RETRACTIONS

Carcinogenesis is retracting the following paper following a joint research misconduct investigation by the Birmingham VA Medical Center and the University of Alabama at Birmingham, AL, USA.


In this paper, splice marks are visible in western blots presented in Figures 3C and 4C. In Figure 3C, a splice line is apparent between the samples from control lanes of skin from UVB-irradiated mice that were treated with and without EGCG. Quantitative data based on these western blots are also presented. There are no original data or records available with which to confirm the experiment or the conclusions that were stated related to these experiments. Thus, the conclusions of this paper could not be substantiated by the available data.

This retraction is based on the unavailability of original data and records, the lack of replicate experiments that validate the published findings, and the ambiguous identification of many of the samples, treatments and controls.

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There are two main concerns. First, the β-actin controls for experimental data displayed in Figures 3E and 5B are identical. However, the experimental treatment for data shown in Figure 3 (increasing concentrations of berberine with or without TPA (Figure 3E) or PGE, (Figure 5B) are different. Thus, there is an ambiguous identification of the control conditions for both experiments. There are no original data or records available with which to confirm the experimental results. Thus, the conclusions of the paper could not be substantiated by the available data.

Second, Figure 1B of this paper contains a photomicrograph of Hs294 cells treated with 10 μg/ml of epigallo-catchin-3-gallate, an extract of green tea. This image is identical to Figure 2A of another paper (Singh, T. and S.K. Katiyar, Green tea catechins reduce invasive potential of human melanoma cells by targeting COX-2, PGE2 receptors and epithelial-to-mesenchymal transition. PLOS ONE, 2011. 6(10): p. e25224 https://doi.org/10.1371/journal.pone.0025224); however, the PLOS ONE image is of control Hs294t cells receiving 0 μM berberine. Examination of the two panels at high magnification indicated that the same micrograph was used in each figure but labeled as belonging to different experimental groups. The lack of available original data and records preclude an unambiguous identification of the cell type and the experimental treatment in either paper.

This retraction is based on the unavailability of original data and records, the lack of replicate experiments that validate the published findings, and the ambiguous identification of many of the samples, treatments and controls.

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In Figure 5, there are several instances in which bands are replicated but labelled to represent different experimental conditions. For example, the bands labeled P16INK4a ‘Ac-Histone H3’ and ‘MDB1’ are the same; the bands labeled P16INK4a ‘Ac-Histone H4 Input’ and ‘MBD1 Input’ and ‘HDAC1 Input’ and RASSF1A ‘HDAC1 Input’ appear to be identical.

In some cases, the bands are flipped (compare MeCp2 and HDAC1 in Panel A). While the Input bands for Panel A are all the same, the Input is identical for the image shown at the bottom of Panel B—which is a different promoter region. There are no original data or records with which to clarify the identities of the input controls or experimental samples that are shown, or with which to validate the findings and conclusions drawn from this experiment.

This retraction is based on the unavailability of original data and records, the lack of replicate experiments that validate the published findings, and the ambiguous identification of many of the samples, treatments and controls.

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