subjects tolerated the procedure with no obvious discomfort. The protocol was approved by the Ethics Committee of National Taiwan University Hospital.

**Immunohistochemistry**

Skin specimens were fixed with 2% paraformaldehyde–lysineperoxidase (PLP) in 0.1 M phosphate-buffered saline, pH 7.4, for 48 h (Hsieh et al., 2000). Sections of 50 µm perpendicular to the dermis were cut on a sliding microtome (model 440E; Microm, Walldorf, Germany). They were quenched with 1% H2O2, blocked with 5% normal goat serum and incubated with rabbit antiserum to protein gene product 9.5 (PGP 9.5; UltraClone, Isle of Wight, UK, diluted 1 : 1000 in 1% normal serum/Tris) at 4°C for 16–24 h. After rinsing in Tris, sections were incubated with biotinylated goat anti-rabbit IgG at room temperature for 1 h, followed by incubation with the avidin–biotin complex (Vector, Burlingame, CA, USA) for another 1 h. The reaction product was demonstrated with chromogen SG (Vector) and counterstained with eosin (Sigma, St Louis, MO, USA).

**Quantification of epidermal innervation**

Epidermal innervation was quantified by trained examiners who were blinded to the clinical information (Chien et al., 2001). PGP 9.5-immunoreactive nerve fibres in the epidermis were counted at a magnification of ×400 with an Olympus BX40 microscope (Tokyo, Japan) through the depth of the entire section. Each individual nerve with branching points inside the epidermis was counted as one. For epidermal nerves with branching points in the dermis, each individual nerve was counted separately. The length of the epidermis along the upper margin of the stratum corneum in each section was measured using Image-Pro PLUS (Media Cybernetics, Silver Spring, MD, USA). IENF density was therefore derived and expressed as fibres/mm. For each tissue, there were 48–50 sections, and all the sections were sequentially labelled. Every fifth section was immunostained and quantified. The mean value of these sections was considered as IENF density of the tissue specimen. In the distal leg, normative values from our laboratory [mean ± SD (5th percentile)] of IENF density were 11.16 ± 3.70 (5.88) fibres/mm for subjects aged <60 years and 7.64 ± 3.08 (2.50) fibres/mm for subjects aged ≥60 years. The cut-off point for IENF density was 5.88 and 2.50 fibres/mm in the two age groups, respectively (Chang et al., 2004).

**Pathological assessment of skin biopsies**

The pathological characteristics were examined on formalin-fixed paraffin-embedded skin sections stained with haematoxylin–eosin. Both perivascular inflammation and vascular injury (extravasation of red blood cells, fibrinoid necrosis or disruption of endothelial cell integrity) were required to diagnose definite vasculitis (Collins et al., 2003; Lee et al., 2005). The presence of only one criterion (perivascular inflammation or vascular injury) was defined as borderline vasculitis. The extent of vasculitis was further quantified by calculating the vasculitic ratio (i.e. the number of vessels with vasculitis to the number of total vessels). Five sections with an interval of 50 µm between each section were examined; all cross-sectioned vessels were scored. The involvement of more than one-third of the vessels in the dermis (i.e. vasculitic ratio >0.33) was designated as extensive vasculitis, and the other subgroup as limited vasculitis. Immunohistochemistry was performed using the avidin–biotin–peroxidase complex technique as described above. Cell surface antigens were examined by the following monoclonal antibodies: T cells with CD3 (Ventana Medical System, Tucson, AZ, USA; 1 : 100), B cells with CD20 (DAKO, Glostrup, Denmark; 1 : 100) and macrophages with CD68 (DAKO; 1 : 200) (Hattori et al., 1999; Lee et al., 2005). The skin pathology and phenotypes of cellular infiltration were examined by a pathologist (C.T.S.) in a blinded fashion.

**Assessment of sensory thresholds**

Quantitative sensory testing (QST) was performed by using a Thermal Sensory Analyser and Vibratory Sensory Analyser (Medoc Advanced Medical System, Minneapolis, MN, USA) to measure sensory thresholds of warm, cold and vibratory sensations following an established protocol (Yarnitsky and Ochoa, 1991; Lin et al., 2005). Briefly, the machine delivered a stimulus of constant intensity which had been determined by the test algorithm. The intensity of the next stimulus was either increased or decreased by a fixed ratio according to the response of the subject, i.e. whether or not the subject perceived the stimulus. Such procedures were repeated until a predetermined difference in the intensity was reached. The mean intensity of the last two stimuli was the threshold for the level method. Thermal thresholds recorded on the toe were expressed as warm threshold temperature and cold threshold temperature. Vibratory thresholds recorded on the lateral malleolus were measured with similar algorithms. These values were compared with normative values for the age, which had been documented previously (Pan et al., 2001, 2003), and are similar to those of the previous reports (Pan et al., 2001, 2003; Shun et al., 2004). The 95th percentile value for warm thresholds and vibratory thresholds and the 5th percentile value for cold thresholds were defined as the cut-off values, and thresholds beyond these values were considered abnormal.

**Electrophysiological assessment**

Nerve conduction studies (NCS) followed standardized techniques using a Viking IV electromyograph (Nicolet, Madison, WI, USA). An abnormal result on NCS was defined as having abnormalities of one or more nerves with reduced amplitude, prolonged distal latency or slowed nerve conduction velocity. The amplitudes of the compound muscle action potential (CMAP) and sensory action potential (SAP) were compared with the normative data (Pan et al., 2003). We examined bilateral median, ulnar, peroneal, tibial and sural nerves. Sensorimotor polyneuropathy based on NCS was defined as having abnormalities in two or more nerves (Dyck et al., 1985).

**Statistical analysis**

Numerical variables following a Gaussian distribution were compared using t-test and are expressed as the mean ± SD; for those variables not following a Gaussian distribution, the data are expressed as the median (range) and were analysed with the non-parametric Mann–Whitney U-test. Regression analysis was performed using the statistical software SPSS (SPSS, Chicago, IL, USA) and GraphPad Prism (GraphPad Software, San Diego, CA, USA). Forward and backward stepwise linear regressions were applied in the multiple linear regression analysis. Results were considered significant if P < 0.05.

**Results**

**Clinical features of SLE patients**

There were 45 SLE patients (4 males and 41 females) fulfilling the criteria, with a mean age of 38.4 ± 13.6 years and mean
disease duration of 56.8 ± 63.0 months (Table 1). Compared with the quiescent lupus group, patients with active lupus had higher SLE Disease Activity Index \( (P < 0.0001) \), shorter SLE durations \( (P = 0.033) \), higher levels of anti-dsDNA antibodies \( (P = 0.018) \) and lower C3 levels \( (P = 0.042) \).

Clinically, 20 patients (44.4%) had various neuropsychiatric syndromes. For patients with no CNS neuropsychiatric syndromes, seven had clinical peripheral neuropathy (Table 2); paraesthesia with a glove-stocking distribution was the major presentation, and two had additional neuropathic pain. In summary, six patients had sensory neuropathies (four with pure sensory neuropathy and two with sensory and motor neuropathies) and one patient had pure motor neuropathy.

The other thirteen patients had CNS neuropsychiatric syndromes: six with myelopathy, five with encephalopathy, five with cerebrovascular disorders, four with psychosis, two with seizures and three with mood disorders. Ten patients (22.2%) had more than one neuropsychiatric syndrome. There were no detectable abnormalities on the neurological examinations in the other 25 patients, and they remained free of neuropsychiatric syndromes during follow-up.

**Skin innervation in SLE patients**

In the leg skin of control subjects, there were abundant PGP 9.5-immunoreactive nerves in the epidermis and dermis. In the epidermis, nerves arose vertically from the subepidermal nerve plexuses with a typical varicose appearance (Fig. 1A). The profiles of individual nerve fibres in dermal nerve bundles were dense with continuous immunoreactivities (Fig. 1C). In contrast, in the skin of SLE patients, epidermal nerves were markedly reduced (Fig. 1B). The immunoreactive pattern of dermal nerve bundles was fragmented, reflecting axonal degeneration (Fig. 1D).

SLE patients had significantly lower IENF densities compared with the age- and gender-matched control subjects \( (P < 0.0001) \) (Fig. 2). As a whole, IENF densities were reduced in 38 patients (82.2%).

**Cutaneous vasculitis and correlation with IENF densities in SLE**

To explore inflammatory vasculopathy, we examined dermal vessels on paraffin-embedded skin sections. Eleven of the forty-five SLE patients (24.4%) had definite vasculitis...
The infiltrating cells were positive for T cells (CD3, Fig. 3C) and macrophages (CD68, Fig. 3D), but negative for B cells (CD20, Fig. 3E).

There were grading differences in IENF densities among the three classes of vasculitic pathology; IENF density was the lowest in the group of definite vasculitis. IENF density of the borderline vasculitis group was higher than that of the definite vasculitis group ($P = 0.025$), but lower than that in the group with no vasculitis ($P = 0.048$) (Fig. 4). There was no statistical difference in age and gender among the three subgroups.

Among the 11 patients in the definite vasculitis group, patients with extensive vasculitis ($n = 6$) had even lower IENF density than those with limited vasculitis ($n = 5$) [0.71 (0.00–1.84) versus 2.24 (0.84–4.70) fibres/mm, $P = 0.017$]. These findings indicated that the severity and extent of cutaneous vasculitis were associated with epidermal denervation.
Correlation of IENF densities with lupus activity

To understand the clinical significance of cutaneous denervation in SLE, we evaluated the association of IENF densities with disease activities. Patients with active lupus had lower IENF densities than those with quiescent lupus ($P = 0.0002$) (Fig. 5A). Even patients with quiescent lupus had lower IENF densities than those of age- and gender-matched control subjects ($4.15 \pm 2.20$ versus $11.35 \pm 4.31$ fibres/mm, $P < 0.0001$).

IENF densities were negatively correlated with SLE Disease Activity Index ($P = 0.0002$) (Fig. 5B). SLE Disease Activity Index remained significantly associated with IENF densities ($r = 0.527$, $P < 0.0001$) on multiple linear regression analysis including age and gender as independent variables. We further clarified the contribution of different components of European Consensus Lupus Activity Measurement to epidermal denervation; three factors were associated with reduced IENF densities: neuropsychiatric syndromes ($P = 0.0011$), evolving changes in anti-dsDNA antibodies ($P = 0.012$) and evolving changes in C3 ($P = 0.041$) (Fig. 5C).

We tested the hypothesis that the aggravation of lupus activities could influence cutaneous denervation. In the 27 SLE patients with at least 2 years of regular follow-up, IENF densities were negatively correlated with the cumulative episodes of lupus flare-ups within the previous 2 years before skin biopsy ($P = 0.0014$) (Fig. 5D); this variable remained significant with IENF densities on the model of multiple linear regression ($r = 0.616$, $P = 0.001$). IENF densities in patients with more than two episodes of lupus flare-ups ($n = 6$) were markedly reduced, i.e. less than the first percentile of the normative IENF density (2.50 fibres/mm).

Taken together, lupus activities were associated with reduced IENF densities, and more episodes of lupus flare-ups were correlated with a greater degree of epidermal denervation.

Clinical presentations and skin innervation with SLE

For patients with clinical sensory neuropathies (Patients 1–6 in Table 2), IENF densities were lower than those in the remaining 26 patients with no CNS neuropsychiatric syndromes ($P = 0.022$) (Table 3). Reduced IENF densities were associated with sensory symptoms ($P = 0.043$) and small-fibre sensory signs ($P = 0.022$), but not with motor symptoms.
Table 3 Clinical presentations and IENF density in SLE patients without CNS neuropsychiatric syndromes (n = 32)

<table>
<thead>
<tr>
<th>Parameter</th>
<th>n</th>
<th>IENF density</th>
<th>P</th>
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<tr>
<td>Sensory neuropathy</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Yes</td>
<td>6</td>
<td>1.99 (0.52–3.12)</td>
<td>0.022*</td>
</tr>
<tr>
<td>No</td>
<td>26</td>
<td>4.07 (0.75–9.58)</td>
<td></td>
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<tr>
<td>Sensory symptoms</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Yes</td>
<td>4</td>
<td>1.99 (0.52–2.53)</td>
<td>0.043*</td>
</tr>
<tr>
<td>No</td>
<td>28</td>
<td>3.63 (0.75–9.58)</td>
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<tr>
<td>Small-fibre sensory sign</td>
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<td></td>
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<tr>
<td>Yes</td>
<td>6</td>
<td>1.99 (0.52–3.12)</td>
<td>0.022*</td>
</tr>
<tr>
<td>No</td>
<td>26</td>
<td>4.07 (0.75–9.58)</td>
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<tr>
<td>Large-fibre sensory sign</td>
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<tr>
<td>Yes</td>
<td>1</td>
<td>1.98</td>
<td></td>
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<tr>
<td>No</td>
<td>31</td>
<td>3.42 (0.52–9.58)</td>
<td></td>
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<tr>
<td>Weakness of the foot</td>
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<td></td>
<td></td>
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<tr>
<td>Yes</td>
<td>3</td>
<td>1.98 (1.38–6.69)</td>
<td>0.747</td>
</tr>
<tr>
<td>No</td>
<td>29</td>
<td>3.42 (0.75–9.58)</td>
<td></td>
</tr>
<tr>
<td>Hyporeflexia (ankle)</td>
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<td></td>
<td></td>
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<tr>
<td>Yes</td>
<td>4</td>
<td>2.26 (1.38–6.69)</td>
<td>0.498</td>
</tr>
<tr>
<td>No</td>
<td>28</td>
<td>3.44 (0.75–9.58)</td>
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IENF density [data are expressed as the median (range)]; sensory neuropathy, defined clinically by symptoms and signs as described in Patients and methods. *P < 0.05 (Mann–Whitney U-test).

weakness (P = 0.747) or ankle hyporeflexia (P = 0.498). As described above, the presence of neuropsychiatric syndromes was associated with epidermal denervation (Fig. 5C). Patients with CNS neuropsychiatric syndromes had lower IENF densities than those with no neuropsychiatric syndromes (P = 0.0009); IENF densities were similar between patients with sensory neuropathy and those with CNS neuropsychiatric syndromes (P = 0.592). These findings suggest that cutaneous denervation was not only present in SLE patients with clinical sensory neuropathy, but also existed in patients with CNS neuropsychiatric syndromes.

Sensory thresholds in SLE

Thirty of the thirty-two patients without the CNS involvement agreed to participate in the QST assessment. Compared with control subjects, these 30 SLE patients had significantly higher warm threshold temperatures (39.51 ± 2.17 versus 37.81 ± 2.09°C, P = 0.003) and lower cold threshold temperatures (28.89 ± 1.71 versus 29.59 ± 0.82°C, P = 0.048). Vibratory thresholds did not significantly differ between SLE patients and control subjects (4.13 ± 4.87 versus 2.43 ± 1.23 µm, P = 0.069). Totally, 10 patients (33.3%) had abnormal thresholds: 6 (18.2%) with elevated warm threshold temperature, 4 (13.3%) with reduced cold threshold temperature and 7 (23.3%) with elevated vibratory threshold. IENF densities were associated with abnormal warm thresholds [1.84 (range 0.74–5.40) versus 4.07 (0.86–9.58) fibres/mm, P = 0.032], but not with abnormal cold thresholds (P = 0.12) and vibratory thresholds (P = 0.28).

Electrophysiological studies of SLE

Compared with control subjects, SLE patients had lower amplitudes of sural SAP (15.94 ± 7.99 versus 21.91 ± 7.50 μV, P = 0.0004), peroneal CMAP (3.74 ± 2.31 versus 5.85 ± 1.67 mV, P < 0.0001) and tibial CMAP (13.77 ± 6.16 versus 17.22 ± 4.14 mV, P = 0.002). Of all the 45 patients, 14 (31.1%) had abnormal results on NCS: 7 with sensorimotor polyneuropathy, 4 with polyradiculopathy, 2 with mononeuropathy multiplex and 1 with mononeuropathy; 12 had electrophysiological features of axonal degeneration, while 2 had demyelinating features. Six patients (13.3%) had reduced SAP amplitudes. In these 14 patients with abnormal results on NCS, 4 had clinical sensory neuropathies (Patients 1, 2, 4 and 5 in Table 2), only 1 was free from neuropsychiatric syndromes and the other 9 patients (64.3%) had various CNS neuropsychiatric syndromes: in total 5 with myelopathy, 5 with cerebrovascular disorders, 3 with encephalopathy and 1 with psychosis. Among the 7 patients with clinical peripheral neuropathy only 2 had reduced SAP amplitudes of sural nerves (Patients 2 and 5 in Table 2). Sensory symptoms were not associated with reduced SAP amplitudes (P = 0.825).

Discussion

Two major findings in this report are (i) skin denervation was present not only in lupus patients with sensory neuropathy, but also in patients with CNS neuropsychiatric syndromes; and (ii) the severity and extent of cutaneous vasculitis were strongly associated with skin denervation. In addition, epidermal denervation was correlated with lupus activities. These findings suggest that the reduction in IENF density reflects lupus activity and neural involvement in SLE.

Skin denervation in SLE

The pathological hallmarks of cutaneous nerve degeneration in SLE are similar to those in metabolic and inflammatory sensory neuropathies (McCarthy et al., 1995; Chiang et al., 2002; Pan et al., 2003; Shun et al., 2004). In SLE, the frequency of skin denervation (82.2%) is much higher than those of QST abnormalities (33.3%) and NCS abnormalities (31.1%). Thermal thresholds parallel IENF densities in SLE, similar to diabetic neuropathies (Malik et al., 2001; Shun et al., 2004). The number of patients with clinical sensory neuropathy could be underestimated in the current analysis. The frequent involvement of CNS in SLE might obscure symptoms of peripheral sensory neuropathies (Ainiala et al., 2001b; Brey et al., 2002). For example, all the six patients with overt myelopathy also had reduced IENF densities and, hence, sensory manifestations owing to peripheral nerve injury could not be ascertained in these patients. The concomitant small-fibre sensory neuropathy could only be demonstrated by cutaneous denervation on skin biopsies.

Cutaneous vasculitis and skin denervation

The present report provides direct evidence that cutaneous vasculitis may underlie skin denervation. Traditionally,
vasculitic neuropathy was evaluated by sural nerve biopsies (Kissel et al., 1985; Dyck et al., 1987; Mellgren et al., 1989; Collins et al., 2003; Mawrin et al., 2004; Seo et al., 2004). Unmyelinated fibres are vulnerable to vasculitis and ischaemic neuropathy (Parry and Brown, 1982; Vital and Vital, 1985; Laconis et al., 1997). Patients with cutaneous vasculitis in systemic vasculitides were reported to have a higher prevalence of peripheral neuropathies (Ramos-Casals et al., 2004). However, the functional correlations with unmyelinated fibre pathology are not well characterized. Extending our previous observations on the skin of patients with systemic vasculitic neuropathy (Lee et al., 2005), the current report indicates that vasculitis is present in a significant proportion of SLE patients despite the absence of active lupus lesions. In SLE, the vasculitic pathology could affect sural nerves (McCombe et al., 1987; Bodi et al., 1998; Mawrin et al., 2003). The current study does not exclude the potential involvement of nerve trunks by vasculitis in lupus, but suggests that cutaneous vasculitis could be a manifestation of systemic vasculitis distributed along the nerve trunks as shown by the finding of reduced SAP amplitude in sural nerves. This examination also establishes the application of skin biopsy as a valuable tool in evaluating inflammatory vasculopathy. The correlation of IENF densities with the severity and extent of cutaneous vasculitic pathology not only supports the idea that vasculitis contributes to the pathogenesis of skin denervation, but also extends the previous understanding of sensory neuropathy in vasculitis.

**Skin denervation and lupus activities**

An intriguing and clinically useful message from the current study is the association of IENF densities with lupus activities. For large-fibre neuropathy in SLE, the associations between electrophysiologic parameters and lupus activity are controversial (Omdal et al., 1991; Sivri et al., 1995; Huynh et al., 1999; Omdal et al., 2001; Harel et al., 2002). A critical issue about cutaneous denervation in SLE is how epidermal nerve degeneration evolves during the long disease course of lupus. Cutaneous denervation in SLE did not correlate simply with the disease duration of SLE; instead, IENF densities were correlated with the disease activities and cumulative episodes of lupus flare-ups. Similar to lupus nephritis (Sidiroopoulos et al., 2005) and corroborated with a recent study of axonal dysfunction in SLE (Appenzeller et al., 2005), this analysis suggests that aggravation of immune dysregulation on a background of lupus activities precipitates epidermal nerve injury in SLE.

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Skin innervation in lupus