Macroautophagy is an intracellular degradation system that involves the de novo formation of membrane structures called autophagosomes, although the detailed process by which membrane lipids are supplied during autophagosome formation is yet to be elucidated. Macroautophagy is thought to be associated with canonical membrane trafficking, but several mechanistic details are still missing. In this review, the current understanding and potential mechanisms by which membrane trafficking participates in macroautophagy are described, with a focus on the enigma of the membrane protein Atg9, for which the proximal mechanisms determining its movement are disputable, despite its key role in autophagosome formation.

Introduction

Autophagy is an intracellular degradation process that contributes to cellular homeostasis. Because autophagy involves highly dynamic membrane organization, it has occasionally been classified in the broad category of membrane trafficking. For example, microautophagy involves canonical membrane trafficking events, as it is simply an invagination of the lysosomal membrane, and the mechanism whereby membrane lipids are mobilized is clearly understood [1]. However, in macroautophagy, the double-membrane structure, the autophagosome, engulfs some of the cellular contents and delivers them to the lysosome for degradation. Thus, macroautophagy involves the de novo formation of membrane structures, i.e. autophagosomes, and the detailed process by which membrane lipids are supplied during autophagosome formation is unclear. The basic dogma of membrane biogenesis is that an organelle is generated from a pre-existing organelle. Therefore, various hypotheses have been proposed to explain the formation of de novo membrane structures, such as the involvement of unknown membrane vesicles and the free transfer of lipid molecules [2]. Here, the current understanding and hypotheses regarding the role of membrane trafficking in macroautophagy are summarized, with a focus on the enigma of Atg9, i.e. the mechanism whereby it arrives at the autophagosome-formation site.

Platform for autophagosome formation: omegasome and pre-autophagosomal structure

The discovery of the omegasome was a major breakthrough in autophagy research with respect to the membrane-trafficking system. The double FYVE domain-containing protein (DFCP-1) binds to phosphatidylinositol 3-phosphate (PI3P) [3]. Specific types of phosphoinositides are exclusively localized to specific organelles and the endoplasmic reticulum (ER) and Golgi are regarded as being devoid of PI3P. These findings seem to contradict the fact that DFCP-1 is localized to the ER and Golgi apparatus. During starvation, DFCP-1 accumulates in or near the ER, forming a structure resembling the Greek character omegasome [4] (Figure 1). This accumulation is entirely dependent on...
the PI3P-binding capacity of the FYVE domain, indicating that PI3P exists in the omegasome. Strikingly, autophagosomes are generated inside, or in close proximity to, the omegasome, suggesting that autophagosomes form in specialized sites within the cell [4]. PI3 kinase, which specifically functions in autophagy and contains the Atg14L subunit, is recruited to the ER, and PI3P generated by the PI3 kinase complex is required for omegasome formation [5]. Detailed electron microscopy utilizing 3D tomography revealed a physical membranous connection between the ER and the forming autophagosome, i.e. the isolation membrane or phagophore [6,7] (Figure 1). More detailed serial and tomographic electron microscopy has shown that a green fluorescence protein–DFCP-1 fusion is localized to the tubular vesicular structure adjacent to the isolation membrane, representing at least part of the omegasome [8] (Figure 1). The tubular vesicular structure was observed as a potentially more nascent seed structure labeled with Atg13 before double-membrane autophagosome formation was initiated [9]. The ER–Golgi-intermediate compartment (ERGIC) is a tubular vesicular organelle that functions in transport from the ER to the Golgi apparatus. Data from an in vitro-reconstitution study suggested that the ERGIC plays a pivotal role in autophagosome formation [10]. ER-exit sites (ERESs) are specialized in the ER involved in the biogenesis of COPII-coated transport vesicles that traffic to the Golgi. In vivo observations by super-resolution microscopy suggest that autophagosome formation takes place, in part, in the vicinity of the ERES and ERGIC, but does not necessarily always occur in this area [9] (Figure 1). FIP200 and a SEC12-binding protein, CTAGE5, participate in remodeling ERESs and may contribute to autophagosome formation [11]. It has recently been reported that specialized regions of the ER are crucial for autophagosome formation. The ER functions in the biosynthesis of membrane lipids, including phosphatidylinositol (PI). The enzyme responsible for PI synthesis (PI synthase) is a transmembrane protein that is mostly localized to the ER, but is enriched in a less well-characterized, highly mobile compartment with an ER origin [12]. Interestingly, the autophagy-related proteins FIP200 and Ulk1, which are now thought to determine the initiation of autophagosome formation, are transiently colocalized with PI synthase in membranes, suggesting a crucial role of this organelle in autophagosome formation [13] (Figure 1). Other reports have shown that autophagosome formation occurs in specialized regions of the ER, such as ER–mitochondria- and ER–plasma membrane-contact sites [14,15] (Figure 1). It is necessary to determine how these specialized regions of the ER are functionally connected to tubular vesicular structure-containing omegasomes.

Unlike mammalian autophagy, autophagosome formation in the yeast *Saccharomyces cerevisiae* is thought to occur primarily in the pre-autophagosomal structure (PAS; alternatively known as the phagophore-assembly site). The PAS is characterized as the site where Atg proteins gather [16,17] (Figure 1). The organization of the PAS has gradually become apparent; the dephosphorylation of Atg13 in response to TORC1 suppression during starvation is a key trigger of PAS organization [18]. The PAS is always associated with vacuoles, corresponding to lysosomes in mammals, although only one or a few vacuoles are present in each cell [16,17].
Autophagosome formation and vacuole fusion are interconnected, at least in some types of autophagy involving the Cvt pathway [19]. More importantly, a physical connection appears to exist between the ER and PAS (Figure 1). Markers of ERESs are frequently observed in the vicinity of the PAS [20,21] (Figure 1). Additionally, in COPII mutants, severe autophagy defects have been observed [22–24]. An ER-resident SNARE protein, Ufe1, is translocated to the autophagosome-formation site during autophagy [25]. Therefore, it is highly possible that autophagosome formation may be linked with the ER, as in mammalian cells.

Atg9 vesicles in yeast: a membrane source of autophagosomes or just a reservoir?

Regarding the relationship between autophagy and membrane trafficking, a key question is related to Atg9, which is a six-transmembrane protein essential for autophagosome formation that was first identified in yeast [26,27] (Figure 2). Atg9 is localized to the outer membrane of double autophagosomal membranes in yeast [28] (Figure 2). In addition to autophagosomes, Atg9 is localized to PASs and numerous cytoplasmic vesicular structures with a 30–60 nm diameter, which move quite rapidly throughout the cytoplasm [28] (Figure 2). Atg9 is thought to cycle between the PAS/autophagosome and the peripheral vesicular pools [29]. Without Atg1 kinase, Atg9 accumulates at the PAS, suggesting that proper autophagosome formation requires this cycling [29]. Atg9 associates with the HORMA domain of Atg13 at the PAS [30] (Figure 2) and then associates with Atg2 and Atg18 [31]. Therefore, Atg9 plays a direct, crucial role in autophagosome formation at the PAS, but its actual function remains unclear.

A key question is how Atg9 arrives at the PAS. Based on the canonical concept that a transmembrane protein does not leave the membrane during membrane trafficking, these cytoplasmic Atg9 proteins in small vesicles and PAS/autophagosomes are expected to be interconnected by canonical membrane-trafficking processes, such as budding and fusion. Two different models have been used to explain the movement of Atg9. One model holds that Atg9-containing vesicles directly arrive at the PAS and the Atg9-vesicular membrane constitutes, at least part of, the autophagosome membrane. An alternative hypothesis is that Atg9 arrives by

![Figure 2. Model whereby Atg9 vesicles serve as a direct source of autophagosome membrane in yeast cells.](image-url)
another route, such as the ER, and this possibility should be carefully considered. Hereafter, these two views will be described in detail.

My interpretation is that the former view stems from the idea that the identities of Atg9-containing vesicles are uniform; these are termed ‘Atg9 vesicles’ [28,32]. It is clear that Atg9 vesicles originate from the Golgi apparatus [28,33] (Figure 2). These vesicles contain factors required for fusion to the nascent autophagosome, such as Trs85 (the sole specific subunit of TRAPP III) [34], which functions as a guanine nucleotide-exchange factors with Ypt1 [35] (Figure 2). Ypt1 is the yeast homolog of the Rab1 small GTPase; it was originally described as functioning in ER-to-Golgi trafficking [35]. Ypt1 recruits Atg1 to the PAS [32] (Figure 2). TRAPP III also interacts with Sec23, a subunit of the COPII coat, during vesicle budding from the ER [36] (Figure 2). Another subunit of COPII, Sec24, is phosphorylated by the Hrr25 protein kinase during autophagy, inducing starvation; thereby, it becomes associated with Atg9 [37] (Figure 2). Thus, it has been proposed that Atg9 vesicles and COPII vesicles fuse and form the autophagosome membranes [37,38] (Figure 2). Other membrane sources, such as COPI vesicles, may well be involved in autophagosome formation, because the estimated number of membrane lipids supplied from Atg9 vesicles (calculated from the number of Atg9 molecules on autophagosomes) is not sufficient for the autophagosome membrane [28].

Another view is based on the idea that the so-called Atg9 vesicles are not homogeneous entities, but consist of a mixture of transport vesicles transiting between the Golgi apparatus and endosomes instead (Figure 3). Historically, the involvement of endosomal trafficking has not received substantial attention in yeast autophagy, because endosomal trafficking mutants exhibited no or highly limited defects in autophagy. However, Atg9 is partially distributed to the early and late endosomes [33]. Ohashi and Munro [39] noted that some double mutants for factors involved in endosomal trafficking show severe defects in autophagy and Atg9 trafficking.

This discovery has since been verified using a simpler model [40]. Specifically, Trs85 mutants showed severe defects in the Cvt pathway, a selective autophagy type that occurs even under nutrient-rich conditions, but showed only partial defects during starvation-induced macroautophagy [41]. GARP is a tethering factor complex that receives vesicles from the endosome at the Golgi [42]. GARP mutants show severe defects in the Cvt pathway, but not in starvation-induced macroautophagy [43]. Two small G proteins (Ypt6 and Arl1) appear to be involved in autophagy, at least in the GARP-dependent step [44] (Figure 3). Importantly, the combination of mutations in the GARP subunits vps51 and trs85 results in almost completely defective starvation-induced macroautophagy, and Atg9 trafficking to PAS is abrogated [40] (Figure 3). Likewise, trs85 mutations result in severe defects in autophagy when combined with mutations in the retromer subunit Vps17, which generates vesicles for late endosome-to-Golgi retrograde trafficking [41] (Figure 3).

Atg24 is a sorting nexin involved in early endosome-to-Golgi trafficking [45]. Atg24 mutant shows severe defects in the Cvt pathway, similar to the trs85 mutant, but does not show defects in starvation-induced autophagy [46]. In contrast with the vps51 mutant, double mutations in atg24 and trs85 do not have synergistic effects on autophagy and Atg9 trafficking [40] (Figure 3). Therefore, Trs85 is thought to function in the same pathway as Atg24, i.e. early endosome-to-Golgi trafficking [40] (Figure 3). Consistent with these findings, Trs85 is localized to the Golgi [40]. TRAPP III is a guanine nucleotide-exchange factor for Ypt1, and Ypt1 and its GTPase activator Gyp1 mutants are also defective in early endosome-to-Golgi trafficking and autophagy [35,47–49]. Therefore, I propose that the defects in autophagy associated with trs85 mutants are due to impaired early endosome-to-Golgi trafficking of Atg9 (Figure 3). These trs85-derived, autophagy-defective phenotypes (under nutrient-rich conditions) are bypassed during starvation by rerouting Atg9 to the Golgi via late endosomes; however, when additional blockages between the late endosome and Golgi occur, Atg9 cannot reach the Golgi and autophagy is defective (Figure 3). A mutant of TRAPPIII, a guanine nucleotide-exchange factor for Ypt31/32, exhibits defective membrane trafficking, particularly exiting the Golgi toward the endosome and plasma membrane [50]. The TRAPPIII mutant is also defective in Atg9 trafficking [51]; therefore, it is highly possible that TRAPPIII-dependent Atg9 trafficking from the Golgi is important (Figure 3). Taking these findings together, I propose that ‘Atg9 vesicles’ are not homogeneous, but form a heterogeneous population representing vesicles found between the Golgi and the endosome (Figure 3) instead. One interpretation of this physiological result is that reserving a membrane protein as cargo of general membrane trafficking may result in less load balancing, compared with reserving in some specialized vesicles.

Atg9 trafficking is supported by other, potentially specific proteins, i.e. Atg27 and Atg23; without these proteins, Atg9 vesicle formation and delivery to the PAS are severely impaired [28,52,53]. Atg23 is a peripheral membrane protein and its localization overlaps substantially with Atg9 peripheral pools, suggesting that it facilitates Atg9 trafficking [33,54]. Atg27 is a single transmembrane protein and localizes to the Golgi, endosomes,
and vacuoles, and travels among these sites [52,55]. Atg9 is phosphorylated by Atg1 at PAS, which affects the association efficiency with Atg23, Atg27, and Atg18 [56,57]. In addition to its role in facilitating Atg9 trafficking in the reservoir pool, Atg27 may be involved in the retrieval of Atg9 from the vacuole [55]. The return of Atg27 from the early endosome to the Golgi largely depends on the Snx41–Snx4 (Atg24)-sorting nexin complex, which is different from the Atg9-related Atg20–Snx4 (Atg24) complex [58]. This distinction may be important in the regulation of Atg9 trafficking.

**Atg9 trafficking in mammalian systems**

The latter model that Atg9 vesicles are heterogeneous is rather consistent with observations of mammalian ATG9 trafficking (Figure 4). Even in mammals, ATG9 clearly plays a crucial role in autophagosome formation [59–62]. Early studies showed that ATG9 is localized to both the Golgi and endosomes [59]. Live cell imaging has revealed that multiple ATG9-containing vesicles move around in the cytoplasm and that the associations between ATG9-positive areas and areas positive for other Atg markers, such as LC3 and DFCP1, are transient [63,64] (Figure 4). It later became clear that at least some ATG9 molecules reside in recycling endosomes in addition to the Golgi [60,63,65] (Figure 4). ATG9 also travels to the plasma membrane and returns via
clathrin-dependent endocytosis using the clathrin coat protein complex AP-2, together with the Rab GAP
protein TBC1D5 [61] (Figure 4). It is unclear whether Atg9 is also transported to the plasma membrane in
yeast, although the involvement of late secretory pathway components in the plasma membrane has been
reported in autophagy [66, 67]. Mammalian AP-2 recognizes tyrosine-based and di-leucine-based sorting motifs
residing in the N-terminal flanking stretch of ATG9 exposed to the cytosolic face; a sorting motif-deficient
ATG9 mutant exhibits accumulation in the recycling endosome [62] (Figure 4). Wild-type ATG9 accumulated
in the Golgi, although an ATG9 mutant lacking the tyrosine- and di-leucine-based sorting motifs did not, sug-
gest that ATG9 returned to the Golgi from the recycling endosome [62]. Sorting nexin 18 functions in the
tubulation of the recycling endosome to promote autophagy [68]. The requirement for an endosomal coatier
in autophagy may also relate to this process [69]. The ATG9-containing vesicle detaches from the recycling
endosome via the membrane fission machinery containing Bif-1 and Dynamin 2 [70] (Figure 4).

TRAPPIII is also involved in mammalian autophagy, and the suppression of its specific subunit TRAPPC8
(Trs85 homolog) or TRAPPC13 results in autophagy defects [62, 71–73]. TRAPPIII defects are related to retro-
grade trafficking of several cargo molecules from the endosome to the Golgi, including Shiga-toxin and ATG9
[62, 72–74] (Figure 4). TRAPPC8 interacts with TBC1D14, a negative regulator of autophagy that controls the
delivery of membranes from RAB11-positive recycling endosomes to forming autophagosomes [60, 72]
(Figure 4). Rab1 is also important for autophagy in mammalian cells [75]. TRAPPIII and TBC1D14 affect
Rab1, which controls ATG9, similar to the yeast counterpart Ypt1 [72]. In addition to ATG9, another
ATG16L-positive vesicle has been identified as a potential autophagosome precursor, but this result may reflect
an artifact due to transient overexpression [76, 77].

Do we need a new model to explain Atg9 trafficking?
Theoretically, both models (i.e. Atg9 vesicles help form autophagosome membranes, and Atg9 vesicles are
reservoirs of Atg9 molecules) are not mutually exclusive, and it is possible that some ‘Atg9 vesicles’ can be di-
rectly targeted to the PAS and omegasome. However, the latter model proposes that most ‘Atg9 vesicles’ moving
around the cytoplasm represent the Atg9 reservoir pooled in transport vesicles before functioning in the PAS
or omegasome. If this is the case, the delivery route from the reservoir to the PAS may be indirect, and several
other pathways should be carefully considered. Another model that explains the route from the Golgi to PAS
and omegasomes should be established. It is quite possible that before arriving at the PAS and omegasome,
Atg9 arrives at the Golgi because the failure to arrive in the Golgi is coupled with autophagy defects [40, 62].
For example, Ypt31/32 are involved in vesicle exit from the Golgi, and their mutants show defective autophagy
without impaired Atg9 delivery to the PAS [66]. These findings suggest that forward traffic from the Golgi is
dispensable, at least for Atg9 delivery to the PAS. Interestingly, the COG complex is also involved in autophagy
via Atg9 trafficking [78–80]. COG function in retrograde traffic within the Golgi may be an effector of Ypt1,
highlighting the possibility that retrograde traffic inside the Golgi and to the ER may be important [81]. In
mammalian cells, COPI-mediated retrograde traffic from the Golgi to the ER is required for autophagy [9].
Calpain is also implicated in playing some role in mobilizing ATG9 from Golgi stacks; therefore,
Calpain-dependent processes may be related to Atg9 delivery to autophagosomes [82]. ATG9, localized on
tubular appendices of the ER, subsequently associates with ATG13-positive particles, which grow into autophagosomes [9]. In both yeast and mammals, Atg9-positive tubular vesicular structures have been observed in the vicinity of nascent autophagosomes [33, 63]. An interesting possibility is that the lipid composition of such structures plays some roles. Autophagosome formation depends on desaturated fatty acid [83] and at least in
yeast, the requisite step is at the Atg9 delivery to PAS [84]. By clarifying the identities of these Atg9-positive
signals involved in autophagosome formation, it will be possible to resolve the enigma regarding the movement
of Atg9 vesicles.

Abbreviations
DFCP-1, double FYVE domain-containing protein; ER, endoplasmic reticulum; ERESs, ER-exit sites; ERGIC,
ER–Golgi-intermediate compartment; PAS, pre-autophagosomal structure; PI, phosphatidylinositol; PI3P,
phosphatidylinositol 3-phosphate.

Competing Interests
The Author declares that there are no competing interests associated with this manuscript.
References


