

## TRANSPLANTATION

# Unrelated donor vs HLA-haploidentical $\alpha/\beta$ T-cell- and B-cell-depleted HSCT in children with acute leukemia

Alice Bertaina,<sup>1,\*</sup> Marco Zecca,<sup>2,\*</sup> Barbara Buldini,<sup>3</sup> Nicoletta Sacchi,<sup>4</sup> Mattia Algeri,<sup>1</sup> Francesco Saglio,<sup>5</sup> Cesare Perotti,<sup>6</sup> Anna Maria Gallina,<sup>4</sup> Valentina Bertaina,<sup>1</sup> Edoardo Lanino,<sup>7</sup> Arcangelo Prete,<sup>8</sup> Walter Barberi,<sup>9</sup> Manuela Tumino,<sup>3</sup> Claudio Favre,<sup>10</sup> Simone Cesaro,<sup>11</sup> Francesca Del Bufalo,<sup>1</sup> Mimmo Ripaldi,<sup>12</sup> Stella Boghen,<sup>2,13</sup> Gabriella Casazza,<sup>14</sup> Marco Rabusin,<sup>15</sup> Adriana Balduzzi,<sup>16</sup> Franca Fagioli,<sup>5</sup> Daria Pagliara,<sup>1,†</sup> and Franco Locatelli<sup>1,17,†</sup>

<sup>1</sup>Department of Pediatric Hematology/Oncology and Cell and Gene Therapy, Istituto di Ricovero e Cura a Carattere Scientifico (IRCCS), Ospedale Bambino Gesù, Rome, Italy; <sup>2</sup>Pediatric Hematology/Oncology, Fondazione IRCCS Policlinico San Matteo, Pavia, Italy; <sup>3</sup>Pediatric Hematology-Oncology Unit, Department of Women's and Children's Health, Azienda Ospedaliera-University of Padova, Padova, Italy; <sup>4</sup>Struttura Complessa Laboratorio di Istocompatibilità/Italian Registry of Bone Marrow Donors, Ente Ospedaliero Ospedali Galliera, Genoa, Italy; <sup>5</sup>Pediatric Onco-Hematology, Stem Cell Transplantation and Cellular Therapy Division, Regina Margherita Children's Hospital, Torino, Italy; <sup>6</sup>Immunohaematology and Transfusion Service, Fondazione IRCCS Policlinico S. Matteo, Pavia, Italy; <sup>7</sup>Hematopoietic Stem Cell Unit, Department of Hematology-Oncology, Istituto G. Gaslini, Genova, Italy; <sup>8</sup>Oncology, Hematology and Hematopoietic Stem Cell Transplant Program, Unità operativa Pediatrics- S. Orsola-Malpighi University of Bologna, Bologna, Italy; <sup>9</sup>Department of Hematology, Azienda Policlinico Umberto I, Sapienza University of Rome, Rome, Italy; <sup>10</sup>Transplantation Unit, Department of Pediatric Oncology, Meyer Children's Hospital, Florence, Italy; <sup>11</sup>Pediatric Hematology Oncology, Azienda Ospedaliera Universitaria Integrata, Verona, Italy; <sup>12</sup>Department of Hemato-Oncology, Santobono-Pausilipon Hospital, Bone Marrow Transplant Unit, Napoli, Italy; <sup>13</sup>Cell Factory, Fondazione IRCCS Policlinico San Matteo, Pavia, Italy; <sup>14</sup>Pediatric Onco-Hematology, Azienda Ospedaliera/Universitaria Pisana, Pisa, Italy; <sup>15</sup>Department of Pediatrics, Institute of Maternal and Child Health, IRCCS Burlo Garofolo Trieste, Trieste, Italy; <sup>16</sup>Bone Marrow Transplantation Unit, Pediatric Department of Milano-Bicocca University, Monza e Brianza per il Bambino e la sua Mamma Foundation, Monza, Italy; and <sup>17</sup>Department of Pediatric Sciences, University of Pavia, Pavia, Italy

## KEY POINTS

- Children with acute leukemia given  $\alpha\beta$ haplo-HSCT have a lower risk for acute and chronic GVHD than those transplanted from an unrelated donor.
- GVHD-free, relapse-free survival after  $\alpha\beta$ haplo-HSCT is better than that observed in unrelated donor HSCT recipients.

**Traditionally, hematopoietic stem cell transplantation (HSCT) from both HLA-matched related and unrelated donors (UD) has been used for treating children with acute leukemia (AL) in need of an allograft. Recently, HLA-haploidentical HSCT after  $\alpha\beta$  T-cell/B-cell depletion ( $\alpha\beta$ haplo-HSCT) was shown to be effective in single-center studies. Here, we report the first multicenter retrospective analysis of 127 matched UD (MUD), 118 mismatched UD (MMUD), and 98  $\alpha\beta$ haplo-HSCT recipients, transplanted between 2010 and 2015, in 13 Italian centers. All these AL children were transplanted in morphological remission after a myeloablative conditioning regimen. Graft failure occurred in 2% each of UD-HSCT and  $\alpha\beta$ haplo-HSCT groups. In MUD vs MMUD-HSCT recipients, the cumulative incidence of grade II to IV and grade III to IV acute graft-versus-host disease (GVHD) was 35% vs 44% and 6% vs 18%, respectively, compared with 16% and 0% in  $\alpha\beta$ haplo-HSCT recipients ( $P < .001$ ). Children treated with  $\alpha\beta$ haplo-HSCT also had a significantly lower incidence of overall and extensive chronic GVHD ( $P < .01$ ). Eight (6%) MUD, 32 (28%) MMUD, and 9 (9%)  $\alpha\beta$ haplo-HSCT patients died of transplant-related complications. With a median follow-up of 3.3 years, the 5-year probability of leukemia-free survival in the 3 groups was 67%, 55%, and 62%, respectively. In the 3 groups, chronic GVHD-free/relapse-free (GRFS) probability of survival was 61%, 34%, and 58%, respectively ( $P < .001$ ). When compared with patients given MMUD-HSCT,  $\alpha\beta$ haplo-HSCT recipients had a lower cumulative incidence of nonrelapse mortality and a better GRFS ( $P < .001$ ). These data indicate that  $\alpha\beta$ haplo-HSCT is a suitable therapeutic option for children with AL in need of transplantation, especially when an allele-matched UD is not available. (*Blood*. 2018;132(24):2594-2607)**

free (GRFS) probability of survival was 61%, 34%, and 58%, respectively ( $P < .001$ ). When compared with patients given MMUD-HSCT,  $\alpha\beta$ haplo-HSCT recipients had a lower cumulative incidence of nonrelapse mortality and a better GRFS ( $P < .001$ ). These data indicate that  $\alpha\beta$ haplo-HSCT is a suitable therapeutic option for children with AL in need of transplantation, especially when an allele-matched UD is not available. (*Blood*. 2018;132(24):2594-2607)

## Introduction

During the last 4 decades, allogeneic hematopoietic stem cell transplantation (HSCT) from an HLA-matched donor, either related or unrelated (UD), has been extensively used to treat patients with both malignant and nonmalignant disorders.<sup>1</sup> However, only 25% of patients who are candidates to receive allogeneic HSCT have an HLA-identical sibling, and a suitable UD can be identified for less than 60% of the remaining patients.<sup>2</sup> The likelihood of finding an optimal donor varies

among racial and ethnic groups, with the probability of identifying an appropriate donor being highest among whites of European descent (75%) and lowest among blacks of South or Central American descent (16%).<sup>3</sup> In the absence of an HLA-matched donor, alternative donor/sources of HSCs, such as unrelated umbilical cord blood and HLA-haploidentical relatives, are being increasingly used.<sup>2,4,5</sup> The majority of patients have a family member with 1 identical HLA-haplotype and the other mismatched (ie, haploidentical).<sup>6</sup> Although mature donor

T cells present in the graft facilitate T-cell reconstitution, they are also responsible for the occurrence of graft-versus-host disease (GVHD).<sup>7,8</sup> Different strategies, based on either pharmacological immunosuppression or T-cell depletion of the graft, have been developed to prevent GVHD after HLA-haploidentical HSCT.<sup>9-12</sup> Pioneering studies in adults have demonstrated that the infusion of megadoses of purified CD34<sup>+</sup> cells can prevent both graft rejection and severe GVHD in haplo-HSCT recipients.<sup>13,14</sup> However, extensive lymphoid cell depletion has resulted in an increased risk for opportunistic infections, especially in the first months after HSCT. In the attempt to reduce the risk for infections, recently, a new strategy of graft manipulation has been implemented, based on the selective elimination of  $\alpha\beta$  T cells and CD19<sup>+</sup> B cells ( $\alpha\beta$ haplo-HSCT).<sup>15</sup> This refined technique of graft engineering reduces the problems associated with delayed immune recovery, which is typical in the CD34<sup>+</sup> cell selection haplo-HSCT approach. Indeed, using  $\alpha\beta$ haplo-HSCT, it is possible to transfer to the recipient not only donor HSCs but also committed hematopoietic progenitors, as well as mature natural killer (NK) and  $\gamma\delta$  T cells.<sup>16,17</sup> These lymphocyte subsets may provide a protective effect against both leukemia relapse and severe infections.<sup>18-21</sup> Using  $\alpha\beta$ haplo-HSCT, we reported promising clinical results in children with life-threatening non-malignant disorders.<sup>22</sup> More recently, single-center experiences in pediatric patients with malignancies have been published, showing the risks for nonrelapse mortality (NRM) and leukemia relapse being comparable to those from HLA-identical siblings or UD-HSCT.<sup>23-25</sup> Moreover, patients receiving  $\alpha\beta$ haplo-HSCT had a lower risk for both acute and chronic GVHD, leading to better GVHD-free/relapse-free survival (GRFS).<sup>25</sup>

Here, we report the first multicenter, retrospective comparative analysis on the outcome of children with acute leukemia (AL) given either UD-HSCT or  $\alpha\beta$ haplo-HSCT. Using data reported by 13 centers affiliated with the Associazione Italiana di Oncoematologia Pediatrica-HSCT network, we evaluated children undergoing a first HSCT between October 2010 and December 2015. These data, obtained in a multicenter setting, show that the outcome of children given  $\alpha\beta$ haplo-HSCT is equivalent to that of patients transplanted from an 8/8 UD-HSCT (MUD), whereas it is superior to that of patients given an allograft from a HLA-mismatched UD-HSCT (7/8 or 6/8, mismatched MUD [MMUD]).

## Patients and methods

This is a retrospective, registry-based analysis conducted on behalf of the Associazione Italiana di Oncoematologia Pediatrica-HSCT network. Table 1 summarizes main patient, donor, and transplant characteristics. Six centers performed both UD-HSCT and  $\alpha\beta$ haplo-HSCT, whereas in the remaining 7 centers, only the former procedure was performed. Seventy of the patients enrolled in the  $\alpha\beta$ haplo-HSCT group have been already published in a recent single-center experience.<sup>25</sup> All consecutive patients younger than 21 years with either acute lymphoblastic leukemia (ALL) or acute myeloid leukemia (AML) who received a first allograft in morphological complete remission after a myeloablative conditioning from an UD or  $\alpha\beta$ haplo-HSCT between October 2010 and December 2015 were enrolled. Written informed consent was obtained from either the patient or parents/legal guardians in accordance with the Helsinki Declaration, and the study was approved by the local Ethical Committee. The UD was selected using high-resolution typing for the HLA-loci A, B,

C, and DRB1. A MUD was employed in 52% of cases; the remaining 48% of the patients were transplanted from a donor with either 1 or 2 HLA disparities (MMUD). Centers performing  $\alpha\beta$ haplo-HSCT offered this approach to children lacking either an HLA-identical sibling or a MUD, or in need of an urgent procedure, according to physician's judgment. Total body irradiation (TBI) was used in patients older than 3 years affected by either ALL or very high risk AML (ie, those with cytogenetic/molecular features predicting high risk for relapse; see Table 1 for further details). Anti-T-lymphocyte globulin (ATLG [Grafalon; Neovii Biotech]) or antithymocyte globulin [Thymoglobulin; Sanofi-Genzyme] was administered to all patients for preventing both graft rejection and GVHD. Moreover, in the  $\alpha\beta$ haplo-HSCT group, for further lowering the risk for Epstein-Barr virus (EBV)-related post-transplant lymphoproliferative disorder (PTLD), on day  $-1$ , patients were given rituximab (200 mg/m<sup>2</sup>) for in vivo depletion of both donor and recipient B cells. No patient in the  $\alpha\beta$ haplo-HSCT group received posttransplant pharmacological GVHD prophylaxis, whereas the post-HSCT combination of calcineurin inhibitor and short-term methotrexate was employed for preventing GVHD occurrence in UD-HSCT recipients (see Table 1).

Minimal residual disease (MRD) evaluated by multicolor flow cytometry within 30 days before HSCT was available in 163 patients (47.8% of the overall cohort). Among these patients, it was less than  $1 \times 10^{-3}$  in 143 (88%) and more than  $1 \times 10^{-3}$  in 20 (12%) cases.

Within the  $\alpha\beta$ haplo-HSCT group, the donor was the mother for 58 patients (59%) and the father for the remaining 40 patients (41%). The HLA-haploidentical donor was selected according to immunological criteria, giving priority to NK alloreactivity (evaluated through the killer immunoglobulin-like receptor [KIR]/ligand model),<sup>26,27</sup> KIR B haplotype,<sup>28</sup> higher B-content score,<sup>29</sup> and size of NK alloreactive subset.<sup>30-32</sup> An NK alloreactive donor was employed in 41 (43%) of 96 evaluable donor/recipient pairs.

$\alpha\beta$ haplo-HSCT donors received granulocyte-colony stimulating factor for 4 days at the total dose of 12  $\mu$ g/kg body weight, and apheresis was performed on the fifth day. When on day 4 the CD34<sup>+</sup> cell count was less than 40/ $\mu$ L and/or the predicted apheresis yields was less than  $12.0 \times 10^6$  CD34<sup>+</sup> HSCs/kg recipient's body weight, a CXCR4 antagonist (Plerixafor, Mozobil) was given for boosting mobilization of HSCs/progenitor cells.<sup>33,34</sup> In all centers, manipulations were performed in a closed system. Clinical-grade reagents, disposable kits, and instrumentation were from Miltenyi Biotec (Bergisch Gladbach, Germany).

In the UD-HSCT group, the median number of total nucleated cells infused was  $5.6 \times 10^8$ /kg, whereas the median number of CD34<sup>+</sup> infused was  $5 \times 10^6$ /kg. Children enrolled in the  $\alpha\beta$ haplo-HSCT cohort received a median number of  $14.4 \times 10^6$ /kg CD34<sup>+</sup> cells. In this latter case, the  $\alpha\beta$  T-cell depletion was homogeneously very efficient, the median number of residual  $\alpha\beta$  T cells infused into the graft being  $0.04 \times 10^6$ /kg. As previously reported, a large number of NK and  $\gamma\delta$  T cells was retained into the graft (see Table 1 for further details).<sup>34</sup>

Chimerism analysis evaluated through the Short Tandem Repeats polymorphism was performed on cells obtained from bone marrow aspirates whenever performed, and on peripheral mononuclear cells, weekly for the first 3 months and monthly thereafter.

**Table 1. Patient, donor, and transplant characteristics**

	<b>MUD</b>	<b>MMUD</b>	<b>Haploidentical</b>	<b>P</b>
Number of patients	127	118	98	
<b>Sex</b>				
Male	74 (58%)	71 (60%)	65 (66%)	.450
Female	53 (42%)	47 (40%)	33 (34%)	
Median age at diagnosis (range), y	5.7 (0.2-17.4)	8.8 (0.3-17.5)	6.6 (0.1-17.3)	.113
<b>Diagnosis</b>				
ALL	84 (66%)	86 (73%)	68 (68%)	.513
AML	43 (34%)	32 (27%)	30 (32%)	
<b>ALL phenotype</b>				
B-cell precursor	71 (85%)	67 (78%)	53 (78%)	.474
T-cell precursor	13 (15%)	19 (22%)	15 (22%)	
<b>AML FAB classification</b>				
AML M0	1 (2%)	2 (6%)	3 (10%)	.623
AML M1	7 (17%)	6 (19%)	4 (13%)	
AML M2	13 (30%)	4 (13%)	6 (20%)	
AML M3	1 (2%)	1 (3%)	1 (3%)	
AML M4	5 (12%)	4 (13%)	5 (17%)	
AML M5	12 (28%)	13 (40%)	8 (27%)	
AML M6	3 (7%)	1 (3%)	0 (0%)	
AML M7	1 (2%)	1 (3%)	3 (10%)	
<b>Disease phase at HSCT</b>				
ALL				
CR1	35 (42%)	39 (45%)	19 (28%)	<.001
CR2 S1-S2	35 (42%)	28 (33%)	22 (32%)	
CR2 S3-S4	14 (16%)	18 (21%)	17 (25%)	
Other CR	0	1 (1%)	10 (15%)	
AML				
CR1	33 (77%)	26 (81%)	23 (77%)	.803
CR2	9 (21%)	6 (19%)	7 (23%)	
Other CR	1 (2%)	0 (0%)	0 (0%)	
Median age at HSCT (range), y	5.7 (1.0-18.0)	10.3 (0.4-18.1)	9.4 (0.9-18.2)	.106
<b>Stem cell source</b>				
BM	97 (76%)	90 (76%)	0 (0%)	<.001
PBSC	30 (24%)	28 (24%)	98 (100%)	
<b>Conditioning regimen</b>				
TBI-based	64 (50%)	68 (58%)	72 (74%)	.016
Busulfan-based	54 (43%)	43 (36%)	18 (18%)	
Treosulfan-based	8 (6%)	6 (5%)	7 (7%)	
Other chemotherapy	1 (1%)	1(1%)	1 (1%)	
<b>GVHD prophylaxis</b>				
Cs-A+MTX	2 (2%)	2 (2%)	0 (0%)	<.001
Cs-A+MTX+ATLG	112 (89%)	97 (82%)	0 (0%)	
Cs-A+MTX+ATLG+MMF	0 (0%)	6 (5%)	0 (0%)	
Tacrolimus+MTX+ATLG	12 (9%)	13 (11%)	0 (0%)	
αβ <sup>+</sup> and CD19 <sup>+</sup> negative selection + ATLG	0 (0%)	0 (0%)	98 (100%)	

Data are expressed as median and range or as absolute number and column percentage, as appropriate. Indications for hematopoietic stem cell transplantation in CR1 of patients with ALL were high level of minimal residual disease at end of induction therapy (ie,  $>1 \times 10^{-3}$  at day +78 after beginning of treatment), high-risk infant ALL, t(4;11), hypodiploidy ( $\leq 43$  chromosomes), and hyperleukocytosis T ALL with poor response to the steroid prephase. Indications for hematopoietic stem cell transplantation in CR1 of patients with AML were t(10;11), t(6;11), t(6;9), t(5;11), complex karyotype ( $\geq 3$  either numeric or structural aberrations), FLT3-ITD with high allelic ratio, M7 AML, CBFA2T3-GLIS2 fusion transcript, and not being in morphological CR after the first of the 2 induction courses.

BM, bone marrow; CR, complete remission; CsA, cyclosporine-A; MMF, mycophenolate mofetil; MTX, methotrexate; PBSC, peripheral blood stem cells.

**Table 1. (continued)**

	MUD	MMUD	Haploidentical	P
<b>Cell dose infused</b>				
Total nucleated cell dose (range), $\times 10^9/\text{kg}$	5.5 (1.7-48)	5.4 (2.2-43)	10.2 (3-52)	<.001
CD34 <sup>+</sup> cells, $\times 10^6/\text{kg}$	5.2 (1.0-40)	4.9 (2.1-39)	14.4 (5.5-56)	<.001
CD3 <sup>+</sup> $\alpha\beta^+$ T lymphocytes, $\times 10^6/\text{kg}$	—	—	0.04 (0.00-0.99)	—
CD3 <sup>+</sup> $\gamma\delta^+$ T lymphocytes, $\times 10^6/\text{kg}$	—	—	8.1 (1.0-64.6)	—

Data are expressed as median and range or as absolute number and column percentage, as appropriate. Indications for hematopoietic stem cell transplantation in CR1 of patients with ALL were high level of minimal residual disease at end of induction therapy (ie,  $>1 \times 10^{-3}$  at day +78 after beginning of treatment), high-risk infant ALL, t(4;11), hypodiploidy ( $\leq 43$  chromosomes), and hyperleukocytosis T ALL with poor response to the steroid prephase. Indications for hematopoietic stem cell transplantation in CR1 of patients with AML were t(10;11), t(6;11), t(6;9), t(5;11), complex karyotype ( $\geq 3$  either numeric or structural aberrations), FLT3-ITD with high allelic ratio, M7 AML, CBFA2T3-GLIS2 fusion transcript, and not being in morphological CR after the first of the 2 induction courses.

BM, bone marrow; CR, complete remission; CsA, cyclosporine-A; MMF, mycophenolate mofetil; MTX, methotrexate; PBSC, peripheral blood stem cells.

### Definitions and statistical analysis

Graft failure was defined as either lack of initial engraftment of donor cells or loss of donor cells after initial engraftment. Time to neutrophil engraftment was defined as time from HSCT to the first of 3 consecutive days with an absolute neutrophil count of at least  $0.5 \times 10^9/\text{L}$ , whereas time to platelet engraftment was defined as time from HSCT to the first of 7 consecutive days with an unsupported platelet count of at least  $50 \times 10^9/\text{L}$ .

Results are reported in the 2 UD and  $\alpha\beta$ haplo groups. Moreover, the effect of the type of donor on the clinical outcome are assessed in 3 different groups: 8/8 UD-HSCT (MUD), 6/8 or 7/8 UD-HSCT (MMUD), and  $\alpha\beta$ haplo-HSCT (supplemental Table 1, available on the *Blood* Web site).

Patients surviving more than 14 and 100 days after HSCT were evaluated for acute and chronic GVHD, respectively, which were diagnosed and graded according to previously published criteria.<sup>35,36</sup> NRM was defined as the probability of death from any cause other than recurrence of leukemia. Overall survival (OS) was defined as the probability of being alive at last follow-up, whereas leukemia-free survival (LFS) was defined as the probability of survival, without evidence of disease at any time after HSCT. In estimating LFS, death and relapse were considered events, whereas patients who were alive, with sustained donor engraftment and disease-free, were censored at last follow-up. We also evaluated the composite end point of chronic GRFS.<sup>37</sup>

Data on patients transplanted from either an HLA-haploidentical relative or a UD were collected in the data warehouse of the Associazione Italiana di Oncoematologia Pediatrica-HSCT group. Both primary and secondary end points were assessed by the statistician of the group (M.Z.). Data were analyzed as of 31 December 2017. Quantitative variables were reported as median value and range, whereas categorical variables were expressed as absolute value and percentage. Demographic and clinical characteristics of the patients were compared using the  $\chi$ -square test or Fisher's exact test for categorical variables, whereas the Mann-Whitney rank sum test or the Student t-test were used for continuous variables, whenever appropriate. Acute and chronic GVHD, rejection, engraftment, OS, LFS, NRM, and relapse incidence were estimated from the date of HSCT to the date of an event or last follow-up. Probabilities of LFS and OS were calculated according to the Kaplan and Meier method.<sup>38</sup> Acute GVHD and chronic GVHD, NRM, and relapse

were calculated as cumulative incidence curves to adjust the estimates for competing risks. All results were expressed as either probability or cumulative incidence (%) with 95% confidence interval (CI).<sup>39,40</sup>

The significance of differences between LFS and OS was estimated by the log-rank test (Mantel-Cox), whereas Gray's test was used to assess, in univariable analyses, differences between cumulative incidences.<sup>41</sup>

Multivariable analysis was performed using the Cox proportional hazard regression model or the method of Fine and Gray, as appropriate.<sup>39,40</sup> All variables significant in univariable analysis were included in the multivariable model.

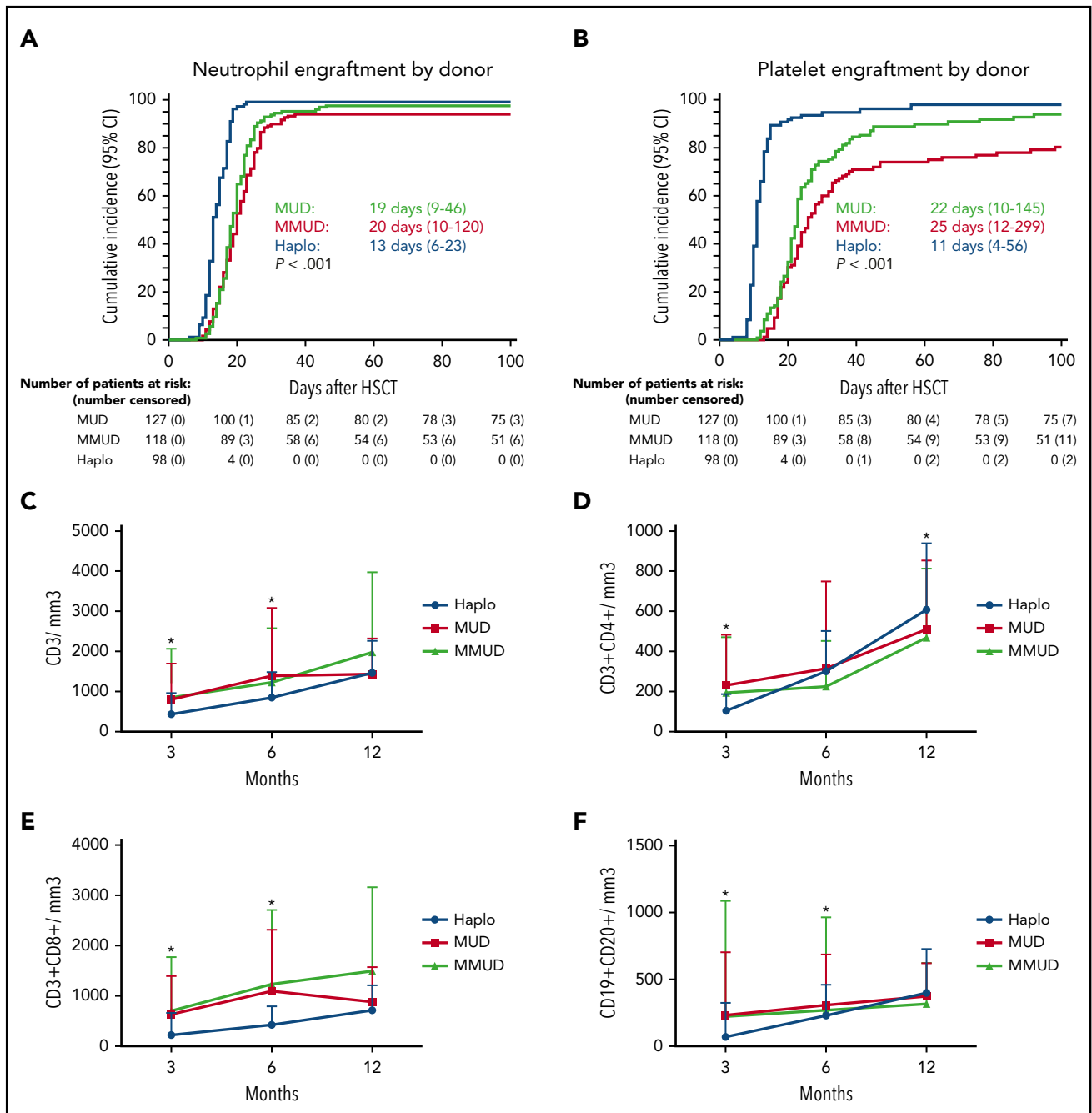
P values  $<.05$  were considered statistically significant. Statistical analyses were performed by means of NCSS (NCSS 10 Statistical Software, <https://www.ncss.com/software/ncss/>) and Stata MP/15 (StataCorp LP, <http://www.stata.com/>).

## Results

### Engraftment and immune recovery

In the entire cohort (343 patients), 6 patients did not engraft: 2 each in the MUD (1.6%), MMUD (1.7%), and  $\alpha\beta$ haplo-HSCT (2%) groups. All children experiencing primary graft failure in the UD-HSCT group had received less than  $4 \times 10^6$  CD34<sup>+</sup>/kg, and 2 of them received less than  $2 \times 10^6$  CD34<sup>+</sup>/kg. Three of these patients died after rejecting the graft because of infectious complications, and the remaining patient relapsed. No correlation between graft composition and graft failure was observed in the  $\alpha\beta$ haplo-HSCT cohort. One of the 2 patients who experienced graft failure in this latter group was successfully retransplanted using the other parent as a donor. The second patient died of a disseminated adenovirus infection. Patients given  $\alpha\beta$ haplo-HSCT had a faster neutrophil and platelet recovery than those transplanted from a MUD or a MMUD ( $P < .001$ ; Figure 1A-B, respectively). Moreover, MMUD-HSCT recipients had a lower probability of platelet recovery at 100 days after the allograft in comparison with patients receiving either  $\alpha\beta$ haplo or MUD-HSCT (Figure 1B). Notably, in the  $\alpha\beta$ haplo-HSCT group, no patient received posttransplant granulocyte-colony stimulating factor to accelerate neutrophil recovery.

Data on immune reconstitution were available in 94 (96%), 95 (75%), and 90 (76%)  $\alpha\beta$ haplo, MUD, and MMUD-HSCT recipients.



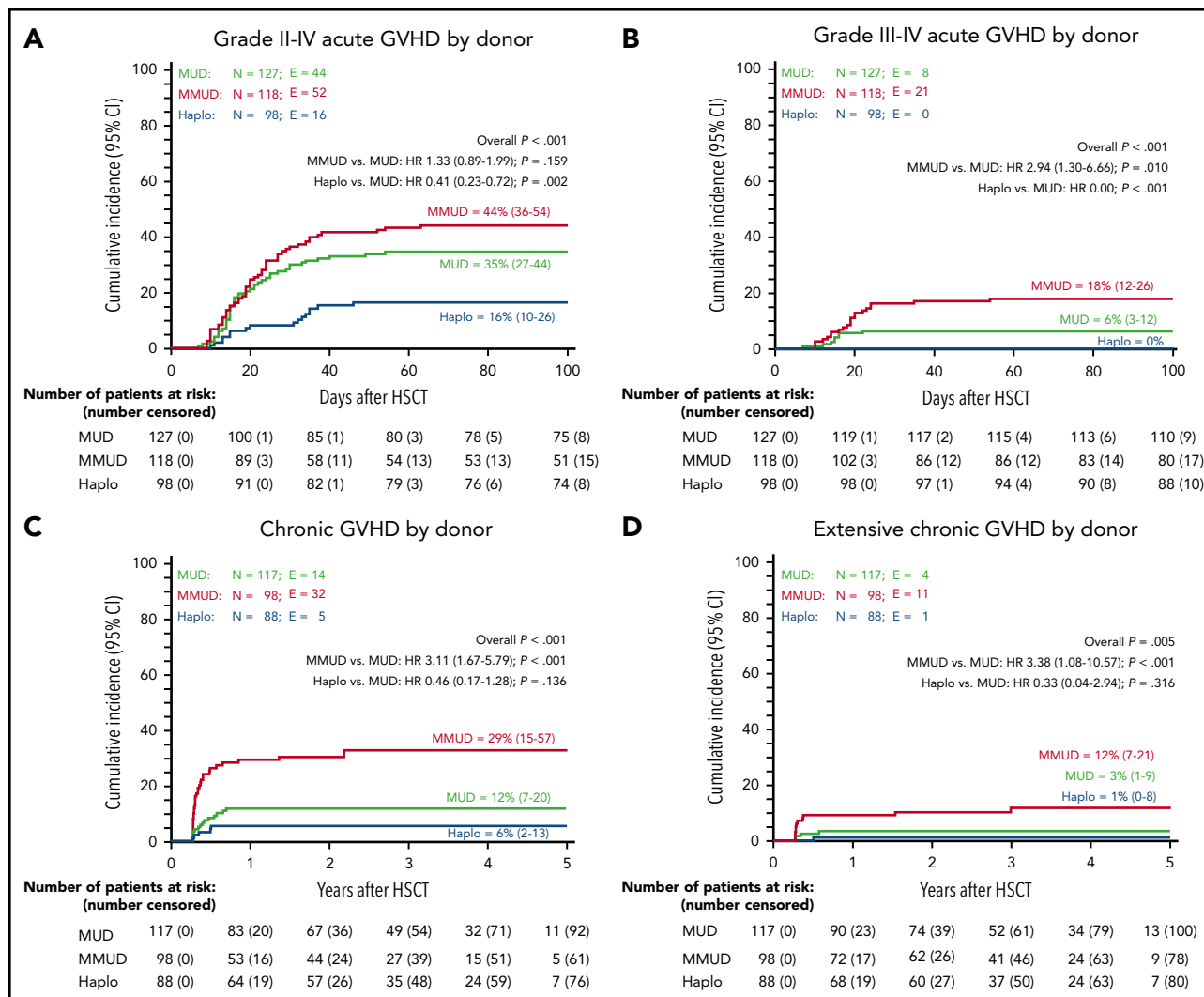
**Figure 1. Engraftment's kinetic and immune recovery in  $\alpha\beta$ haplo-HSCT, MUD-HSCT, and MMUD-HSCT recipients.** Cumulative incidence of neutrophil (A) and platelet engraftment (B), recovery of CD3<sup>+</sup> cells (C), CD3<sup>+</sup>/CD4<sup>+</sup> cells (D), CD3<sup>+</sup>/CD8<sup>+</sup> cells (E), and B cells (F), according to the type of donor employed.

Recovery of CD3<sup>+</sup> and of CD3<sup>+</sup>/CD8<sup>+</sup> cells was faster in patients given MUD and  $\beta$ haplo-HSCT in comparison with that of patients given  $\alpha\beta$ haplo-HSCT until 6 months after the allograft (Figure 1C,E, respectively). Reconstitution of CD3<sup>+</sup>/CD4<sup>+</sup> was better in the MUD and MMUD groups only at 3 months, whereas at 12 months it was better in  $\alpha\beta$ haplo-HSCT than in the other 2 groups. B-cell recovery mimicked that of CD3<sup>+</sup> and CD3<sup>+</sup>/CD8<sup>+</sup> cells (Figure 1F).

### Acute and chronic GVHD

The cumulative incidence of grade II to IV and grade III to IV acute GVHD in patients given MUD vs MMUD-HSCT was 35% vs

44% and 6% vs 18%, respectively, as compared with 16% and 0% in  $\alpha\beta$ haplo-HSCT recipients ( $P < .001$ ; Figure 2A-B). Remarkably, no patients enrolled in this latter group developed visceral involvement of acute GVHD. Children treated with  $\alpha\beta$ haplo-HSCT benefited also from a lower incidence of both overall and extensive chronic GVHD. Indeed, although the cumulative incidence of overall and extensive chronic GVHD in patients given MUD vs MMUD-HSCT was 12% vs 29% and 3% vs 12%, respectively, in recipients of  $\alpha\beta$ haplo-HSCT, it was 6% and 1% ( $P < .001$  and  $P < .005$ , respectively; Figure 2C-D; see supplemental Table 1 for more details). The only patient experiencing extensive chronic GVHD in the  $\alpha\beta$ haplo-HSCT



**Figure 2. Cumulative incidence of acute and chronic GVHD in  $\alpha\beta$ haplo-HSCT, MUD-HSCT, and MMUD-HSCT recipients.** Cumulative incidence of grade II to IV acute GVHD (A), grade III to IV acute GVHD (B), chronic GVHD (C), and extensive chronic GVHD (D), according to the type of donor employed. All results were expressed as either probability or cumulative incidence (%) with 95% CI.

group received a total number of  $\alpha\beta^+$  T cells equal to  $0.025 \times 10^6/\text{kg}$ .

### Nonrelapse mortality and relapse

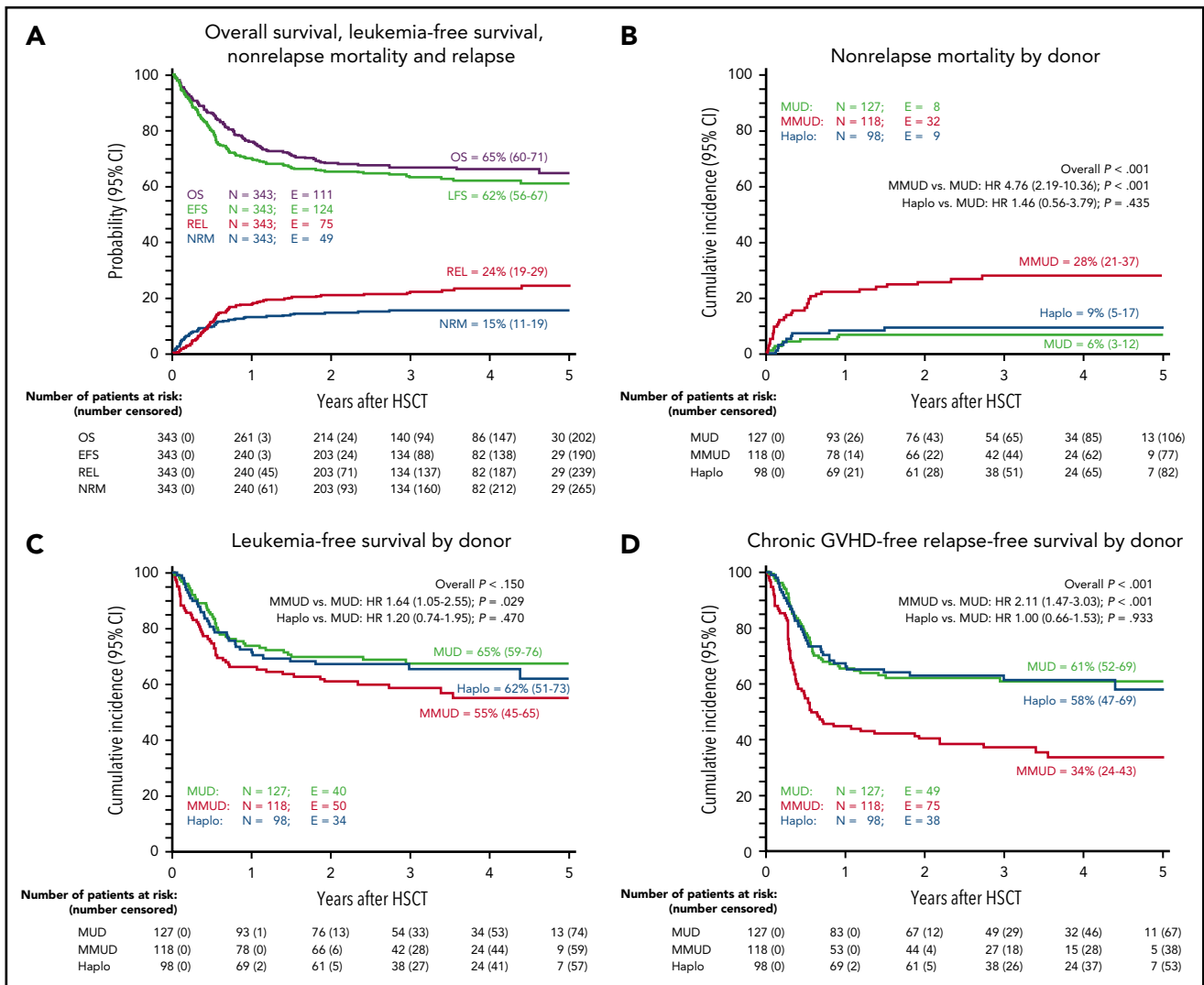
Forty-nine patients died of transplant-related complications (Figure 3A; Table 2): 40 in the UD-HSCT group (17%) and 9 who had received  $\alpha\beta$ haplo-HSCT (9%). Although the probability of NRM was superimposable in MUD-HSCT (6%) and  $\alpha\beta$ haplo-HSCT (9%), MMUD-HSCT was associated with a higher risk for mortality (28%;  $P < .001$ ; Figure 3B), as confirmed in the multivariable analysis (hazard ratio [HR], 4.35; 95% CI, 1.97-9.60;  $P < .001$ ), after adjusting for age, which was also associated with NRM (age,  $>13$  years vs  $<13$  years: HR, 1.88; 95% CI, 1.05-3.35;  $P = .033$ ; Table 3). Overall, infection was the main cause of nonleukemia death, occurring in 4 (12%), 15 (33%), and 5 (16%) MUD, MMUD, and  $\alpha\beta$ haplo-HSCT recipients, respectively (supplemental Table 2). Although bacterial infections were significantly higher in MMUD recipients (34%) in comparison with MUD (17%) and  $\alpha\beta$ haplo-HSCT (8%), the incidence of fungal and viral infections was superimposable

( $P = \text{N.S.}$  (not significant); supplemental Table 3). Remarkably, none of the patients who received  $\alpha\beta$ haplo-HSCT developed EBV-related PTLD in comparison with 2 UD-HSCT recipients (1 each in the MUD and MMUD subgroup).

Seventy-five (24%) children relapsed at a median time of 190 days (range, 40-1603 days) after the allograft (Figure 3A). No statistically significant difference for the cumulative incidence of disease recurrence was observed among MUD-HSCT, MMUD-HSCT, and  $\alpha\beta$ haplo-HSCT recipients (26% vs 17% vs 29%, respectively; see supplemental Table 4 for more details). A poor disease risk (HR, 3.43; 95% CI, 1.69-6.94;  $P = .001$ ) and a high pre-HSCT MRD level (HR, 2.97; 95% CI, 1.32-6.67;  $P = .008$ ) were associated with increased risk for relapse in the multivariable analysis, as detailed in Table 3.

### Survival, LFS, and GFRS

With a median follow-up of 3.3 years (range, 1.5-7 years for surviving patients), the 5-year probability of OS for patients given either UD-HSCT or  $\alpha\beta$ haplo-HSCT is 64% and 68%,



**Figure 3. Probability of OS, LFS, NRM, relapse incidence (REL), and chronic GVHD-free, relapse-free survival (GFRS).** OS, LFS, NRM, and REL of the overall cohort of patients included in the study (A). Cumulative incidence of NRM according to the type of donor employed (B). Five-year probability of LFS according to the type of donor employed (C). Five-year probability of GFRS according to the type of donor employed (D). All results were expressed as either probability or cumulative incidence (%) with 95% CI.

respectively ( $P = \text{N.S.}$ ; supplemental Table 1). The 5-year probability of LFS is 65%, 55%, and 62% in MUD, MMUD, and  $\alpha\beta$ haplo-HSCT, respectively ( $P = \text{N.S.}$ ; Figure 3C). In univariable analysis, an advanced disease phase at HSCT, MRD level higher than  $1 \times 10^{-3}$  30 days before HSCT and a Treosulfan-based conditioning regimen in ALL were associated with poorer outcome ( $P < .0001$ ,  $P = .003$ , and  $P = .004$ , respectively; see Table 4 for details). In multivariable analysis, the disease risk stratification (HR, 3.43; 95% CI, 1.69-6.94;  $P = .001$ ) and MRD level before HSCT remained significant (HR, 2.97; 95% CI, 1.32-6.67;  $P = .008$ , Table 3). In the  $\alpha\beta$ haplo-HSCT recipients, the 5-year LFS of patients transplanted from either an NK-alloreactive or nonalloreactive donor was superimposable (60% vs 64%, respectively;  $P = \text{N.S.}$ ). The 5-year chronic GFRS in the UD-HSCT and  $\alpha\beta$ haplo-HSCT groups was 48% and 59%, respectively ( $P = .03$ ; supplemental Table 1). Notably, chronic GFRS of MUD-HSCT and  $\alpha\beta$ haplo-HSCT recipients was superimposable (61% and 58%, respectively), whereas the choice of MMUD-HSCT had a detrimental effect on this composite end point (34%;  $P < .001$ ; Figure 3D).

## Discussion

This study represents the first multicenter, retrospective comparative analysis on AL children receiving either UD-HSCT or  $\alpha\beta$ haplo-HSCT. Indeed, no comparative data are available for pediatric patients, although, in adults, some studies have compared the outcome of UD-HSCT with that of unmanipulated HLA-haploidentical HSCT.<sup>42-44</sup>

Recently, several single-center reports demonstrated the safety and efficacy of the  $\alpha\beta$  T-cell/CD19<sup>+</sup> B-cell depletion as a graft-engineering approach in children receiving HLA-haploidentical HSCT.<sup>23-25</sup> The present analysis definitely establishes on a large/multisite scale that  $\alpha\beta$ haplo-HSCT represents an equally effective option to MUD-HSCT for children with AL lacking a sibling donor.

As reported in Table 1, the majority of patients enrolled in our study had ALL (59%). Remarkably,  $\alpha\beta$ haplo-HSCT recipients were transplanted in a more advanced disease phase and more

**Table 2. Univariate analysis of nonrelapse mortality in the cohort of patients analyzed**

	Number of patients	Events	Cumulative incidence	95% CI	P	HR*	95% CI	P
All patients	343	49	15%	11-19				
<b>Sex</b>								
Male	210	25	12%	8-17	.116			
Female	133	24	18%	13-27		1.57	0.90-2.74	.116
<b>Age at HSCT, y</b>								
<9.5	172	15	9%	6-15	.003			
≥9.5	171	34	20%	15-27		2.42	1.32-4.45	.004
<6	94	10	11%	6-20	.013			
6-9.5	78	5	6%	3-15		0.59	0.20-1.72	.334
9.5-13	80	13	16%	10-27		1.59	0.69-3.65	.274
≥13	91	21	24%	16-34		2.30	1.09-4.87	.029
<13	252	28	13%	8-16	.006			
≥13	91	21	24%	16-34		2.95	1.25-3.84	.006
<b>Diagnosis</b>								
ALL	237	37	16%	12-21	.281			
AML	106	12	12%	7-20		0.79	0.37-1.34	.280
<b>ALL phenotype</b>								
B-cell precursor	191	29	16%	11-22				
T-cell precursor	47	8	17%	9-32	.744	1.15	0.52-2.53	.732
<b>Disease phase at HSCT</b>								
Only ALL patients								
CR1†	93	16	18%	11-28	.850			
CR2 S1-S2†	85	11	13%	8-23		0.72	0.34-1.56	.405
CR2 S3-S4‡	49	8	16%	9-31		0.93	0.40-2.17	.865
Other CR‡	11	2	18%	5-64		1.98	0.24-4.83	.920
Only AML patients								
CR1†	82	8	10%	5-19	.531			
CR2†	22	4	18%	8-45		1.94	0.59-6.35	.273
<b>Prognosis†‡</b>								
Good prognosis†	282	39	14%	11-19	.581			
Poor prognosis‡	61	10	16%	9-29		1.21	0.60-2.44	.583
<b>MRD, all patients</b>								
<1 × 10 <sup>-3</sup>	143	16	11%	7-18	.855			
≥1 × 10 <sup>-3</sup>	20	2	10%	3-37		0.87	0.20-3.73	.855
Neg	119	14	12%	7-19	.842			
Pos <1 × 10 <sup>-3</sup>	24	2	8%	2-31		0.66	0.16-2.80	.578
Pos ≥1 × 10 <sup>-3</sup>	20	2	10%	3-37		0.82	0.19-3.55	.792
<b>MRD, only patients with ALL</b>								
<1 × 10 <sup>-3</sup>	121	15	12%	8-20	.743			
≥1 × 10 <sup>-3</sup>	11	1	9%	1-59		0.72	0.10-5.33	.744
Neg	98	13	13%	8-22	.756			
Pos <1 × 10 <sup>-3</sup>	23	2	9%	2-33		0.61	0.14-2.59	.503
Pos ≥1 × 10 <sup>-3</sup>	11	1	9%	1-59		0.66	0.09-4.99	.678

Subgroups including fewer than 5 patients were not shown in the table.

\*The Cox regression model was used to estimate the HR, and the first subgroup of patients of each independent variable was used as reference value.

†The good prognosis group included patients affected by ALL in first CR, ALL in second CR after a relapse belonging to the S1-S2 risk groups and patients with AML in first CR.

‡The poor prognosis group included patients affected by ALL in second CR after a relapse belonging to the S3-S4 risk groups, ALL in third or subsequent CR, and patients with AML in second CR.



Table 2. (continued)

	Number of patients	Events	Cumulative incidence	95% CI	P	HR*	95% CI	P
<b>MRD, only patients with AML</b>								
<1 × 10 <sup>-3</sup>	22	1	5%	1-31	.524			
≥1 × 10 <sup>-3</sup>	9	1	11%	2-71		2.39	0.16-35.82	.529
Neg	21	1	5%	1-32	.802			
Pos ≥1 × 10 <sup>-3</sup>	9	1	11%	2-71		2.28	0.15-34.09	
<b>Donor</b>								
MUD	245	40	17%	13-22	.085			
Haploidentical	98	9	9%	5-17		0.54	0.26-1.11	.092
<b>HLA compatibility</b>								
MUD	127	8	6%	3-12	<.001			
MMUD	118	32	28%	21-37		4.76	2.19-10.36	<.001
Haploidentical	98	9	9%	5-17		1.46	0.56-3.79	.435
<b>Stem cell source</b>								
BM	187	30	16%	12-23	.267			
PBSC	156	19	12%	8-19		0.72	0.41-1.28	.268
<b>Conditioning regimen</b>								
TBI-based	204	31	16%	11-21	.827			
Busulfan-based	115	16	14%	9-22		0.92	0.50-1.67	.774
Treosulfan-based	21	2	10%	3-36		0.64	0.15-2.79	.553
<b>Conditioning regimen, ALL patients</b>								
TBI-based	189	29	16%	11-22	.904			
Busulfan-based	34	6	18%	9-36		0.21	0.49-2.99	.673
Treosulfan-based	12	2	17%	5-59		0.16	0.25-5.19	.846
<b>Conditioning regimen, AML patients</b>								
TBI-based	15	2	13%	4-48	.733			
Busulfan-based	81	10	13%	7-23		0.93	0.21-4.24	
Treosulfan-based	9	0	0%	—		0.00	—	<.001

Subgroups including fewer than 5 patients were not shown in the table.

\*The Cox regression model was used to estimate the HR, and the first subgroup of patients of each independent variable was used as reference value.

†The good prognosis group included patients affected by ALL in first CR, ALL in second CR after a relapse belonging to the S1-S2 risk groups and patients with AML in first CR.

‡The poor prognosis group included patients affected by ALL in second CR after a relapse belonging to the S3-S4 risk groups, ALL in third or subsequent CR, and patients with AML in second CR.

frequently received a TBI-based conditioning regimen (74%) as compared with the MUD (50%) and MMUD-HSCT groups (58%; Table 1). In fact,  $\alpha\beta$ haplo-HSCT was offered to children who lacked either an HLA-identical sibling or a MUD, or in case of a rapidly progressive or unstable disease not allowing the time to identify a suitable MUD. Remarkably, the incidence of graft failure in the  $\alpha\beta$ haplo-HSCT did not differ from that of patients given UD-HSCT. Consistent with what we reported in a recent single-center trial,<sup>25</sup> none of the  $\alpha\beta$ haplo-HSCT recipients experienced either grade III to IV or visceral acute GVHD (Figure 2B). The advantage of  $\alpha\beta$ haplo-HSCT is even more evident if we compare the cumulative incidence of the more severe form of acute GVHD in recipients of  $\alpha\beta$ haplo-HSCT and that in recipients of MMUD-HSCT (Figure 2B). Moreover, in the  $\alpha\beta$ haplo-HSCT cohort, all cases but 1 of chronic GVHD were of limited severity (Figure 2D). The low risk of developing severe acute GVHD likely contributed to a lower risk for NRM in MUD and  $\alpha\beta$ haplo-HSCT recipients in comparison with MMUD-HSCT ( $P < .001$ ; Figure 3B). Notably, the median number of residual  $\alpha\beta$  T cells in the haploidentical grafts was very low, being

$0.04 \times 10^6/\text{kg}$  body weight. In light of the direct correlation between  $\alpha\beta$  T cells and GVHD, we believe that an optimal graft composition is crucial for the outcome of the  $\alpha\beta$ haplo-HSCT. Thus, we strongly recommend maintaining a threshold of residual  $\alpha\beta$  T cells lower than  $1 \times 10^5/\text{kg}$  recipient body weight. In the  $\alpha\beta$ haplo-HSCT group, the ex vivo CD19<sup>+</sup> depletion coupled with the in vivo CD20<sup>+</sup> depletion obtained through the pre-transplant use of rituximab could explain the total absence of EBV-related PTLD, a fearsome complication typical of immunocompromised patients. Moreover, we can speculate that the use of rituximab on day -1 helped prevent the occurrence not only of EBV-PTLD but also that of GVHD.<sup>45</sup>

In the last 20 years, multiple strategies of ex vivo and in vivo T-cell depletion techniques have been reported to be effective in HLA-haploidentical HSCT.<sup>46,47</sup> In addition to the pioneering studies using CD34<sup>+</sup> positive selection,<sup>13,14</sup> some centers reported data on combined CD3<sup>+</sup> and CD19<sup>+</sup> negative depletion.<sup>48-50</sup> Both these approaches resulted in the loss of some cell subsets playing a crucial role in infection defense and leukemia

**Table 3. Multivariable analysis of factors affecting NRM, cumulative incidence of relapse and LFS**

Variable	Risk ratio	95% CI	P
<b>Nonrelapse mortality</b>			
Age at HSCT			
≥13 y vs <13 y	1.88	1.05-3.35	.033
Donor			
MMUD vs MUD	4.35	1.97-9.60	<.001
Haploidentical vs MUD	1.37	0.52-3.56	.523
<b>Relapse</b>			
Diagnosis			
AML vs ALL	1.20	0.52-3.46	.737
Prognosis			
Poor vs good	3.43	1.69-6.94	.001
MRD			
≥1 × 10 <sup>-3</sup> vs <1 × 10 <sup>-3</sup>	2.97	1.32-6.67	.008
Conditioning regimen			
Busulfan vs TBI	1.51	0.51-4.45	.452
Treosulfan vs TBI	1.46	0.64-3.56	.370
Donor			
MMUD vs MUD	0.52	0.25-1.10	.088
Haploidentical vs MUD	0.71	0.33-1.54	.383
<b>Leukemia-free-survival</b>			
Sex			
Female vs male	1.63	0.97-2.75	.065
Age at HSCT			
≥13 y vs <13 y	0.94	0.52-1.72	.842
Diagnosis			
AML vs ALL	0.81	0.33-1.98	.641
Prognosis			
Poor vs good	3.00	1.69-5.31	<.001
MRD level			
≥1 × 10 <sup>-3</sup> vs <1 × 10 <sup>-3</sup>	2.90	1.47-5.71	.002
Conditioning regimen			
Busulfan vs TBI	1.22	0.52-2.86	.648
Treosulfan vs TBI	1.45	0.64-3.27	.372
Donor			
MMUD vs MUD	0.93	0.51-1.69	.800
Haploidentical vs MUD	0.69	0.36-1.32	.258

Subgroups including fewer than 5 patients were not shown in the table.

recurrence prevention, translating into a higher risk for NRM and disease relapse. After 2010, several centers, mostly European, started to use the αβ T-cell/CD19<sup>+</sup> B-cell depletion approach.<sup>15</sup> The goal of this new method of graft manipulation is to leave in the graft not only HSCs but also cells pertaining to innate immunity, which might facilitate engraftment and reduce risk for both infections and leukemia recurrence.<sup>16,17</sup> In fact, although T cells carrying the αβ T-cell receptor (TCR) are responsible for GVHD,<sup>51,52</sup> γδ T cells have no alloreactive capacity.<sup>53</sup> However, they can contribute to an important anti-infectious<sup>54</sup> and antileukemia effect.<sup>21,55-57</sup> Thanks to this technique, patients can also immediately benefit from donor NK cells contained in this type of graft that can fully exert their activity, filling the 6- to 8-week gap documented in previous studies before mature KIR<sup>+</sup> NK cells differentiating from CD34<sup>+</sup> cells are consistently detected.<sup>19,32</sup> Altogether, the infusion with the graft of these different lymphoid cell subsets could explain our

finding that NRM and LFS were superimposable when results of αβhaplo HSCT and MUD-HSCT are compared. Notably, the incidence of fatal events resulting from severe infections was equal in the MUD and αβhaplo-HSCT group, highlighting the role of both NK and γδ cells in protecting the host in the very early phase after the procedure in the αβhaplo-HSCT group. Conversely, in the MMUD-HSCT cohort, most deaths (33%) were a result of infections, which were mainly bacterial (supplemental Tables 2 and 3). In addition, the infusion with the graft of a large number of such effector cells, along with a megadose of HSCs and with the use of a fully myeloablative conditioning regimen, might explain the remarkably low incidence of graft failure (2%) observed in our αβhaplo-HSCT group. Moreover, the graft composition granted a faster engraftment of neutrophils and platelets in the αβhaplo-HSCT group. This last finding can, at least partly, be responsible of the significantly lower incidence of bacterial infections in the αβhaplo-HSCT (8%) group in comparison with the MUD (17%) and the MMUD-HSCT (34%) groups (*P* < .001; supplemental Table 3).

Ultimately, the absence of pharmacological post-HSCT GVHD prophylaxis in our haploidentical cohort and the use of a fully myeloablative conditioning regimen could explain the lower incidence of relapse in comparison with previously published single-center reports. Indeed, with a median follow-up of 1.6 years, Lang et al showed a 41.4% cumulative incidence of leukemia recurrence in a cohort of 41 pediatric patients transplanted after a reduced-intensity, TBI-free conditioning regimen.<sup>23</sup> Maschan et al analyzed the outcome of children with high-risk AML receiving αβ T-cell-depleted HSCT either from an UD or a haploidentical donor.<sup>24</sup> The majority of patients received different combination of post-HSCT pharmacological prophylaxis. The 2-year cumulative incidence of relapse in their haplo cohort was 40%. Although even in our study the main cause of treatment failure is relapse after αβhaplo-HSCT (cumulative incidence, 24%; Figure 2A; supplemental Table 4 for more details), we can speculate that NK cells and other innate lymphoid cells not exposed to immunosuppressive treatment could display a better antileukemia effect leading to superior LFS.

In contrast to what was observed with the CD34<sup>+</sup> positive selection approach, but consistent with our single-center experience data recently published,<sup>25</sup> in our αβhaplo-HSCT cohort, the use of an NK alloreactive donor did not affect the leukemia-free survival. Likely, we can speculate that infusion of γδ T cells with the graft can obscure the role played by NK alloreactivity in a contest of extensive and prolonged T-cell depletion.

In multivariable analysis, belonging to a good-prognosis risk group and an MRD level lower than 1 × 10<sup>-3</sup> before HSCT had a positive effect on LFS (HR, 3.43 [95% CI, 1.69-6.94]; *P* = .001); HR, 2.97 [95% CI, 1.32-6.67; *P* = .008], respectively). These findings confirm the importance of obtaining the best possible leukemia control already reported from different groups, regardless of donor type or graft-manipulation strategy.<sup>58,59</sup> The recent implementation of next-generation sequencing approaches for evaluating MRD may further affect the success of HSCT, allowing us to implement strategies aimed at preventing recurrence of leukemia.<sup>60,61</sup> In this multicenter analysis, contrary to what was observed in the single-center experience we previously reported,<sup>25</sup> the type of conditioning regimen employed (TBI-based or chemotherapy-based) did not influence LFS in multivariable analysis (Table 4). As

**Table 4. Univariable analysis of LFS in the cohort of patients analyzed**

	Number of patients	Events	Probability	95% CI	P	HR*	(95% CI)	P
All patients	343	124	62%	56-67				
<b>Sex</b>								
Male	210	66	66%	58-73	.015			
Female	133	58	55%	47-64		1.54	1.07-2.20	.016
<b>Age at HSCT, y</b>								
<9.5	172	57	65%	58-73	.218			
≥9.5	171	67	58%	50-66		1.25	0.88-1.78	.218
<6	94	33	64%	54-74	.165			
6-9.5	78	24	66%	54-78		0.82	0.48-1.39	.460
9.5-13	80	26	65%	53-77		0.91	0.54-1.51	.705
≥13	91	41	52%	41-63		1.37	0.87-2.17	.178
<13	252	83	65%	59-72	.032			
≥13	91	41	52%	41-63		1.50	1.03-2.18	.033
<b>Diagnosis</b>								
ALL	237	93	59%	52-66	.064			
AML	106	31	67%	57-78		0.68	0.45-1.03	.066
<b>ALL phenotype</b>								
B-cell precursor	191	74	59%	50-66	.713			
T-cell precursor	47	19	59%	45-73		0.91	0.55-1.50	.713
<b>Disease phase at HSCT</b>								
Only ALL patients								
CR1†	93	31	66.7%	56-76	.0005			
CR2 S1-S2†	85	26	69.4%	58-79		0.89	0.53-1.49	.651
CR2 S3-S4‡	49	29	40.8%	24-53		2.11	1.27-3.51	.004
Other CR‡	11	7	36.4%	8-64		2.74	1.21-6.24	.016
Only AML patients								
CR1†	82	23	72%	56-80	.084			
CR2†	22	7	68.2%	48-88		1.25	0.54-2.91	.607
<b>Prognosis†‡</b>								
Good prognosis†	282	87	67%	62-73	.0001			
Poor prognosis‡	61	37	37%	23-50		2.54	1.73-3.74	.0001
<b>MRD, all patients</b>								
<1 × 10 <sup>-3</sup>	143	52	63%	54-71				
≥1 × 10 <sup>-3</sup>	20	13	20%	0-49	.003	2.48	1.35-4.56	.004
Neg	119	39	66%	58-75				
Pos <1 × 10 <sup>-3</sup>	24	13	42%	20-63		1.68	0.90-3.16	.105
Pos ≥1 × 10 <sup>-3</sup>	20	16	20%	0-49	.003	2.76	1.47-5.19	.002
<b>MRD, only ALL patients</b>								
<1 × 10 <sup>-3</sup>	121	46	61%	52-70				
≥1 × 10 <sup>-3</sup>	11	7	36%	8-65	.046	2.20	0.99-4.88	.052
Neg	98	33	66%	57-75				
Pos <1 × 10 <sup>-3</sup>	23	13	39%	17-61		1.70	0.89-3.23	.110
Pos ≥1 × 10 <sup>-3</sup>	11	7	36%	8-65	.040	2.49	1.10-5.64	.028

Subgroups including fewer than 5 patients were not shown in the table.

\*The Cox regression model was used to estimate the HR, and the first subgroup of patients of each independent variable was used as reference value.

†The good prognosis group included patients affected by ALL in first CR, ALL in second CR after a relapse belonging to the S1-S2 risk groups and patients with AML in first CR.

‡The poor-prognosis group included patients affected by ALL in second CR after a relapse belonging to the S3-S4 risk groups, ALL in third or subsequent CR, and patients with AML in second CR.

**Table 4. (continued)**

	Number of patients	Events	Probability	95% CI	P	HR*	(95% CI)	P
<b>MRD, only AML patients</b>								
<1 × 10 <sup>-3</sup>	22	6	72%	52-91				
≥1 × 10 <sup>-3</sup>	9	6	0%	—	.010	4.14	1.28-13.38	.017
Neg	21	6	70%	50-90				
Pos ≥1 × 10 <sup>-3</sup>	9	9	0%	—	.033	3.92	1.22-12.65	.022
<b>Donor</b>								
MUD	245	90	63.3%	65-68	.650			
Haploidentical	98	34	65.3%	51-73		1095	0.74-1.63	.650
<b>HLA compatibility</b>								
MUD	127	40	67%	59-76	.150			
MMUD	118	50	55%	45-65		1.48	0.98-2.25	.063
Haploidentical	98	34	62%	51-73		1.11	0.71-1.76	.642
<b>Stem cell source</b>								
BM	187	68	64%	55-69	.731			
PBSC	156	56	64%	53-70		1.064	0.75-1.52	.731
<b>Conditioning regimen</b>								
TBI-based	204	73	64%	54-69	.673			
Busulfan-based	115	40	65%	53-72		0.95	0.64-1.39	.783
Treosulfan-based	21	10	52%	28-72		1.45	0.75-2.81	.270
<b>Conditioning regimen, ALL patients</b>								
TBI-based	189	69	61%	53-69	.004			
Busulfan-based	34	14	59%	42-75		1.20	0.68-2.13	.532
Treosulfan-based	12	10	17%	0-38		3.11	1.60-6.06	.001
<b>Conditioning regimen, AML patients</b>								
TBI-based	15	4	73%	51-96	.155			
Busulfan-based	81	26	65%	53-76		1.14	0.40-3.27	.805
Treosulfan-based	9	0	100%	—		0.00	—	<.001

Subgroups including fewer than 5 patients were not shown in the table.

\*The Cox regression model was used to estimate the HR, and the first subgroup of patients of each independent variable was used as reference value.

†The good prognosis group included patients affected by ALL in first CR, ALL in second CR after a relapse belonging to the S1-S2 risk groups and patients with AML in first CR.

‡The poor-prognosis group included patients affected by ALL in second CR after a relapse belonging to the S3-S4 risk groups, ALL in third or subsequent CR, and patients with AML in second CR.

chemotherapy, we mainly employed a Busulfan-based conditioning regimen. Only a very small proportion of patients received Treosulfan as a part of the preparative regimen (6%, 5%, and 7% in the MUD, MMUD, and αβhaplo-HSCT groups, respectively). Although the relapse incidence of patients with ALL receiving Treosulfan is higher as compared with the other regimens (supplemental Table 4), in view of the low number of patients treated according to this chemotherapy, no definitive conclusion can be drawn.

In parallel to different graft-engineering approaches, in vivo strategies of T-cell depletion based on the posttransplant infusion of Cyclophosphamide have been developed in the setting of haplo-HSCT.<sup>62</sup> Although largely used in adults, little is known on the use of this method in children.<sup>63,64</sup> The few studies so far available reported a risk for leukemia recurrence higher than the one observed in this multicenter analysis.<sup>64</sup>

In conclusion, our study, the first in a multicenter setting, confirms that αβ T-cell- and B-cell-depleted HLA-haploidentical

HSCT is currently a valuable option for children with AL lacking a sibling donor. First, this approach is associated with a cumulative incidence of NRM and disease recurrence comparable to that of children transplanted from a MUD. Second, αβ T-cell and B-cell depletion abrogates the risk of developing severe acute and chronic GVHD and is also associated with faster neutrophil and platelet recovery than UD-HSCT. Finally, when compared with MMUD-HSCT, αβhaplo-HSCT is clearly superior, showing a significantly lower cumulative incidence of NRM and a better chronic GFRS. As noted previously, an alternative treatment using posttransplant Cyclophosphamide as an in vivo T-cell depletion approach have shown some encouraging results,<sup>62,65</sup> although published work, to date, has focused primarily on adults. Prospective trials comparing this approach with αβ T-cell and B-cell depletion are needed in children to establish which alternative represents the optimal treatment under which conditions. In the meantime, data from the present study support the argument that αβhaplo-HSCT should be considered a transplant option of equal efficacy vs MUD-HSCT and better than MMUD-HSCT for AL children in need of an allograft and lacking a fully matched related donor.

## Acknowledgments

This work was supported by grants from AIRC (Special Grant "5xmille"-9962 to F.L.; investigator grant IG 17200 to F.L.; "My first AIRC" grant 15925 to A. Bertaina), Ministero della Salute (RF-2010-2316606 to F.L.; Ricerca Corrente to F.L.), Regione Lazio (Grant FILAS to F.L.), and Ministero dell'Istruzione, Università e Ricerca (Grant Progetto di Rilevante Interesse Nazionale, PRIN 2010, to F.L.).

## Authorship

Contribution: A. Bertaina, M.Z., and F.L. provided conception and design; A. Bertaina, M.Z., M.A., F.S., C.P., E.L., A.P., W.B., M.T., C.F., S.C., F.D.B., M.R., S.B., G.C., M.R., A. Balduzzi, F.F., D.P., and F.L. provided transplantation of patients and collection of data; N.S. and A.M.G. provided unrelated donor search and selection; B.B. provided minimal residual disease monitoring; V.B. provided immune monitoring; A. Bertaina, M.Z., D.P., and F.L. provided analysis and interpretation of data (eg, statistical analysis, computational analysis); and all authors provided writing, review and/or revision of the manuscript.

Conflict-of-interest disclosure: The authors declare no competing financial interests.

ORCID profiles: A.B., 0000-0002-3729-436X; V.B., 0000-0002-9028-7767.

Correspondence: Alice Bertaina, Division of Stem Cell Transplant and Regenerative Medicine, Department of Pediatrics, School of Medicine, Stanford University, 1000 Welch Rd, Stanford, CA, 94305; e-mail: aliceb1@stanford.edu.

## Footnotes

Submitted 5 July 2018; accepted 3 October 2018. Prepublished online as *Blood* First Edition paper, 22 October 2018; DOI 10.1182/blood-2018-07-861575.

\*A. Bertaina and M.Z. are joint first authors.

†D.P. and F.L. are joint senior authors.

The online version of this article contains a data supplement.

The publication costs of this article were defrayed in part by page charge payment. Therefore, and solely to indicate this fact, this article is hereby marked "advertisement" in accordance with 18 USC section 1734.

## REFERENCES

- Copelan EA. Hematopoietic stem-cell transplantation. *N Engl J Med*. 2006;354(17):1813-1826.
- Rocha V, Locatelli F. Searching for alternative hematopoietic stem cell donors for pediatric patients. *Bone Marrow Transplant*. 2008;41(2):207-214.
- Gragert L, Eapen M, Williams E, et al. HLA match likelihoods for hematopoietic stem-cell grafts in the U.S. registry. *N Engl J Med*. 2014;371(4):339-348.
- Gluckman E. Cord blood transplantation. *Biol Blood Marrow Transplant*. 2006;12(8):808-812.
- Locatelli F. Improving cord blood transplantation in children. *Br J Haematol*. 2009;147(2):217-226.
- Martelli MF, Reisner Y. Haploidentical 'megadose' CD34+ cell transplants for patients with acute leukemia. *Leukemia*. 2002;16(3):404-405.
- Ferrara JLM, Levine JE, Reddy P, Holler E. Graft-versus-host disease. *Lancet*. 2009;373(9674):1550-1561.
- Petersdorf EW. Which factors influence the development of GVHD in HLA-matched or mismatched transplants? *Best Pract Res Clin Haematol*. 2017;30(4):333-335.
- Pulsipher MA. Haplo is the new black. *Blood*. 2014;124(5):675-676.
- Bertaina A, Pitisci A, Sinibaldi M, Algeri M. T cell-depleted and T cell-replete HLA-haploidentical stem cell transplantation for non-malignant disorders. *Curr Hematol Malig Rep*. 2017;12(1):68-78.
- Reisner Y, Hagin D, Martelli MF. Haploidentical hematopoietic transplantation: current status and future perspectives. *Blood*. 2011;118(23):6006-6017.
- Martelli MF, Aversa F. Haploidentical transplants using ex vivo T-cell depletion. *Semin Hematol*. 2016;53(4):252-256.
- Aversa F, Tabilio A, Velardi A, et al. Treatment of high-risk acute leukemia with T-cell-depleted stem cells from related donors with one fully mismatched HLA haplotype. *N Engl J Med*. 1998;339(17):1186-1193.
- Aversa F, Tabilio A, Terenzi A, et al. Successful engraftment of T-cell-depleted haploidentical "three-loci" incompatible transplants in leukemia patients by addition of recombinant human granulocyte colony-stimulating factor-mobilized peripheral blood progenitor cells to bone marrow inoculum. *Blood*. 1994;84(11):3948-3955.
- Chaleff S, Otto M, Barfield RC, et al. A large-scale method for the selective depletion of alphabeta T lymphocytes from PBSC for allogeneic transplantation. *Cytotherapy*. 2007;9(8):746-754.
- Locatelli F, Bauquet A, Palumbo G, Moretta F, Bertaina A. Negative depletion of alpha/beta T cells and of CD19+ B lymphocytes: a novel frontier to optimize the effect of innate immunity in HLA-mismatched hematopoietic stem cell transplantation. *Immunol Lett*. 2013;155(1-2):21-23.
- Locatelli F, Pende D, Mingari MC, et al. Cellular and molecular basis of haploidentical hematopoietic stem cell transplantation in the successful treatment of high-risk leukemias: role of alloreactive NK cells. *Front Immunol*. 2013;4:15.
- Moretta L, Pietra G, Vacca P, et al. Human NK cells: From surface receptors to clinical applications. *Immunol Lett*. 2016;178:15-19.
- Locatelli F, Moretta F, Brescia L, Merli P. Natural killer cells in the treatment of high-risk acute leukaemia. *Semin Immunol*. 2014;26(2):173-179.
- Airoidi I, Bertaina A, Prigione I, et al. gamma delta T-cell reconstitution after HLA-haploidentical hematopoietic transplantation depleted of TCR-alpha beta+/CD19+ lymphocytes. *Blood*. 2015;125(15):2349-2358.
- Lamb LS Jr, Henslee-Downey PJ, Parrish RS, et al. Increased frequency of TCR gamma delta + T cells in disease-free survivors following T cell-depleted, partially mismatched, related donor bone marrow transplantation for leukemia. *J Hematother*. 1996;5(5):503-509.
- Bertaina A, Merli P, Rutella S, et al. HLA-haploidentical stem cell transplantation after removal of alpha beta+ T and B cells in children with nonmalignant disorders. *Blood*. 2014;124(5):822-826.
- Lang P, Feuchtinger T, Teltschik HM, et al. Improved immune recovery after transplantation of TCRalpha beta/CD19-depleted allografts from haploidentical donors in pediatric patients. *Bone Marrow Transplant*. 2015;50(suppl 2):S6-S10.
- Maschan M, Shelikhova L, Ilushina M, et al. TCR-alpha/beta and CD19 depletion and treosulfan-based conditioning regimen in unrelated and haploidentical transplantation in children with acute myeloid leukemia. *Bone Marrow Transplant*. 2016;51(5):668-674.
- Locatelli F, Merli P, Pagliara D, et al. Outcome of children with acute leukemia given HLA-haploidentical HSCT after alpha beta T-cell and B-cell depletion. *Blood*. 2017;130(5):677-685.
- Ruggeri L, Mancusi A, Capanni M, et al. Donor natural killer cell allorecognition of missing self in haploidentical hematopoietic transplantation for acute myeloid leukemia: challenging its predictive value. *Blood*. 2007;110(1):433-440.
- Pende D, Marcenaro S, Falco M, et al. Anti-leukemia activity of alloreactive NK cells in KIR ligand-mismatched haploidentical HSCT for pediatric patients: evaluation of the functional role of activating KIR and redefinition of inhibitory KIR specificity. *Blood*. 2009;113(13):3119-3129.
- Oevermann L, Michaelis SU, Mezger M, et al. KIR B haplotype donors confer a reduced risk for relapse after haploidentical transplantation in children with ALL. *Blood*. 2014;124(17):2744-2747.

29. Cooley S, Weisdorf DJ, Guethlein LA, et al. Donor selection for natural killer cell receptor genes leads to superior survival after unrelated transplantation for acute myelogenous leukemia. *Blood*. 2010;116(14):2411-2419.
30. Alicata C, Pende D, Meazza R, et al. Hematopoietic stem cell transplantation: Improving alloreactive Bw4 donor selection by genotyping codon 86 of KIR3DL1/S1. *Eur J Immunol*. 2016;46(6):1511-1517.
31. Moretta L, Locatelli F, Pende D, Marcenaro E, Mingari MC, Moretta A. Killer Ig-like receptor-mediated control of natural killer cell alloreactivity in haploidentical hematopoietic stem cell transplantation. *Blood*. 2011;117(3):764-771.
32. Pende D, Marcenaro S, Falco M, et al. Anti-leukemia activity of alloreactive NK cells in KIR ligand-mismatched haploidentical HSCT for pediatric patients: evaluation of the functional role of activating KIR and redefinition of inhibitory KIR specificity. *Blood*. 2009;113(13):3119-3129.
33. Rutella S, Filippini P, Bertaina V, et al. Mobilization of healthy donors with plerixafor affects the cellular composition of T-cell receptor (TCR)- $\alpha\beta$ /CD19-depleted haploidentical stem cell grafts. *J Transl Med*. 2014;12(1):240.
34. Li Pira G, Malaspina D, Girolami E, et al. Selective depletion of  $\alpha\beta$  T cells and B cells for human leukocyte antigen-haploidentical hematopoietic stem cell transplantation: a three-year follow-up of procedure efficiency. *Biol Blood Marrow Transplant*. 2016;22(11):2056-2064.
35. Glucksberg H, Storb R, Fefer A, et al. Clinical manifestations of graft-versus-host disease in human recipients of marrow from HL-A-matched sibling donors. *Transplantation*. 1974;18(4):295-304.
36. Lee SJ, Vogelsang G, Flowers MED. Chronic graft-versus-host disease. *Biol Blood Marrow Transplant*. 2003;9(4):215-233.
37. Holtan SG, DeFor TE, Lazaryan A, et al. Composite end point of graft-versus-host disease-free, relapse-free survival after allogeneic hematopoietic cell transplantation. *Blood*. 2015;125(8):1333-1338.
38. Kaplan EL, Meier P. Nonparametric estimation from incomplete observations. *J Am Stat Assoc*. 1958;53(282):457-481.
39. Klein JP, Rizzo JD, Zhang MJ, Keiding N. Statistical methods for the analysis and presentation of the results of bone marrow transplants. Part 1: unadjusted analysis. *Bone Marrow Transplant*. 2001;28(10):909-915.
40. Klein JP, Rizzo JD, Zhang MJ, Keiding N. Statistical methods for the analysis and presentation of the results of bone marrow transplants. Part 2: Regression modeling. *Bone Marrow Transplant*. 2001;28(11):1001-1011.
41. Gray RJ. A class of K-sample tests for comparing the cumulative incidence of a competing risk. *Ann Stat*. 1988;16(3):1141-1154.
42. Bashey ZA, Zhang X, Brown S, et al. Comparison of outcomes following transplantation with T-replete HLA-haploidentical donors using post-transplant cyclophosphamide to matched related and unrelated donors for patients with AML and MDS aged 60 years or older. *Bone Marrow Transplant*. 2018;53(6):756-763.
43. Sestili S, Labopin M, Ruggeri A, et al. T-cell-depleted haploidentical stem cell transplantation results improve with time in adults with acute leukemia: A study from the Acute Leukemia Working Party of the European Society of Blood and Marrow Transplantation (EBMT). *Cancer*. 2018;124(10):2142-2150.
44. Devillier R, Legrand F, Rey J, et al. HLA-matched sibling versus unrelated versus haploidentical related donor allogeneic hematopoietic stem cell transplantation for patients aged over 60 years with acute myeloid leukemia: a single-center donor comparison. *Biol Blood Marrow Transplant*. 2018;24(7):1449-1454.
45. Christopheit M, Schütte V, Theurich S, Weber T, Grothe W, Behre G. Rituximab reduces the incidence of acute graft-versus-host disease. *Blood*. 2009;113(13):3130-3131.
46. Mancusi A, Ruggeri L, Velardi A. Haploidentical hematopoietic transplantation for the cure of leukemia: from its biology to clinical translation. *Blood*. 2016;128(23):2616-2623.
47. Liu JH, Kanakry CG, Luznik L. Have haploidentical transplants replaced umbilical cord transplants for acute leukemias? *Curr Opin Hematol*. 2018;25(2):103-111.
48. Lang P, Teltschik HM, Feuchtinger T, et al. Transplantation of CD3/CD19 depleted allografts from haploidentical family donors in paediatric leukaemia. *Br J Haematol*. 2014;165(5):688-698.
49. Huenecke S, Bremm M, Cappel C, et al. Optimization of individualized graft composition: CD3/CD19 depletion combined with CD34 selection for haploidentical transplantation. *Transfusion*. 2016;56(9):2336-2345.
50. Bethge WA, Faul C, Bornhäuser M, et al. Haploidentical allogeneic hematopoietic cell transplantation in adults using CD3/CD19 depletion and reduced intensity conditioning: an update. *Blood Cells Mol Dis*. 2008;40(1):13-19.
51. Korngold R, Sprent J. T cell subsets and graft-versus-host disease. *Transplantation*. 1987;44(3):335-339.
52. Shlomchik WD. Graft-versus-host disease. *Nat Rev Immunol*. 2007;7(5):340-352.
53. Handgretinger R, Schilbach K. The potential role of  $\gamma\delta$  T cells after allogeneic HCT for leukemia. *Blood*. 2018;131(10):1063-1072.
54. Bonneville M, O'Brien RL, Born WK. Gammadelta T cell effector functions: a blend of innate programming and acquired plasticity. *Nat Rev Immunol*. 2010;10(7):467-478.
55. Dolstra H, Fredrix H, van der Meer A, de Witte T, Figdor C, van de Wiel-van Kemenade E. TCR gamma delta cytotoxic T lymphocytes expressing the killer cell-inhibitory receptor p58.2 (CD158b) selectively lyse acute myeloid leukemia cells. *Bone Marrow Transplant*. 2001;27(10):1087-1093.
56. Godder KT, Henslee-Downey PJ, Mehta J, et al. Long term disease-free survival in acute leukemia patients recovering with increased gammadelta T cells after partially mismatched related donor bone marrow transplantation. *Bone Marrow Transplant*. 2007;39(12):751-757.
57. Bertaina A, Zorzoli A, Petretto A, et al. Zoledronic acid boosts  $\gamma\delta$  T-cell activity in children receiving  $\alpha\beta^+$  T and CD19 $^+$  cell-depleted grafts from an HLA-haplo-identical donor. *Oncoimmunology*. 2016;6(2):e1216291.
58. Lovisa F, Zecca M, Rossi B, et al. Pre- and post-transplant minimal residual disease predicts relapse occurrence in children with acute lymphoblastic leukaemia. *Br J Haematol*. 2018;180(5):680-693.
59. Bader P, Kreyenberg H, von Stackelberg A, et al. Monitoring of minimal residual disease after allogeneic stem-cell transplantation in relapsed childhood acute lymphoblastic leukemia allows for the identification of impending relapse: results of the ALL-BFM-SCT 2003 trial. *J Clin Oncol*. 2015;33(11):1275-1284.
60. Getta BM, Devlin SM, Levine RL, et al. Multicolor flow cytometry and multigene next-generation sequencing are complementary and highly predictive for relapse in acute myeloid leukemia after allogeneic transplantation. *Biol Blood Marrow Transplant*. 2017;23(7):1064-1071.
61. Kotrova M, van der Velden VHJ, van Dongen JJM, et al. Next-generation sequencing indicates false-positive MRD results and better predicts prognosis after SCT in patients with childhood ALL. *Bone Marrow Transplant*. 2017;52(7):962-968.
62. Luznik L, Bolaños-Meade J, Zahurak M, et al. High-dose cyclophosphamide as single-agent, short-course prophylaxis of graft-versus-host disease. *Blood*. 2010;115(16):3224-3230.
63. Berger M, Lanino E, Cesaro S, et al. Feasibility and outcome of haploidentical hematopoietic stem cell transplantation with post-transplant high-dose cyclophosphamide for children and adolescents with hematologic malignancies: an AIEOP-GITMO retrospective multicenter study. *Biol Blood Marrow Transplant*. 2016;22(5):902-909.
64. Klein OR, Buddenbaum J, Tucker N, et al. Nonmyeloablative Haploidentical Bone Marrow Transplantation with Post-Transplantation Cyclophosphamide for Pediatric and Young Adult Patients with High-Risk Hematologic Malignancies. *Biol Blood Marrow Transplant*. 2017;23(2):325-332.
65. Luznik L, Fuchs EJ. High-dose, post-transplantation cyclophosphamide to promote graft-host tolerance after allogeneic hematopoietic stem cell transplantation. *Immunol Res*. 2010;47(1-3):65-77.