



IRON METABOLISM AND ITS DISORDERS

Iron deficiency

Clara Camaschella

Division of Genetics and Cell Biology, San Raffaele Scientific Institute, Milan, Italy

Iron deficiency anemia affects >1.2 billions individuals worldwide, and iron deficiency in the absence of anemia is even more frequent. Total-body (absolute) iron deficiency is caused by physiologically increased iron requirements in children, adolescents, young and pregnant women, by reduced iron intake, or by pathological defective absorption or chronic blood loss. Adaptation to iron deficiency at the tissue level is controlled by iron regulatory proteins to increase iron uptake and retention; at the systemic level, suppression of the iron hormone hepcidin increases iron release to plasma by absorptive enterocytes and recycling macrophages. The diagnosis of absolute iron deficiency is easy unless the condition is masked by inflammatory conditions. All cases of iron deficiency should

be assessed for treatment and underlying cause. Special attention is needed in areas endemic for malaria and other infections to avoid worsening of infection by iron treatment. Ongoing efforts aim at optimizing iron salts-based therapy by protocols of administration based on the physiology of hepcidin control and reducing the common adverse effects of oral iron. IV iron, especially last-generation compounds administered at high doses in single infusions, is becoming an effective alternative in an increasing number of conditions because of a more rapid and persistent hematological response and acceptable safety profile. Risks/benefits of the different treatments should be weighed in a personalized therapeutic approach to iron deficiency. (Blood. 2019;133(1):30-39)

Introduction

Iron balance is essential for all cell life. Iron homeostatic mechanisms evolved to avoid iron excess and the generation of harmful reactive oxygen species by reutilizing body iron and limiting its uptake from the environment. The inevitable other side of the coin is the easy development of iron deficiency.

Iron deficiency is the depletion of total-body iron, especially of macrophage and hepatocyte iron stores. Because the largest amount of iron is consumed for hemoglobin (Hb) synthesis to produce 200 billion erythrocytes daily, anemia is the more evident sign of iron deficiency, and iron deficiency anemia is often considered synonymous with iron deficiency. However, iron deficiency is a broader condition that often precedes the onset of anemia or indicates deficiency in organs/tissues other than those involved in erythropoiesis, such as skeletal muscles and the heart, the latter highly iron dependent for myoglobin and energy production to sustain mechanical contraction.

This article reviews the mechanisms of adaptation to iron deficiency and related anemia, examines how improved knowledge is influencing treatment, and discusses areas that remain uncertain from the biological and clinical perspectives.

Epidemiology

According to the Global Burden of Disease Study 2016, iron deficiency anemia is 1 of the 5 leading causes of years lived with disability burden and is the first cause in women.¹ Adopting the World Health Organization–recommended cutoff for anemia

(Hb <13 g/dL in males, <12 g/dL in females, <11g/dL during pregnancy), a worldwide survey showed that in 2010, anemia still affected one third of the population, with approximately half of the cases resulting from iron deficiency. The estimate is that ~1.24 billion individuals experience iron deficiency anemia, although with huge variations from low- to high-income countries.² The global prevalence of iron deficiency without anemia remains elusive, although the suggested figure is at least double that of iron deficiency anemia. The problem becomes even more relevant if we take into account functional iron deficiency, which occurs when iron is hardly mobilized from stores, as in chronic inflammations/infections or when the vigorous erythropoietic expansion by exogenous or endogenous erythropoietin (EPO) causes an acute disproportion between iron demand and supply.

Globally, iron deficiency anemia has relevant medical and social impacts, accounting for impairment of cognitive performance in young children,³ adverse outcomes of pregnancy for both mothers and newborns,⁴ decreased physical and working capacities in adults, and cognitive decline in the elderly.^{5,6} From available data, the relative contribution of iron deficiency to these negative outcomes is difficult to disassociate from that of anemia.

Pathophysiology of iron deficiency

Iron deficiency deeply affects iron homeostasis, inducing adaptive mechanisms on the hepcidin-ferroportin (FPN) axis, the iron regulatory protein (IRP)/iron responsive element (IRE)

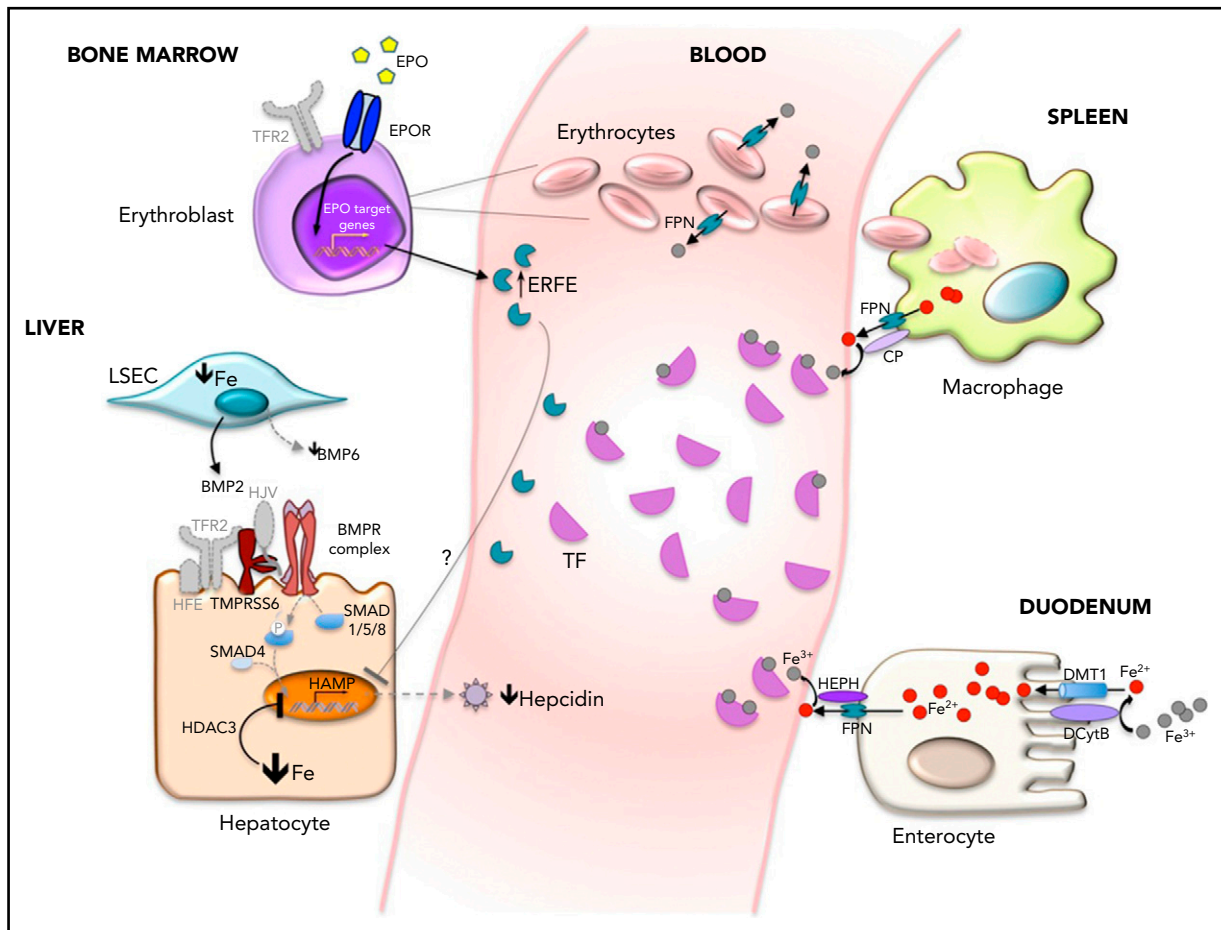


Figure 1. Mechanisms of hepcidin inhibition in iron deficiency anemia. Main cells/organs involved in hepcidin (HAMP) inhibition in iron deficiency are illustrated. In the hepatocytes, bone morphogenetic protein (BMP)-SMAD signaling, the main activator of hepcidin, is low because low levels of BMP6 are produced by liver sinusoidal endothelial cells (L-SEC), the BMP coreceptor hemojuvelin (HJV) is cleaved from the hepatocyte surface by the transmembrane serine protease 6 (TMPRSS6), and the second transferrin receptor (TFR2) is not stabilized on the cell surface in the absence of the ligand diferric transferrin (TF). Low hepcidin levels increase iron absorption by enterocytes and recycling by macrophages through increased activity of the iron exporter FPN. In mild iron deficiency in the absence of hypoxia, increased EPO sensitivity is due to the loss of TFR2 on erythroblast surfaces. Histone deacetylase 3 (HDAC3) participates in hepcidin suppression by erasing markers of activation at the hepcidin locus. In iron deficiency anemia, hypoxia increases EPO. Increased ERFE fully blocks the hepcidin pathway, although the molecular mechanism of hepcidin inhibition by ERFE remains unknown (?). BMPR, BMP receptor; CP, ceruloplasmin; DCYTB, duodenal cytochrome B; DMT1, divalent metal transporter 1; EPOR, EPO receptor; HEPH, hephestin.

machinery, and other regulators. The aim is to optimize iron usage by erythropoiesis and to counteract the physiological inhibition of iron absorption.

Mechanisms of adaptation

Systemic regulation Liver hepcidin is the master hormone that physiologically limits iron entry into plasma. Binding to its receptor FPN, hepcidin blocks iron export both by occluding the exporter central cavity⁷ and by inducing its degradation.⁸ Because of the high FPN expression on professional iron exporter cells, such as enterocytes and macrophages, hepcidin suppression in iron deficiency enhances both iron absorption and its release from macrophages to plasma. Multiple factors downregulate hepcidin transcription (Figure 1). The BMP-SMAD signaling pathway is repressed, because in iron deficiency, expression of BMP6 ligand is low,⁹ the BMP coreceptor HJV is cleaved by TMPRSS6,¹⁰ and TFR2 is removed from the cell surface.¹¹ In addition, the histone deacetylase HDAC3 erases activation markers from the hepcidin locus,¹² providing an epigenetic contribution to hormone suppression. The function of ERFE, released by erythroid cells stimulated by erythropoietin,¹³ is less

relevant in iron deficiency without anemia, because hepcidin is downregulated when iron deficiency is induced in *Erfe*^{-/-} mice.¹² However, ERFE plays a role in the presence of anemia and hypoxia.¹³

Local mechanisms increase intestinal iron absorption. Hypoxia-inducible factor 2 α (HIF2 α) upregulates the expression of both the brush border machinery (DMT1 and DCYTB) that uptakes iron from the lumen and the iron exporter FPN at the basolateral membrane by binding hypoxia-responsive elements of these gene promoters.¹⁴

Macrophages rapidly recycle iron derived from the phagocytosis of senescent red cells (Figure 1). However, the absolute amount of iron recycled from hypochromic erythrocytes by heme-oxygenase 1 decreases in parallel with the severity of iron deficiency, because Hb content per cell (mean corpuscular Hb [MCH]) is reduced. A novel mechanism related to erythrocyte FPN, which is highly expressed in iron deficiency, may contribute to maintaining circulating iron levels.¹⁵ Low serum hepcidin levels ultimately determine the amount of iron entering the circulation (Figure 1).

Cellular regulation Cellular iron content is controlled by IRPs that in iron deficiency bind stem-loop sequences (IREs) in the untranslated regions (UTRs) of iron genes to posttranscriptionally coordinate proteins of iron absorption, export, use, and storage.¹⁶ Binding to 3'UTR IREs, IRPs stabilize the messenger RNA of *TFRC* and *DMT1*; binding to 5'UTR IREs, they repress translation of ferritin, FPN, 5'-aminolevulinic synthase 2 (*ALAS2*), and *HIF2 α* . To avoid deleterious iron retention in iron-deficient enterocytes and maturing erythroblasts, an alternative isoform of FPN lacking 5'UTR IRE escapes IRP control¹⁵ while remaining sensitive to the hepcidin effect.

Other IRP-independent mechanisms optimize iron use in low-iron states. mTOR inhibition activates tristetraprolin, which reduces both TFR1 and FPN expression to save iron for tissue metabolic needs.¹⁷ Cells may recover their own iron stored in ferritin. In iron deficiency, ferritin is delivered to autophagosomes for degradation (ferritinophagy) by nuclear receptor coactivator 4, which in contrast is proteasome degraded in iron-replete cells.¹⁸ Reduced ferritinophagy makes *NcoA4* knockout mice susceptible to hypoferremia and iron deficiency.¹⁹ Ferritinophagy has been shown to provide iron for erythroid differentiation *in vitro*²⁰ and in zebrafish.²¹ It has not been assessed whether ferritin is reduced in plasma when it undergoes ferritinophagy. Although serum ferritin is the best biomarker of iron deficiency, the mechanisms of its release as well as its function in the circulation remain mysterious.²²

Iron-restricted erythropoiesis Iron restriction limits the expansion of early erythropoiesis and optimizes iron use by terminal erythropoiesis. *In vitro* iron deprivation blunts the EPO responsiveness of early progenitors through inactivation of iron-dependent aconitase, which suppresses isocitrate production.²³ Accordingly, iron or isocitrate treatment restores erythroid lineage differentiation.²⁴ EPO is not elevated in mice with iron deficiency without anemia.^{25,26} However, in the same condition, terminal erythropoiesis is modified, with decreased apoptosis and increased number of late erythroblasts. The same phenotype, expression of increased EPO sensitivity, is recapitulated by the genetic loss of the EPOR partner TFR2 in mice; this condition mimics iron deficiency,²⁷ because TFR2 is lost from the membrane when diferric TF is reduced.^{11,28}

With the development of anemia and hypoxia, EPO levels increase exponentially, and multiple mediators, such as erythropoietin,¹³ GDF15,²⁹ and PDGF-BB,³⁰ suppress hepcidin to enhance iron supply. In this process, a role of soluble TFR (sTFR), an accepted biomarker of iron deficiency,³¹ although reasonable, remains unproven.

Because of the increased number of erythroblasts and limited iron supply, heme content per cell is reduced. Globin translation is also impaired by low heme; the stress sensor heme-regulated inhibitor (HRI) phosphorylates the elongation initiation factor 2a (eIF2A) to block translation, concomitantly increasing ATF4, which inhibits the translation regulator mTOR.³² The heme/globin coordination improves erythropoiesis, producing microcytic (low mean corpuscular volume)/hypochromic (low MCH) erythrocytes. The optimization of erythropoiesis might preserve iron for vital functions within a global body economy. However, the mechanism is not fully effective, because even in the absence of anemia, other organs may become iron deficient.

Etiology of iron deficiency

Individuals at risk

Reflecting high iron requirements, infants, preschool children (age <5 years), young menstruating women, and women in the second/third trimester of pregnancy and postpartum are the most affected groups.^{33,34} Adolescents also are susceptible to iron deficiency because of rapid growth.

In Western countries, other healthy individuals may be at risk. These include vegetarians, especially vegans, because of diet restriction and blood donors.³⁵ The RISE study, which evaluated the iron status of >2000 frequent blood donors in the United States, showed that two thirds of women and half of men were iron deficient.³⁶ Elite endurance athletes are at risk because of inflammation-induced increased hepcidin and blood losses. Females are more affected in all the groups listed here.

Iron deficiency with or without anemia may be isolated or secondary to a causative disorder or occur in the context of multiple pathological conditions (eg, in the elderly). Iron deficiency is usually acquired and exceptionally inherited.

Acquired iron deficiency

In developing countries, iron deficiency anemia is nutritional, resulting from reduced intake of bioavailable iron (Table 1), and often associated with infections causing hemorrhages, such as hookworm infestation or schistosomiasis. In Western societies, other than in individuals at risk, iron depletion results from chronic bleeding and/or reduced iron absorption, disorders that may be more relevant than anemia itself (Table 1). For this reason, considering age, sex, clinical history, and symptoms, identification of the underlying cause is an essential part of the patient's workup.^{33,34}

Absolute iron deficiency may be masked by comorbidities (eg, in the elderly, and in the setting of renal failure). Anemia in the elderly has multiple causes.³⁷ Iron deficiency accounts for ~30% of cases, resulting from low intake, reduced absorption (atrophic gastritis, use of proton pump inhibitors), gastrointestinal blood losses (antithrombotic drugs, angiodysplasia, peptic ulcer, hemorrhoids, and even colorectal cancer). Unfortunately, being obscured by comorbidities, it often remains undiagnosed,³⁸ while even mild anemia worsens the outcome of associated disorders and influences mortality.³⁹ Patients with chronic kidney disease (CKD) are prone to absolute iron deficiency because of reduced absorption⁴⁰ and blood loss at dialysis, at an estimated rate of up to 2 to 3 g per year.⁴¹ However, high hepcidin levels and inflammation, which reduce iron mobilization from stores, may mask absolute deficiency. A recognized cause of dysregulation of iron metabolism is obesity, which may lead to iron deficiency, especially after bariatric surgery because of global absorption impairment (Table 1).⁴² In Western countries, as a result of increased life expectancy, these types of iron deficiency are expected to increase in coming years.

Considering the need for balancing iron demand and supply, specific clinical settings are characterized by acute restriction of iron for erythropoiesis. The best-known example is treatment with erythropoiesis-stimulating agents. Another example is postoperative anemia that follows major surgery. Recovery from anemia may be limited or delayed because of preexisting

Table 1. Main causes of absolute iron deficiency/iron deficiency anemia

Type of cause	Condition	Pathophysiologic mechanism
Increased iron requirements	Infants, preschool children, adolescents	Rapid growth
	Pregnant women: second and third trimesters	Expansion of maternal and fetal erythroid mass
	ESA treatment	Acute expansion of erythroid mass
Low iron intake	Malnutrition*	Insufficient dietary iron: low heme iron or scarcely bioavailable iron (eg, chelated by phytates)
	Vegetarians, vegans	
Decreased intestinal iron absorption	Gastrectomy, duodenal bypass, bariatric surgery	Decreased absorptive surface
	Gluten-induced enteropathy	
	Autoimmune atrophic gastritis	Increased pH
	<i>Helicobacter pylori</i> infection	Increased pH and blood loss
	Drugs: proton pump inhibitors, H ₂ blockers	Blocking of gastric acid secretion
	Genetic IRIDA†	High serum hepcidin levels
Chronic blood loss	Hookworm infestation*	Bleeding from gastrointestinal tract
	Gastrointestinal benign and malignant lesions	
	Salicylates, corticosteroids, nonsteroidal anti-inflammatory drugs	
	Heavy menses, hematuria	Bleeding from genitourinary system
	Intravascular hemolysis (PNH, march hemoglobinuria)	Urinary loss of hemoglobin (iron)
	Drugs: anticoagulants, antiplatelet compounds	Systemic bleeding
	Defects of hemostasis (hereditary hemorrhagic telangiectasia, von Willebrand disease)	
	Frequent blood donors	Repeated blood letting
Multiple causes (absolute iron deficiency associated with inflammation)	Chronic infections in malnutrition*	Reduced intake, increased proinflammatory cytokines
	Chronic kidney disease	Decreased iron absorption, increased blood loss, reduced hepcidin excretion and increased production, drugs, ESAs
	Chronic systolic heart failure	Decreased iron absorption, increased inflammation, blood loss
	Inflammatory bowel diseases	Decreased iron absorption, increased blood loss, high hepcidin
	Postoperative anemia of major surgery	Blood loss, increased proinflammatory cytokines

ESA, erythropoiesis-stimulating agent; H₂ antagonists, histamine receptor blockers; IRIDA, iron-refractory iron deficiency anemia; PNH, paroxysmal nocturnal hemoglobinuria.

*More common in developing countries.

†Rarely resulting from gene mutations other than *TMPRSS6*.¹⁰⁰

unrecognized iron deficiency that becomes evident after surgery and/or cytokine-induced defective iron mobilization.

Genetics of iron deficiency

IRIDA⁴³ is a rare recessive condition resulting from mutations of *TMPRSS6*,^{44,45} leading to an inability to cleave the BMP coreceptor HJV and inhibit hepcidin.¹⁰ High hepcidin in IRIDA patients impairs iron absorption, counteracting an essential

compensatory mechanism to sustain erythropoiesis. IRIDA patients are refractory to oral iron supplementation.^{46,47} IV iron is indicated when anemia is severe, but it may be only partially effective. In adults, especially men, anemia may be less evident than in children, while iron deficiency and microcytosis persist.⁴⁸

Populations studies suggest that susceptibility to iron deficiency is in part influenced by genetics. Studies of blood donors

have strengthened the hypothesis that genetic variants of iron genes, especially *TMPRSS6* and *HFE*, reported to influence iron parameters^{49,50} and hepcidin,⁵¹ may predispose to or protect individuals from iron deficiency.⁵²

In a novel murine model, genetic iron deficiency anemia was caused by loss of the enzyme of the sulfur assimilation pathway bisphosphate-3'-nucleotidase (*Bpnt1*). Iron deficiency anemia characterizes both germinal and intestinal conditional *Bpnt1* knockout mice, establishing a novel link between sulfur and iron homeostasis.⁵³

Diagnosing iron deficiency

Clinical signs and symptoms of iron deficiency anemia are limited and often neglected. The most important, fatigue, is unspecific. Alterations of epithelial cells such as dry mouth, cheilitis, atrophic glossitis, Plummer-Vinson pharyngeal webs, and hair loss are observed in longstanding deficiency. Restless leg syndrome reveals iron deficiency in a proportion of cases.^{54,55} In the elderly, iron deficiency anemia may cause heart failure or angina. For a detailed discussion of symptoms in iron deficiency anemia, readers are referred elsewhere.^{34,56}

A correct diagnosis requires laboratory tests. Low serum ferritin levels are the hallmark of absolute iron deficiency, reflecting exhausted stores. Levels <30 mg/L are the accepted threshold that identifies mild cases; in the presence of anemia, ferritin levels are usually lower (<10-12 mg/L). In the absence of inflammations/infections, serum ferritin shows the best correlation with bone marrow stainable iron, once the gold standard in assessing depletion of iron stores.^{33,34}

Measuring TF saturation (<16%) is unnecessary for diagnosis, although it has diagnostic value in functional deficiency when serum ferritin is unreliable. Hepcidin levels, which are low/undetectable in absolute iron deficiency, are also unnecessary. Exceptions are the rare IRIDA patients who show low TF saturation and normal/high hepcidin and serum ferritin levels, reflecting increased macrophage iron. Measuring serum hepcidin may be diagnostic of this atypical iron deficiency, provided that inflammation is excluded.⁵⁷

sTFR and its relationship to ferritin (sTFR/logFt index) are good indicators of iron-deficient erythropoiesis,^{58,59} but tests to measure these indicators are scarcely available in clinics. Reduction of MCH and mean corpuscular volume and increased (>6% in CKD) hypochromic red cells (with MCH <28 pg) occur relatively late because of the erythrocyte lifespan. Reticulocyte Hb content may reveal rapid changes in erythropoietic activity. Early reduction (<26 pg) may occur after erythropoiesis-stimulating agent treatment and early increase after iron supplementation.⁶⁰ When heme is low, zinc is incorporated into protoporphyrin-IX, levels of which become elevated and measurable in mature erythrocytes.⁶⁰

All tissues are assumed to be iron deficient when ferritin is low. No specific test assesses tissue (eg, cardiac or muscle) iron deficiency when ferritin is unreliable, such as in inflammation. Perception of this deficiency by patients is highly variable. Clinical diagnosis relies on deterioration of the specific organ (eg, heart) function or on unspecific symptoms, the most popular

being fatigue. Alternatively, the diagnosis is based on a positive outcome after iron supplementation, such as in heart failure.⁶¹

To correctly diagnose iron deficiency in the context of multiple comorbidities, such as in inflammation, ferritin threshold <100 mg/L or even higher values are suggested, in combination with low (<20%) TF saturation.⁶² Although these arbitrary cutoffs likely overestimate iron deficiency, they are largely used for therapeutic decisions. The diagnosis of absolute iron deficiency is also challenging in the elderly; proposed cutoffs between >30 and <100 mg/L are based on small studies.^{37,38} This supports the need for well-designed prospective clinical trials and development of biomarkers for tissue iron deficiency.

Advances and controversies in the treatment of iron deficiency

The etiological cause of iron deficiency should be addressed in all cases and, whenever possible, eliminated. Iron treatment should be started immediately, even in the absence of anemia, especially in symptomatic patients.^{63,64} A systematic review of the efficacy of iron supplementation in iron-deficient nonanemic individuals concluded that treatment (any type) increased Hb and ferritin and reduced self-reported fatigue but did not improve physical performance or maximal oxygen consumption.⁶⁵

The choice of iron compound and the route of administration are largely dependent on the presence and degree of anemia, reversibility of the underlying cause, clinical status (age, sex, longstanding vs recent onset), and in some instances patient preference.

Oral iron supplementation

Iron salts such as iron sulfate, fumarate, and gluconate remain a mainstay of therapy in absolute iron deficiency. Mounting evidence indicates that low doses are more effective and better tolerated than the traditionally recommended 100 to 200 mg of elementary iron per day. Because absorption of nonheme iron is modest (5% to 28% at the fastest),⁶⁶ high doses may result in ROS-mediated toxicity of nonabsorbed iron on intestinal mucosa. Common adverse effects, such as nausea, vomiting, constipation, or diarrhea, may lead to noncompliance with therapy in 30% to 70% of cases⁶⁷ and jeopardize the prolonged (several months) treatment planned. Importantly, even a mild increase in serum iron activates hepcidin to limit iron absorption. This physiological response was exploited to design the most appropriate dose and schedule of oral iron administration in iron-deficient nonanemic women. In short-term studies that used stable iron isotopes, supplementation with iron sulfate (60-240 mg) induced hepcidin increase for up to 48 hours, limiting the absorption of the subsequent doses.⁶⁸ In another trial in which participants were randomly assigned to receive 60 mg of iron per day for 14 days or on alternate day for 28 days, fractional iron absorption was significantly greater in the latter group (21.8% vs 16.3%). In a study comparing 2 groups of women who were receiving 120 mg of iron sulfate per day either as a single or 2 divided doses, the first group showed smaller serum hepcidin increases.⁶⁹ Altogether these elegant studies indicate that changing the administration from daily to alternate-day schedules and from divided to single doses increases the efficacy of treatment in iron-deficient nonanemic individuals and has the potential to improve tolerability. An ongoing study in

Table 2. Indication for IV iron therapy

Condition	Reason
Oral iron intolerance	Persistent gastrointestinal adverse effects
Oral iron refractoriness	Defective absorption: gastrectomy, duodenal bypass, bariatric surgery
	Intestinal disorders (selected cases): IBD, atrophic gastritis, <i>Helicobacter pylori</i> infection, gluten enteropathy
	Genetic forms (IRIDA)
	No Hb improvement after 4 wk of oral therapy
Severe anemia (Hb <7-8 g/dL)	Need for rapid Hb improvement
Second and third trimesters of pregnancy	Need for rapid Hb increase; often intolerance to oral preparations
ESA treatment	More effective than oral iron in CKD
Chronic blood loss difficult to manage with oral iron	Heavy uterine bleeding
	Hereditary disorders of hemostasis
Other	Postoperative anemia of major surgery
	Chronic systolic heart failure

IBD, inflammatory bowel disease.

women with iron deficiency anemia⁷⁰ is assessing whether the alternate-day protocol should also be recommended in the presence of anemia,⁷¹ when hypoxia further increases intestinal iron absorption and fully suppresses hepcidin.¹⁴

Other adverse effects of unabsorbed iron include alterations in the composition of the gut microbiome, with reduction of beneficial *Lactobacillus* and *Bifidobacterium* bacteria, enhancement of potential pathogens (*Enterobacteriaceae*), and increased inflammation and diarrhea, as shown in African children.^{72,73}

The minimal dose used for iron supplementation is 60 mg per day. Lower doses (37.5 mg per day) of oral iron have proven useful in blood donors to limit deferrals from donations.³⁶

A prophylactic treatment with iron sulfate (60 mg in adults and 30 mg in children) has been recommended in world areas characterized by high prevalence of iron deficiency anemia.⁷⁴ However, the validity of universal supplementation in countries with high prevalence of malaria and/or other infections is controversial. Epidemiological⁷⁵ and in vitro studies have shown that iron deficiency is an adaptation process protecting from *Plasmodium* virulence and that its correction may increase infection severity.^{76,77} Recent evidence shows that FPN expressed in erythrocytes is functional and reduced by the high hepcidin levels induced by iron supplementation. This would increase erythrocyte iron content, favoring the parasite growth.¹⁵ In these cases, iron supplementation should occur in association with antimalarial treatment.⁷⁸ Another problem is related supplemented iron causing gut dysbiosis and diarrhea. To avoid the latter effects, a future solution is the development of iron compounds bioavailable only to humans and not to pathogens.

There is great interest in the development of compounds better tolerated than iron salts; numerous compounds have been proposed (eg, sucrosomial iron, heme iron polypeptide, iron containing nanoparticles), but studies are limited.⁷⁹ Sucrosomial

iron has been tested in patients with CKD,⁸⁰ but the mechanism of absorption and the real benefits are uncertain. In the same condition, the phosphate binder iron ferric citrate simultaneously corrects both hyperphosphatemia and iron deficiency; its double effect is being tested in a clinical trial in CKD.⁸¹ A phase 3 trial of ferric maltol provided positive results on iron deficiency anemia in inflammatory bowel diseases.⁸² Rigorously designed clinical trials are needed to confirm the efficacy of these iron preparations.

The natural compound extracted from the bark of the Taiwanese tree hinokitiol restores iron transport in cells lacking transporters, such as DMT1 or FPN.⁸³ Exploiting the iron gradient that, in the absence of the transporter, is formed across membranes, hinokitiol restores transport direction both in vitro and in zebrafish, but no data are available on its chronic use in mice.

IV iron

The alternative for patients intolerant or unresponsive to oral compounds is IV iron.⁴⁷ Once limited by the risk of severe hypersensitivity reactions, this route of administration is currently more widely used as a result of the improved safety profile of last-generation compounds. Established indications to IV iron are reduced absorption capacity in the presence of gastrointestinal disorders or bariatric surgery, severe anemia (Hb <7-8 g/dL), high hepcidin resulting from concomitant inflammation, and rarely IRIDA and when a fast recovery is desirable (Table 2). Advantages are the more rapid effect and the negligible gastrointestinal toxicity.⁶⁷ IV iron is more effective than oral iron in CKD patients treated with erythropoiesis-stimulating agents^{41,84} and avoids oxidative damage to the intestinal mucosa in active inflammatory bowel diseases.⁸⁵ In the latter disorders, IV iron preserves the normal microbiome, which would be disrupted by oral iron.⁴⁰ The European Crohn's and Colitis Organization recommends IV iron as a first-line therapy for patients with active disease and Hb <10 g/dL and oral iron in inactive disease/mild anemia patients,⁸⁶ the latter being more likely to have absolute iron deficiency.

IV iron is available in different forms; iron gluconate and iron sucrose require repeated infusions, whereas ferric carboxymaltose, ferumoxytol, low molecular weight iron dextran, and iron isomaltoside may be administered in high doses to rapidly replace the total iron deficit (usually 1-1.5 g) in 1 or 2 infusions.⁵⁶ The stable carbohydrate shell of the latter compounds prevents free iron release, a feature that increases their safety.⁵⁶ The high-dose schedule avoids repeated hospital visits (eg, for patients with reduced mobility, such as the elderly, for whom oral therapy can be particularly disturbing)³⁸ and is convenient when a fast recovery is needed (such as in the second and third trimesters of pregnancy or in postpartum anemia⁸⁷ and the prevention of repeated cycles of therapy [eg, in heavy uterine bleeding]).⁸⁸ In addition to the prompt Hb increase, this protocol rapidly reconstitutes stores,⁸⁹ making the advantages (single access, accelerated recovery, limited need for blood tests) outweigh the disadvantages (cost, invasiveness, risk of reactions). However, this decision should be carefully made on an individual basis.

High-dose IV iron may increase Hb or iron stores before surgery predicted to induce heavy bleeding. This is a kind of prevention of acute postoperative anemia and an alternative to blood transfusions, which are associated with several postoperative complications, including infections. Patient blood management programs that limit blood transfusions by perioperative iron use reduce morbidity and negative prognoses in high-risk interventions.⁹⁰ A randomized trial of IV iron administration at postoperative day 1 vs standard care showed less anemia and reduction of transfusions and infections in the iron arm in major orthopedic and abdominal surgeries.⁹¹ The iron infusion approach might be especially valuable in surgery candidates prone to iron deficiency, such as young women or patients with colorectal cancer.⁹²

An important issue concerning IV iron is safety. Because iron is a growth factor for several pathogens, iron therapy is contraindicated in infections. The risk of infection after IV iron is still a matter of controversy. Increased risk was found in a meta-analysis evaluating trials of IV iron to spare transfusions,⁹³ and caution was suggested in dialysis patients.⁹⁴ Another meta-analysis of >10 000 patients receiving different IV compounds or oral iron or placebo did not find different risks of infection.⁹⁵ Long-term studies are needed in patients with different disorders.

Hypophosphatemia after ferric carboxymaltose is usually transient and reversible, although rarely, severe cases have been reported after repeated infusions.⁹⁶ Minor/moderate infusion reactions (nausea, pruritus, urticaria, flushing, back or thoracic pain), often self-limited, may be observed in 1:200 infusions and more serious reactions (hypotension, dyspnea) in 1:200 000.⁹⁵ Although globally IV treatment seems safe, the number of reported patients is usually too limited to detect extremely rare

anaphylactic reactions, such as those once caused by high molecular weight iron dextran. Their unclear pathogenesis is ascribed to the release of iron particles in the circulation and has been interpreted as a "complement activation-related pseudo-allergy."^{97(p5029)} Personnel who administer IV iron must be prepared to manage any type of reaction, including exceptionally severe ones.⁹⁸

The superior efficacy of IV vs oral iron is undisputable and expected; the long-term adverse effects of ROS generation in cases of therapy-induced positive iron balance have been scarcely explored, although overtreatment might occur in functional rather than in absolute iron deficiency. A recent analysis in CKD concluded that patients seemed to tolerate positive iron balance, because iron that was not used was safely stored in reticulo-endothelial cells.⁹⁹ However, in the absence of data and iron toxicity tests, it is advisable to regularly assess iron status when high doses are repeatedly administered.

Future perspectives

Although advances in understanding iron metabolism and regulation are systematically providing novel insights, additional studies are needed before iron therapy becomes a personalized approach in all cases. These studies should aim at discovering markers of tissue iron deficiency, investigate novel schedules of iron administration based on iron physiology, provide clearer indications to high-dose IV iron, and contribute long-term evaluations of treatment outcomes.

Acknowledgments

The author thanks Domenico Girelli for his valuable advice and criticism and Alessia Pagani for help with the figure.

Authorship

Contribution: C.C. conceived, wrote, and reviewed the paper.

Conflict-of-interest disclosure: C.C. is an advisor for Vifor Iron Core and has received honoraria from Vifor Pharma.

ORCID profile: C.C., 0000-0003-1427-0143.

Correspondence: Clara Camaschella, Division of Genetics and Cell Biology, San Raffaele Scientific Institute, Via Olgettina, 58, 20132 Milan, Italy; e-mail: camaschella.clara@hsr.it.

Footnote

Submitted 30 May 2018; accepted 16 July 2018. Prepublished online as *Blood* First Edition paper, 6 November 2018; DOI 10.1182/blood-2018-05-815944.

REFERENCES

- GBD 2016 Disease and Injury Incidence and Prevalence Collaborators. Global, regional, and national incidence, prevalence, and years lived with disability for 328 diseases and injuries for 195 countries, 1990-2016: a systematic analysis for the Global Burden of Disease Study 2016. *Lancet*. 2017;390(10100):1211-1259.
- Kassebaum NJ, Jasrasaria R, Naghavi M, et al. A systematic analysis of global anemia burden from 1990 to 2010. *Blood*. 2014;123(5):615-624.
- McCann JC, Ames BN. An overview of evidence for a causal relation between iron deficiency during development and deficits in cognitive or behavioral function. *Am J Clin Nutr*. 2007;85(4):931-945.
- Achebe MM, Gafter-Gvili A. How I treat anemia in pregnancy: iron, cobalamin, and folate. *Blood*. 2017;129(8):940-949.
- Falkingham M, Abdelhamid A, Curtis P, Fairweather-Tait S, Dye L, Hooper L. The effects of oral iron supplementation on cognition in older children and adults: a systematic review and meta-analysis. *Nutr J*. 2010;9:4.
- Andro M, Le Squere P, Estivin S, Gentic A. Anaemia and cognitive performances in the elderly: a systematic review. *Eur J Neurol*. 2013;20(9):1234-1240.

7. Aschemeyer S, Qiao B, Stefanova D, et al. Structure-function analysis of ferroportin defines the binding site and an alternative mechanism of action of hepcidin. *Blood*. 2018;131(8):899-910.
8. Nemeth E, Tuttle MS, Powelson J, et al. Hepcidin regulates cellular iron efflux by binding to ferroportin and inducing its internalization. *Science*. 2004;306(5704):2090-2093.
9. Kautz L, Meynard D, Monnier A, et al. Iron regulates phosphorylation of Smad1/5/8 and gene expression of Bmp6, Smad7, Id1, and Atoh8 in the mouse liver. *Blood*. 2008;112(4):1503-1509.
10. Silvestri L, Pagani A, Nai A, De Domenico I, Kaplan J, Camaschella C. The serine protease matrilysin-2 (TMPRSS6) inhibits hepcidin activation by cleaving membrane hemoujuvelin. *Cell Metab*. 2008;8(6):502-511.
11. Pagani A, Vieillevoeye M, Nai A, et al. Regulation of cell surface transferrin receptor-2 by iron-dependent cleavage and release of a soluble form. *Haematologica*. 2015;100(4):458-465.
12. Pasricha SR, Lim PJ, Duarte TL, et al. Hepcidin is regulated by promoter-associated histone acetylation and HDAC3. *Nat Commun*. 2017;8(1):403.
13. Kautz L, Jung G, Valore EV, Rivella S, Nemeth E, Ganz T. Identification of erythroferrone as an erythroid regulator of iron metabolism. *Nat Genet*. 2014;46(7):678-684.
14. Mastrogiannaki M, Matak P, Peyssonnaud C. The gut in iron homeostasis: role of HIF-2 under normal and pathological conditions. *Blood*. 2013;122(6):885-892.
15. Zhang DL, Wu J, Shah BN, et al. Erythrocytic ferroportin reduces intracellular iron accumulation, hemolysis, and malaria risk. *Science*. 2018;359(6383):1520-1523.
16. Zhang DL, Ghosh MC, Rouault TA. The physiological functions of iron regulatory proteins in iron homeostasis – an update. *Front Pharmacol*. 2014;5:124.
17. Bayeva M, Khechaduri A, Puig S, et al. mTOR regulates cellular iron homeostasis through tristetraprolin. *Cell Metab*. 2012;16(5):645-657.
18. Mancias JD, Wang X, Gygi SP, Harper JW, Kimmelman AC. Quantitative proteomics identifies NCOA4 as the cargo receptor mediating ferritinophagy. *Nature*. 2014;509(7498):105-109.
19. Bellelli R, Federico G, Matte' A, et al. NCOA4 deficiency impairs systemic iron homeostasis. *Cell Reports*. 2016;14(3):411-421.
20. Ryu MS, Zhang D, Protchenko O, Shakoury-Elizeh M, Philpott CC. PCBP1 and NCOA4 regulate erythroid iron storage and heme biosynthesis. *J Clin Invest*. 2017;127(5):1786-1797.
21. Mancias JD, Pontano Vaites L, Nissim S, et al. Ferritinophagy via NCOA4 is required for erythropoiesis and is regulated by iron-dependent HERC2-mediated proteolysis. *eLife*. 2015;4:e10308.
22. Truman-Rosentsvit M, Berenbaum D, Spektor L, et al. Ferritin is secreted via 2 distinct nonclassical vesicular pathways. *Blood*. 2018;131(3):342-352.
23. Bullock GC, Delehanty LL, Talbot AL, et al. Iron control of erythroid development by a novel aconitase-associated regulatory pathway. *Blood*. 2010;116(1):97-108.
24. Richardson CL, Delehanty LL, Bullock GC, et al. Isocitrate ameliorates anemia by suppressing the erythroid iron restriction response. *J Clin Invest*. 2013;123(8):3614-3623.
25. Ghosh MC, Zhang DL, Jeong SY, et al. Deletion of iron regulatory protein 1 causes polycythemia and pulmonary hypertension in mice through translational derepression of HIF2 α . *Cell Metab*. 2013;17(2):271-281.
26. Anderson SA, Nizzi CP, Chang YI, et al. The IRP1-HIF-2 α axis coordinates iron and oxygen sensing with erythropoiesis and iron absorption. *Cell Metab*. 2013;17(2):282-290.
27. Nai A, Lidonnici MR, Rausa M, et al. The second transferrin receptor regulates red blood cell production in mice. *Blood*. 2015;125(7):1170-1179.
28. Johnson MB, Chen J, Murchison N, Green FA, Enns CA. Transferrin receptor 2: evidence for ligand-induced stabilization and redirection to a recycling pathway. *Mol Biol Cell*. 2007;18(3):743-754.
29. Tanno T, Bhanu NV, Oneal PA, et al. High levels of GDF15 in thalassemia suppress expression of the iron regulatory protein hepcidin. *Nat Med*. 2007;13(9):1096-1101.
30. Sonnweber T, Nachbaur D, Schroll A, et al. Hypoxia induced downregulation of hepcidin is mediated by platelet derived growth factor BB. *Gut*. 2014;63(12):1951-1959.
31. Beguin Y, Clemons GK, Pootrakul P, Fillet G. Quantitative assessment of erythropoiesis and functional classification of anemia based on measurements of serum transferrin receptor and erythropoietin. *Blood*. 1993;81(4):1067-1076.
32. Zhang S, Macias-Garcia A, Velazquez J, Paltrinieri E, Kaufman RJ, Chen JJ. HRI coordinates translation by eIF2 α P and mTORC1 to mitigate ineffective erythropoiesis in mice during iron deficiency. *Blood*. 2018;131(4):450-461.
33. Camaschella C. Iron-deficiency anemia. *N Engl J Med*. 2015;372(19):1832-1843.
34. Lopez A, Cacoub P, Macdougall IC, Peyrin-Biroulet L. Iron deficiency anaemia. *Lancet*. 2016;387(10021):907-916.
35. Kiss JE, Vassallo RR. How do we manage iron deficiency after blood donation? *Br J Haematol*. 2018;181(5):590-603.
36. Cable RG, Brambilla D, Glynn SA, et al; National Heart, Lung, and Blood Institute Recipient Epidemiology and Donor Evaluation Study-III (REDS-III). Effect of iron supplementation on iron stores and total body iron after whole blood donation. *Transfusion*. 2016;56(8):2005-2012.
37. Stauder R, Valent P, Theurl I. Anemia at older age: etiologies, clinical implications, and management. *Blood*. 2018;131(5):505-514.
38. Girelli D, Marchi G, Camaschella C. Anemia in the elderly. *HemaSphere*. 2018;2(3):e40.
39. Culleton BF, Manns BJ, Zhang J, Tonelli M, Klarenbach S, Hemmelgarn BR. Impact of anemia on hospitalization and mortality in older adults. *Blood*. 2006;107(10):3841-3846.
40. Lee T, Clavel T, Smirnov K, et al. Oral versus intravenous iron replacement therapy distinctly alters the gut microbiota and metabolome in patients with IBD. *Gut*. 2017;66(5):863-871.
41. Macdougall IC, Bircher AJ, Eckardt KU, et al; Conference Participants. Iron management in chronic kidney disease: conclusions from a "Kidney Disease: Improving Global Outcomes" (KDIGO) Controversies Conference. *Kidney Int*. 2016;89(1):28-39.
42. Steenackers N, Van der Schueren B, Mertens A, et al. Iron deficiency after bariatric surgery: what is the real problem? *Proc Nutr Soc*. 2018;77(4):445-455.
43. Finberg KE, Heeney MM, Campagna DR, et al. Mutations in TMPRSS6 cause iron-refractory iron deficiency anemia (IRIDA). *Nat Genet*. 2008;40(5):569-571.
44. Du X, She E, Gelbart T, et al. The serine protease TMPRSS6 is required to sense iron deficiency. *Science*. 2008;320(5879):1088-1092.
45. Folgueras AR, de Lara FM, Pendás AM, et al. Membrane-bound serine protease matrilysin-2 (Tmprss6) is an essential regulator of iron homeostasis. *Blood*. 2008;112(6):2539-2545.
46. Camaschella C, Nai A. Ineffective erythropoiesis and regulation of iron status in iron loading anaemias. *Br J Haematol*. 2016;172(4):512-523.
47. Hershko C, Camaschella C. How I treat unexplained refractory iron deficiency anemia. *Blood*. 2014;123(3):326-333.
48. De Falco L, Silvestri L, Kannengiesser C, et al. Functional and clinical impact of novel TMPRSS6 variants in iron-refractory iron-deficiency anemia patients and genotype-phenotype studies. *Hum Mutat*. 2014;35(11):1321-1329.
49. Soranzo N, Spector TD, Mangino M, et al. A genome-wide meta-analysis identifies 22 loci associated with eight hematological parameters in the HaemGen consortium. *Nat Genet*. 2009;41(11):1182-1190.
50. Benyamin B, Esko T, Ried JS, et al; InterAct Consortium. Novel loci affecting iron homeostasis and their effects in individuals at risk for hemochromatosis [published correction appears in *Nat Commun*. 2015;6:6542]. *Nat Commun*. 2014;5:4926.
51. Traglia M, Girelli D, Biino G, et al. Association of HFE and TMPRSS6 genetic variants with iron and erythrocyte parameters is only in part dependent on serum hepcidin concentrations. *J Med Genet*. 2011;48(9):629-634.
52. Sørensen E, Rigas AS, Thømer LW, et al. Genetic factors influencing ferritin levels in 14,126 blood donors: results from the Danish Blood Donor Study. *Transfusion*. 2016;56(3):622-627.

53. Hudson BH, Hale AT, Irving RP, Li S, York JD. Modulation of intestinal sulfur assimilation metabolism regulates iron homeostasis. *Proc Natl Acad Sci USA*. 2018; 115(12):3000-3005.
54. Allen RP, Auerbach S, Bahrain H, Auerbach M, Earley CJ. The prevalence and impact of restless legs syndrome on patients with iron deficiency anemia. *Am J Hematol*. 2013; 88(4):261-264.
55. Connor JR, Ponnuru P, Wang XS, Patton SM, Allen RP, Earley CJ. Profile of altered brain iron acquisition in restless legs syndrome. *Brain*. 2011;134(Pt 4):959-968.
56. Auerbach M, Adamson JW. How we diagnose and treat iron deficiency anemia. *Am J Hematol*. 2016;91(1):31-38.
57. Girelli D, Nemeth E, Swinkels DW. Hepcidin in the diagnosis of iron disorders. *Blood*. 2016;127(23):2809-2813.
58. Prentice AM, Doherty CP, Abrams SA, et al. Hepcidin is the major predictor of erythrocyte iron incorporation in anemic African children. *Blood*. 2012;119(8):1922-1928.
59. Pfeiffer CM, Looker AC. Laboratory methodologies for indicators of iron status: strengths, limitations, and analytical challenges. *Am J Clin Nutr*. 2017;106(Suppl 6):1606S-1614S.
60. Archer NM, Brugnara C. Diagnosis of iron-deficient states. *Crit Rev Clin Lab Sci*. 2015; 52(5):256-272.
61. Anker SD, Comin Colet J, Filippatos G, et al; FAIR-HF Trial Investigators. Ferric carboxymaltose in patients with heart failure and iron deficiency. *N Engl J Med*. 2009;361(25):2436-2448.
62. Weiss G, Goodnough LT. Anemia of chronic disease. *N Engl J Med*. 2005;352(10):1011-1023.
63. Krayenbuehl PA, Battagay E, Breyman C, Furrer J, Schulthess G. Intravenous iron for the treatment of fatigue in nonanemic, premenopausal women with low serum ferritin concentration. *Blood*. 2011;118(12):3222-3227.
64. Camaschella C. New insights into iron deficiency and iron deficiency anemia. *Blood Rev*. 2017;31(4):225-233.
65. Houston BL, Hurrie D, Graham J, et al. Efficacy of iron supplementation on fatigue and physical capacity in non-anaemic iron-deficient adults: a systematic review of randomised controlled trials. *BMJ Open*. 2018;8(4):e019240.
66. Cook JD, Reddy MB. Efficacy of weekly compared with daily iron supplementation. *Am J Clin Nutr*. 1995;62(1):117-120.
67. Tolkien Z, Stecher L, Mander AP, Pereira DI, Powell JJ. Ferrous sulfate supplementation causes significant gastrointestinal side-effects in adults: a systematic review and meta-analysis. *PLoS One*. 2015;10(2):e0117383.
68. Moretti D, Goede JS, Zeder C, et al. Oral iron supplements increase hepcidin and decrease iron absorption from daily or twice-daily doses in iron-depleted young women. *Blood*. 2015;126(17):1981-1989.
69. Stoffel NU, Cercamondi CI, Brittenham G, et al. Iron absorption from oral iron supplements given on consecutive versus alternate days and as single morning doses versus twice-daily split dosing in iron-depleted women: two open-label, randomised controlled trials. *Lancet Haematol*. 2017;4(11):e524-e533.
70. Stoffel N, Zimmermann MB, Zeder C, Brittenham G, Moretti D. In anaemic women, daily oral iron supplementation increases hepcidin and decreases iron absorption compared to alternate day dosing. In: Proceedings from the 2018 European Iron Club Annual Meeting; 8-11 February 2018; Zurich, Switzerland. Abstract 61.
71. Schrier SL. So you know how to treat iron deficiency anemia. *Blood*. 2015;126(17):1971.
72. Zimmermann MB, Chassard C, Rohner F, et al. The effects of iron fortification on the gut microbiota in African children: a randomized controlled trial in Cote d'Ivoire. *Am J Clin Nutr*. 2010;92(6):1406-1415.
73. Jaeggi T, Kortman GA, Moretti D, et al. Iron fortification adversely affects the gut microbiome, increases pathogen abundance and induces intestinal inflammation in Kenyan infants. *Gut*. 2015;64(5):731-742.
74. World Health Organization. Use of Multiple Micronutrient Powders for Point-of-Use Fortification of Foods Consumed by Infants and Young Children Aged 6-23 Months and Children Aged 2-12 Years. Geneva, Switzerland: World Health Organization; 2016.
75. Gwamaka M, Kurtis JD, Sorensen BE, et al. Iron deficiency protects against severe *Plasmodium falciparum* malaria and death in young children. *Clin Infect Dis*. 2012;54(8):1137-1144.
76. Sazawal S, Black RE, Ramsan M, et al. Effects of routine prophylactic supplementation with iron and folic acid on admission to hospital and mortality in preschool children in a high malaria transmission setting: community-based, randomised, placebo-controlled trial. *Lancet*. 2006;367(9505):133-143.
77. Clark MA, Goheen MM, Fulford A, et al. Host iron status and iron supplementation mediate susceptibility to erythrocytic stage *Plasmodium falciparum*. *Nat Commun*. 2014;5:4446.
78. Neuberger A, Okebe J, Yahav D, Paul M. Oral iron supplements for children in malaria-endemic areas. *Cochrane Database Syst Rev*. 2016;2:CD006589.
79. Girelli D, Ugolini S, Busti F, Marchi G, Castagna A. Modern iron replacement therapy: clinical and pathophysiological insights. *Int J Hematol*. 2018;107(1):16-30.
80. Pisani A, Riccio E, Sabbatini M, Andreucci M, Del Rio A, Visciano B. Effect of oral liposomal iron versus intravenous iron for treatment of iron deficiency anaemia in CKD patients: a randomized trial. *Nephrol Dial Transplant*. 2015;30(4):645-652.
81. Isaka Y, Fujii H, Tsujimoto Y, Teramukai S, Hamano T. Rationale, design, and characteristics of a trial to evaluate the new phosphate iron-based binder sucroferri oxyhydroxide in dialysis patients with the goal of advancing the practice of E.B.M. (EPISODE). *Clin Exp Nephrol*. 2018;22(4):967-972.
82. Gasche C, Ahmad T, Tulassay Z, et al; AEGIS Study Group. Ferric maltol is effective in correcting iron deficiency anemia in patients with inflammatory bowel disease: results from a phase-3 clinical trial program. *Inflamm Bowel Dis*. 2015;21(3):579-588.
83. Grillo AS, SantaMaria AM, Kafina MD, et al. Restored iron transport by a small molecule promotes absorption and hemoglobinization in animals. *Science*. 2017;356(6338):608-616.
84. Macdougall IC, Bock AH, Carrera F, et al; FIND-CKD Study Investigators. FIND-CKD: a randomized trial of intravenous ferric carboxymaltose versus oral iron in patients with chronic kidney disease and iron deficiency anaemia. *Nephrol Dial Transplant*. 2014; 29(11):2075-2084.
85. Nielsen OH, Ainsworth M, Coskun M, Weiss G. Management of iron-deficiency anemia in inflammatory bowel disease: a systematic review. *Medicine (Baltimore)*. 2015;94(23):e963.
86. van Rheenen PF, Aloï M, Biron IA, et al. European Crohn's and Colitis Organisation topical review on transitional care in inflammatory bowel disease. *J Crohn's Colitis*. 2017;11(9):1032-1038.
87. Holm C. Intravenous iron treatment in the puerperium. *Dan Med J*. 2018;65(4):B5471.
88. Breyman C, Auerbach M. Iron deficiency in gynecology and obstetrics: clinical implications and management. *Hematology Am Soc Hematol Educ Program*. 2017;2017:152-159.
89. Auerbach M, Ballard H. Clinical use of intravenous iron: administration, efficacy, and safety. *Hematology Am Soc Hematol Educ Program*. 2010;2010:338-347.
90. Meybohm P, Froessler B, Goodnough LT, et al. "Simplified International Recommendations for the Implementation of Patient Blood Management" (SIR4PBM). *Perioper Med (Lond)*. 2017;6:5.
91. Khalafallah AA, Yan C, Al-Badri R, et al. Intravenous ferric carboxymaltose versus standard care in the management of post-operative anaemia: a prospective, open-label, randomised controlled trial. *Lancet Haematol*. 2016;3(9):e415-e425.
92. Laso-Morales M, Jericó C, Gómez-Ramírez S, et al. Preoperative management of colorectal cancer-induced iron deficiency anemia in clinical practice: data from a large observational cohort. *Transfusion*. 2017; 57(12):3040-3048.
93. Litton E, Xiao J, Ho KM. Safety and efficacy of intravenous iron therapy in reducing requirement for allogeneic blood transfusion: systematic review and meta-analysis of randomised clinical trials. *BMJ*. 2013;347:f4822.

94. Agarwal R. Iron deficiency anemia in chronic kidney disease: uncertainties and cautions. *Hemodial Int.* 2017;21(suppl 1):S78-S82.
95. Avni T, Bieber A, Grossman A, Green H, Leibovici L, Gafter-Gvili A. The safety of intravenous iron preparations: systematic review and meta-analysis. *Mayo Clin Proc.* 2015;90(1):12-23.
96. Schaefer B, Würtinger P, Finkenstedt A, et al. Choice of high-dose intravenous iron preparation determines hypophosphatemia risk. *PLoS One.* 2016;11(12):e0167146.
97. Szebeni J, Fishbane S, Hedenus M, et al. Hypersensitivity to intravenous iron: classification, terminology, mechanisms and management. *Br J Pharmacol.* 2015;172(21):5025-5036.
98. Rampton D, Folkersen J, Fishbane S, et al. Hypersensitivity reactions to intravenous iron: guidance for risk minimization and management. *Haematologica.* 2014;99(11):1671-1676.
99. Wish JB, Aronoff GR, Bacon BR, et al. Positive iron balance in chronic kidney disease: how much is too much and how to tell? *Am J Nephrol.* 2018;47(2):72-83.
100. Pagani A, Colucci S, Bocciardi R, et al. A new form of IRIDA due to combined heterozygous mutations of *TMPRSS6* and *ACVR1A* encoding the BMP receptor ALK2. *Blood.* 2017;129(25):3392-3395.