



The role of ADAMTS13 testing in the diagnosis and management of thrombotic microangiopathies and thrombosis

Camila Masias and Spero R. Cataland

Department of Medicine, Ohio State University, Columbus, OH

ADAMTS13 (a disintegrin and metalloproteinase with thrombospondin type 1 motif, 13) is a metalloprotease responsible for cleavage of ultra-large von Willebrand factor (VWF) multimers. Severely deficient activity of the protease can trigger an acute episode of thrombotic thrombocytopenic purpura (TTP). Our understanding of the pathophysiology of TTP has allowed us to grasp the important role of ADAMTS13 in other thrombotic microangiopathies (TMAs) and thrombotic disorders, such as ischemic stroke and coronary artery disease. Through its action on VWF, ADAMTS13 can have prothrombotic and proinflammatory properties, not only

when its activity is severely deficient, but also when it is only moderately low. Here, we will discuss the biology of ADAMTS13 and the different assays developed to evaluate its function in the context of TTP, in the acute setting and during follow-up. We will also discuss the latest evidence regarding the role of ADAMTS13 in other TMAs, stroke, and cardiovascular disease. This information will be useful for clinicians not only when evaluating patients who present with microangiopathic hemolytic anemia and thrombocytopenia, but also when making clinical decisions regarding the follow-up of patients with TTP. (*Blood*. 2018;132(9):903-910)

Introduction

The term microangiopathic hemolytic anemia (MAHA) is used to describe the mechanical destruction of red blood cells that may occur in the setting of hemodynamic turbulence or in thrombotic microangiopathies (TMAs), as the red blood cells are sheared by microthrombi in the arterioles. It is the hallmark of diseases such as thrombotic thrombocytopenic purpura (TTP) and hemolytic uremic syndrome (HUS), but it can also be seen in many other acute illnesses, such as sepsis, malignant hypertension, and preeclampsia. These diseases may have similar clinical presentations yet very different treatments and prognoses. Whereas a majority of patients with immune-mediated TTP (iTTP) will respond to plasma exchange (PEX) and steroids, only 30% of patients with complement-mediated HUS (cm-HUS) will respond to PEX, and most will require complement-inhibition therapy with eculizumab. The treatment of other diseases presenting with TMA findings is directed toward treating the underlying illness.¹

TTP can be distinguished from other causes of MAHA by the finding of severely deficient ADAMTS13, typically <10% of normal.² ADAMTS13 is a plasma protease responsible for the cleavage of von Willebrand factor (VWF), preventing the accumulation of ultra-large VWF (ULVWF) multimers that can spontaneously interact with platelets, leading to microvascular thrombosis.

The role of ADAMTS13 goes beyond TTP. Endothelial activation resulting from inflammation can induce the release of VWF

into the circulation with subsequent ADAMTS13 consumption, leading to moderately decreased ADAMTS13 activity. This can be seen in other forms of TMA and thrombotic disorders, such as myocardial infarction and ischemic stroke. The ADAMTS13 protease has also been the focus of recent research toward the development of novel agents,³⁻⁸ with exciting new potential therapeutic options for patients with TTP. In the same way, a better characterization of the function of ADAMTS13 in maintaining normal vascular homeostasis has opened the door for potential new treatments in patients with inflammatory and thrombotic disorders but only moderately decreased ADAMTS13.

This article will discuss the structure and function of ADAMTS13, its role in the pathophysiology of TMAs, and potential future directions and clinical utilities of ADAMTS13 activity testing in the diagnosis and treatment of TMAs and other thrombotic conditions (Figure 1).

Biology of ADAMTS13

ADAMTS13 has a structure similar to that of other members of the ADAMTS family, composed of 14 different domains: a metalloprotease, a disintegrin, a first thrombospondin type 1 repeat (TSP1), cys-rich, and spacer domains. The distal part of the protease contains 7 additional TSP1 repeats and 2 CUB domains.^{9,10} Each domain has a different role in the function of ADAMTS13 activity, with the CUB and spacer domains being

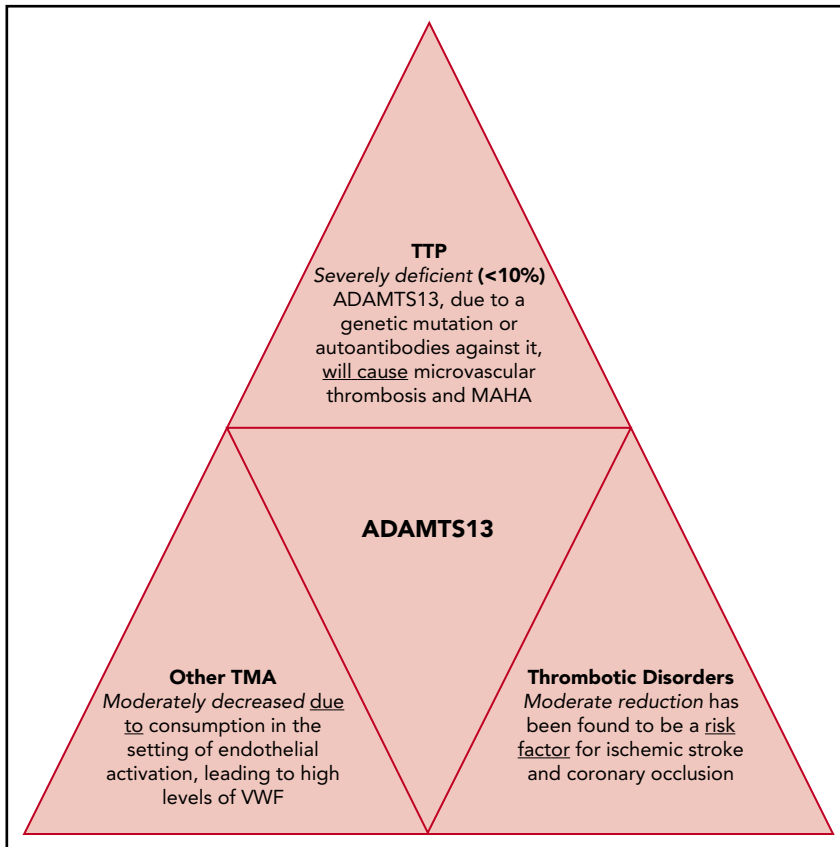


Figure 1. ADAMTS13 deficiency. ADAMTS13 deficiency is the cause of TTP but also has a significant role in the pathophysiology, diagnosis, and prognosis of other TMAs and thrombotic disorders, such as stroke and myocardial infarction.

particularly important in TTP. Even though ADAMTS13 is released as an active enzyme,^{11,12} it circulates in a closed conformation through the interaction between CUB and spacer domains.^{13,14} When VWF binds to ADAMTS13, the protease changes its configuration and becomes activated. Roose et al¹⁵ used antibodies that recognized cryptic epitopes in the spacer domain to further elucidate the conformational changes in the protease. Although ADAMTS13 was found to circulate in a folded conformation in all healthy donors and in 78% of patients in TTP remission, it was found to have an open conformation (defined as a conformation index of >0.5) in 92% of patients with acute TTP. Additional studies are needed to determine the role of these conformational changes of ADAMTS13 in the diagnosis and prognosis of TTP.

ADAMTS13 is synthesized primarily in the interstitial area of hepatic stellate cells,^{10,16,17} but it is also produced in trace amounts in the endothelial cells,^{11,18} megakaryocytes, platelets,^{19,20} renal podocytes, and tubular epithelial cells.^{21,22} Studies in rats and humans have shown that ADAMTS13 activity can drop 30% to 40% after a partial hepatectomy, suggesting that ADAMTS13 produced in other tissues has a small but important contribution to normal plasma protease levels. Endothelial-derived ADAMTS13 may cleave newly formed ULVWF multimers, contributing to the maintenance of a cell surface that is free of hyperadhesive VWF.^{12,23} Although the amount of ADAMTS13 produced outside the liver is small, it may still be biologically important because of its role in thrombus formation, although more studies are needed to better understand this relationship.

ADAMTS13 has a plasma half-life of 2 to 3 days,²⁴ with 3% to 5% of ADAMTS13 circulating bound to VWF.²⁵ There are no data regarding the clearance of the protease, nor any known inhibitor of its function. Therefore, the regulation of ADAMTS13 is likely to be at the substrate level (VWF), with 3 factors known to regulate its activity: (1) fluid shear stress, found in the microcirculation, which allows VWF to unfold and expose its A2 domain for ADAMTS13 binding; (2) factor VIII, which enhances the ADAMTS13 proteolytic activity by affecting the A1A2A3 domain-domain interaction when ADAMTS13 binds VWF under shear forces^{26,27}; and (3) platelet glycoprotein 1b α (GP1b α), which increases ADAMTS13 function under static or shear conditions by exposing the VWF A2 domain when it binds to the A1 domain.²⁸ The main function of the ADAMTS13 protease is the binding and cleavage of cell-bound ULVWF strings at the Tyr1605-Met1606 bond.^{29,30} Under shear stress, as seen in the microcirculation or after thrombus formation, VWF undergoes conformational changes that unfold its A2 domain, making the ADAMTS13 cleavage site accessible.³¹⁻³⁴

Assays of ADAMTS13 activity

There are several ADAMTS13 biomarker assays that are either available for routine clinical use or under study for their potential clinical applications. The most common types of these ADAMTS13 assays measure in vitro functional ADAMTS13 activity.

ADAMTS13 activity assays

Measuring ADAMTS13 activity has become increasingly important in the evaluation and treatment of patients with TTP.

One of the primary limitations of ADAMTS13 activity assays remains the need for nonphysiologic assay conditions (guanidine hydrochloride to expose the VWF cleavage site, low potential of hydrogen and sodium chloride concentrations) to simulate the high shear forces important for in vivo ADAMTS13 function. This limitation has been overcome using a VWF substrate with the VWF cleavage site instead of the full-length VWF molecule.

There have been multiple assays developed in the past 20 years, with the FRETs-WWF73–based assay³⁵ being the most commonly used in clinical settings given the relative ease to perform it, although it is still primarily done in reference laboratories.

ADAMTS13 antigen assay

The levels of the ADAMTS13 antigen can be measured by an enzyme-linked immunosorbent assay (ELISA). Although this test is not yet part of routine testing for patients with TTP, Awan et al³⁶ recently showed that patients with the ADAMTS13 antigen at presentation in the lowest quartile (<1.5%) had a higher TTP-related mortality rate, compared with those with higher levels of the antigen, suggesting a prognostic value of the test.

ADAMTS13 autoantibody and inhibitor assays

Patients with iTTP will have an autoantibody that targets the spacer domain of the protease.³⁷ The detection of an acquired inhibitor is essential to differentiate iTTP from congenital TTP (cTTP). Most of the autoantibodies against ADAMTS13 are of the immunoglobulin G (IgG) isotype (with IgG4 being the most common, followed by IgG1), although IgM and IgA have also been reported.^{38,39} However, quantifying the amount of anti-ADAMTS13 antibodies through an ELISA does not confirm the functional activity of the antibody. Although most of the autoantibodies will have an inhibitory function (which can be measured by a Bethesda-like assay), ~10% to 25% of patients with iTTP have autoantibodies that do not neutralize the protease, but rather are thought to be involved in its in vivo clearance.^{40,41} In the evaluation of patients with TTP, the use of the Bethesda-like assay to demonstrate inhibitory function is important to confirm the diagnosis of iTTP and exclude cTTP.

ADAMTS13-specific CICs

Anti-ADAMTS13 autoantibodies can circulate freely in plasma or bind to ADAMTS13, forming a complex⁴²⁻⁴⁴ that can be detected by an ELISA. Circulating immune complexes (CICs) are found in patients with TTP, during acute episodes and in remission. Although the role of CICs in the pathophysiology of TTP is not entirely clear, Mancini et al⁴⁵ noted a higher risk of recurrence in those patients who had CICs detected during acute episodes of TTP.

Assays of ADAMTS13 in TTP

When a patient presents with thrombocytopenia and MAHA without an alternative clinical explanation, ADAMTS13 activity of <10% confirms the diagnosis of TTP. Additional studies to detect the presence of an inhibitor of the protease with a Bethesda-like assay, as described in “ADAMTS13 autoantibody and inhibitor assays,” will help to differentiate between iTTP or cTTP. If no inhibitor is detected, and the clinical picture is consistent with cTTP (first manifestation in childhood, more rapid initial recovery, persistently deficient ADAMTS13 activity over time with a full recovery of activity with plasma infusion), then an ADAMTS13 mutational analysis should be ordered to confirm the diagnosis of cTTP.

Once a patient with iTTP has achieved clinical response (platelet count $>150 \times 10^9/L$ and normal lactate dehydrogenase, with stabilization or improvement of symptoms), 3 scenarios can occur²: (1) clinical remission (continuous clinical response after cessation of PEX, maintained for >30 days); (2) exacerbation, defined as a recurrent thrombocytopenia and the need to restart PEX within 30 days of the last PEX procedure, which can occur in 40% to 50% of cases⁴⁶; and (3) relapse, defined as a recurrence of TTP >30 days after the last PEX procedure (incidence of ~30%-50%).⁴⁶⁻⁴⁸ Interestingly, neither the normalization of ADAMTS13 activity nor the variability during remission has been incorporated into the clinical definitions of TTP.

Although there is no question regarding the diagnostic role of ADAMTS13 testing at the time of clinical presentation,^{47,49-52} there are uncertainties regarding the role of these assays at different points of the disease (Table 1).

ADAMTS13 activity during the acute episode

Wu et al⁵³ reported the prognostic value of ADAMTS13 activity when measured daily during PEX in 19 patients with iTTP who were severely deficient (<10%) at presentation. The first PEX did not significantly change the ADAMTS13 activity levels. Even after 3 PEX procedures, 14 of the 19 patients still had ADAMTS13 activity <10% (measured immediately before the next PEX procedure). After a week of PEX procedures, 12 of the 19 patients had not achieved ADAMTS13 activity >10%.

Compared with those who recovered their activity before day 7, these patients took a longer time to achieve a clinical response ($P = .001$). There was also a trend toward more exacerbations in the group with more delayed recovery of ADAMTS13 activity in the first week, but the difference was not statistically significant. These data confirm that even in situations in which ADAMTS13 activity is not measured before initiating PEX, there is still a diagnostic role for the test, with a sensitivity of 89%, 83%, and 78% after days 1, 2, and 3 of PEX, respectively.

ADAMTS13 at clinical response of TTP

A retrospective study at our institution evaluated the role of ADAMTS13 activity measured at clinical response to predict exacerbations in 44 acute episodes from 26 patients with iTTP.⁵⁴ In this study, African American race and lower pretreatment ADAMTS13 activity were both associated with an increased risk for exacerbation.

ADAMTS13 activity measured in the first week after stopping PEX was significantly lower in those patients who experienced an exacerbation (1.2% vs 22.5%) compared with those who did not, but the difference was no longer significant after accounting for race as a covariable.

Two recent studies evaluating caplacizumab, a nanobody that binds to the A1 of VWF, preventing the interaction between platelets and VWF, also provide important information regarding the prognostic value of ADAMTS13 to predict exacerbation. In the TITAN study,⁵⁵ 75 patients with a clinical diagnosis of iTTP were randomly assigned to receive caplacizumab or placebo daily with PEX and to continue 30 days after the last PEX. In this study, 11 patients (28%) experienced an exacerbation in the placebo arm, compared with 3 (8%) in the caplacizumab group, supporting the hypothesis that caplacizumab prevents

Table 1. Prediction of relapse using different ADAMTS13-related biomarkers at different points of disease activity

Biomarker	Measured at beginning of acute episode	Measured at clinical response	Measured in remission
ADAMTS13 activity <10%	Diagnostic of TTP	Predictor of relapse	Predictor of relapse ^{38,57,58}
ADAMTS13 Ag	No association ⁸¹	Higher levels associated with sustained remission ⁸²	Associated with relapse vs no association ⁵⁸
ADAMTS13 inhibitor	No association ⁸¹	No association	Associated with relapse
Anti-ADAMTS13 Ab	IgG associated with relapse		IgG ³⁹ associated with relapse (IgG4)
VWF Ag	No association	No association	No association

Ab, antibody; Ag, antigen.

exacerbations of iTTP. Importantly, ADAMTS13 activity was <10% at the time of clinical response in 13 of the 14 patients who experienced an exacerbation. In the follow-up phase 3 Hercules study, a similar decrease in exacerbations was seen in the caplacizumab arm compared with placebo (3% vs 28%), with the presumption that persistently severely deficient ADAMTS13 activity was a significant factor in the development of exacerbations.⁵⁶ In this study, physicians were encouraged (but not required) to continue caplacizumab until there was clear improvement and correction of the severe ADAMTS13 deficiency to avoid recurrences of TTP after stopping caplacizumab.

These data provide the rationale for the use of ADAMTS13 at the time of PEX discontinuation to identify those patients who may be more prone to exacerbations and would benefit from additional immune suppressive therapy.

The French Clinical and Biological Network on Adult TMA published its experience regarding the prognostic value of ADAMTS13 activity and anti-ADAMTS13 antibody characteristics (Ig isotype, titer, and inhibitory effect) in a cohort of 35 patients.³⁸ Samples were obtained within 7 days of achieving clinical response. Thirteen patients (41%) had undetectable activity at the time of clinical response. These patients had a higher relapse rate (38.5%) when compared with those who recovered their activity at clinical response (5%). A high IgG autoantibody level at clinical presentation was also associated with persistently deficient ADAMTS13 activity at clinical response.

Although presently the treatment of acute TTP is directed only at the correction of the thrombocytopenia and any end-organ injury present, these results suggest that weekly monitoring of ADAMTS13 activity after PEX is discontinued may be an equally important biomarker of treatment response, with the goal of correcting the severely deficient ADAMTS13 activity with immune suppressive therapy to prevent exacerbations and relapse of iTTP.

ADAMTS13 monitoring during remission

There are uncertainties surrounding the measurement of ADAMTS13 activity during remission, including how often should it be monitored, the level of ADAMTS13 activity that would prompt the use of preemptive rituximab or another

immune suppressive agent, and the correlation of the known variability of ADAMTS13 activity during remission with relapse risk. Jin et al⁵⁷ found an association between lower ADAMTS13 activity measured in remission and an increased risk of relapse in 24 patients with iTTP followed with serial ADAMTS13 monitoring during remission for an average of 23 months. There was no association found between the risk of relapse and the ADAMTS13 IgG levels. Another study involving 109 patients with iTTP with samples obtained during remission (most commonly 1 sample per patient, at variable times after achieving clinical response; range, 1-96 months) found that ADAMTS13 activity was severely deficient in 32% of patients at remission. Of these 109 patients, 46 had a recurrence. The prevalence of severe ADAMTS13 deficiency was higher in patients with recurrent TTP (46% vs 22%; $P = .01$). The likelihood of recurrence associated with severe ADAMTS13 deficiency when adjusted in the multivariate analysis was statistically significant (odds ratio, 2.9; 95% confidence interval [CI], 1.3-6.8). Despite lower ADAMTS13 antigen levels in the relapse group (median, 36% vs 58%; $P = .003$), the odds ratio did not predict relapse. Finally, regarding the anti-ADAMTS13 antibody analysis, autoantibodies were present in 64% of patients in the relapse group, compared with 36% in the nonrelapse group.⁵⁸

These studies and others (Table 1) form the basis to consider prophylactic rituximab to prevent relapses in patients who are found to have severely deficient ADAMTS13 activity during remission.⁵⁹⁻⁶¹ However, there are also data from patients with iTTP and persistently deficient ADAMTS13 activity in remission who do not uniformly experience relapse, with some patients having an isolated ADAMTS13 activity <10% that improves spontaneously.⁶² We recently calculated the sensitivity and specificity of ADAMTS13 activity measured during remission to predict relapses in the following 30 and 90 days.⁶³ The assays were obtained every 3 months in 39 patients (total of 557 samples). There was a significant variability in ADAMTS13 activity, with 145 (26%) of the samples showing severely deficient activity of <10%; however, only 11 of these were followed by a relapse in the subsequent 90 days. We estimated that the sensitivity and specificity of ADAMTS13 activity <10% to predict a relapse in the next 30 and 90 days were 40.74% and 74.72%, respectively (likelihood ratio, 1.61; CI, 1-2.6). When we used 20% as a threshold, the sensitivity and specificity were both 66% (likelihood ratio, 1.97; CI, 1.47-2.64).

In summary, although severely deficient ADAMTS13 activity in remission may be a risk factor for relapse, it does not mean a relapse will absolutely occur. For decisions regarding prophylactic treatment with rituximab, ADAMTS13 activity <20% may be a more useful threshold than ADAMTS13 <10%. Better models for a more accurate prediction of relapse, incorporating additional biomarkers, are needed. The most appropriate use of these ADAMTS13 activity data would be to have an informed discussion with patients to balance the risk of relapse with the risks of prophylactic rituximab therapy. Our approach to the use of ADAMTS13 testing and prophylactic rituximab therapy is shown in Figure 2. We follow patients after their first acute TTP episode indefinitely, not only to try to prevent relapses, but also to monitor the long-term complications described in these patients.⁶⁴⁻⁶⁶

ADAMTS13 activity in other TMAs

Although severely deficient ADAMTS13 activity (<10%) is unique to TTP, there is ample evidence showing that ADAMTS13 is moderately decreased in other TMAs. Martin et al⁶⁷ reported moderately decreased ADAMTS13 activity in 30 patients with severe sepsis (43.2%; interquartile range, 32.7-67.0) when compared with 29 patients with organ failure resulting from other causes (67.8%; interquartile range, 57.4-87.9; $P < .05$) and 30 healthy participants (105.6%; interquartile range, 87.2-125.6; $P < .001$). A study in 53 patients with TMAs associated with different systemic illnesses, including sepsis, solid organ transplantation, malignancy, and autoimmune diseases, showed decreased ADAMTS13 activity (median activity, 33.5%; range, 16%-47%) when compared with control patients and healthy participants.⁶⁸ Similar findings have been reported in patients with malignant hypertension, in which the median ADAMTS13 activity was 64%.⁶⁹ Increased release of VWF in the setting of endothelial stimulation seen in these disorders is thought to be the cause of the lower ADAMTS13.⁷⁰ These studies and many others highlight the delicate balance between the ADAMTS13 protease and VWF to permit physiologic hemostasis while at the same time preventing pathological thrombosis. Although low ADAMTS13 activity is not the cause of TMAs other than TTP, it seems to have an important prognostic role.

ADAMTS13 in thrombotic disorders

Via its cleavage of VWF, ADAMTS13 is thought to have antithrombotic properties. It is no surprise then that the protease may also have a role in the pathophysiology of cardiovascular disease (CVD). In animal models, *ADAMTS13*^{-/-} mice showed significantly larger infarctions after middle cerebral artery occlusion compared with wild-type (WT) mice.⁷¹ Fujioka et al⁷² found similar results, noting progressively decreased regional cerebral blood flow in *ADAMTS13*^{-/-} mice compared with WT mice after reperfusion after an induced middle cerebral artery occlusion. VWF is released from Weibel-Palade bodies in endothelial cells during episodes of ischemia, making the function of ADAMTS13 even more important, not only before the event but also after reperfusion. Another animal model showed a significant reduction in microvascular length and area, as well as decreased perfusion, in *ADAMTS13*^{-/-} mice 14 days after stroke. *ADAMTS13*^{-/-} mice had significantly reduced endothelial proliferation 2 weeks after stroke, with decreasing neo-vascularization. They also had an 82% increase in blood-brain barrier permeability compared with WT mice. The mechanism

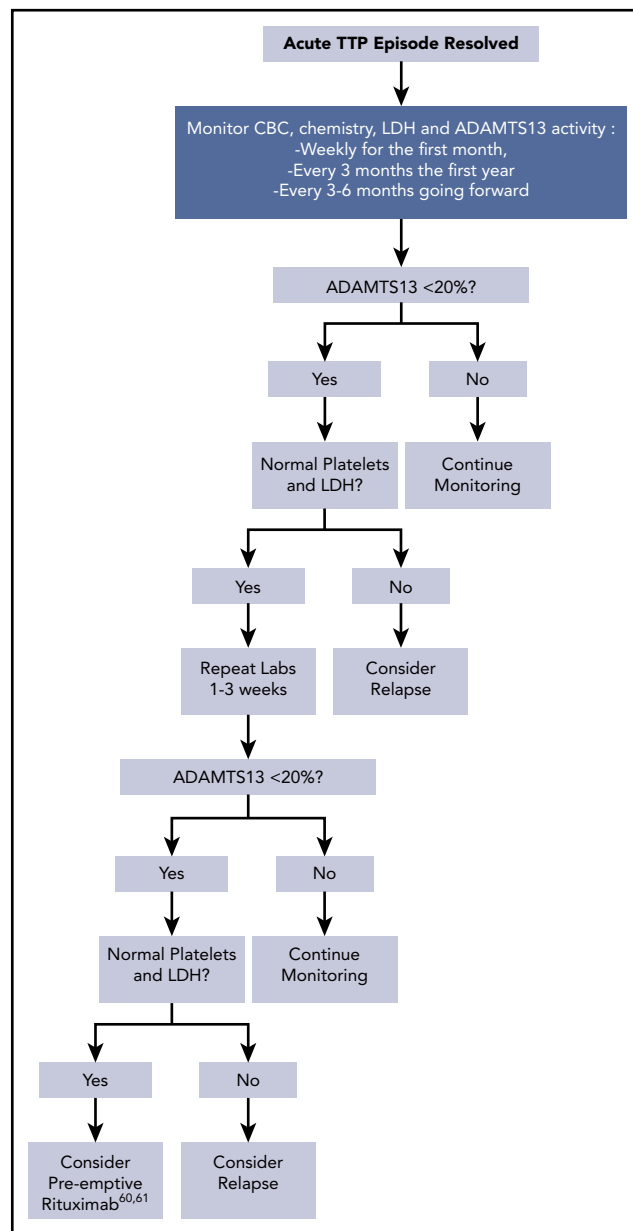


Figure 2. Approach to patient follow-up after initial diagnosis of iTTP. CBC, complete blood count; LDH, lactate dehydrogenase.

involved in this process may be related to downregulation of angiotensin-1 and galectin-3, as well as a decrease in phosphorylation of VEGFR-2 in *ADAMTS13*-deficient mice.⁷³ These studies noted abnormal findings in *ADAMTS13*^{-/-} mice, but mice that were also deficient in VWF behaved similarly to WT mice, implying that the role of ADAMTS13 in stroke is dependent on its action on VWF.

Although severely deficient ADAMTS13 activity in TTP may lead to microthrombi that can be clinically evident as seizures, focal neurologic deficits, and coma, a moderate reduction in ADAMTS13 activity has been found to be a risk factor for ischemic stroke and coronary occlusion. The most complete information in humans comes from the Rotterdam study, a population-based cohort study in which almost 6000 individuals

had their ADAMTS13 activity measured at enrollment to evaluate the association between ADAMTS13 activity, VWF/Ag level, and stroke. Over a median follow-up of 10.7 years, 461 participants had a stroke, 306 of which were ischemic, and 315 individuals had a transient ischemic attack. Patients were divided into 4 quartiles based on their ADAMTS13 activity. Comparing the group in the lowest quartile (ADAMTS13 activity <80%) with those with the highest activity (>102.27%), the former had higher risk of ischemic stroke (absolute risk, 7.3% vs 5.3%), transient ischemic attack (absolute risk, 6.8% vs 4%), and all cerebrovascular events (absolute risk, 17.8% vs 9.3%). Moreover, adding ADAMTS13 to the traditional risk factors used to predict a stroke improved the performance of the model. The investigators also observed a 5.68% decrease in ADAMTS13 activity per 10-year increase in age and an 8.6% higher activity in women compared with men. Diabetes and smoking were also associated with decreased ADAMTS13 activity. These findings were all statistically significant in the population studied.⁷⁴ A case-control study measuring ADAMTS13 antigen and plasma VWF levels in 85 healthy volunteers, 104 patients with an acute ischemic stroke, and 112 patients with chronic cerebrovascular disease found that patients with an acute ischemic stroke had significantly lower levels of ADAMTS13 antigen than healthy volunteers (82.6% ± 21.0% vs 110.6% ± 26.9%; $P < .0001$). Patients with chronic cerebrovascular disease also had lower ADAMTS13 activity than healthy volunteers (99.6% ± 24.5%; $P < .03$), but not as low as those with an acute stroke ($P < .0001$). Furthermore, a higher VWF/ADAMTS13 ratio was associated with stroke severity.⁷⁵

Moderately decreased ADAMTS13 is seen not only in patients with neurovascular disorders but also in those with CVD. A study of 27 patients undergoing emergency cardiac catheterization for ST-elevation myocardial infarction showed moderately decreased ADAMTS13 activity (median, 75%). The study also showed significantly reduced ADAMTS13 activity in coronary blood compared with peripheral blood.⁷⁶ Another subset of the Rotterdam study evaluated the risk for coronary artery disease (CAD) based on ADAMTS13 activity. Of almost 6000 individuals followed for a median of 9.7 years, 456 had CAD; 156 of these cases occurred in the group with ADAMTS13 activity in the lowest quartile (mean, 70.5%), compared with only 85 cases in the group in the highest quartile of ADAMTS13 activity (mean, 114.3%),^{77,78} suggesting a 42% higher risk of developing CAD with moderately deficient ADAMTS13 activity. A follow-up study also reported an association with low ADAMTS13 activity and higher cardiovascular mortality (hazard ratio, 1.46;

95% CI, 1.09-1.96).⁷⁷ Taken together, these findings should be the basis for clinical trials that may lead to new prognostic tools and treatment options for patients with CAD and CVD.

Conclusions and future directions

In the evaluation of a patient with suspected TTP, severely deficient ADAMTS13 <10% will confirm the diagnosis of TTP and differentiate it from other forms of TMA. A better understanding of the role of ADAMTS13 TTP has led to insights regarding its role in CVD, as a regulator of thrombi formation and neovascularization through its action on VWF.

Recombinant ADAMTS13 has shown promising results in a phase 1 study of patients with cTTP in whom the drug was well tolerated, without ADAMTS13 antibody detection, and a good clinical response was seen.³ Moreover, the drug has shown preclinical activity in iTTP via the neutralization of autoantibodies in vitro⁸ and ADAMTS13 immune complex formation in animal models,^{6,79} with reconstitution of ADAMTS13 activity. In thrombotic disorders, infusion of rADAMTS13 in animal models of ischemic stroke has been shown to reduce stroke volume,⁷¹ increase endothelial cell proliferation and perfusion, and decrease the permeability of the blood-brain barrier.⁷³ A model using occlusive VWF-rich thrombi in the middle cerebral arteries of mice showed dissolution of the thrombi with rhADAMTS13 but not with tissue plasminogen activator.⁸⁰ These results suggest a role for rADAMTS13 beyond cTTP and comprise the rationale for future studies.

Authorship

Contribution: C.M. and S.R.C. both wrote, edited, and reviewed the manuscript.

Conflict-of-interest disclosure: S.R.C. serves as a consultant to Ablynx and Shire Pharmaceuticals. The remaining author declares no competing financial interests.

Correspondence: Spero R. Cataland, Department of Hematology, Ohio State University, A361 Starling Loving Hall, 320 W. 10th Ave Columbus, OH 43210; e-mail: spero.cataland@osumc.edu.

Footnote

Submitted 2 February 2018; accepted 3 July 2018. Prepublished online as *Blood* First Edition paper, 13 July 2018; DOI 10.1182/blood-2018-02-791533.

REFERENCES

- Masias C, Vasu S, Cataland SR. None of the above: thrombotic microangiopathy beyond TTP and HUS. *Blood*. 2017;129(21):2857-2863.
- Scully M, Cataland S, Coppo P, et al; International Working Group for Thrombotic Thrombocytopenic Purpura. Consensus on the standardization of terminology in thrombotic thrombocytopenic purpura and related thrombotic microangiopathies. *J Thromb Haemost*. 2017;15(2):312-322.
- Scully M, Knobl P, Kentouche K, et al. Recombinant ADAMTS-13: first-in-human pharmacokinetics and safety in congenital thrombocytopenic purpura. *Blood*. 2017;130(19):2055-2063.
- Antoine G, Zimmermann K, Plaimauer B, et al. ADAMTS13 gene defects in two brothers with constitutional thrombotic thrombocytopenic purpura and normalization of von Willebrand factor-cleaving protease activity by recombinant human ADAMTS13. *Br J Haematol*. 2003;120(5):821-824.
- Schiviz A, Wuersch K, Piskernik C, et al. A new mouse model mimicking thrombotic thrombocytopenic purpura: correction of symptoms by recombinant human ADAMTS13. *Blood*. 2012;119(25):6128-6135.
- Tersteeg C, Schiviz A, De Meyer SF, et al. Potential for recombinant ADAMTS13 as an effective therapy for acquired thrombotic thrombocytopenic purpura. *Arterioscler Thromb Vasc Biol*. 2015;35(11):2336-2342.
- Jian C, Xiao J, Gong L, et al. Gain-of-function ADAMTS13 variants that are resistant to autoantibodies against ADAMTS13 in patients with acquired thrombotic thrombocytopenic purpura. *Blood*. 2012;119(16):3836-3843.
- Plaimauer B, Kremer Hovinga JA, Juno C, et al. Recombinant ADAMTS13 normalizes von Willebrand factor-cleaving activity in plasma of acquired TTP patients by overriding

- inhibitory antibodies. *J Thromb Haemost.* 2011;9(5):936-944.
9. Shelat SG, Ai J, Zheng XL. Molecular biology of ADAMTS13 and diagnostic utility of ADAMTS13 proteolytic activity and inhibitor assays. *Semin Thromb Hemost.* 2005;31(6):659-672.
 10. Zheng XL. Structure-function and regulation of ADAMTS-13 protease. *J Thromb Haemost.* 2013;11(suppl 1):11-23.
 11. Shang D, Zheng XW, Niiya M, Zheng XL. Apical sorting of ADAMTS13 in vascular endothelial cells and Madin-Darby canine kidney cells depends on the CUB domains and their association with lipid rafts. *Blood.* 2006;108(7):2207-2215.
 12. Turner NA, Nolasco L, Ruggeri ZM, Moake JL. Endothelial cell ADAMTS-13 and VWF: production, release, and VWF string cleavage. *Blood.* 2009;114(24):5102-5111.
 13. South K, Luken BM, Crawley JT, et al. Conformational activation of ADAMTS13. *Proc Natl Acad Sci USA.* 2014;111(52):18578-18583.
 14. Tao Z, Peng Y, Nolasco L, et al. Recombinant CUB-1 domain polypeptide inhibits the cleavage of ULVWF strings by ADAMTS13 under flow conditions. *Blood.* 2005;106(13):4139-4145.
 15. Roose E, Schelpe AS, Joly BS, et al. An open conformation of ADAMTS13 is a hallmark of acute acquired thrombotic thrombocytopenic purpura. *J Thromb Haemost.* 2018;16(2):378-388.
 16. Uemura M, Tatsumi K, Matsumoto M, et al. Localization of ADAMTS13 to the stellate cells of human liver. *Blood.* 2005;106(3):922-924.
 17. Levy GG, Nichols WC, Lian EC, et al. Mutations in a member of the ADAMTS gene family cause thrombotic thrombocytopenic purpura. *Nature.* 2001;413(6855):488-494.
 18. Turner N, Nolasco L, Tao Z, Dong JF, Moake J. Human endothelial cells synthesize and release ADAMTS-13. *J Thromb Haemost.* 2006;4(6):1396-1404.
 19. Liu L, Choi H, Bernardo A, et al. Platelet-derived VWF-cleaving metalloprotease ADAMTS-13. *J Thromb Haemost.* 2005;3(11):2536-2544.
 20. Suzuki M, Murata M, Matsubara Y, et al. Detection of von Willebrand factor-cleaving protease (ADAMTS-13) in human platelets. *Biochem Biophys Res Commun.* 2004;313(1):212-216.
 21. Manea M, Kristoffersson A, Schneppenheim R, et al. Podocytes express ADAMTS13 in normal renal cortex and in patients with thrombotic thrombocytopenic purpura. *Br J Haematol.* 2007;138(5):651-662.
 22. Manea M, Tati R, Karlsson J, Békássy ZD, Karpman D. Biologically active ADAMTS13 is expressed in renal tubular epithelial cells. *Pediatr Nephrol.* 2010;25(1):87-96.
 23. Turner N, Nolasco L, Dong JF, Moake J. ADAMTS-13 cleaves long von Willebrand factor multimeric strings anchored to endothelial cells in the absence of flow, platelets or conformation-altering chemicals. *J Thromb Haemost.* 2009;7(1):229-232.
 24. Furlan M, Robles R, Morselli B, Sandoz P, Lämmle B. Recovery and half-life of von Willebrand factor-cleaving protease after plasma therapy in patients with thrombotic thrombocytopenic purpura. *Thromb Haemost.* 1999;81(1):8-13.
 25. Feys HB, Anderson PJ, Vanhoorelbeke K, Majerus EM, Sadler JE. Multi-step binding of ADAMTS-13 to von Willebrand factor. *J Thromb Haemost.* 2009;7(12):2088-2095.
 26. Cao W, Sabatino DE, Altnova E, et al. Light chain of factor VIII is sufficient for accelerating cleavage of von Willebrand factor by ADAMTS13 metalloprotease. *J Biol Chem.* 2012;287(39):32459-32466.
 27. Bonazza K, Rottensteiner H, Schrenk G, et al. Shear-dependent interactions of von Willebrand factor with factor VIII and protease ADAMTS 13 demonstrated at a single molecule level by atomic force microscopy. *Anal Chem.* 2015;87(20):10299-10305.
 28. Skipwith CG, Cao W, Zheng XL. Factor VIII and platelets synergistically accelerate cleavage of von Willebrand factor by ADAMTS13 under fluid shear stress. *J Biol Chem.* 2010;285(37):28596-28603.
 29. Dong JF, Moake JL, Nolasco L, et al. ADAMTS-13 rapidly cleaves newly secreted ultralarge von Willebrand factor multimers on the endothelial surface under flowing conditions. *Blood.* 2002;100(12):4033-4039.
 30. Dong JF. Cleavage of ultra-large von Willebrand factor by ADAMTS-13 under flow conditions. *J Thromb Haemost.* 2005;3(8):1710-1716.
 31. Tsai HM, Sussman II, Nagel RL. Shear stress enhances the proteolysis of von Willebrand factor in normal plasma. *Blood.* 1994;83(8):2171-2179.
 32. Wu T, Lin J, Cruz MA, Dong JF, Zhu C. Force-induced cleavage of single VWFA1A2A3 tridomains by ADAMTS-13. *Blood.* 2010;115(2):370-378.
 33. Siedlecki CA, Lestini BJ, Kottke-Marchant KK, Eppell SJ, Wilson DL, Marchant RE. Shear-dependent changes in the three-dimensional structure of human von Willebrand factor. *Blood.* 1996;88(8):2939-2950.
 34. Crawley JT, de Groot R, Xiang Y, Luken BM, Lane DA. Unraveling the scissile bond: how ADAMTS13 recognizes and cleaves von Willebrand factor. *Blood.* 2011;118(12):3212-3221.
 35. Kokame K, Nobe Y, Kokubo Y, Okayama A, Miyata T. FRET-VWF73, a first fluorogenic substrate for ADAMTS13 assay. *Br J Haematol.* 2005;129(1):93-100.
 36. Alwan F, Vendramin C, Vanhoorelbeke K, et al. Presenting ADAMTS13 antibody and antigen levels predict prognosis in immune-mediated thrombotic thrombocytopenic purpura. *Blood.* 2017;130(4):466-471.
 37. Klaus C, Plaimauer B, Studt JD, et al. Epitope mapping of ADAMTS13 autoantibodies in acquired thrombotic thrombocytopenic purpura. *Blood.* 2004;103(12):4514-4519.
 38. Ferrari S, Scheiflinger F, Rieger M, et al; French Clinical and Biological Network on Adult Thrombotic Microangiopathies. Prognostic value of anti-ADAMTS 13 antibody features (Ig isotype, titer, and inhibitory effect) in a cohort of 35 adult French patients undergoing a first episode of thrombotic microangiopathy with undetectable ADAMTS 13 activity. *Blood.* 2007;109(7):2815-2822.
 39. Ferrari S, Mudde GC, Rieger M, Veyradier A, Kremer Hovinga JA, Scheiflinger F. IgG subclass distribution of anti-ADAMTS13 antibodies in patients with acquired thrombotic thrombocytopenic purpura. *J Thromb Haemost.* 2009;7(10):1703-1710.
 40. Scheiflinger F, Knöbl P, Trattner B, et al. Nonneutralizing IgM and IgG antibodies to von Willebrand factor-cleaving protease (ADAMTS-13) in a patient with thrombotic thrombocytopenic purpura. *Blood.* 2003;102(9):3241-3243.
 41. Thomas MR, de Groot R, Scully MA, Crawley JT. Pathogenicity of Anti-ADAMTS13 Autoantibodies in Acquired Thrombotic Thrombocytopenic Purpura. *EBioMedicine.* 2015;2(8):942-952.
 42. Lotta LA, Valsecchi C, Pontiggia S, et al. Measurement and prevalence of circulating ADAMTS13-specific immune complexes in autoimmune thrombotic thrombocytopenic purpura. *J Thromb Haemost.* 2014;12(3):329-336.
 43. Ferrari S, Palavra K, Gruber B, et al. Persistence of circulating ADAMTS13-specific immune complexes in patients with acquired thrombotic thrombocytopenic purpura. *Haematologica.* 2014;99(4):779-787.
 44. Rieger M, Ferrari S, Kremer Hovinga JA, et al. Relation between ADAMTS13 activity and ADAMTS13 antigen levels in healthy donors and patients with thrombotic microangiopathies (TMA). *Thromb Haemost.* 2006;95(2):212-220.
 45. Mancini I, Ferrari B, Valsecchi C, et al; Italian Group of TTP Investigators. ADAMTS13-specific circulating immune complexes as potential predictors of relapse in patients with acquired thrombotic thrombocytopenic purpura. *Eur J Intern Med.* 2017;39:79-83.
 46. Page EE, Kremer Hovinga JA, Terrell DR, Vesely SK, George JN. Thrombotic thrombocytopenic purpura: diagnostic criteria, clinical features, and long-term outcomes from 1995 through 2015. *Blood Adv.* 2017;1(10):590-600.
 47. Vesely SK, George JN, Lämmle B, et al. ADAMTS13 activity in thrombotic thrombocytopenic purpura-hemolytic uremic syndrome: relation to presenting features and clinical outcomes in a prospective cohort of 142 patients. *Blood.* 2003;102(1):60-68.
 48. Scully M, McDonald V, Cavenagh J, et al. A phase 2 study of the safety and efficacy of rituximab with plasma exchange in acute acquired thrombotic thrombocytopenic purpura. *Blood.* 2011;118(7):1746-1753.
 49. Furlan M, Robles R, Galbusera M, et al. von Willebrand factor-cleaving protease in thrombotic thrombocytopenic purpura and the hemolytic-uremic syndrome. *N Engl J Med.* 1998;339(22):1578-1584.
 50. Tsai HM, Lian EC. Antibodies to von Willebrand factor-cleaving protease in acute thrombotic thrombocytopenic purpura. *N Engl J Med.* 1998;339(22):1585-1594.

51. Zheng XL, Kaufman RM, Goodnough LT, Sadler JE. Effect of plasma exchange on plasma ADAMTS13 metalloprotease activity, inhibitor level, and clinical outcome in patients with idiopathic and nonidiopathic thrombotic thrombocytopenic purpura. *Blood*. 2004; 103(11):4043-4049.
52. Veyradier A, Obert B, Houllier A, Meyer D, Girma JP. Specific von Willebrand factor-cleaving protease in thrombotic microangiopathies: a study of 111 cases. *Blood*. 2001;98(6):1765-1772.
53. Wu N, Liu J, Yang S, et al. Diagnostic and prognostic values of ADAMTS13 activity measured during daily plasma exchange therapy in patients with acquired thrombotic thrombocytopenic purpura. *Transfusion*. 2015;55(1):18-24.
54. Cataland SR, Yang SB, Witkoff L, et al. Demographic and ADAMTS13 biomarker data as predictors of early recurrences of idiopathic thrombotic thrombocytopenic purpura. *Eur J Haematol*. 2009;83(6):559-564.
55. Peyvandi F, Scully M, Kremer Hovinga JA, et al; TITAN Investigators. Caplacizumab for acquired thrombotic thrombocytopenic purpura. *N Engl J Med*. 2016;374(6):511-522.
56. Scully M, Cataland S, Peyvandi F, et al. Results of the randomized, double-blind, placebo-controlled, phase 3 Hercules study of caplacizumab in patients with acquired thrombotic thrombocytopenic purpura [abstract]. *Blood*. 2017;130(suppl 1). Abstract LBA-1.
57. Jin M, Casper TC, Cataland SR, et al. Relationship between ADAMTS13 activity in clinical remission and the risk of TTP relapse. *Br J Haematol*. 2008;141(5):651-658.
58. Peyvandi F, Lavoretano S, Palla R, et al. ADAMTS13 and anti-ADAMTS13 antibodies as markers for recurrence of acquired thrombotic thrombocytopenic purpura during remission. *Haematologica*. 2008;93(2):232-239.
59. Hie M, Gay J, Galicier L, et al; French Thrombotic Microangiopathies Reference Centre. Preemptive rituximab infusions after remission efficiently prevent relapses in acquired thrombotic thrombocytopenic purpura. *Blood*. 2014;124(2):204-210.
60. Westwood JP, Thomas M, Alwan F, et al. Rituximab prophylaxis to prevent thrombotic thrombocytopenic purpura relapse: outcome and evaluation of dosing regimens. *Blood Adv*. 2017;1(15):1159-1166.
61. Joly BS, Coppo P, Veyradier A. Thrombotic thrombocytopenic purpura. *Blood*. 2017; 129(21):2836-2846.
62. Page EE, Kremer Hovinga JA, Terrell DR, Vesely SK, George JN. Clinical importance of ADAMTS13 activity during remission in patients with acquired thrombotic thrombocytopenic purpura. *Blood*. 2016;128(17): 2175-2178.
63. Masias C, Stevens C, Jay L, et al, eds. Prediction of relapse based on remission ADAMTS13 activity in patients with immune-mediated thrombotic thrombocytopenic purpura. Paper presented at Thrombosis and Hemostasis Societies of North America Meeting. 8 March 2018. San Diego, CA.
64. Cataland SR, Scully MA, Paskavitz J, et al. Evidence of persistent neurologic injury following thrombotic thrombocytopenic purpura. *Am J Hematol*. 2011;86(1):87-89.
65. Saultz JN, Wu HM, Cataland S. Headache prevalence following recovery from TTP and aHUS. *Ann Hematol*. 2015;94(9):1473-1476.
66. Deford CC, Reese JA, Schwartz LH, et al. Multiple major morbidities and increased mortality during long-term follow-up after recovery from thrombotic thrombocytopenic purpura. *Blood*. 2013;122(12):2023-2029, quiz 2142.
67. Martin K, Borgel D, Lerolle N, et al. Decreased ADAMTS-13 (A disintegrin-like and metalloprotease with thrombospondin type 1 repeats) is associated with a poor prognosis in sepsis-induced organ failure. *Crit Care Med*. 2007;35(10):2375-2382.
68. Farkas P, Csuka D, Mikes B, et al. Complement activation, inflammation and relative ADAMTS13 deficiency in secondary thrombotic microangiopathies. *Immunobiology*. 2017;222(2):119-127.
69. Khanal N, Dahal S, Upadhyay S, Bhatt VR, Bierman PJ. Differentiating malignant hypertension-induced thrombotic microangiopathy from thrombotic thrombocytopenic purpura. *Ther Adv Hematol*. 2015;6(3): 97-102.
70. van den Born BJ, van der Hoeven NV, Groot E, et al. Association between thrombotic microangiopathy and reduced ADAMTS13 activity in malignant hypertension. *Hypertension*. 2008;51(4):862-866.
71. Zhao BQ, Chauhan AK, Canault M, et al. von Willebrand factor-cleaving protease ADAMTS13 reduces ischemic brain injury in experimental stroke. *Blood*. 2009;114(15): 3329-3334.
72. Fujioka M, Hayakawa K, Mishima K, et al. ADAMTS13 gene deletion aggravates ischemic brain damage: a possible neuroprotective role of ADAMTS13 by ameliorating post-ischemic hypoperfusion. *Blood*. 2010;115(8): 1650-1653.
73. Xu H, Cao Y, Yang X, et al. ADAMTS13 controls vascular remodeling by modifying VWF reactivity during stroke recovery. *Blood*. 2017;130(1):11-22.
74. Sonneveld MA, de Maat MP, Portegies ML, et al. Low ADAMTS13 activity is associated with an increased risk of ischemic stroke. *Blood*. 2015;126(25):2739-2746.
75. Denorme F, Kraft P, Pareyn I, et al. Reduced ADAMTS13 levels in patients with acute and chronic cerebrovascular disease. *PLoS One*. 2017;12(6):e0179258.
76. Pedrazzini G, Biasco L, Sulzer I, et al. Acquired intracoronary ADAMTS13 deficiency and VWF retention at sites of critical coronary stenosis in patients with STEMI. *Blood*. 2016; 127(23):2934-2936.
77. Sonneveld MA, Franco OH, Ikram MA, et al. Von Willebrand factor, ADAMTS13, and the risk of mortality: the Rotterdam study. *Arterioscler Thromb Vasc Biol*. 2016;36(12): 2446-2451.
78. Sonneveld MA, Kavousi M, Ikram MA, et al. Low ADAMTS-13 activity and the risk of coronary heart disease - a prospective cohort study: the Rotterdam Study. *J Thromb Haemost*. 2016;14(11):2114-2120.
79. Plaimauer B, Schiviz A, Kaufmann S, Höllriegel W, Rottensteiner H, Scheiflinger F. Neutralization of inhibitory antibodies and restoration of therapeutic ADAMTS-13 activity levels in inhibitor-treated rats by the use of defined doses of recombinant ADAMTS-13. *J Thromb Haemost*. 2015;13(11):2053-2062.
80. Denorme F, Langhauser F, Desender L, et al. ADAMTS13-mediated thrombolysis of t-PA-resistant occlusions in ischemic stroke in mice. *Blood*. 2016;127(19):2337-2345.
81. Bettoni G, Palla R, Valsecchi C, et al. ADAMTS-13 activity and autoantibodies classes and subclasses as prognostic predictors in acquired thrombotic thrombocytopenic purpura. *J Thromb Haemost*. 2012;10(8): 1556-1565.
82. Yang S, Jin M, Lin S, Cataland S, Wu H. ADAMTS13 activity and antigen during therapy and follow-up of patients with idiopathic thrombotic thrombocytopenic purpura: correlation with clinical outcome. *Haematologica*. 2011;96(10):1521-1527.