

## Review Article

# LRRK2 in peripheral and central nervous system innate immunity: its link to Parkinson's disease

Heyne Lee, William S. James and Sally A. Cowley

Sir William Dunn School of Pathology, University of Oxford, South Parks Road, Oxford OX1 3RE, U.K.

Correspondence: Heyne Lee (heyne.lee@path.ox.ac.uk)



Mutations in the *leucine-rich repeat kinase 2 (LRRK2)* gene are found in familial and idiopathic cases of Parkinson's disease (PD), but are also associated with immune-related disorders, notably Crohn's disease and leprosy. Although the physiological function of LRRK2 protein remains largely elusive, increasing evidence suggests that it plays a role in innate immunity, a process that also has been implicated in neurodegenerative diseases, including PD. Innate immunity involves macrophages and microglia, in which endogenous LRRK2 expression is precisely regulated and expression is strongly up-regulated upon cell activation. This brief report discusses the current understanding of the involvement of LRRK2 in innate immunity particularly in relation to PD, critically examining its role in myeloid cells, particularly macrophages and microglia.

## Parkinson's disease

Parkinson's disease (PD) is a complex, multifactorial neurodegenerative disease. In North America, it affects 1.5% of the population over the age of 65. Patients gradually develop motor impairments, caused by a slow and progressive degeneration of dopaminergic neurones in the *substantia nigra pars compacta* (SNpc). The aetiology of PD is largely unknown, involving a complex interaction between various genetic and environmental factors. To date, 17 distinctive chromosomal locations, named *parkin (PARK) 1–18*, have been identified in association with inherited PD. Although only ~10% of PD cases are identified as familial PD, genome-wide association studies (GWAS) have also detected a role for genetic variants in idiopathic PD (reviewed in ref. [1]). Understanding the roles of PD-associated genes, therefore, has increasing significance as this would provide valuable insights into shared pathological mechanisms underlying both inherited and idiopathic PD pathogenesis.

Interestingly, key PD-associated genes, *α-synuclein (SNCA)*, *PARK2*, *deglycase (DJ-1)*, *leucine-rich repeat kinase 2 (LRRK2)*, and *glucocerebrosidase (GBA)*, are all expressed in immune cells, implying their potential role in immunity (reviewed in ref. [2]). Neuronal injuries commonly elicit activation of innate immune responses in the central nervous system (CNS), and inflammation-driven neurotoxicity has been suggested to play a central role in progression of various neurodegenerative diseases, including PD (reviewed in ref. [3]). PD-associated genes may have distinct cellular functions in immune cells, and it can also be hypothesized that mutations in these genes commonly contribute to abnormal immune responses, which in turn may act as a driving force to exacerbate the progression of inflammation-mediated neurodegeneration.

## Systemic and CNS inflammation in PD

The PD brain displays numerous signs of ongoing inflammatory processes. Both PD patients and animal models of PD display higher levels of activated microglia, which remain phagocytic for a prolonged period of time [4,5]. Elevated levels of inflammatory cytokines, especially tumour necrosis factor- $\alpha$  (TNF- $\alpha$ ), interleukin-1 $\beta$  (IL-1 $\beta$ ), IL-2, IL-6, IL-8, and interferon- $\gamma$  (IFN- $\gamma$ ), are detected in the brain, cerebrospinal fluid, and blood of PD patients (reviewed in ref. [2]). Infiltration of peripheral immune cells, notably CD4+ and CD8+ T lymphocytes, is found as a consequence of abnormal

Received: 23 August 2016

Revised: 9 October 2016

Accepted: 12 October 2016

Version of Record published:  
15 February 2017

permeability of the blood–brain barrier, which normally keeps the CNS in an immune-privileged state [6]. The involvement of humoral immunity has also been implicated: Lewy bodies and dopaminergic neurones in the SNpc show strong immunolabelling for immunoglobulin G [7]. Collectively, all of these ongoing inflammatory processes involve activation of microglia, suggesting its relevance to the pathophysiology of PD.

Microglia, the resident macrophages in the CNS, ensure a healthy environment for neurones by conducting a suite of homeostatic functions. These include clearing cell debris, extracellular protein aggregates, and excess neurotransmitters. In response to pathological stimuli, microglia become activated, proliferate, and accumulate at the site of injury, where they phagocytose dead cells and secrete inflammatory mediators and a myriad of cytotoxic factors, especially reactive oxygen species (ROS) and nitric oxide [8]. Activation and subsequent down-regulation of microglial activity are strictly controlled, as exaggerated inflammatory responses can be harmful to neurones [9].

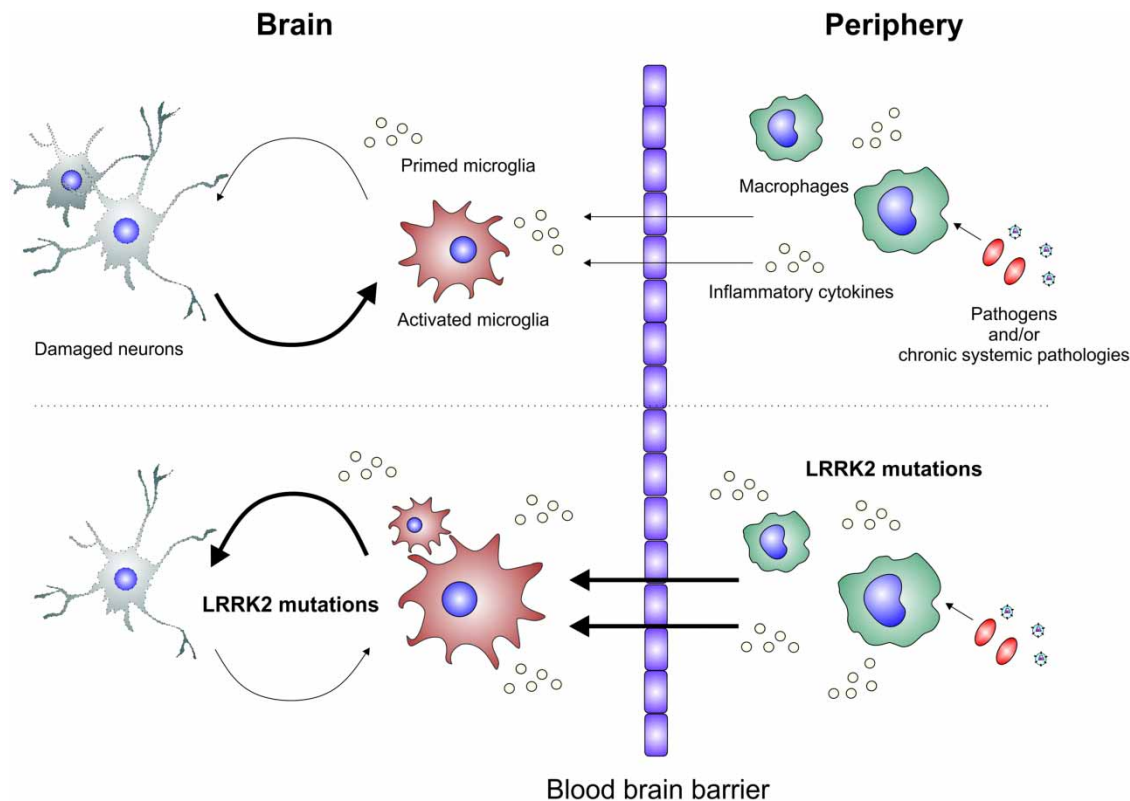
Chronic pathological factors (including repeated exposure to environmental toxins, genetic predispositions, and abnormal immune responses) may prolong the activated state of microglia, potentially instigating a feedforward cycle of chronic degeneration of neurones and inflammation. This self-perpetuating cycle of microglia-mediated neurotoxicity is particularly relevant in PD. First, dopaminergic neurones in the SNpc are intrinsically vulnerable to metabolic stress, particularly that caused by dopamine oxidation or mitochondrial dysfunction. High cytosolic dopamine levels can be dangerous, as dopamine metabolites (e.g. 6-hydroxydopamine) are toxic to neurones. Furthermore, compared with mesolimbic dopaminergic neurones, dopaminergic neurones in the SNpc exhibit a much larger  $\text{Ca}^{2+}$  influx, which requires the endoplasmic reticulum (ER)–mitochondrial system to clear excess  $\text{Ca}^{2+}$  (reviewed in ref. [10]). Secondly, the SNpc is one of the brain regions with the highest density of microglia and a relatively low density of astrocytes [11,12]. These factors suggest that the feedforward cycle of chronic activation of microglia and chronic damage of dopaminergic neurones would be particularly detrimental in the SNpc.

Systemic inflammation may also contribute to this cycle by amplifying microglia activation. The concept of microglia priming was proposed to describe exaggerated and prolonged inflammatory phenotypes displayed by microglia upon exposure to subsequent stimuli (reviewed in ref. [9]). For example, infection with *Salmonella* has been shown to activate microglia, and subsequent injection of lipopolysaccharide (LPS) to the brain triggered much higher microglial responses [13]. In the context of PD, this has been experimentally shown in rats, where prenatal exposure to LPS makes the brain more susceptible to subsequent LPS injection in midlife, leading to progressive loss of nigral dopaminergic neurones [14]. Since PD-associated genes are found in both peripheral and CNS immune cells, pathological interplay between the two immune systems carrying PD mutations could predispose the brain to reach a critical threshold of inflammation, triggering a self-perpetuating cycle of inflammation and neuronal death (Figure 1).

## Leucine-rich repeat kinase 2

LRRK2 is a large, multidomain protein, displaying both GTPase and kinase activities. Most PD-causing mutations, notably R1441C/G and G2019S, cluster within these two enzymatic sites, which are surrounded by large protein–protein interacting domains (reviewed in ref. [15]). *LRRK2* mutations are one of the most common genetic causes of PD: mutations can account for as much as 40% of familial PD [16] and its variants are also found within idiopathic cases [17]. Unlike other PD-associated genes, *LRRK2* Parkinsonism manifests similar clinical phenotypes to idiopathic PD, displaying strong age-dependent development of PD symptoms [18]. Deciphering the role of LRRK2 in PD pathogenesis may reveal common pathological mechanisms underlying idiopathic PD and is consequently of great research interest.

Despite intense research effort over the past decade, the physiological function of LRRK2 and the contribution of mutations to PD remain largely elusive. This is at least in part because earlier research has mainly focussed on the role of LRRK2 in neurones, in which endogenous expression is low [19]. Many studies have, therefore, relied on overexpression of LRRK2 in non-physiologically relevant cell lines or animal models, but these approaches generate results that do not necessarily reflect the normal physiological interactome of LRRK2. This, together with the complex, multidomain structure of LRRK2, with protein–protein interaction domains at the N- and C-terminal segments, has led to LRRK2 being reported to interact with numerous molecules in a wide variety of cellular pathways, including endosome vesicle trafficking, cytoskeleton reorganization, mitochondrial function, regulation of ER/Golgi retromer complex, autophagy, and various signalling pathways, including wingless/int, TNF- $\alpha$ /Fas ligand (FasL)/Fas-associated protein with death domain, mitogen-activated protein kinase, and nuclear factor  $\kappa$ -light-chain-enhancer of activated B cells pathways [20–23]. Future investigations should examine LRRK2 expressed at physiological levels from its endogenous promoter at its normal



**Figure 1. LRRK2 and microglia-driven neuroinflammation in PD.**

(Top panel) In this scenario, mutations in PD genes exert their effects directly in neurons, leading to chronic neuronal damage. This triggers microglial activation, leading to a vicious cycle of neuronal death and chronically activated microglia, with inflammatory cytokines causing collateral damage. The threshold for microglial activation may be lowered by peripheral inflammation, which ‘primes’ microglia. (Bottom panel) In this scenario, mutations in PD genes (in this case LRRK2) also exert effects by expression in macrophages and microglia, leading to dysfunctional immune responses. LRRK2-mutant microglia may exhibit exaggerated responses to neuronal damage, causing an amplified vicious cycle. The threshold for microglia activation could be further lowered by LRRK2 mutations within peripheral immune cells contributing to chronic microglial priming.

human chromosomal location in authentic, relevant human cells, to discern which of these cellular pathways represent the bona fide function(s) of LRRK2.

## LRRK2 expression is precisely regulated in myeloid cells

Although the existence of LRRK2 protein in microglia and astrocytes has been reported in the past [24], and *LRRK2* variants have been linked through GWAS to Crohn’s disease [25,26] and leprosy [27], it was not until 2010 that researchers found that LRRK2 expression is precisely up-regulated by inflammatory signals in myeloid cells, strongly implicating its potential role as a regulator of immune responses [21].

Although the level of endogenous LRRK2 is low in resting leukocytes, upon stimulation with IFN- $\gamma$ , robust up-regulation of endogenous LRRK2 has been consistently detected across various subsets of myeloid cells and lymphocytes, human peripheral blood mononuclear cell-derived CD11b<sup>+</sup> monocytes, CD3<sup>+</sup> T lymphocytes, CD19<sup>+</sup> B lymphocytes [21], human primary monocyte-derived macrophages, mouse primary microglia [28], and transformed cell lines, including human THP-1 monocytic leukaemia cells [29,30], and murine RAW264.7 macrophage-like cells [21]. IFN- $\gamma$  activation has a direct effect on the *LRRK2* promoter region, which contains binding sites for IFN-response factors [21]. Janus kinase/signal transducers and activators of transcription and the extracellular signal-regulated kinase 5 mediate IFN- $\gamma$ -induced LRRK2 up-regulation in macrophages, although the exact signalling cascades are yet to be elucidated [29].

LRRK2 is also moderately inducible by other inflammatory mediators, namely IFN- $\beta$ , TNF- $\alpha$ , and IL-6 [31], whereas the Toll-like receptor 4 (TLR4) agonist, LPS, is found to have an inconsistent effect. Some groups have reported significant up-regulation of LRRK2 protein expression by LPS in primary mouse microglia or in THP-1 cells [28,32], whereas others did not detect any changes in murine immortalised microglia (BV-2), primary mouse microglia [33,34], or mouse bone marrow-derived macrophages (BMDMs) [35]. This discrepancy could be attributed to many factors, such as cell types, experimental conditions, or technical variations. Regardless, activation of TLR4 reproducibly leads to phosphorylation at Ser910/935 residues of LRRK2 in all myeloid cell lineages [19,35,36]. Phosphorylation at Ser910/935 determines its cellular localization, interaction with 14-3-3 protein, dimerization, and translocation from cytosol to the membrane [19,37–39]. However, further experimental investigation is needed to understand the direct physiological consequences of phosphorylation at Ser910/935 residues in myeloid cells.

## Roles of LRRK2 in macrophages and microglia

Microglia and macrophages are both classified as mononuclear phagocytes, sharing common functions of various maintenance and protective roles, but can be distinguished by their ontogeny and transcription profiles (reviewed in ref. [40]). Since biochemical changes in LRRK2 upon inflammatory cues are identical in both systems [19], efforts have been made to inspect the role of LRRK2 in various aspects of innate immunity, summarized in Table 1. These studies imply opposing roles of LRRK2 in peripheral and CNS innate immunity. However, it should be noted that none of these studies has directly compared microglia and macrophages under the same experimental conditions, so direct evidence for opposing roles is still lacking. Moreover, most data are from mouse models that do not faithfully recapitulate all aspects of the human immune system [41].

### Cytokine release

In mouse primary microglia, R1441G mutation leads to an increase in LPS-driven inflammatory cytokine release [28], and abolishing LRRK2 protein expression has the opposite effect [32,33,42]. However, in mouse primary macrophages, neither R1441G [43] nor G2019S mutations [36], nor *LRRK2* knockout (KO) [35,43,44], cause any change in LPS-driven cytokine release. Only Dectin-1 activation by zymosan (yeast) has been shown to produce higher levels of cytokine release in *LRRK2* KO mouse macrophages [45]. Therefore, in macrophages, LRRK2 may be dispensable in TLR4-mediated cytokine release, but may serve an important role in responding to other inflammatory stimuli. A recent report has shown that higher levels of peripheral inflammatory cytokines were found in the sera of both asymptomatic *LRRK2* G2019S carriers and PD patients carrying *LRRK2* G2019S [46], suggesting pathological contributions from *LRRK2* mutations within peripheral immune cells. Further studies are merited to establish which specific inflammatory pathways are mediated by LRRK2 in macrophages or microglia. Additionally, the role of LRRK2 in peripheral cytokine release is still equivocal, and it will help the field to have studies that directly compare cytokine release in microglia and macrophages from the same animals under the same experimental conditions, comparing all the key *LRRK2* mutations and KO together. Finally, LRRK2-mediated immune responses by more disease-relevant immunogenic agonists, such as aggregates of SNCA, should be explored further, as there is evidence for close interaction between  $\alpha$ -synuclein and LRRK2 (reviewed in ref. [47]).

### Migration

The ability to respond and migrate to the site of injury or infection is a key feature of innate immunity, and recent evidence suggests the involvement of LRRK2 in this process. Abolishing LRRK2 expression in BV-2 microglial-like cells [48] or mouse primary microglia [49] leads to significantly higher migration versus wild type, and the G2019S mutation in mouse primary microglia diminishes ADP-induced migration [48]. However, in mouse primary macrophages, the same mutation has been shown to have the opposite effect, enhancing motility towards ADP [36].

### Phagocytosis

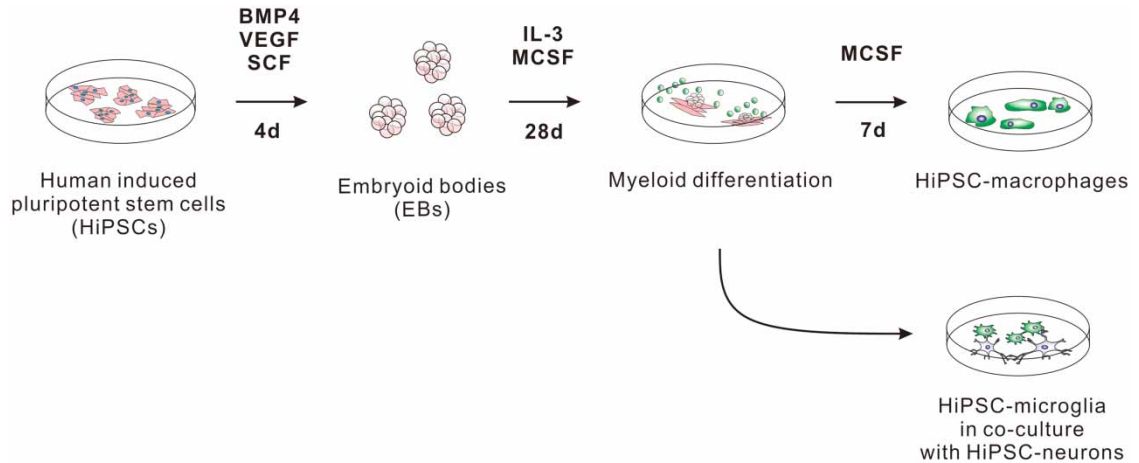
The absence of LRRK2 activity in BV-2 microglial-like cells or RAW 264.7 macrophage-like cells has been reported to have no effect on phagocytosis [19]; likewise, G2019S mutation was not observed in mouse primary macrophages [36]. However, it has been reported that LRRK2 localizes to phagosomes upon bacterial infection, and that lack of LRRK2 expression reduces ROS production and enhances bacterial survival in RAW 264.7 cells [21]. Therefore, LRRK2 may have a specific role during phagocytosis, which may not be detected by simple

**Table 1 Summary of reports on the role of LRRK2 in innate immunity**

Species	Cell types	Methods	Results	References
LPS-mediated cytokine and chemokine release				
Mouse	BV-2	LRRK2 knockdown (KD) shRNA	↓TNF- $\alpha$ , IL-6, Nitrite	[42]
Mouse	Primary microglia	LRRK2 KD RNAi Kinase inhibition: sunitinib LRRK2-in-1	↓TNF- $\alpha$	[32]
Mouse	Primary microglia	LRRK2 KO Kinase inhibition: LRRK2-in-1 GSK2578215A	↓IL-1 $\beta$ , cyclooxygenase-2 mRNA	[33]
Mouse	Primary microglia	R1441G	↑TNF- $\alpha$ ; ↓IL-10	[28]
Mouse	BMDMs	R1441C LRRK2 KO	No difference in IL-6 or keratinocyte chemokine (KC)	[43]
Mouse	BMDMs	LRRK2 KO	No difference in TNF- $\alpha$ , IL-6, KC, IL-1 $\beta$ , IL-10, IL-12	[35]
Mouse	Thioglycollate-elicited peritoneal macrophages (TEPMs)	LRRK2 KO	No difference in IL-1 $\beta$ , IL-10, IL-1 $\alpha$ , TNF- $\alpha$ , IL-6, KC, granulocyte colony-stimulating factor, monocyte chemoattractant protein-1	[44]
Mouse	TEPMs	G2019S	No difference in TNF- $\alpha$	[36]
Migration				
Mouse	Primary microglia BV-2	G2019S LRRK2 KD shRNA	↓ADP-induced migration ↑ADP-induced migration	[48]
Mouse	Primary microglia	LRRK2 KO	↑Fractalkine-induced migration	[49]
Mouse	Primary TEPMs	G2019S Kinase inhibition SRI 29451 HG-10-102-01	↑ADP-induced migration ↓G2019S-enhanced ADP-induced migration	[36]
Phagocytosis				
Mouse	BV-2 RAW264.7	LRRK2 KD shRNA Kinase inhibition LRRK2-in-1 GSK2578215A HG-10-102-01	No difference in uptake of FITC-conjugated beads	[19]
Mouse	Primary TEPMs	G2019S	No difference in uptake of fluorescent zymosan bioparticles	[36]
Mouse	RAW264.7	<i>Salmonella typhimurium</i> infection LRRK2 KD siRNA	LRRK2 localization to phagosomes ↓ROS with zymosan ↑Survival of intracellular <i>S. typhimurium</i>	[21]

phagocytosis assays measuring initial uptake of bioparticles. Indeed, LRRK2 is implicated in autophagy in myeloid cells ([19]; reviewed in ref. [50]), a pathway that shares common features with phagocytosis and is also involved in innate immunity (reviewed in ref. [51]). Further exploration of the role of LRRK2 during specific stages of phagocytosis pathways is merited.





**Figure 2. Schematic of macrophage and microglia differentiation from HiPSCs.**

For the detailed protocol for HiPSC-derived macrophage differentiation, see ref. [52]. BMP4, bone morphogenetic protein 4; VEGF, vascular endothelial growth factor; SCF, stem cell factor; IL-3, interleukin 3; MCSF, macrophage colony-stimulating factor; d, days.

## Human-induced pluripotent stem cell macrophages and microglia as tools to study LRRK2 biology

It is clear from above that the data on the role of LRRK2 in myeloid cells has so far been collected primarily from murine *ex vivo* models or from transformed murine cell lines, under a wide variety of experimental conditions. While murine systems are extremely useful, they do not precisely replicate all human cellular and biochemical pathways [41]. Moreover, transformed cell lines poorly reproduce the cellular physiology of authentic primary human macrophages and microglia, which are terminally differentiated cells. Human-induced pluripotent stem cell (HiPSC)-derived macrophages provide an attractive and highly authentic model to study LRRK2 biology (Figure 2). HiPSC-derived macrophages are genetically tractable, can be generated efficiently, and, at scale, become terminally differentiated and accurately recapitulate macrophage functionality [52]. Together with clustered regularly interspaced short palindromic repeats (CRISPR)/Cas-9 gene editing of iPSCs, one can investigate LRRK2 protein at the endogenous level by generating KO lines, correcting and introducing mutations, creating reporter lines, or tagging endogenous proteins. The applicability of this system has been shown already in other diseases, including HIV and chronic granulomatous disease [53,54]. Methods for skewing HiPSC-derived macrophages to microglia are currently under development and will enable direct comparisons of LRRK2 function in human macrophages and microglia.

### Concluding remarks

Recent evidence supports the idea that pathological interplay between peripheral and CNS innate immunity probably contributes to the progression of PD. LRRK2 may be involved in this interplay, as expression of LRRK2 is tightly regulated in both systems and evidence reviewed here implicates LRRK2 in both peripheral and CNS innate immunity. Although the current literature appears to suggest that LRRK2 plays distinct roles in microglia and macrophages, more work needs to be done to unequivocally establish the *bona fide* function (s) of LRRK2 in human macrophages and microglia, and the role of LRRK2 mutations in these cells in PD. To achieve this, macrophages/microglia differentiated from HiPSCs provide a powerful tool to better understand LRRK2-mediated pathology in PD and also other LRRK2-mediated immune disorders.

### Abbreviations

BMDMs, bone marrow-derived macrophages; CNS, central nervous system; CRISPR, clustered regularly interspaced short palindromic repeats; ER, endoplasmic reticulum; FasL, Fas ligand; GWAS, genome-wide association studies; HiPSC, human-induced pluripotent stem cell; IFN- $\gamma$ , interferon- $\gamma$ ; IL-1 $\beta$ , interleukin-1 $\beta$ ; KO, knockout; LPS, lipopolysaccharide; LRRK2, leucine-rich repeat kinase 2; PARK2, *parkin*; PBMC, peripheral blood

mononuclear cell; PD, Parkinson's disease; ROS, reactive oxygen species; *SNCA*,  $\alpha$ -synuclein; SNpc, *substantia nigra pars compacta*; TLR, Toll-like receptor; TNF- $\alpha$ , tumour necrosis factor- $\alpha$ .

## Funding

We acknowledge financial support from the Wellcome Trust [WTISSF121302], the Oxford Martin School [LC0910-004], the Oxford Parkinson's Disease Centre (OPDC) Monument Trust Discovery Award from Parkinson's UK, a charity registered in England and Wales [2581970] and in Scotland [SC037554], with the support of the National Institute for Health Research (NIHR) Oxford Biomedical Research Centre based at Oxford University Hospitals NHS Trust and University of Oxford, and the NIHR Comprehensive Local Research Network, and European Federation of Pharmaceutical Industries and Associations (EFPIA), European Union Innovative Medicines Initiatives (EU IMI) "StemBANCC", who provide the following statement: The research leading to these results has received support from the Innovative Medicines Initiative Joint Undertaking under grant agreement no. 115439, resources of which are composed of financial contribution from the European Union's Seventh Framework Programme (FP7/2007–2013), and EFPIA companies in kind contribution. This publication reflects that only the author's views and neither the IMI JU ([www.imi.europa.eu](http://www.imi.europa.eu)) nor EFPIA, nor the European Commission, are liable for any use that may be made of the information contained therein.

## Competing Interests

The Authors declare that there are no competing interests associated with the manuscript.

## References

- 1 Ferreira, M. and Massano, J. (2016). An updated review of Parkinson's disease genetics and clinicopathological correlations. *Acta Neurol. Scand.* PMID:27273099
- 2 Dzamko, N., Geczy, C.L. and Halliday, G.M. (2015) Inflammation is genetically implicated in Parkinson's disease. *Neuroscience* **302**, 89–102 doi:10.1016/j.neuroscience.2014.10.028
- 3 Amor, S. Peferoen, L.A.N., Vogel, D.Y.S., Breur, M., van der Valk, P., Baker D. et al. (2014) Inflammation in neurodegenerative diseases — an update. *Immunology* **142**, 151–166 doi:10.1111/imm.12233
- 4 Barcia, C., Ros, C.M., Ros-Bernal, F., Gómez, A., Annese, V., Carrillo-de Sauvage, M.A. et al. (2013) Persistent phagocytic characteristics of microglia in the substantia nigra of long-term Parkinsonian macaques. *J. Neuroimmunol.* **261**, 60–66 doi:10.1016/j.jneuroim.2013.05.001
- 5 McGeer, P.L., Itagaki, S., Boyes, B.E. and McGeer, E.G. (1988) Reactive microglia are positive for HLA-DR in the substantia nigra of Parkinson's and Alzheimer's disease brains. *Neurology* **38**, 1285 doi:10.1212/WNL.38.8.1285
- 6 Brochard, V., Combadière, B., Prigent, A., Laouar, Y., Perrin, A., Beray-Berthet, V. et al. (2009) Infiltration of CD4+ lymphocytes into the brain contributes to neurodegeneration in a mouse model of Parkinson disease. *J. Clin. Invest.* **119**, 182–192 PMID:19104149
- 7 Orr, C., Rowe, D.B., Mizuno, Y., Mori, H. and Halliday, G.M. (2005) A possible role for humoral immunity in the pathogenesis of Parkinson's disease. *Brain* **128**, 2665–2674 doi:10.1093/brain/awh625
- 8 Sanchez-Guajardo, V., Barnum, C.J., Tansey, M.G. and Romero-Ramos, M. (2013) Neuroimmunological processes in Parkinson's disease and their relation to  $\alpha$ -synuclein: microglia as the referee between neuronal processes and peripheral immunity. *ASN Neuro.* **5**, 113–139 doi:10.1042/AN20120066
- 9 Perry, V.H. and Teeling, J. (2013) Microglia and macrophages of the central nervous system: the contribution of microglia priming and systemic inflammation to chronic neurodegeneration. *Semin. Immunopathol.* **35**, 601–612 doi:10.1007/s00281-013-0382-8
- 10 Gonzalez-Hernandez, T., Cruz-Muros, I., Alfonso-Oramas, D., Salas-Hernandez, J. and Castro-Hernandez, J. (2010) Vulnerability of mesostriatal dopaminergic neurons in Parkinson's disease. *Front. Neuroanat.* **4**, 140 doi:10.3389/fnana.2010.00140
- 11 Kim, W.G., Mohney, R.P., Wilson, B., Jeohn, G.H., Liu, B. and Hong, J.S. (2000) Regional difference in susceptibility to lipopolysaccharide-induced neurotoxicity in the rat brain: role of microglia. *J. Neurosci.* **20**, 6309–6316 PMID:10934283
- 12 Savchenko, V.L., McKanna, J.A., Nikonenko, I.R. and Skibo, G.G. (2000) Microglia and astrocytes in the adult rat brain: comparative immunocytochemical analysis demonstrates the efficacy of lipocortin 1 immunoreactivity. *Neuroscience* **96**, 195–203 doi:10.1016/S0306-4522(99)00538-2
- 13 Püntener, U., Booth, S.G., Perry, V.H. and Teeling, J.L. (2012) Long-term impact of systemic bacterial infection on the cerebral vasculature and microglia. *J. Neuroinflammation* **9**, 146 doi:10.1186/1742-2094-9-146
- 14 Ling, Z., Zhu, Y., Tong, C.W., Snyder, J.A., Lipton, J.W. and Carvey, P.M. (2006) Progressive dopamine neuron loss following supra-nigral lipopolysaccharide (LPS) infusion into rats exposed to LPS prenatally. *Exp. Neurol.* **199**, 499–512 doi:10.1016/j.expneurol.2006.01.010
- 15 Greggio, E. (2012) Role of LRRK2 kinase activity in the pathogenesis of Parkinson's disease. *Biochem. Soc. Trans.* **40**, 1058–1062 doi:10.1042/BST20120054
- 16 Lesage, S., Dürr, A., Tazir, M., Lohmann, E., Leutenegger, A.-L., Janin, S. et al. (2006) *LRRK2* G2019S as a cause of Parkinson's disease in North African Arabs. *N. Engl. J. Med.* **354**, 422–423 doi:10.1056/NEJMc055540
- 17 Satake, W., Nakabayashi, Y., Mizuta, I., Hirota, Y., Ito, C., Kubo, M. et al. (2009) Genome-wide association study identifies common variants at four loci as genetic risk factors for Parkinson's disease. *Nat. Genet.* **41**, 1303–1307 doi:10.1038/ng.485
- 18 Healy, D.G., Falchi, M., O'Sullivan, S.S., Bonifati, V., Durr, A., Bressman, S. et al. (2008) Phenotype, genotype, and worldwide genetic penetrance of LRRK2-associated Parkinson's disease: a case-control study. *Lancet Neurol.* **7**, 583–590 doi:10.1016/S1474-4422(08)70117-0

- 19 Schapansky, J., Nardozi, J.D., Felizia, F. and LaVoie, M.J. (2014) Membrane recruitment of endogenous LRRK2 precedes its potent regulation of autophagy. *Hum. Mol. Genet.* **23**, 4201–4214 doi:10.1093/hmg/ddu138
- 20 Wallings, R., Manzoni, C. and Bandopadhyay, R. (2015) Cellular processes associated with LRRK2 function and dysfunction. *FEBS J.* **282**, 2806–2826 doi:10.1111/febs.13305
- 21 Gardet, A., Benita, Y., Li, C., Sands, B.E., Ballester, I., Stevens, C. et al. (2010) LRRK2 is involved in the IFN- $\gamma$  response and host response to pathogens. *J. Immunol.* **185**, 5577–5585 doi:10.4049/jimmunol.1000548
- 22 Ho, C.C.-Y., Rideout, H.J., Ribe, E., Troy C.M. and Dauer, W.T. (2009) The Parkinson disease protein leucine-rich repeat kinase 2 transduces death signals via Fas-associated protein with death domain and caspase-8 in a cellular model of neurodegeneration. *J. Neurosci.* **29**, 1011–1016 doi:10.1523/JNEUROSCI.5175-08.2009
- 23 Berwick, D. and Harvey, K. (2014) The regulation and deregulation of Wnt signaling by PARK genes in health and disease. *J. Mol. Cell. Biol.* **6**, 3–12 doi:10.1093/jmcb/mjt037
- 24 Miklossy, J., Arai, T., Guo, J.-P., Klegeris, A., Yu, S., McGeer, E.G. et al. (2006) LRRK2 expression in normal and pathologic human brain and in human cell lines. *J. Neuropathol. Exp. Neurol.* **65**, 953–963 doi:10.1097/01.jnen.0000235121.98052.54
- 25 Franke, A., McGovern, D.P.B., Barrett, J.C., Wang, K., Radford-Smith, G.L., Ahmad, T. et al. (2010) Genome-wide meta-analysis increases to 71 the number of confirmed Crohn's disease susceptibility loci. *Nat. Genet.* **42**, 1118–1125 doi:10.1038/ng.717
- 26 Barrett, J., Hansoul, S., Nicolae, D.L., Cho, J.H., Duerr, R.H., Rioux, J.D. et al. (2008) Genome-wide association defines more than 30 distinct susceptibility loci for Crohn's disease. *Nat. Genet.* **40**, 955–962 doi:10.1038/ng.175
- 27 Zhang, F.-R., Huang, W., Chen, S.-M., Sun, L.-D., Liu, H., Li, Y. et al. (2009) Genomewide association study of leprosy. *N. Engl. J. Med.* **361**, 2609–2618 doi:10.1056/NEJMoa0903753
- 28 Gillardon, F., Schmid, R. and Draheim, H. (2012) Parkinson's disease-linked leucine-rich repeat kinase 2(R1441G) mutation increases proinflammatory cytokine release from activated primary microglial cells and resultant neurotoxicity. *Neuroscience* **208**, 41–48 doi:10.1016/j.neuroscience.2012.02.001
- 29 Kuss, M., Adamopoulou, E. and Kahle, P. (2014) Interferon- $\gamma$  induces leucine-rich repeat kinase LRRK2 via extracellular signal-regulated kinase ERK5 in macrophages. *J. Neurochem.* **129**, 980–987 doi:10.1111/jnc.12668
- 30 Tsuchiya, S. et al. (1980) Establishment and characterization of a human acute monocytic leukemia cell line (THP-1). *Int. J. Cancer* **26**, 171–176
- 31 Thévenet, J., Pescini Gobert, R., Hooft van Huijsduijnen, R., Wiessner, C. and Sagot, Y.J. (2011) Regulation of LRRK2 expression points to a functional role in human monocyte maturation. *PLoS ONE* **6**, e21519 doi:10.1371/journal.pone.0021519
- 32 Moehle, M.S., Webber, P.J., Tse, T., Sukar, N., Standaert, D.G., DeSilva, T.M. et al. (2012) LRRK2 inhibition attenuates microglial inflammatory responses. *J. Neurosci.* **32**, 1602–1611 doi:10.1523/JNEUROSCI.5601-11.2012
- 33 Russo, I., Berti, G., Plogher, N., Bernardo, G., Filograna, R., Bubacco, L. et al. (2015) Leucine-rich repeat kinase 2 positively regulates inflammation and down-regulates NF- $\kappa$ B p50 signaling in cultured microglia cells. *J. Neuroinflammation* **12**, 230 doi:10.1186/s12974-015-0449-7
- 34 Blasi, E. et al. (1990) Immortalization of murine microglial cells by a v-raf/v-myc carrying retrovirus. *J. Neuroimmunol.* **27**, 229–237
- 35 Dzamko, N., Inesta-Vaquera, F., Zhang, J., Xie, C., Cai, H., Arthur, S. et al. (2012) The I $\kappa$ B kinase family phosphorylates the Parkinson's disease kinase LRRK2 at Ser935 and Ser910 during Toll-like receptor signaling. *PLoS ONE* **7**, e39132 doi:10.1371/journal.pone.0039132
- 36 Moehle, M., Daher, J.P.L., Hull, T.D., Boddu, R., Abdelmotilib, H.A., Mobley, J. et al. (2015) The G2019S LRRK2 mutation increases myeloid cell chemotactic responses and enhances LRRK2 binding to actin-regulatory proteins. *Hum. Mol. Genet.* **24**, 4250–4267 doi:10.1093/hmg/ddv157
- 37 Dzamko, N., Deak, M., Hentati, F., Reith, A.D., Prescott, A.R., Alessi, D.R. et al. (2010) Inhibition of LRRK2 kinase activity leads to dephosphorylation of Ser<sup>910</sup>/Ser<sup>935</sup>, disruption of 14-3-3 binding and altered cytoplasmic localization. *Biochem. J.* **430**, 405–413 doi:10.1042/BJ20100784
- 38 Nichols, R.J., Dzamko, N., Morrice, N.A., Campbell, D.G., Deak, M., Ordureau, A. et al. (2010) 14-3-3 binding to LRRK2 is disrupted by multiple Parkinson's disease-associated mutations and regulates cytoplasmic localization. *Biochem. J.* **430**, 393–404 doi:10.1042/BJ20100483
- 39 Berger, Z., Smith, K.A. and Lavoie, M.J. (2010) Membrane localization of LRRK2 is associated with increased formation of the highly active LRRK2 dimer and changes in its phosphorylation. *Biochemistry* **49**: 5511–5523 doi:10.1021/bi100157u
- 40 Davies, L.C., Jenkins, S.J., Allen, J.E. and Taylor, P.R. (2013) Tissue-resident macrophages. *Nat. Immunol.* **14**, 986–995 doi:10.1038/ni.2705
- 41 Mestas, J. and Hughes, C.C.W. (2004). Of mice and not men: differences between mouse and human immunology. *J. Immunol.* **172**, 2731–2738 doi:10.4049/jimmunol.172.5.2731
- 42 Kim, B., Yang, M.-S., Choi, D., Kim, J.-H., Kim, H.-S., Seol, W. et al. (2012) Impaired inflammatory responses in murine Lrrk2-knockdown brain microglia. *PLoS ONE* **7**, e34693 doi:10.1371/journal.pone.0034693
- 43 Hakimi, M., Selvanantham, T., Swinton, E., Padmore, R.F., Tong, Y., Kabbach, G. et al. (2011) Parkinson's disease-linked LRRK2 is expressed in circulating and tissue immune cells and upregulated following recognition of microbial structures. *J. Neural. Transm.* **118**, 795–808 doi:10.1007/s00702-011-0653-2
- 44 Wandu, W.S., Tan, C., Ogbeifun, O., Vistica, B.P., Shi, G., Hinshaw, S.J.H. et al. (2015) Leucine-rich repeat kinase 2 (Lrrk2) deficiency diminishes the development of experimental autoimmune uveitis (EAU) and the adaptive immune response. *PLoS ONE* **10**, e0128906 doi:10.1371/journal.pone.0128906
- 45 Liu, Z., Lee, J., Krummey, S., Lu, W., Cai, H. and Lenardo, M.J. (2011) The kinase LRRK2 is a regulator of the transcription factor NFAT that modulates the severity of inflammatory bowel disease. *Nat. Immunol.* **12**, 1063–1070 doi:10.1038/ni.2113
- 46 Dzamko, N., Rowe, D.B. and Halliday, G.M. (2016) Increased peripheral inflammation in asymptomatic leucine-rich repeat kinase 2 mutation carriers. *Mov. Disord.* **31**, 889–897 doi:10.1002/mds.26529
- 47 Schapansky, J., Nardozi, J.D. and LaVoie, M.J. (2014) The complex relationships between microglia, alpha-synuclein, and LRRK2 in Parkinson's disease. *Neuroscience* PMID: 25284317
- 48 Choi, I., Kim, B., Byun, J.-W., Baik, S.H., Huh, Y.H. and Kim, J.-H. et al. (2015) LRRK2 g2019s mutation attenuates microglial motility by inhibiting focal adhesion kinase. *Nat. Commun.* **6**, 8255 doi:10.1038/ncomms9255
- 49 Ma, B., Xu, L., Pan, X., Sun, L., Ding, J., Xie, C. et al. (2016) LRRK2 modulates microglial activity through regulation of chemokine (C-X3-C) receptor 1-mediated signaling pathways. *Hum. Mol. Genet.* PMID:27378696
- 50 Esteves, A.R. and Cardoso, S.M. (2016) LRRK2 at the crossroad between autophagy and microtubule trafficking: Insights into Parkinson's disease. *Neuroscientist* PMID:26740081



- 51 Deretic, V., Saitoh, T. and Akira, S. (2013) Autophagy in infection, inflammation and immunity. *Nat. Rev. Immunol.* **13**, 722–737 doi:10.1038/nri3532
- 52 van Wilgenburg, B., Browne, C., Vowles, J. and Cowley, S.A. (2013) Efficient, long term production of monocyte-derived macrophages from human pluripotent stem cells under partly-defined and fully-defined conditions. *PLoS ONE* **8**, e71098 doi:10.1371/journal.pone.0071098
- 53 van Wilgenburg, B., Moore, M.D., James, W.S. and Cowley, S.A. (2014) The productive entry pathway of HIV-1 in macrophages is dependent on endocytosis through lipid rafts containing CD4. *PLoS ONE* **9** doi:10.1371/journal.pone.0086071
- 54 Flynn, R., Grundmann, A., Renz, P., Hänseler, W. and James, W.S. (2015) CRISPR-mediated genotypic and phenotypic correction of a chronic granulomatous disease mutation in human iPS cells. *Exp. Hematol.* **43**, 838–848.e3 doi:10.1016/j.exphem.2015.06.002