

- lymphocyte leukemia. *Blood Rev.* 2014;28(3): 87-94.
3. Zhang J, Xu X, Liu Y. Activation-induced cell death in T cells and autoimmunity. *Cell Mol Immunol.* 2004;1(3):186-192.
  4. Loughran TP Jr, Kadin ME, Starkebaum G, et al. Leukemia of large granular lymphocytes: association with clonal chromosomal abnormalities and autoimmune neutropenia, thrombocytopenia, and hemolytic anemia. *Ann Intern Med.* 1985; 102(2):169-175.
  5. Lamy T, Moignet A, Loughran TP Jr. LGL leukemia: from pathogenesis to treatment. *Blood.* 2017;129(9):1082-1094.
  6. Isabelle C, Boles A, Chakravarti N, Porcu P, Brammer J, Mishra A. Cytokines in the pathogenesis of large granular lymphocytic leukemia. *Front Oncol.* 2022;12:849917.
  7. Hodge DL, Yang J, Buschman MD, et al. Interleukin-15 enhances proteasomal degradation of bid in normal lymphocytes: implications for large granular lymphocyte leukemias. *Cancer Res.* 2009;69(9): 3986-3994.
  8. Epling-Burnette PK, Liu JH, Catlett-Falcone R, et al. Inhibition of STAT3 signaling leads to apoptosis of leukemic large granular lymphocytes and decreased Mcl-1 expression. *J Clin Invest.* 2001;107(3): 351-362.
  9. Jerez A, Clemente MJ, Makishima H, et al. STAT3 mutations unify the pathogenesis of chronic lymphoproliferative disorders of NK cells and T-cell large granular lymphocyte leukemia. *Blood.* 2012;120(15):3048-3057.
  10. Waldmann TA, Conlon KC, Stewart DM, et al. Phase 1 trial of IL-15 trans presentation blockade using humanized Mikbeta1 mAb in patients with T-cell large granular lymphocytic leukemia. *Blood.* 2013;121:476-484.
- <https://doi.org/10.1182/blood.2023021476>  
© 2023 by The American Society of Hematology

## RED CELLS, IRON, AND ERYTHROPOIESIS

Comment on [Xiao et al](#), page 1312

# BMP5: a novel tile of the hepcidin regulatory pathway

**Antonella Nai** | IRCCS Ospedale San Raffaele and Vita-Salute San Raffaele University

**In this issue of *Blood*, [Xiao et al](#)<sup>1</sup> describe, for the first time, a role for bone morphogenetic protein 5 (BMP5) in the control of hepcidin, the master regulator of systemic iron homeostasis.**

Hepcidin is a peptide hormone produced by the liver, which limits iron absorption from the diet and iron release from stores. Hepcidin acts by occluding and degrading the sole iron exporter, ferroportin. Hepcidin is transcriptionally regulated by different pathways, among which a major role is played by the BMP suppressor of mothers against decapentaplegic (SMAD) system. On BMP's binding, the BMP type II receptors (BMP receptor 2 and activin A receptor type 2A (ACVR2A)) phosphorylate type I receptors (activin receptor-like kinase 2 (ALK2) and activin receptor-like kinase 3 (ALK3)), which, in turn, phosphorylate SMAD1/5/8. These proteins bind the cargo SMAD4; then, the complex translocates into the nucleus to activate the transcription of genes carrying a BMP-responsive element, including hepcidin.<sup>2</sup>

Two BMP ligands mainly produced by liver sinusoidal endothelial cells (LSECs), BMP2<sup>3,4</sup> and BMP6,<sup>5,6</sup> are relevant in hepcidin modulation. BMP2 maintains basal

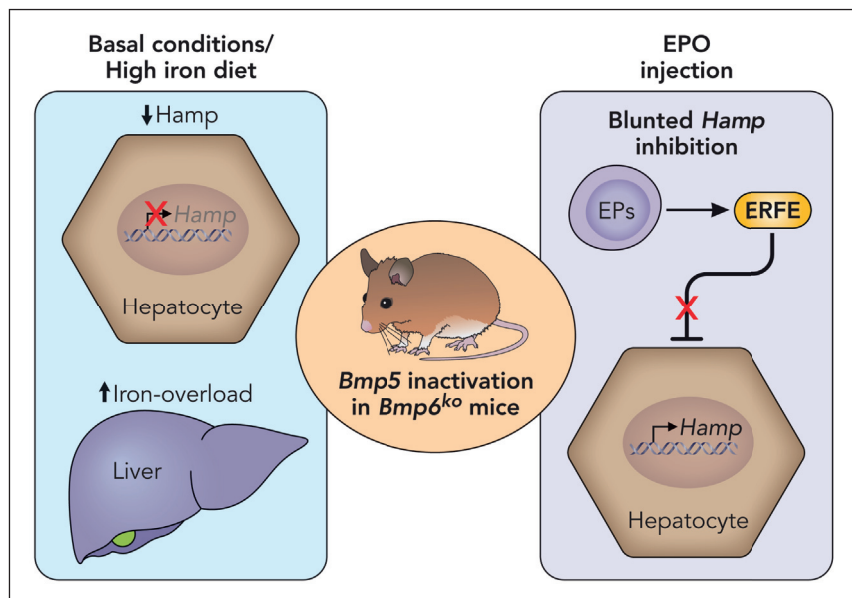
hepcidin levels binding ALK3, whereas BMP6, upregulated by liver iron, activates the pathway in conditions of tissue iron accumulation, preferentially via ALK2.<sup>2</sup> Despite the crucial and nonredundant roles of these 2 ligands, iron is still able to activate the BMP-SMAD-hepcidin pathway in double LSEC *Bmp2* knockout (KO) and global *Bmp6* KO mice, suggesting that at least 1 additional ligand is involved in the upregulation of hepcidin. In the present study, [Xiao et al](#) demonstrate that BMP5 is likely the missing piece of the puzzle.

The role of BMP5 in the physiological regulation of hepcidin is marginal compared with that of BMP2 and BMP6. Indeed, mice lacking *Bmp5* in the whole organism do not show dysregulation of the iron regulatory system, with the sole exception of a modest decrease in hepcidin levels when 10 days old. This is potentially reminiscent of a defect during fetal life, which is resolved in adulthood. However, when these animals are

challenged with an iron-poor or a high-iron diet, they accumulate more iron in the liver and maintain inappropriately low hepcidin levels. This finding proves that BMP5 contributes to the transcriptional activation of hepcidin in response to both low and high iron, when *Bmp6* is virtually absent or maximally induced. In agreement, *Bmp5* inactivation in mice lacking *Bmp6* (both globally or specifically in LSECs) dramatically reduces hepcidin levels, worsening both hepatic and extra-hepatic iron overload, an effect exacerbated by feeding animals a high-iron diet (see [figure](#)). In *Bmp6* total KO, the loss of *Bmp5* causes an increased mortality, irrespective of the degree of iron overload, which is comparable in both *Bmp6* global and LSEC-specific KO, but likely related to a redundant nonendothelial developmental role of BMP6 and BMP5, which remains to be elucidated.

At difference with BMP2 and BMP6, which are mainly produced by LSECs in response to iron levels, *Bmp5* is not preferentially expressed in any liver cells and is not transcriptionally activated by iron. However, its activity appears strongly dependent on iron availability, raising the possibility that the metal controls BMP5 at the posttranscriptional level. However, the lack of specific antibodies currently precludes the possibility of addressing this point. Also, how BMP5 works, which receptors it uses, whether it can form heterodimers with BMP2, BMP6, and/or other BMPs, its potential involvement in the fetal iron homeostasis, and why BMP6 levels are limiting for the function of BMP5 are still unsolved issues, which likely will be the focus of further studies from the same and other groups in the field.

BMPs are crucial not only for the iron-mediated regulation of hepcidin, but also for hepcidin control by the expanded erythropoiesis. In response to erythropoietin (EPO) stimulation, developing erythroblasts produce the erythroid regulator erythroferrone (ERFE), which inhibits hepcidin transcription to increase iron supply needed for hemoglobin production.<sup>7</sup> ERFE functions by binding and sequestering BMPs, mainly BMP6, but also BMP5 and BMP7.<sup>8</sup> However, EPO injection is still able to suppress hepcidin in *Bmp6* global KO,<sup>9</sup> proving that other BMPs likely contribute to the ERFE-mediated hepcidin downregulation. The current study demonstrates that BMP5 is involved in this



The loss of *Bmp5* severely impacts the phenotype of *Bmp6*-deficient (*Bmp6*<sup>ko</sup>) mice. In basal conditions and following a high-iron diet, *Bmp5* deficiency further decreases hepcidin (*Hamp*) levels and worsens hepatic and extrahepatic iron overload in mice lacking *Bmp6* both in the whole organism and selectively in LSECs. Conversely, following an erythropoietic stimulus, erythroferrone (ERFE) fails in downregulating hepcidin transcription in mice lacking both BMP5 and BMP6. EP, erythroid progenitor; EPO, erythropoietin. Professional illustration by Patrick Lane, ScEYence Studios.

regulation. Indeed, EPO injection, although properly inhibiting hepcidin in *Bmp5*-deficient mice, is completely ineffective in male mice lacking both *Bmp5* and *Bmp6* (see figure). However, residual hepcidin suppression is observed in double-mutant female mice, which have higher basal hepcidin levels, raising the possibility that other BMPs might play a role.

Overall, this work reveals BMP5 as a novel player in the complex regulation of hepcidin and systemic iron homeostasis, and it paves the way for more compre-

hensive studies on the function of this, and potentially other, BMP ligand. From a translational perspective, interfering with their function might potentially become a novel therapeutic opportunity for iron and erythroid disorders due to excessive hepcidin production, in line with recent results obtained with anti-BMP6 agents.<sup>10</sup>

*Conflict-of-interest disclosure:* The author declares no competing financial interests. ■

## REFERENCES

1. Xiao X, Xu Y, Moschetta GS, et al. BMP5 contributes to hepcidin regulation and

systemic iron homeostasis in mice. *Blood*. 2023;142(15):1312-1322.

2. Silvestri L, Pettinato M, Furiosi V, Bavuso Volpe L, Nai A, Pagani A. Managing the dual nature of iron to preserve health. *Int J Mol Sci*. 2023;24(4):3995.
3. Canali S, Wang CY, Zumbrennen-Bullough KB, Bayer A, Babbitt JL. Bone morphogenetic protein 2 controls iron homeostasis in mice independent of *Bmp6*. *Am J Hematol*. 2017;92(11):1204-1213.
4. Koch PS, Olsavsky V, Ulbrich F, et al. Angiocrine *Bmp2* signaling in murine liver controls normal iron homeostasis. *Blood*. 2017;129(4):415-419.
5. Andriopoulos B Jr, Corradini E, Xia Y, et al. BMP6 is a key endogenous regulator of hepcidin expression and iron metabolism. *Nat Genet*. 2009;41(4):482-487.
6. Meynard D, Kautz L, Darnaud V, Canonne-Hergaux F, Coppin H, Roth MP. Lack of the bone morphogenetic protein BMP6 induces massive iron overload. *Nat Genet*. 2009; 41(4):478-481.
7. Kautz L, Jung G, Valore EV, Rivella S, Nemeth E, Ganz T. Identification of erythroferrone as an erythroid regulator of iron metabolism. *Nat Genet*. 2014;46(7): 678-684.
8. Arezes J, Foy N, McHugh K, et al. Erythroferrone inhibits the induction of hepcidin by BMP6. *Blood*. 2018;132(14):1473-1477.
9. Nai A, Rubio A, Campanella A, et al. Limiting hepatic *Bmp-Smad* signaling by matriptase-2 is required for erythropoietin-mediated hepcidin suppression in mice. *Blood*. 2016; 127(19):2327-2336.
10. Petzer V, Tymoszuk P, Asshoff M, et al. A fully human anti-BMP6 antibody reduces the need for erythropoietin in rodent models of the anemia of chronic disease. *Blood*. 2020; 136(9):1080-1090.

<https://doi.org/10.1182/blood.2023021643>

© 2023 by The American Society of Hematology